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ARYLATION CUPRO-CATALYSÉE DE COURTS PEPTIDES CONTENANT L'HISTIDINE
AVEC DES TRIARYLBISMUTHINES

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HWAI-CHIEN CHAN

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DÉDICACE

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TABLE DES MATIÈRES

REMERCIEMENTS	iii
DÉDICACE.....	v
LISTE DES FIGURES.....	ix
LISTE DES TABLEAUX.....	xii
LISTE DES ABRÉVIATIONS, DES SIGLES ET DES ACRONYMES	xiii
RÉSUMÉ.....	xvi
ABSTRACT	xvii
INTRODUCTION.....	1
CHAPITRE 1 L'HISTIDINE, DE L'ACIDE AMINE AUX PEPTIDES	3
1.1 L'histidine au quotidien.....	3
1.1.1 Chez les êtres vivants.....	3
1.1.2 Chez les chimistes.....	9
1.2 L'histidine, catalyseur de la réaction d'aldolisation	9
1.3 Les modifications synthétiques de l'histidine.....	11
1.3.1 Alkylations et groupements protecteurs	11
1.3.2 Oxydation de la chaîne latérale de l'histidine.....	17
1.3.3 Méthodes d'arylations de la chaîne latérale de l'histidine.....	20
CHAPITRE 2 LES BISMUTHINES, UN NOUVEL OUTIL	29
2.1 Les généralités	29
2.2 Les organobismuthines, leurs synthèses et applications.....	30
2.2.1 Synthèse des organobismuthines trivalents	30
2.2.2 Synthèse des organobismuthines pentavalents	33
2.2.3 Réaction d'alkylation par les alkylbismuthines.....	35
2.2.4 Réaction d'arylation par des arylbismuthines trivalentes.....	37
2.2.5 Réactions d'arylation par des arylbismuthines pentavalentes	39
LE BISMUTH AU SERVICE DES MODIFICATIONS DE PEPTIDES.....	46
CHAPITRE 3 COPPER-PROMOTED N-ARYLATION OF THE IMIDAZOLE SIDE CHAIN OF PROTECTED HISTIDINE BY USING TRIARYLBISMUTH REAGENTS	47
3.1 Introduction.....	47
3.2 Article issu de ces travaux	47
3.3 Informations supplémentaires.....	55
3.4 Contributions des auteurs à l'article	55

CHAPITRE 4 ON THE COPPER-PROMOTED BACKBONE ARYLATION OF THE HISTIDINE-CONTAINING PEPTIDES USING TRIARYLBISMUTHINES	57
4.1 Introduction.....	57
4.2 Article issu de ces travaux	57
4.3 Informations supplémentaires.....	63
4.4 Contributions des auteurs à l'article	63
CONCLUSION.....	64
BIBLIOGRAPHIE	66
Annexe A	79
Annexe B.....	80

LISTE DES FIGURES

Figure 1-1 Biosynthèse générale de l’histidine. Adaptée de ^[5]	4
Figure 1-2 <i>Histidine Utilization system</i> : voie métabolique de l’histidine chez les bactéries. Adaptée de ^[11]	5
Figure 1-3 Complexes de métal avec l'histidine obtenus par cristallographie	6
Figure 1-4 Les voies cataboliques de l'histidine. MT : Méthyl transférase ; CS : Carnosine synthétase. Adaptée de ^[17]	7
Figure 1-5 A – Histidine cationique et neutre ainsi que ses tautomères. B – Propension catalytique de 10 acides aminés selon les familles d’enzymes. C – Mécanisme d’action de la triade Ser-His-Asp de la chymotrypsine.	8
Figure 1-6 Synthèse de l’histidine à partir d’érythrose	9
Figure 1-7 A – Aldolisation catalysée par une proline ; B – Versatilité de l’aldolisation catalysée par l’histidine	10
Figure 1-8 Mécanismes de l'aldolisation catalysée par la proline (gauche) et par l'histidine (droite)	11
Figure 1-9 A – Alkylation de la position N ^π de l’histidine ; B – Mécanisme de la racémisation de l’histidine	12
Figure 1-10 Étude asymétrique de l'alkylation des azotes de l'imidazole de l'histidine	13
Figure 1-11 A – Structure du Théonellamide A ; B – Synthèse d’un lipo-peptidomimétique	13
Figure 1-12 A – Alkylation à la position N ^π de l’histidine sur un pentapeptide ; B – Propriétés inhibitrice de Plk1 des peptides contenant l’histidine N ^π alkylé et N ^τ macrocyclisés	14
Figure 1-13 A – Phototrifluorométhylation de l’histidine protégée ; B – Phototrifluorométhylation du tripeptide NH ₂ -Pro-His-Glp	15
Figure 1-14 C ² -Alkylation de l’imidazole de l’histidine protégée dans les conditions de Minisci	15
Figure 1-15 Biosynthèse de la diphthamide. Adapté de ^[61]	16
Figure 1-16 A – C ² -Alkylation sélective de l'histidine dans le saralasin par lumière bleue et DHP ^[62] ; B – C ² -Alkylation sélective de l'histidine d’un peptide par l’utilisation de sels de sulfonates et de TBHP ^[63]	17
Figure 1-17 Synthèse de la L-ergothionéine : A – Thiocyanate comme source de soufre ; B – Chlorothioformate de phényle comme source de soufre	18
Figure 1-18 A – Voie biosynthétique de l'ergothionéine ; B – Synthèse de l'ergothionéine à travers l'utilisation de la cystéine ; C – C ⁵ -thiolisation à partir de l’hercynine et d’acide thioacétique ; D – Mécanisme proposé de la sélectivité C ² /C ⁵	19

Figure 1-19 Mécanisme proposé de l'oxydation de l'histidine observée sur l'anticorps monoclonal IgG1. Image tirée de ^[70]	20
Figure 1-20 C ² -Arylation de l'imidazole de l'histidine protégée dans les conditions radicalaires de Minisci	21
Figure 1-21 A – C ⁵ -Arylation sélective de l'imidazole de l'histidine protégée ; B – C ² -Arylation sélective de l'imidazole de l'histidine protégée par C–H activation.....	22
Figure 1-22 Mécanismes proposés tentant d'expliquer la régiosélectivité de l'arylation par C–H activation	22
Figure 1-23 C ⁵ -Arylation indirecte par couplage Suzuki-Miyaura.....	23
Figure 1-24 A – N ¹ -Arylation par S _N Ar ; B – N ¹ -Arylation par attaque nucléophile sur un pro-aromatique ; C – N ¹ -Arylation par couplage croisé de Barton	24
Figure 1-25 A – Transposition des conditions d'arylation de l'imidazole dans le cadre de l'histidine ; B – N ¹ -Arylations de l'imidazole de l'histidine protégée par couplage de type Ullman-Goldberg chauffées par micro-onde	25
Figure 1-26 N ¹ -Arylation de l'imidazole de l'histidine protégée par un couplage Chan-Lam-Evans	26
Figure 1-27 N-Arylations par réaction photochimique	26
Figure 1-28 Double arylation de l'imidazole par réactions successives régiosélectives.....	27
Figure 2-1 Exemples de composés contenant du Bismuth aux propriétés thérapeutiques	30
Figure 2-2 Structures cristallographiques de quelques organobismuthines trivalents	31
Figure 2-3 Voies de synthèse des halogénures de bismuths et leurs transformations	33
Figure 2-4 Méthodes de synthèse des halogénures de triarylbismuthines et pentaarylbismuthines	34
Figure 2-5 Synthèses d'halogénures de trialkylbismuthines et du pentaméthylbismuthine.....	35
Figure 2-6 A – Trifluorométhylation du chlorure de benzoyle, d'énamines et d'aniline ; B – Méthylation d'halogénure d'aryles ; C – Cyclopropylation d'halogénures d'aryles et N-cyclopropylation de lactames, d'azoles et d'indoles	36
Figure 2-7 Alkylations d'halogénures d'aryles pallado-catalysées.....	37
Figure 2-8 Méthylation de l'acide p-toluènesulfonique par le dicarboxylate de méthylbismacyle.....	37
Figure 2-9 A – Arylations pallado-catalysées de chlorures d'acyles ; B – Arylations pallado-catalysées d'halogénures d'aryles ; C – O-Arylations cupro-catalysées de phénols et d'alcools ; D – N-Arylations cupro-catalysées d'azotes	39
Figure 2-10 A – Influence de la nature du bismuth pentavalent sur l'arylation régiosélective de dérivés phénols selon le pH ; B – Mécanismes proposées expliquant la régiosélectivité	41

Figure 2-11 α -Arylation de carbonyles par des bismuths pentavalents en milieu neutre et basique et <i>O</i> -arylation en milieu acide. Milieu : ^A = acide, ^N = neutre, ^B = basique.....	42
Figure 2-12 <i>C</i> -Arylations régiosélectives par l'usage de bismacycles pentavalents	43
Figure 2-13 A – <i>O</i> -Arylation d'alcools et de diols à partir de bismuths pentavalents ; B – <i>N</i> -Arylations d'amide, d'imides et d'indole à partir du trifluoroacétate tétraphénylbismuth.	44

LISTE DES TABLEAUX

Tableau 2-1 Organométaux compatibles à la synthèse d'organobismuthines.....	32
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LISTE DES ABRÉVIATIONS, DES SIGLES ET DES ACRONYMES

ADN : Acide désoxyribonucléique

AIRCAR : Ribonucléotide 5-aminoimidazole-4-carboxamide

ATP : Adénosine triphosphate

CMD : Déprotonation métallique concertée

CS : Carnosine synthétase

DbA : Dibenzylidèneacétone

DCM : Dichlorométhane

DHP : Dihydropyridine

DIPEA : *N,N*-Diisopropyléthylamine

DMF : *N,N*-Diméthylformamide

DMSO : Diméthylsulfoxyde

EF2 : Facteur d'élongation 2

FAO : Food and Agriculture Organization of the United Nations (Organisation des Nations Unies pour l'Alimentation et l'Agriculture)

FDA : US Food Drug Administration (Administration américaine des denrées alimentaires et des médicaments)

FG : Formyl glutamate

FIG : Formimino glutamate

Hol-P : Histidinol phosphate

IAP : Imidazoleacetol phosphate

IGP : Imidazoleglycérol phosphate

MT : Méthyl transférase

m-CPBA : Acide *m*-chloroperbenzoïque

NAD : Nicotinamide adénine dinucléotide

NBS : N-bromosuccinimide

NMP : *N*-méthyl-pyrrolidone

PRATP : Phosphoribosyl-ATP

PRFAR : Ribonucléotide phosphoribosyle formimino-5-aminoimidazole-4-carboxamide

ProFAR : Ribonucléotide prophosphoribosyle formimino-5-aminoimidazole-4-carboxamide

PRPP : Phosphoribosylpyrophosphate

RPECV : Répulsion des paires d'électrons de la couche de valence

SAM : S-adénosylméthionine

SPSS : Synthèse de Peptides sur Support Solide

TBHP : Hydroperoxyde de tertbutyle

TFA : Acide trifluoroacétique

Tp : Température pièce

RÉSUMÉ

L'histidine est un membre du groupe très fermé des acides aminés essentiels. Essentiel parce qu'elle est cruciale dans bon nombre de processus biologiques, mais également parce qu'elle n'est pas synthétisée par l'organisme. Ses fonctions s'étendent sur un large panel d'actions : elles peuvent par exemple passer par une simple dégradation métabolique afin de fournir les produits utiles dans d'autres processus ou alors être impliquées dans la structure complexe des chaînes peptidiques aboutissant à des protéines aux implications diverses. Ses différentes caractéristiques font qu'une quelconque modification structurale de l'histidine peut avoir des conséquences considérables sur l'ensemble de l'organisme. Toutefois, si bien contrôlée et maîtrisée, la modification de l'histidine peut avoir des effets bénéfiques et dont toute une discipline s'est développée autour de ce but unique.

En parallèle, depuis ses débuts dans les années 80, la chimie du bismuth a vu son implication croître dans de nombreuses réactions. Une diversité de bon augure, puisque comparé à ses homologues métalliques, le bismuth reste bon marché et présente en plus une toxicité bien moindre. Ces aspects font du bismuth un métal de choix pour l'incorporer dans des réactions chimiques, notamment dans des modifications de l'histidine.

Cette thèse s'inscrit dans la lignée des modifications spécifiques catalysées au cuivre de l'histidine à travers l'usage d'organobismuthines. L'essence de ces modifications repose sur l'arylation de la chaîne latérale de l'histidine, qu'elle soit isolée ou au sein d'une séquence peptidique. À la suite du développement de la méthode, une observation surprenante d'arylation de la chaîne principale a été découverte et sera présentée en détail à travers ce modeste manuscrit.

Mots clés : Histidine • Triarylbismuthines • Arylations sélectives • Petits peptides • Catalysée au cuivre

ABSTRACT

Histidine belongs to the very select group of essential amino acids. Essential because it is crucial in many biological processes but also because it is not synthesized by the organism. Its functions extend over a wide range of actions: it can for instance, be involved into a simple metabolic degradation in order to provide useful products for other processes or be involved in the complex structure of peptide chains leading to proteins with various implications. These different characteristics mean that any structural modification of histidine can have considerable consequences for the whole organism. However, if well controlled and mastered, modification of histidine can have beneficial effects and an entire discipline has been developed around this single aim.

At the same time, since its beginnings in the 80s, bismuth chemistry has seen its involvement grow in numerous reactions. This diversity augurs well since compared to its metallic counterparts, bismuth remains in one hand, inexpensive and in the other hand, has much lower toxicity. These aspects make bismuth a metal of choice for incorporating it in chemical reactions, especially in histidine modifications.

This thesis is in line with the specific copper-catalyzed modifications of histidine through the use of organobismuthines. The essence of these modifications relies on the arylation of the side chain of histidine, whether isolated or within a peptide sequence. Following the development of the method, a surprising observation of main chain arylation was discovered and will be presented in detail throughout this modest manuscript.

Keywords : Histidine • Triarylbiorganobismuthines • Selective arylations • Small peptides • Copper catalyzed

INTRODUCTION

La chimie des peptides a connu plusieurs grandes révolutions au cours de l'histoire. Une première a eu lieu à l'université de Toronto dans les années 1920, grâce aux travaux remarquables des Nobélisés Frederick Banting et John J. R. Macleod, ainsi que de leurs étudiants Charles H. Best et James B. Collip. En effet, leurs travaux portant sur l'insuline ont permis d'apporter une réponse majeure aux malades du diabète. Leur découverte est d'autant plus importante lorsqu'on s'aperçoit des chiffres endémiques du nombre de diabétiques dans le monde. Ce n'est qu'après une trentaine d'années, que l'insuline a pu être complètement séquencée grâce aux travaux de Frederick Sanger, prix Nobel de chimie en 1958.^[1] Une petite révolution dans la chimie des peptides mais à l'impact considérable puisqu'il amorcera le séquençage de l'ADN ainsi que les travaux du Robert B. Merrifield avec sa synthèse de peptides sur support solide (SPSS). Cette invention de 1963 le récompensera du prix Nobel de chimie en 1984 et marquera une deuxième révolution majeure en chimie des peptides.^[2] En effet, la SPSS est d'une importance et d'un avantage si grand qu'après 60 ans d'évolution technologique, le principe même de cette méthode de synthèse artificielle reste inchangé et est tout autant utilisé. À la suite de ces grandes découvertes et inventions, la chimie peptidique vit actuellement sa troisième révolution. Effectivement, depuis ces dernières décennies, le nombre de peptides à usage thérapeutique ne cesse de croître au fil des années en passant de l'insuline, unique peptide à caractère pharmaceutique, à plus de 80 peptides à ce jour approuvés par la FDA (US Food Drug Administration), ce qui représente 8% des médicaments mis sur le marché.^[3, 4] Il est évident qu'une telle présence ne peut être due par le simple caractère naturel des acides aminés que composent les peptides, mais également aux astucieuses modifications chimiques de ces derniers. En effet, l'utilisation de peptides non-naturels a joué un rôle crucial dans l'amélioration et l'optimisation des effets thérapeutiques des peptides. Dans la littérature, il existe à ce jour une quantité appréciable de méthodes permettant la synthèse de peptides non naturels. Ces derniers peuvent être obtenus selon trois grandes catégories de méthodes, à savoir par la synthèse classique non initiée par des acides aminés, par le recours à des acides aminés non-naturels ou alors par la modification post-synthétique du peptide.

Par le passé, notre groupe de recherche a rapporté de nouvelles méthodes de modification peptidique par l'arylation chemosélective de la chaîne latérale des peptides du tryptophane ou de la tyrosine. Au cours de ces précédents travaux, différents acides aminés ont été soumis aux conditions d'arylation nouvellement établies et l'histidine a présenté une affinité remarquable. C'est dans cette optique que s'est inscrite l'essence même de mes travaux de recherches. En effet, dans le but de fournir une suite logique et complémentaire à ces travaux, mes recherches se sont appuyées comme précédemment, sur l'utilisation de triarylbiisothiazolones catalysée au cuivre afin de modifier des peptides contenant l'histidine.

Les résultats obtenus à l'issue de ces recherches seront décrits après une mise en contexte de l'importance de l'histidine au sein des peptides naturels, de l'état de l'art des modifications de l'histidine et de ses applications dans différents domaines. Un bref historique de la chimie du bismuth et ses applications dans les modifications de peptides sera également présenté avant l'insertion des dites publications qui formeront le cœur central de cette thèse.

CHAPITRE 1

L'HISTIDINE, DE L'ACIDE AMINE AUX PEPTIDES

Dans ce chapitre, il sera question de l'importance de l'histidine dans sa globalité. Comprendre comment l'histidine est synthétisée chez les différentes espèces tout en décrivant son rôle à travers les bactéries, les plantes ainsi que les animaux. On y énumérera quelques particularités de sa réactivité biochimique et chimique tout en apportant les détails qu'ont pu découvrir les chimistes à son égard.

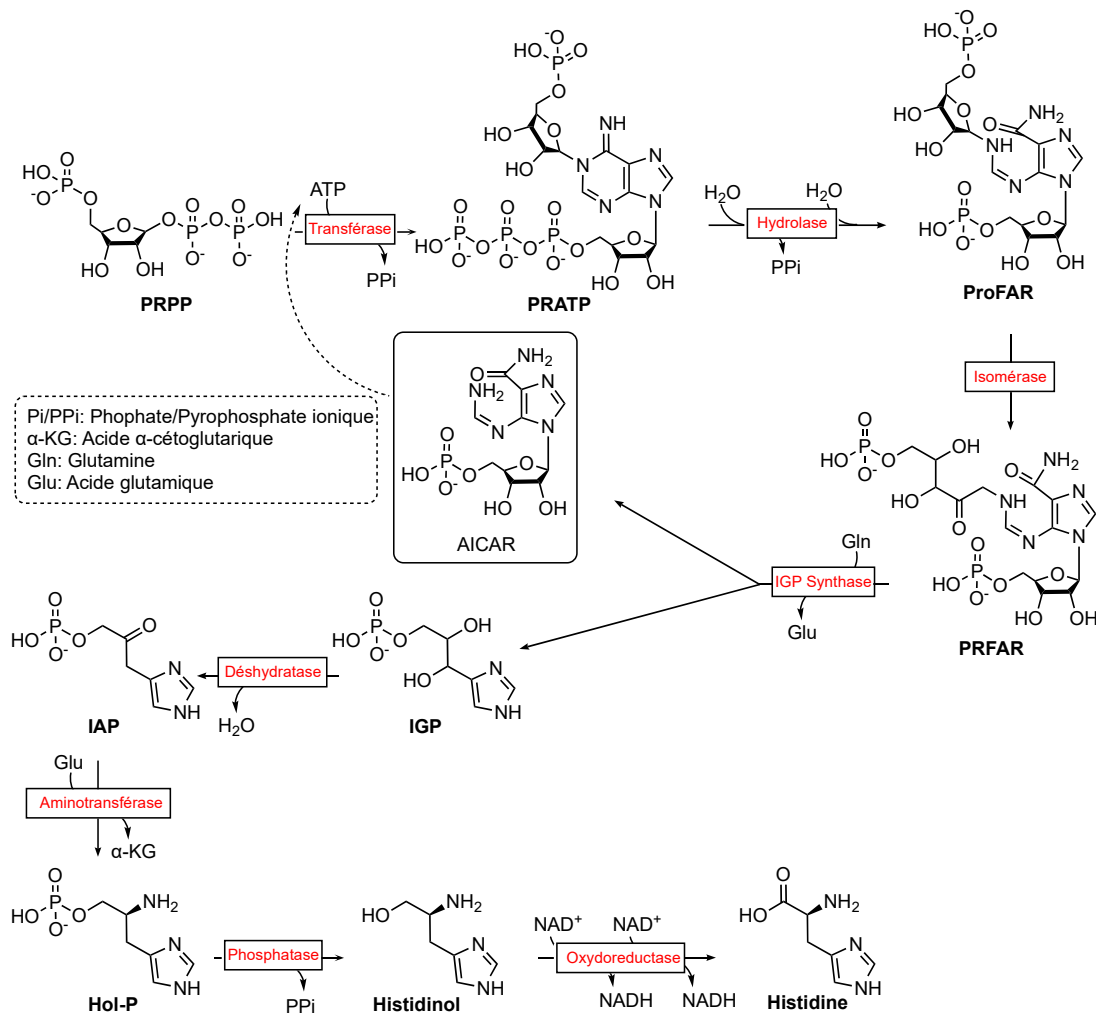
1.1 L'histidine au quotidien

Découvert indépendamment pour la première fois par Albrecht Kossel et Sven G. Hedin en 1896, l'acide 2-amino-3-(1H-imidazol-4-yl)propanoïque, plus communément appelé histidine, est l'acide aminé dont sa chaîne latérale est pourvue d'un imidazole. Parce qu'elle participe dans une multitude de mécanismes biologiques, l'histidine est d'une importance cruciale chez tout être vivant, mais semble être d'une importance tout aussi fondamentale en chimie aux vues de ses propriétés chimiques particulières.

1.1.1 Chez les êtres vivants

L'alimentation chez les êtres humains régit son unique apport en histidine car elle a la particularité d'être synthétisée chez tous les procaryotes, champignons et plantes, mais pas chez les animaux. La biosynthèse de l'histidine se déroule en 10 étapes et engage une multitude d'enzymes propres à chaque espèce. Elle débute par la condensation du phosphoribosylpyrophosphate (PRPP) avec de l'ATP (Adénosine triphosphate) pour former le phosphoribosyl-ATP (PRATP) avant d'être hydrolysé en ribonucléotide phosphoribosyle formimino-5-aminoimidazole-4carboxamide (ProFAR) sous l'action de deux hydrolases successives. Une isomérase initie l'ouverture du phosphoribosyle conduisant au ribonucléotide phosphoribosyle formimino-5-aminoimidazole-4-carboxamide (PRFAR) qui entraîne, grâce à l'IGP synthase et une glutamine, la formation de ribonucléotide 5-aminoimidazole-4-carboxamide (AICAR) et de l'imidazoleglycérol phosphate (IGP). Le AICAR est recyclé en ATP alors que l'IGP subit une déshydratation sous l'effet d'une enzyme conduisant à l'imidazoleacetyl phosphate (IAP). Par la présence d'une aminotransférase et de glutamate, l'IAP passe sous la forme d'histidinol phosphate (Hol-P) qui est rapidement déphosphorylé en histidinol. Catalysé par l'histidinol déshydrogénase, ce dernier est finalement oxydé par deux NAD^+ en histidine (Figure 1-1).^[5, 6]

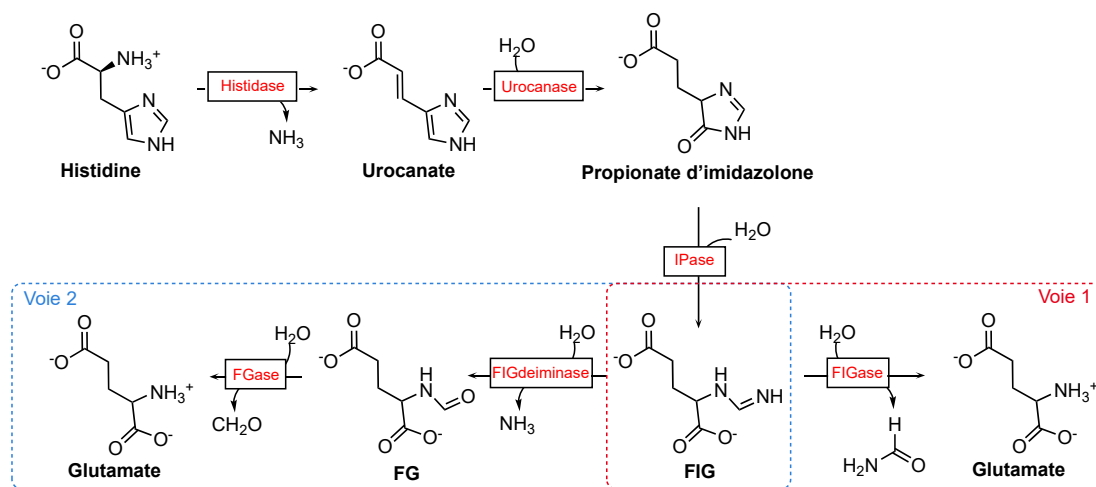
Figure 1-1 Biosynthèse générale de l'histidine. Adaptée de [5]



Chez les bactéries la biosynthèse de l'histidine est hautement conservée et se classe au quatrième rang des processus amino-synthétiques les plus gourmands en énergie, après la synthèse du tryptophane, de la phénylalanine et de la tyrosine.^[7] Ce processus est tellement énergivore que la plupart des bactéries ont su développer une capacité à se servir uniquement de l'histidine présente dans son environnement dans un but totalement économique. L'évolution a même abouti à la mutation de certaines souches de bactéries en espèces auxotrophes, c'est-à-dire, incapables de synthétiser l'histidine par leur propre moyen et de devoir dépendre d'un hôte. Ces derniers ont la particularité d'être souvent pathogène pour l'humain, on peut y trouver par exemple, le *Mycobacterium genitalium* responsable d'urétrites,^[8, 9] l'*Helicobacter pylori* responsable de gastrites et d'ulcères à l'estomac ou bien encore l'*Haemophilus influenzae*, qui lui est responsable de pharyngites.^[9] Bien entendu, bien que la biosynthèse de l'histidine soit un processus hautement conservé, chaque espèce de bactérie possède une affinité aux acides aminés qui lui est propre afin de croître et/ou de développer une résistance à son milieu. Par exemple, *Escherichia coli* peut n'utiliser

que la serine, l'aspartate, la cystéine, la glycine, le glutamate et l'alanine comme unique source d'azotes et de carbones pour se développer et essaye d'éviter les milieux trop concentrés en valine et en leucine. Par opposition, l'espèce *Staphylococcus aureus* a souvent besoin de valine, de proline, d'arginine et de cystéine pour proliférer.^[10] De manière complémentaire à la biosynthèse, l'*Histidine Utilization* (Hut) *system* est la voie catabolique de l'histidine chez les bactéries. Ce processus est tout autant conservé qu'universel et débute par l'action de l'histidase sur l'histidine conduisant à l'urocanate en libérant de l'ammoniac. L'urocanase va ensuite catalyser la formation du propionate d'imidazolone avant l'ouverture du cycle imidazolone par l'IPase pour former du formimino glutamate (FIG). À cette étape, deux voies sont possibles en fonction des espèces. Une première voie permet la formation directe en glutamate et d'un formamide par l'action d'une FIGase. Alors qu'une deuxième voie passe d'abord par la formation du formyl glutamate (FG) avant d'aboutir au glutamate tout en éliminant de l'ammoniac et du formaldéhyde (Figure 1-2).^[11]

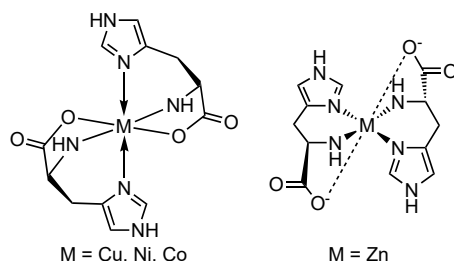
Figure 1-2 *Histidine Utilization system* : voie métabolique de l'histidine chez les bactéries. Adaptée de ^[11]



Chez les plantes, la production d'histidine s'effectue dans les chloroplastes et joue un rôle crucial dans la croissance et le développement de la plante. Des travaux effectués sur l'arabette de *Thalium* ont permis de démontrer qu'en faisant des *knockdowns* de différents gènes des enzymes responsables de la biosynthèse de l'histidine, il était possible d'observer différents phénotypes de silique (fruit de la plante). En effet, le désamorçage du gène *hisn2-1* conduit à la formation de siliques blanches et/ou atrophiées, tandis que celui du gène *hisn8* mène à la présence de siliques immatures. L'apport en solution riche en histidine a permis de sauver partiellement les siliques juvéniles, concluant ainsi de l'importance de l'histidine dans le développement et la croissance des plantes.^[12] L'histidine, de par la nature de ses fonctions chimiques, possède une forte affinité au zinc, cobalt, cuivre et nickel et joue un rôle supplémentaire crucial dans la régulation et le transport de ces métaux lourds potentiellement toxiques. Effectivement, face à la présence

de ces métaux provenant de sols souillés, les plantes ont su développer une réponse défensive qui consiste en la synthèse accrue d'histidine, qui envoyée dans les racines vont complexer les métaux avant de les transporter jusqu'aux feuilles afin de les stocker sans danger.^[13, 14] Ces complexes sont formés par la coordination de deux histidines avec le métal, dont l'azote de l'amine, l'azote de l'imidazole ainsi que l'oxygène du carbonyle de chacun des résidus formant le trident chélatant. La nature du métal influence le caractère cristallographique du complexe. La chélation d'un atome de cuivre, de nickel ou de cobalt par l'histidine suivra un système cristallin orthorhombique alors qu'un atome de zinc préférera un système tétragonal (Figure 1-3).^[15, 16]

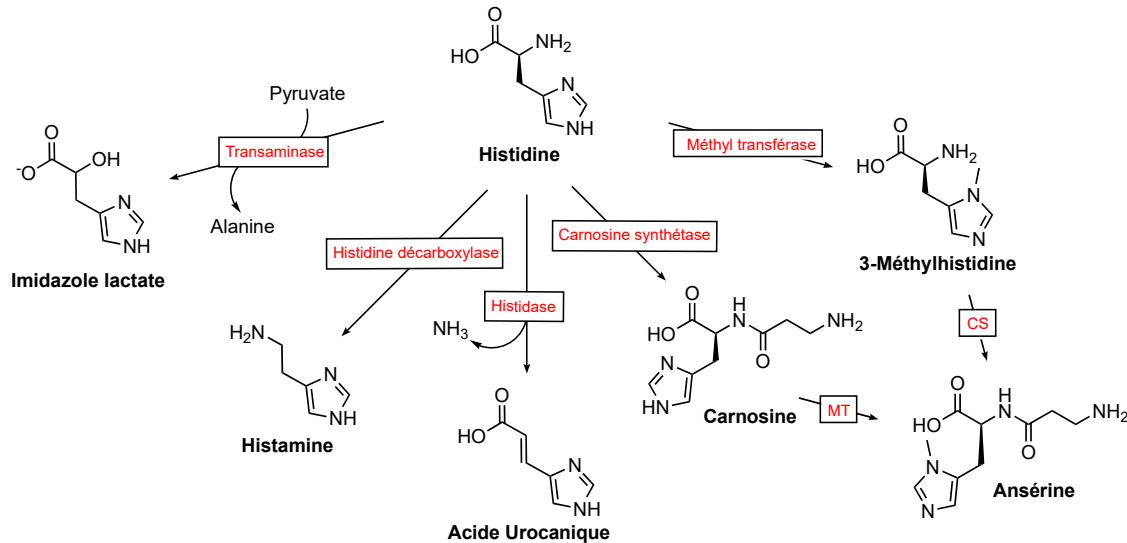
Figure 1-3 Complexes de métal avec l'histidine obtenus par cristallographie



Chez les animaux, de manière assez similaire, l'histidine va jouer un rôle fondamental à la fois dans la croissance et le développement, mais également dans le transport et la séquestration des différents métaux. Par ailleurs, d'après l'Organisation des nations unies pour l'alimentation et l'agriculture (FAO), il est recommandé pour un adulte de consommer de 8 à 12 mg/kg d'histidine par jour.^[17] Il est possible de la retrouver en grande quantité dans la viande rouge et dans le poisson sous forme libre ou protéique. L'importance de l'histidine peut également être soulignée à travers différentes voies cataboliques. Une première voie permet l'élimination de l'excès d'histidine par l'action d'une transaminase et d'un pyruvate pour la transformer en imidazole lactate, afin d'être éliminée par les urines plus facilement. Une deuxième voie consiste en la décarboxylation de l'histidine en histamine, un neurotransmetteur responsable des réponses immunitaires ainsi que de la régulation physiologique dans l'intestin. Une troisième voie possible est sa transformation en acide urocanique, un chromophore s'accumulant dans l'épiderme jouant un rôle protecteur dans la photocarcinogénèse.^[18] Et enfin, les deux dernières voies possibles seraient sa transformation en ansérine soit par l'intermédiaire du méthylhistidine, soit par l'intermédiaire du carnosine. À savoir que l'ansérine et la carnosine sont des actrices dans les réactions inflammatoires du sang et

pourraient retarder la maladie d'Alzheimer chez les personnes âgées si consommées en plus grande quantité (Figure 1-4).^[19]

Figure 1-4 Les voies cataboliques de l'histidine. MT : Méthyl transférase ; CS : Carnosine synthétase. Adaptée de ^[17]

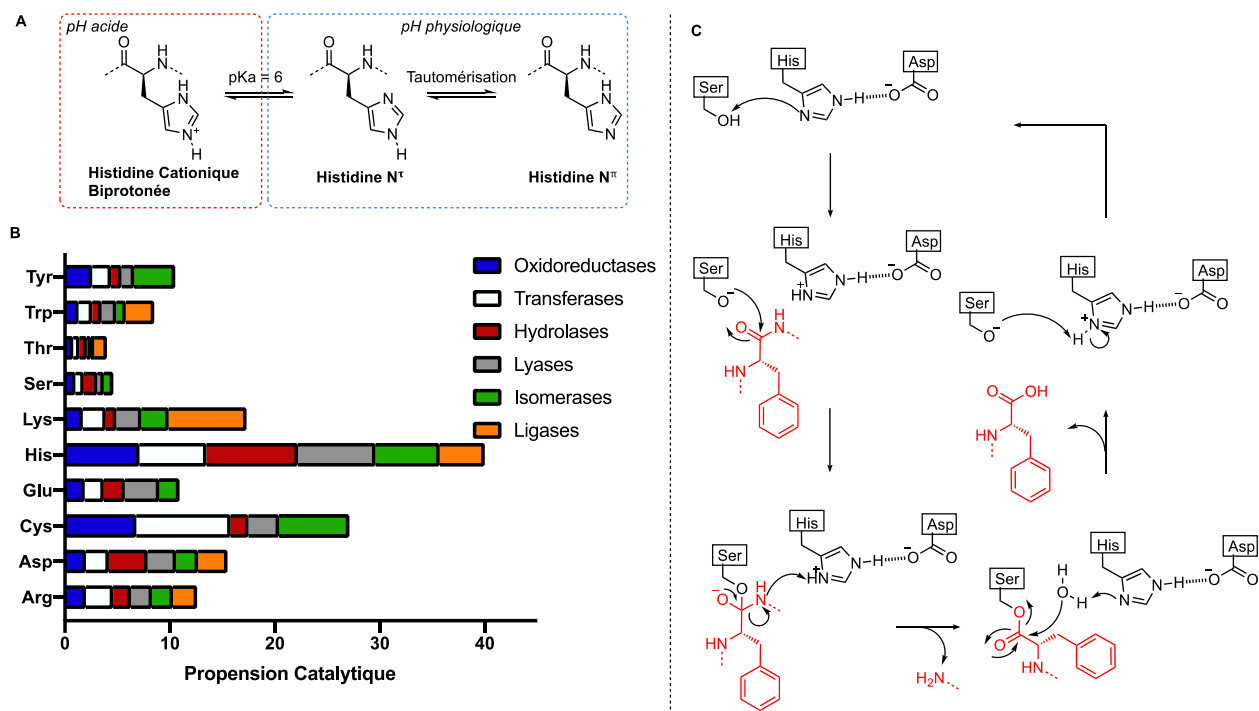


D'autres recherches sur différentes espèces ont également démontré l'utilité de l'histidine. Notamment chez les poissons, une alimentation riche en histidine va non seulement avoir un impact sur la taille des spécimens mais également sur la prévention de la cataracte, une maladie souvent observée dans les piscicultures.^[20-22] Chez les rongeurs, une quantité accrue d'histidine ingérée conduit à une satiété plus rapide sûrement due à l'action neurologique de l'histamine.^[23, 24] Chez les poules d'élevage, un régime enrichi en histidine induit une augmentation du taux en carnosine et en ansérine dans les muscles, ce qui peut être perçu comme une amélioration de la qualité de la viande.^[25] Finalement chez les vaches laitières, un tel régime augmenterait la production de lait.^[26]

Au-delà de l'importance de l'histidine en tant qu'acide aminé libre, l'histidine joue un rôle plus important encore dans le domaine des peptides et des protéines. Ceci est facilement compréhensible lorsqu'on s'aperçoit de sa forte sensibilité au pH. En effet, le pKa de sa chaîne latérale d'environ 6 permet à l'histidine d'endosser les caractéristiques d'un ampholyte ; puisqu'à bas pH elle sera donneuse de protons sous sa forme cationique doublement protonnée, et à pH physiologique elle sera accepteuse de protons sous sa forme neutre mono-protonnée. Petite particularité à pH physiologique, l'histidine peut se présenter sous la forme de deux tautomères, le tautomère protoné N^+ et le tautomère protoné N^π (Figure 1-5A).^[27] Ainsi, l'histidine se retrouve souvent au niveau des sites actifs des protéines ou des sites catalytiques de différents enzymes et se classe première en tant que résidu le plus enclin à être impliqué dans les mécanismes catalytiques. Elle

intervient le plus souvent dans les hydrolases et oxydoréductases, moins souvent dans le cadre des lyases et transférases et plus rarement dans les cas d'isomérases et ligases (Figure 1-5B).^[28] Au niveau des sites catalytiques, l'histidine est souvent accompagnée par deux autres acides aminés pour ainsi former ce qu'on appelle une triade catalytique. Il en existe plusieurs types, mais la triade Ser-His-Asp de la chymotrypsine est certainement la plus connue. Elle catalyse des protéolyses en aval des résidus tyrosine, tryptophane, phénylalanine, méthionine et leucine. Son mécanisme d'action débute à la suite de l'augmentation du pKa de l'histidine par l'aspartate par le biais d'un pont hydrogène. S'en suit l'activation de la serine qui va alors attaquer le carbonyle du substrat formant ainsi l'hémiacétal anionique. L'ester se forme en cassant le lien amide et l'histidine cationique se retrouve de nouveau neutre. Cette dernière va donc pouvoir activer une molécule d'eau qui va effectuer une réaction de saponification de l'ester, rétablissant ainsi la triade catalytique de départ (Figure 1-5C).^[29] Les autres triades impliquant l'histidine, telles que les triades Cys-His-Asp, Cys-His-Ser et Ser-His-His, constituent également le site catalytique d'autres protéases et possèdent un mécanisme d'action similaire.

Figure 1-5 A – Histidine cationique et neutre ainsi que ses tautomères. B – Propension catalytique de 10 acides aminés selon les familles d'enzymes. C – Mécanisme d'action de la triade Ser-His-Asp de la chymotrypsine.

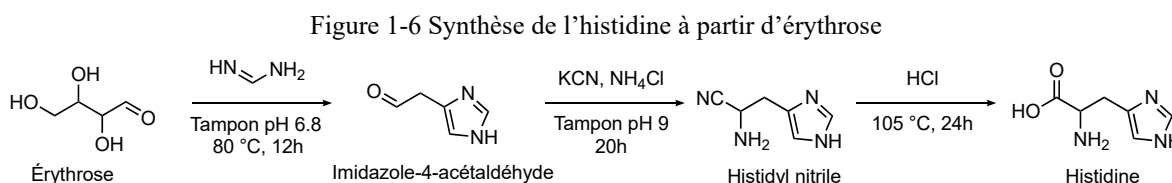


L'histidine est donc un acide aminé essentiel, car indispensable au développement chez tous les êtres vivants, non seulement par elle-même, mais également parce qu'elle est le précurseur de plusieurs composés à caractère immunologique. De plus, de par sa richesse chimique, elle se retrouve souvent au cœur de

mécanismes catalytiques dans de nombreux enzymes aux rôles diversifiés. Du fait de ce rôle bien précis, l'absence ou la modification structurale de l'histidine conduirait à une perte certaine de l'activité de l'enzyme. En chimie médicinale, il n'est pas rare de souhaiter réduire l'activité d'une protéine et pour cela il est inévitable de passer par des modifications chimiques.

1.1.2 Chez les chimistes

Avant de voir comment modifier l'histidine, il est intéressant d'abord de pouvoir la synthétiser. Dans le but de mieux comprendre l'origine de la vie, la synthèse de l'histidine a été décrite pour la première fois en 1987 par Shen qui a fait condenser de l'érythrose avec du formamidine pour former tout d'abord de l'imidazole-4-acétaldéhyde qui va par la suite le transformer en histidine à travers la réaction de Strecker avec un rendement global de 4% (Figure 1-6).^[30]



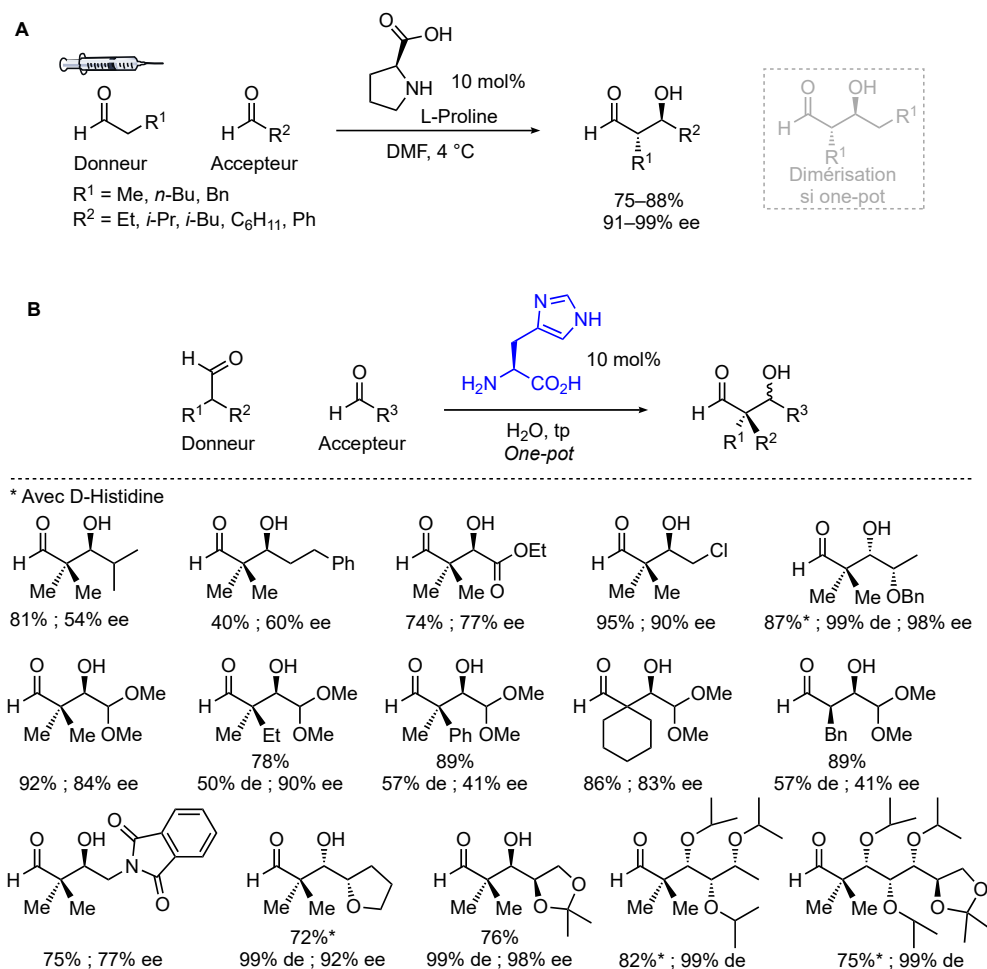
La demande en histidine dans le monde ne cesse de grandir et son marché global a été évalué à 252 millions de dollars en 2022 et pourrait valoir 354 millions d'ici 2028.^[31] Il est évident que la synthèse classique d'histidine n'est pas assez rentable à l'échelle industrielle et c'est à ce titre que les méthodes actuelles de production d'histidine se font majoritairement par fermentation de souches bactériennes.^[32, 33] De plus, c'est parce que l'histidine est souvent utilisée en tant qu'additif alimentaire que les groupes agroalimentaires sont les plus grands demandeurs en histidine. L'industrie pharmaceutique participe également à la prospérité du marché, mais pour des objectifs cette fois-ci tout autre, comme son utilisation en tant que catalyseur mais également de sa modification structurale.

1.2 L'histidine, catalyseur de la réaction d'aldolisation

L'utilisation d'acides aminés comme catalyseur dans la réaction d'aldolisation a remarquablement été étudié par MacMillan. Les acides aminés permettent de se passer à la fois de l'utilisation de métaux de transition et de devoir préactiver le carbonyle en énolate (Figure 1-7A).^[34] Outre la proline, plusieurs groupes de recherche ont activement exploré l'activité catalytique des différents acides aminés, incluant l'histidine. Bien qu'efficaces, ces différentes recherches montrent cependant une profonde limitation dans la diversité des aldéhydes utilisables et n'abrogent pas des limitations pratiques et chimiques.^[35-39] En effet, les grandes limites persistantes sont la nécessité d'ajouter lentement le carbonyle nucléophile, dit donneur, par le biais

d'un pousse-seringue afin d'éviter les sous-produits d'homoaldolisation et qu'un carbonyle non-énolisable ne puisse jouer uniquement le rôle d'accepteur. Ce n'est que grâce aux études de Mahrwald que l'histidine a pu se différencier des autres acides aminés et enfin apporter une réponse valable à ces deux problématiques, car elle permet de réaliser la réaction en *one-pot* et de synthétiser des aldols à centre quaternaire jusque-là impossibles (Figure 1-7B).^[40-42]

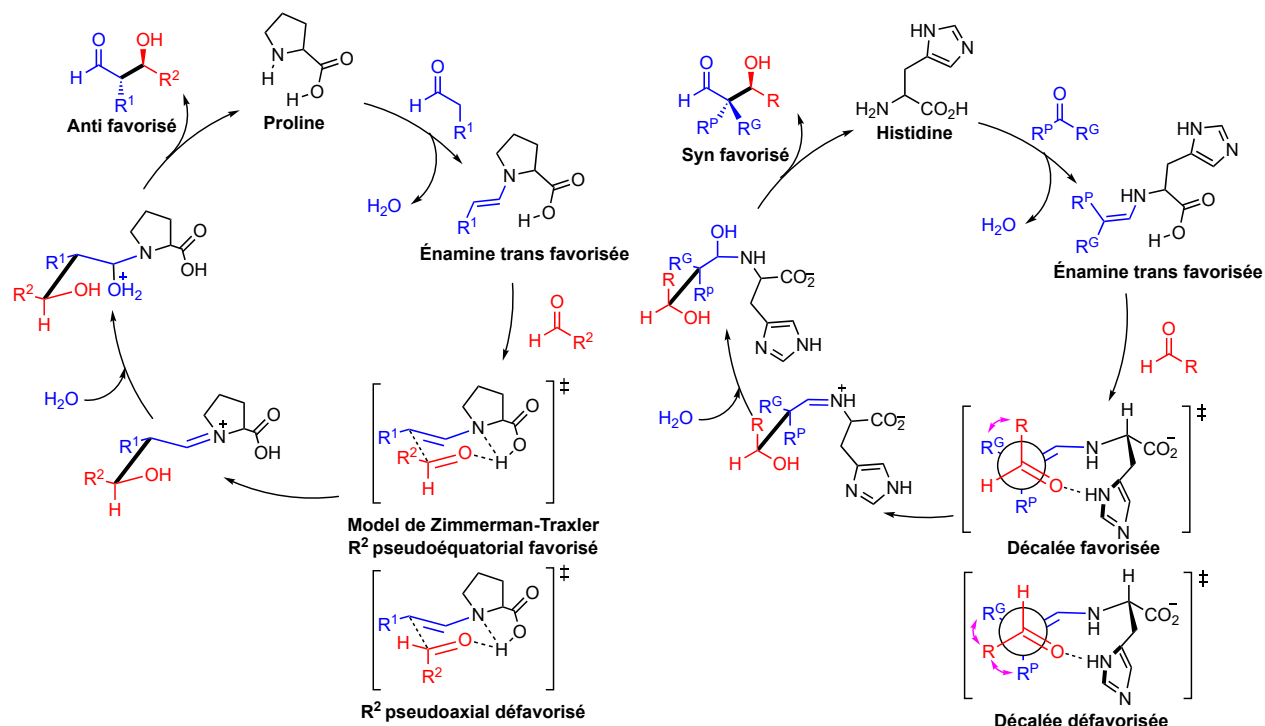
Figure 1-7 A – Aldolisation catalysée par une proline ; B – Versatilité de l'aldolisation catalysée par l'histidine



Si jusque-là l'emploi de l'histidine semble être l'équivalent pratique de la proline, une différence sur la stéréochimie du produit formé existe selon l'acide aminé utilisé. En effet, de son côté la proline favorise la formation d'un produit anti parce que son mécanisme passe par un état de transition chaise de Zimmerman-Traxler favorisant l'attaque nucléophile sur la face Ré du carbonyle accepteur.^[43] Tandis que dans le cadre de l'histidine, l'état de transition de type « Zimmerman-Traxler » provient du pont hydrogène formé avec l'imidazole bloquant le carbonyle dans une configuration décalée minimisant les interactions gauches. Le

carbonyle accepteur ainsi bloqué subit une attaque nucléophile sur sa face Ré conduisant à la formation du produit syn (Figure 1-8).^[44]

Figure 1-8 Mécanismes de l'aldolisation catalysée par la proline (gauche) et par l'histidine (droite)



Apporter des modifications structurales à l'histidine, comme un groupement protecteur sur l'azote terminal, inhibe totalement la réaction d'aldolisation. Ceci démontre à un certain point l'importance de chaque fonction chimique pour une activité donnée. Sur ce principe même, plusieurs groupes ont mené des études sur la modification spécifique de l'histidine et parfois son impact sur son environnement.

1.3 Les modifications synthétiques de l'histidine

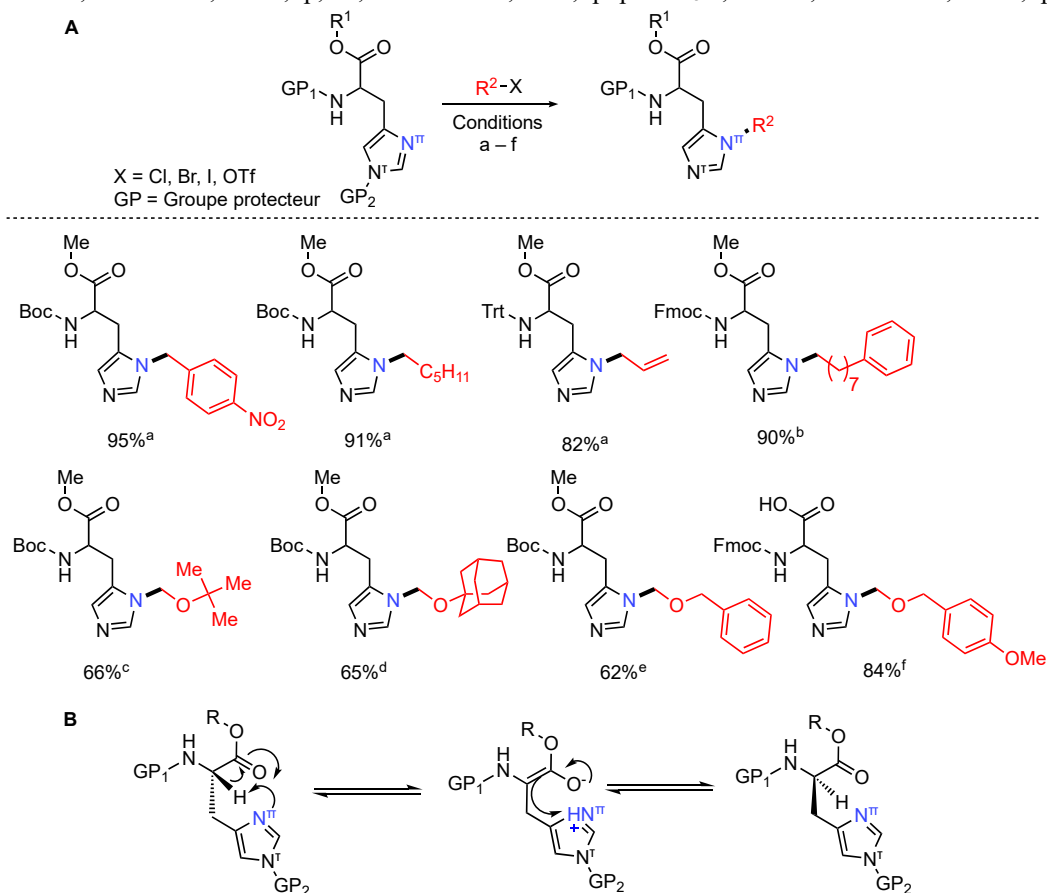
1.3.1 Alkylations et groupements protecteurs

Les premières modifications que l'on peut apporter sur l'histidine sont l'ajout de groupements dits protecteurs au niveau de l'acide carboxylique, de l'amine terminale ainsi que de la position N^ε. Ces modifications sont assez triviales puisque l'histidine se retrouve le plus souvent reliée à d'autres acides aminés et la position N^ε est suffisamment nucléophile pour être facilement occupée par divers groupements par simples réactions de substitution. À l'inverse, les groupements protecteurs ou la présence d'un groupement alkyle à la position N^ε sont pour le coup beaucoup moins fréquents et méritent d'être soulignés. Une méthode robuste largement utilisée permet l'obtention d'un groupement alkyle à cette position, mais il

est néanmoins nécessaire de protéger la position N^τ au préalable. Ainsi les alkyles, allyles ainsi que les benzyles sont tous compatibles avec des rendements excellents.^[45, 46] De manière tout à fait similaire, des groupements protecteurs d'éthers peuvent également être installés (Figure 1-9A).^[47-50] Ces groupements protecteurs ont non seulement l'avantage d'être compatibles à la synthèse peptidique sur support solide, mais confèrent également le pouvoir d'empêcher totalement la racémisation de la molécule. En effet, l'azote N^π est parfaitement situé pour arracher un proton en alpha du carbonyle et un groupement quelconque à ce niveau inhiberait totalement ce genre de phénomène (Figure 1-9B).^[51]

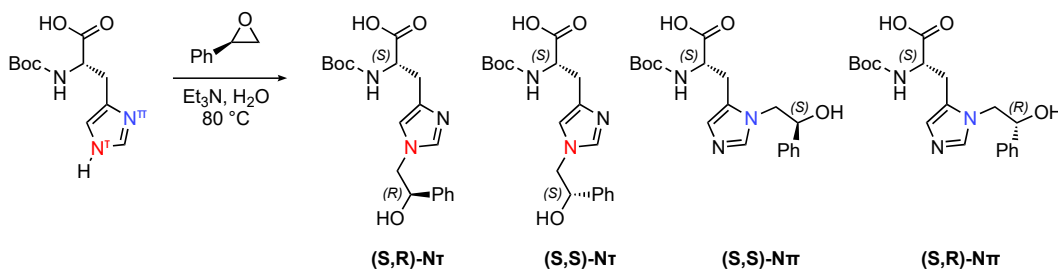
Figure 1-9 A – Alkylation de la position N^π de l'histidine ; B – Mécanisme de la racémisation de l'histidine

^a R₂-Br ou R₂-I, Et₃N, DMF, tp ou 50 °C, 36–72h; ^b R₂-OH, Tf₂O, DIPEA, DCM, -75 °C à tp, 16h; ^c Me₃OCH₂Cl, CCl₄/DCM; ^d Adom-Cl, DCM, tp, 8h; ^e BnOCH₂Cl, Et₂O, tp. puis Et₃N, MeOH; ^f MBom-Cl, DCM, tp, 4h



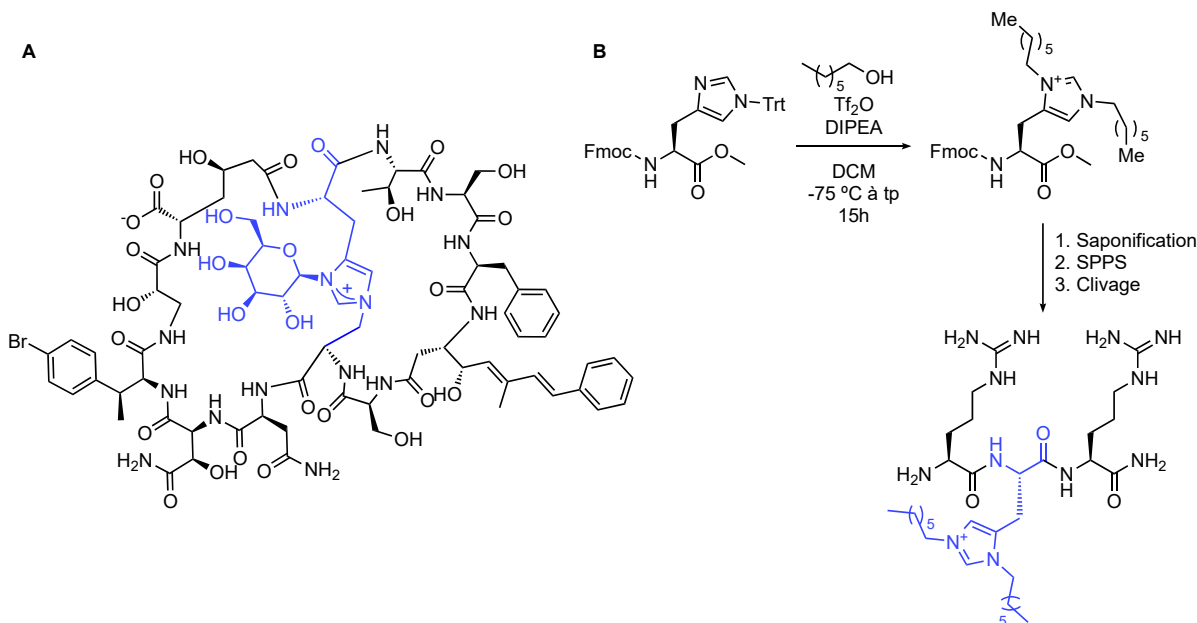
Dans le cadre d'une étude de synthèse asymétrique, une équipe a réussi à additionner un alkyle chiral sur la position N^π par le biais d'une ouverture d'époxydes. Mais n'ayant pas protégé la position N^τ, les résultats obtenus démontrent que l'alkylation de l'azote N^π reste grandement défavorisée sans trop de surprise avec un ratio (S,R)-N^τ:(S,S)-N^τ:(S,S)-N^π:(S,R)-N^π de 28:18:1:5 (Figure 1-10).^[52]

Figure 1-10 Étude asymétrique de l'alkylation des azotes de l'imidazole de l'histidine



Dans ce cadre bien précis, l'espèce dialkylée n'est qu'un intermédiaire obligatoire à l'obtention du produit N^π -monoalkylé désiré. Cependant, il existe dans la nature des théonellamides, une famille de peptides aux propriétés antifongiques isolée d'éponges de mer, qui comportent une histidine cationique dont ses deux azotes de sa chaîne latérale sont alkylés (Figure 1-11A).^[53] Dans une optique similaire, le groupe de Bang a réussi à synthétiser et isoler l'histidine doublement alkylée et de l'utiliser par la suite dans la synthèse d'un lipo-peptidomimétique, un peptide aux propriétés antimicrobiennes (Figure 1-11B).^[54]

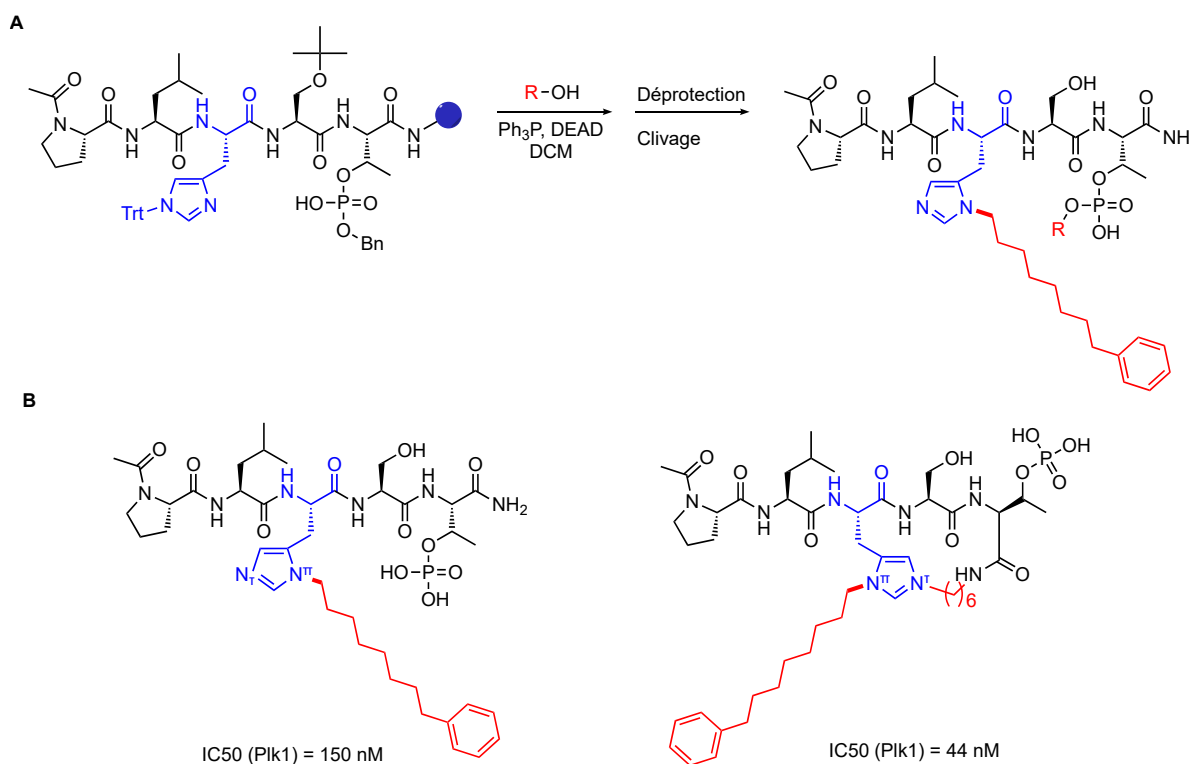
Figure 1-11 A – Structure du Théonellamide A ; B – Synthèse d'un lipo-peptidomimétique



L'alkylation chimiosélective de l'imidazole à la position N^π au sein d'un petit peptide est possible et a été découverte par hasard par le groupe de Burke aux États-Unis. En mettant en présence l'histidine N^π -protégée d'un pentapeptide sur support solide dans les conditions réactionnelles de Mistunobu, ils ont découvert que l'azote N^π s'est retrouvé alkylé.^[55] Ils ont tout d'abord pensé que la présence de la thréonine O-phosphorylée située à un résidu de distance jouait un rôle dans le mécanisme réactionnel, mais la nécessité d'un deuxième

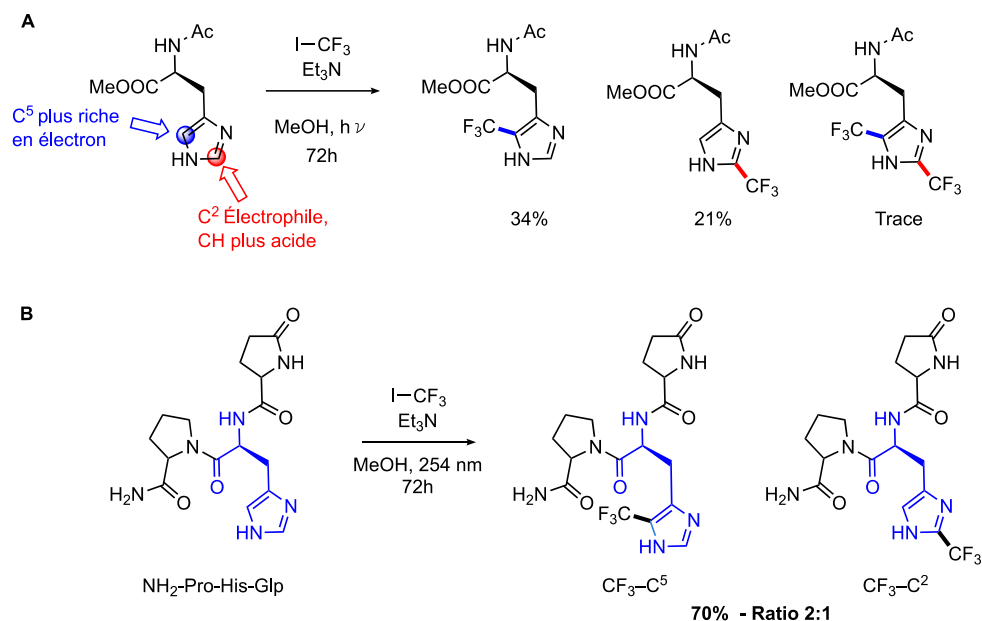
équivalent de « réactifs de Mistunobu » pour que l'azote N^π se retrouve alkylé ainsi que la présence de l'alkyle sur le groupement phosphate sur le produit final, ont permis de démontrer le contraire (Figure 1-12A).^[46] Le groupe ne s'est pas arrêté à l'étude mécanistique et a poursuivi leurs travaux en étudiant l'activité inhibitrice de cette nouvelle molécule sur la polo-like kinase 1 (Plk1), une protéine jouant un rôle dans la mitose et la méiose. En ce faisant, ils ont découvert qu'en ajoutant un macrocycle au peptide reliant l'imidazole de l'histidine à l'amide terminal il était possible d'améliorer son caractère inhibiteur contre Plk1 en passant d'un IC_{50} de 150 à 44 nM (Figure 1-12B).^[56]

Figure 1-12 A – Alkylation à la position N^π de l'histidine sur un pentapeptide ; B – Propriétés inhibitrice de Plk1 des peptides contenant l'histidine N^π alkylé et N^π macrocyclisés



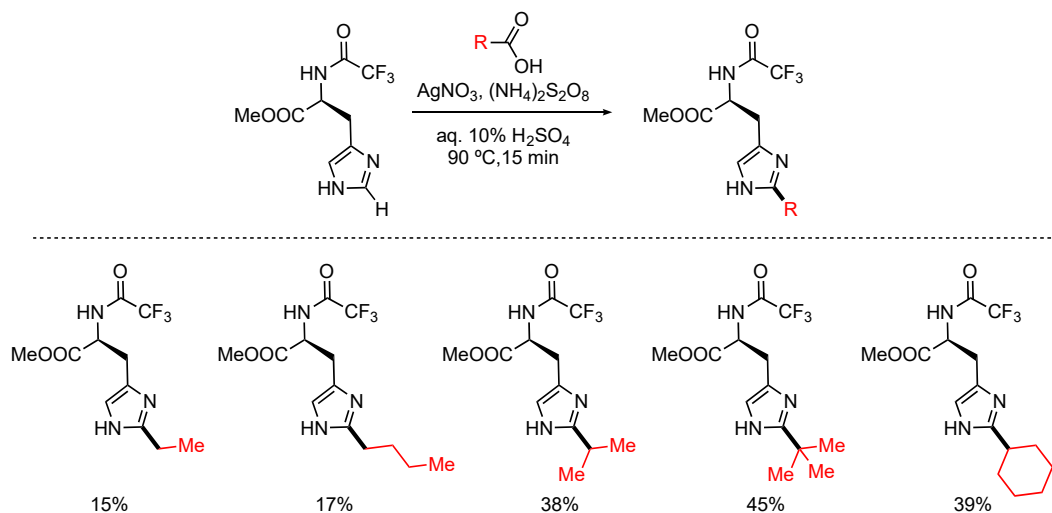
Bien que peu manifestes, les carbones C^2 et C^5 de l'imidazole peuvent être également alkylés. En 1984, Cohen a rapporté les premières réactions d'alkylation qui ont été effectuées par photochimie et de ce fait, un mélange inévitable de produits d'alkylation C^2 et C^5 a été obtenu. Toutefois, les résultats obtenus montrent que l'alkylation sur le carbone C^5 se trouve plus favorisée par rapport au carbone C^2 dû au fait de la différence de densité électronique et des traces de produit doublement alkylé.^[57] Un peu plus tard, la réaction sur un tripeptide a donné des résultats assez similaires avec un rendement total de 70% donnant toujours le produit d'alkylation C^5 majoritaire (Figure 1-13).^[58]

Figure 1-13 A – Phototrifluométhylation de l’histidine protégée ; B – Phototrifluométhylation du tripeptide NH₂-Pro-His-Glp



Ce n’est que sept années plus tard, qu’en s’inspirant des travaux du groupe de Minisci,^[59] ce même groupe de recherche a réussi à installer un plus large panel d’alkyles de manière régiosélective sur le carbone C² le plus électrophile. Ces nouvelles conditions de réaction permettent de se passer de lumière et d’utiliser le nitrate d’argent comme agent radicalaire d’acides carboxyliques (Figure 1-14).^[60]

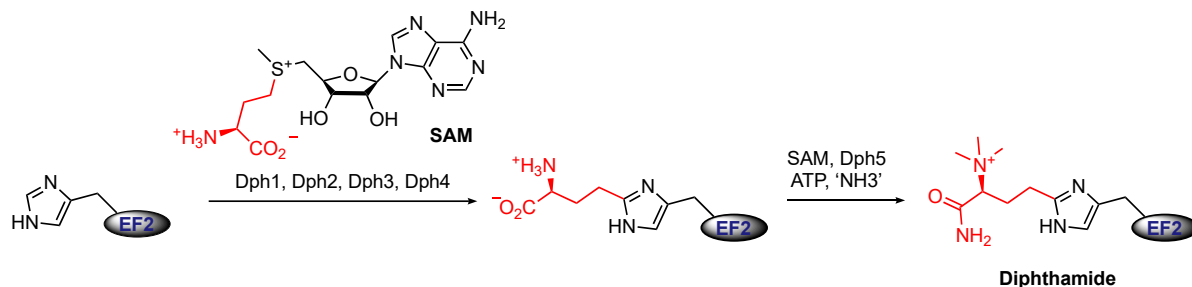
Figure 1-14 C²-Alkylation de l’imidazole de l’histidine protégée dans les conditions de Minisci



Chez les archaeas et les eucaryotes, la biosynthèse de la diphtamide s’effectue à travers une réaction d’alkylation C²-sélective de l’histidine du facteur d’élongation 2 (EF2), une protéine jouant un rôle crucial

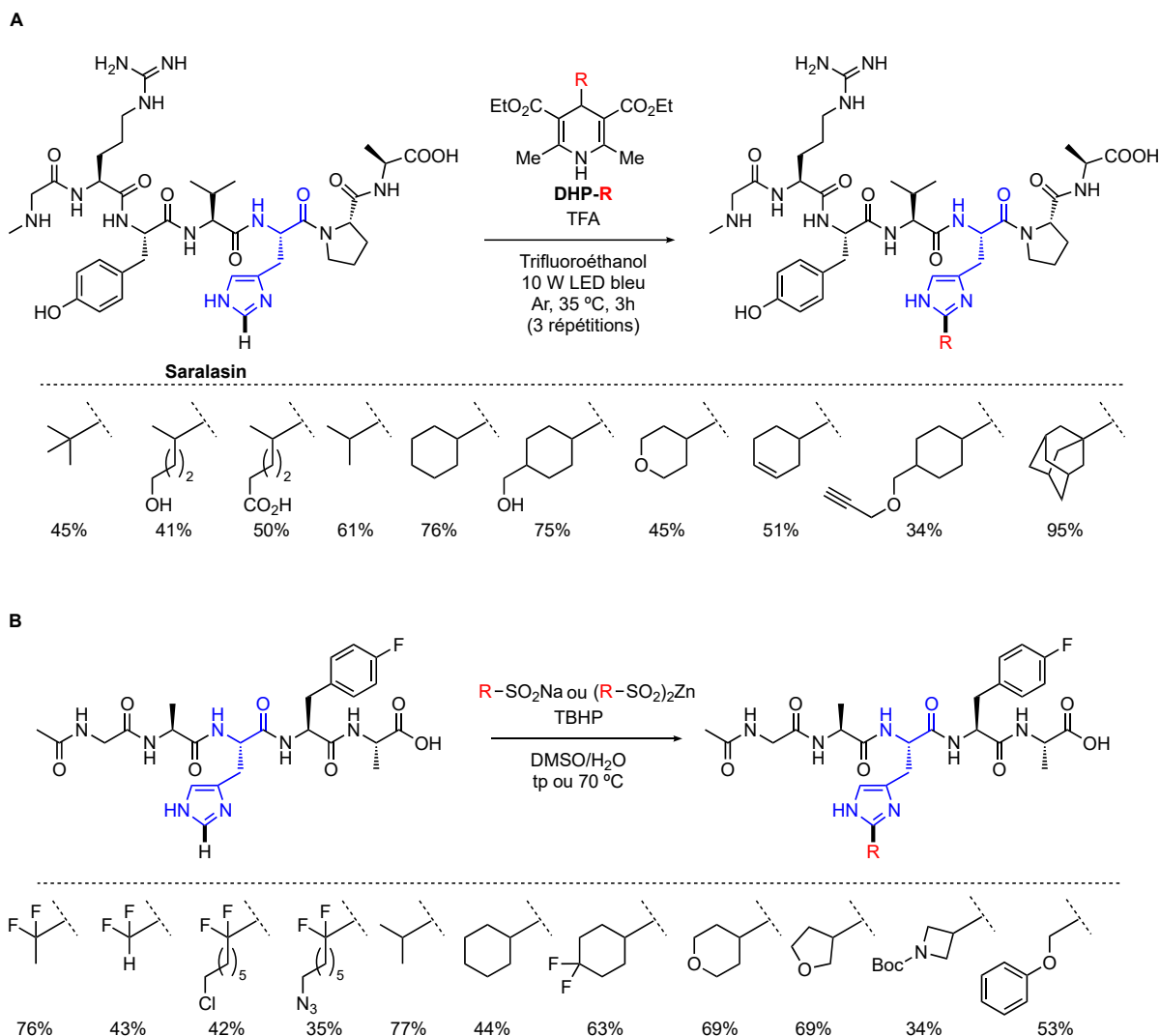
dans la synthèse de protéines. Il est accepté que la réaction se fasse de manière radicalaire et soit catalysée par cinq protéines Dph où la S-adénosylméthionine (SAM) fournit le groupement alkyle (Figure 1-15).^[61]

Figure 1-15 Biosynthèse de la diphthamide. Adapté de ^[61]



Ce n'est que très récemment, à la même année, que deux groupes de recherche sont parvenus à alkyler sélectivement l'imidazole de l'histidine à la position C² sur des peptides plus ou moins longs. Ces travaux impressionnent tant par leur versatilité que par leur robustesse mais adoptent toutefois des stratégies différentes. L'un utilise de la lumière bleue et des dérivés d'alkyles de dihydropyridine (DHP) comme couple initiateur radicalaire,^[62] alors que l'autre utilise des sels d'alkylsulfonates et de l'hydroperoxyde de *tert*-butyle (TBHP) (Figure 1-16).^[63] Leurs travaux remarquables sur l'alkylation de l'histidine montrent qu'elle fonctionne sur plus de dix peptides non-protégés différents, mais ne seront pas mentionnés ici par souci de concision.

Figure 1-16 A – C²-Alkylation sélective de l'histidine dans le saralasin par lumière bleue et DHP [62]; B – C²-Alkylation sélective de l'histidine d'un peptide par l'utilisation de sels de sulfonates et de TBHP [63]



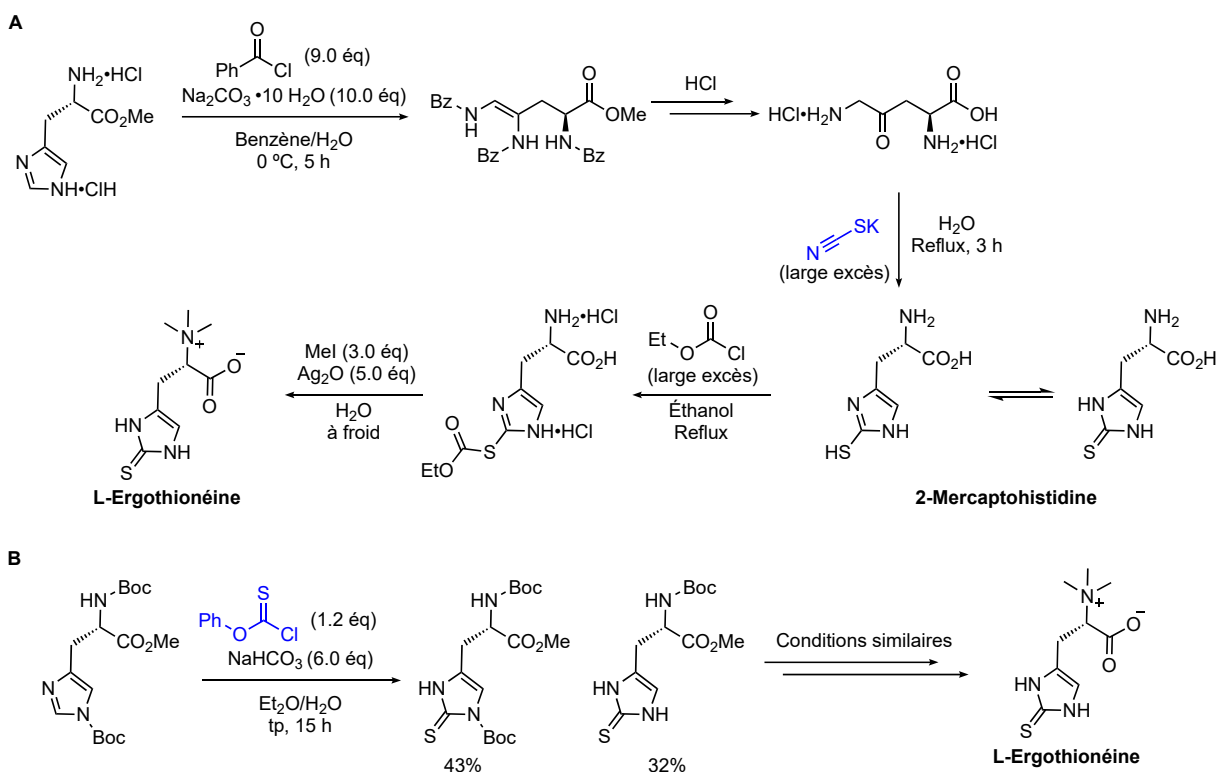
À ce jour, aucun exemple d'alkylation sélective à la position C⁵ de l'imidazole de l'histidine n'a été rapporté dans la littérature. Cependant, bien que sporadique, la modification de cette position par oxydation peut être mentionnée.

1.3.2 Oxydation de la chaîne latérale de l'histidine

Isolé pour la première fois par Charles Joseph Tanret en 1909, l'ergothionéine est un acide aminé naturel non-protéino-gène synthétisé par le champignon *Claviceps purpurea*.^[64] Sa structure chimique est simplement une histidine oxydée par une fonction thione à sa position C² de l'imidazole et une triméthylamine sur son azote terminal. Sa synthèse très instable et difficile à isoler, il a longtemps été l'objet d'étude pour les chimistes dans la quête de sa préparation en laboratoire. Ce n'est qu'en 1951, que sa

synthèse a été rapportée pour la première fois par le groupe de Heath.^[65] Une longue voie synthétique de six étapes qui à partir de méthyle ester histidine passe par une ouverture de l'imidazole par du chlorure de benzoyle, suivit de sa fermeture et de l'installation du thione grâce au thiocyanate de potassium. L'ergothionéine est par la suite obtenue par méthylation de l'azote terminal sur l'intermédiaire carboéthoxythiohistidine suivie d'une étape de déprotection (Figure 1-17A). Beaucoup plus tard, le groupe de Yadan a eu l'idée d'incorporer la source de soufre directement à la première étape, à travers l'utilisation du chlorothioformate de phényle, ce qui a permis d'obtenir, sans intermédiaire, le mercaptohistidine et d'écourter la synthèse de deux étapes (Figure 1-17B).^[66]

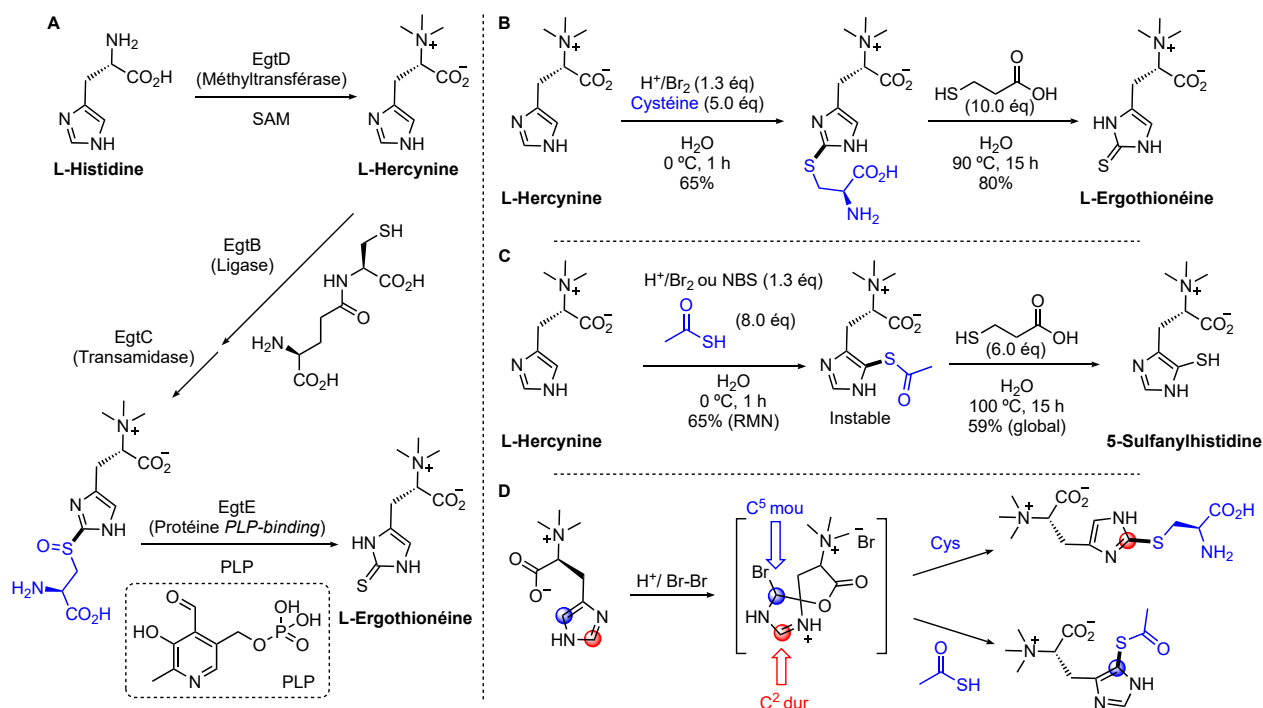
Figure 1-17 Synthèse de la L-ergothionéine : A – Thiocyanate comme source de soufre ; B – Chlorothioformate de phényle comme source de soufre



Récemment, en s'inspirant de la voie biosynthétique de l'ergothionéine, ce même groupe de recherche a proposé une nouvelle voie de synthèse de l'ergothionéine. En effet, ce dernier est produit par les champignons à partir de l'histidine en quatre étapes dont la première serait la triméthylation de l'amine terminale grâce à une molécule de SAM catalysée par une méthyltransférase, l'EgtD. L'hercynine ainsi formée, est convertie en hercynine cystéinesulfoxyde à l'aide de deux enzymes (EgtB et EgtC) et du γ -glutamylcystéine. Cet intermédiaire est finalement clivé en ergothionéine par une dernière enzyme, le EgtE (Figure 1-18A). Ainsi, en partant directement de l'hercynine, en remplaçant le γ -glutamylcystéine et

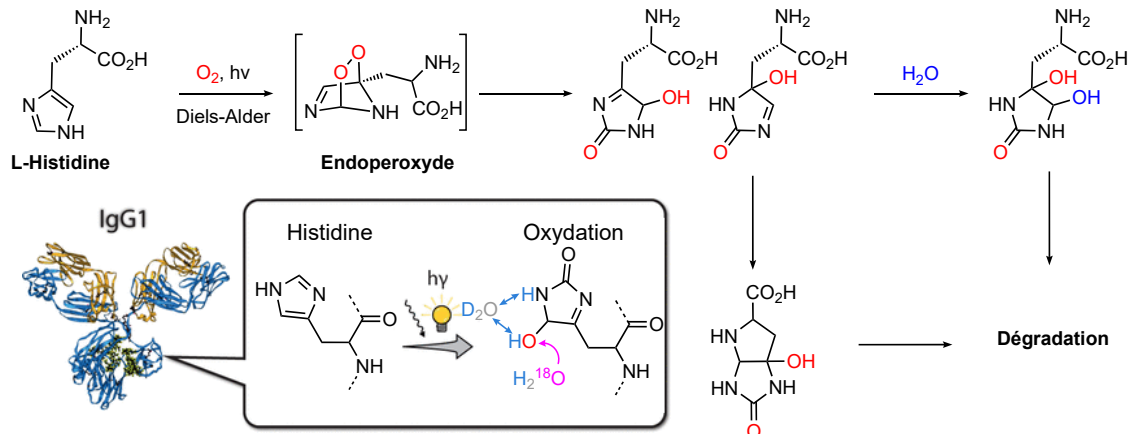
la protéine EgtE par une simple cystéine en milieu acide et bromé, et par de l'acide mercaptopropionique respectivement, ils ont réussi à mimer la biosynthèse de l'ergothionéine avec un rendement global de 52% (Figure 1-18B).^[67] Curieusement, en employant les mêmes conditions de réaction mais en remplaçant cette fois la cystéine par de l'acide thioacétique, la position C² est épargnée et a laissé place à la thiocyclation sélective de la position C⁵ de l'imidazole (Figure 1-18C). Ce phénomène est encore mal compris à ce jour, mais ils proposent un mécanisme à travers lequel l'état de transition bromospirolactone pourrait se faire attaquer préférentiellement à la position C² par un nucléophile dit dur comme la cystéine, et en position C⁵, par un nucléophile dit mou tel que l'acide thioacétique (Figure 1-18D).^[68]

Figure 1-18 A – Voie biosynthétique de l'ergothionéine ; B – Synthèse de l'ergothionéine à travers l'utilisation de la cystéine ; C – C⁵-thiolisation à partir de l'hercynine et d'acide thioacétique ; D – Mécanisme proposé de la sélectivité C²/C⁵



Présentant un intérêt essentiellement limité à la compréhension biomécanistique, l'histidine peut également s'oxyder par la présence d'oxygène via une réaction de Diels-Alder photocatalysée par le biais d'un intermédiaire endoperoxyde.^[69] En effet, des preuves d'histidines oxydées ont été observées sur l'anticorps monoclonal IgG1^[70] et il semblerait que les efforts scientifiques ont été placés sur l'étude mécanistique du phénomène et non sur l'exploitation éventuelle de ce genre de composés dans le domaine synthétique (Figure 1-19).

Figure 1-19 Mécanisme proposé de l'oxydation de l'histidine observée sur l'anticorps monoclonal IgG1. Image tirée de [70]

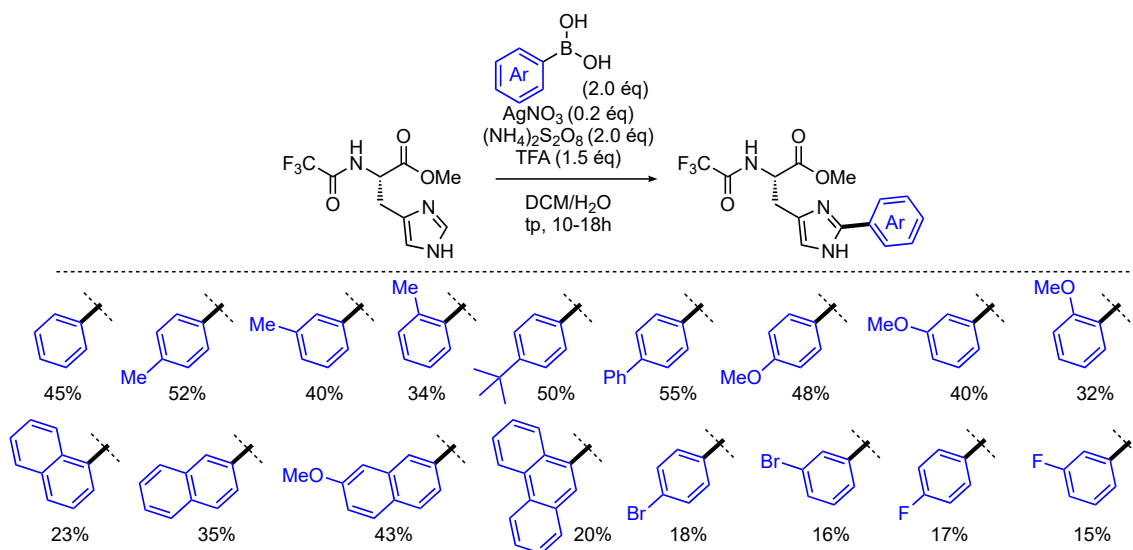


Les modifications de l'histidine par alkylation ont largement été rapportées dans la littérature, mais celles-ci restent toutefois modestes face aux méthodes d'arylation d'histidine décrites dans les revues scientifiques.

1.3.3 Méthodes d'arylation de la chaîne latérale de l'histidine

Comme précédemment, l'imidazole de la chaîne latérale peut être arylé soit sur les azotes N^π et N^τ , soit sur les carbones C^2 et C^5 . Dans le cadre de ces deux dernières, à ce jour uniquement deux méthodes d'arylation directe ont été rapportées dans la littérature : une par réaction radicalaire et une autre par activation C–H. La méthode radicalaire s'inspire des conditions de Minisci vues plus tôt, mais en utilisant cette fois-ci l'acide boronique à la place de l'acide carboxylique. De la même façon, l'arylation est sélective sur le carbone le plus électrophile C^2 et sans trop de surprise, les groupements aromatiques appauvris en électrons donnent des rendements nettement moins bons que leurs homologues riches en électrons (Figure 1-20).^[71] Aujourd'hui, aucune autre méthode d'arylation, que ce soit à la position C^2 ou C^5 par voie radicalaire n'a encore été rapportée.

Figure 1-20 C²-Arylation de l'imidazole de l'histidine protégée dans les conditions radicalaires de Minisci



Plus tard, ce même groupe de recherche a élaboré la deuxième méthode d'arylation par C–H activation, plus robuste et versatile avec des rendements bien plus appréciables. La position C⁵ a pu être sélectivement arylée en mettant en présence l'histidine protégée avec une quantité catalytique de palladium et de ligand phosphoré, de carbonate de potassium et d'acide pivalique comme le couple de déprotonation métallique concertée (CMD) dans du diméthylformamide (DMF) aux micro-ondes à 140 °C (Figure 1-21A).^[72] De manière assez surprenante, ils ont par la suite découvert qu'en ajoutant du cuivre(I) dans les conditions, cela permettait de changer complètement la régiosélectivité de l'arylation, passant cette fois sur le carbone C² de l'imidazole (Figure 1-21B).^[73] La régiosélectivité peut s'expliquer par le mécanisme de la réaction. Sans la présence de cuivre, un mécanisme classique d'activation C–H dans lequel se succède une addition oxydante du palladium, une CMD et une élimination réductrice, pourrait expliquer la sélectivité de l'arylation du carbone C⁵. Dans le cas opposé, le cuivre formerait d'abord l'organocuprate au niveau du carbone le plus acide suivi d'une transmétallation qui aboutirait à l'élimination réductrice, donnant le produit C²-arylé (Figure 1-22).

Figure 1-21 A – C⁵-Arylation sélective de l'imidazole de l'histidine protégée ; B – C²-Arylation sélective de l'imidazole de l'histidine protégée par C–H activation

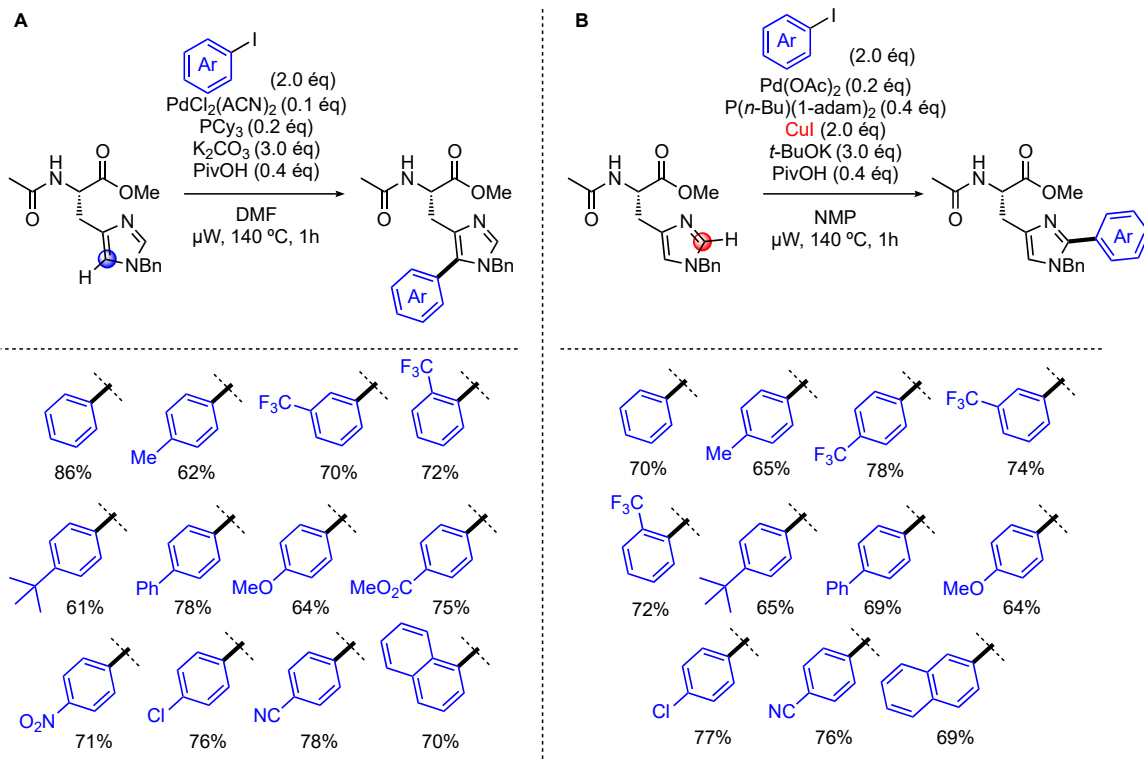
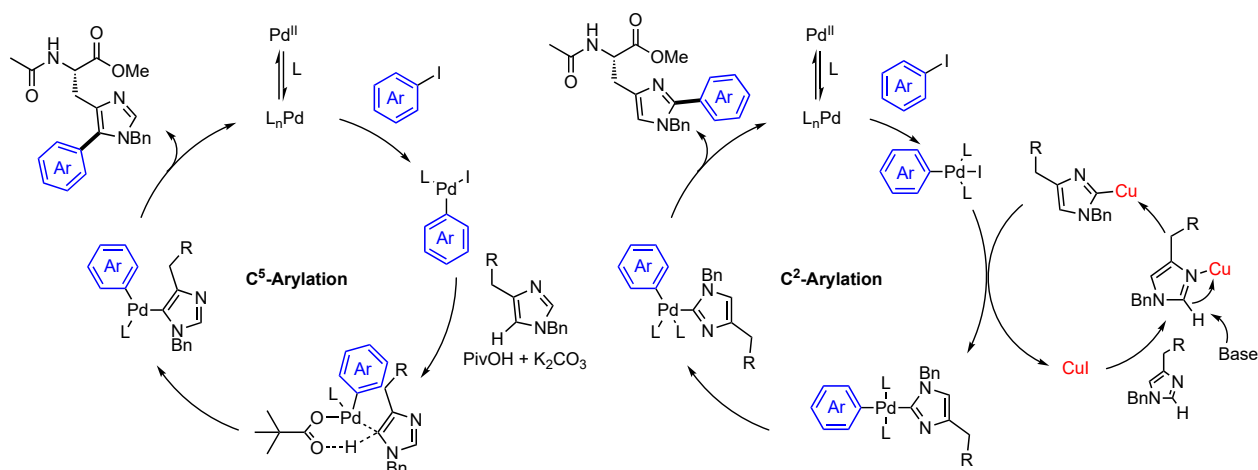
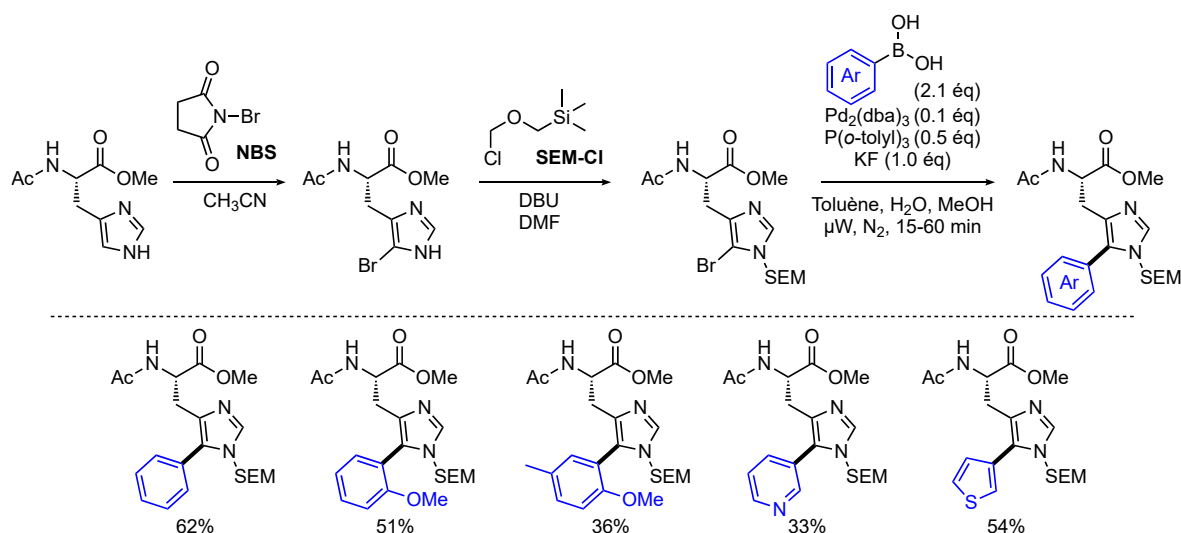


Figure 1-22 Mécanismes proposés tentant d'expliquer la régiosélectivité de l'arylation par C–H activation



Une autre méthode indirecte, plus ancienne, était effectuée par couplage de Suzuki-Miyaura sur l'histidine préalablement bromée en position C⁵ à l'aide du N-bromosuccinimide (NBS). Malgré le fait que cette méthode soit bien moins explorée, elle permet néanmoins d'apporter une réponse favorable sur la tolérance d'un groupement hétéroaromatique (Figure 1-23).^[74]

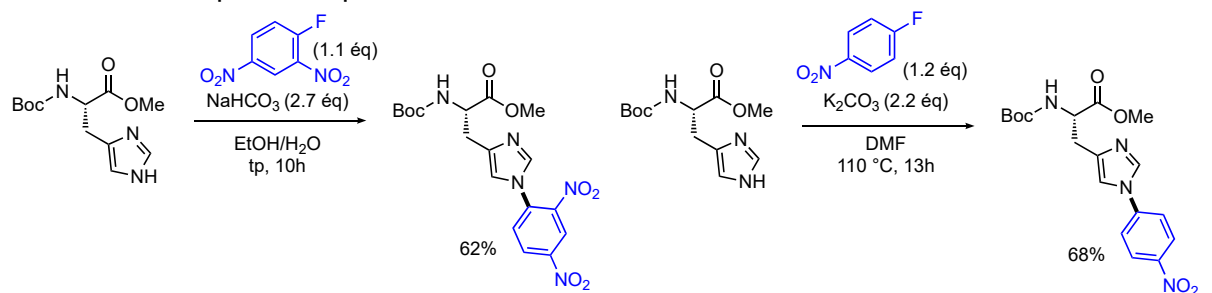
Figure 1-23 C⁵-Arylation indirecte par couplage Suzuki-Miyaura



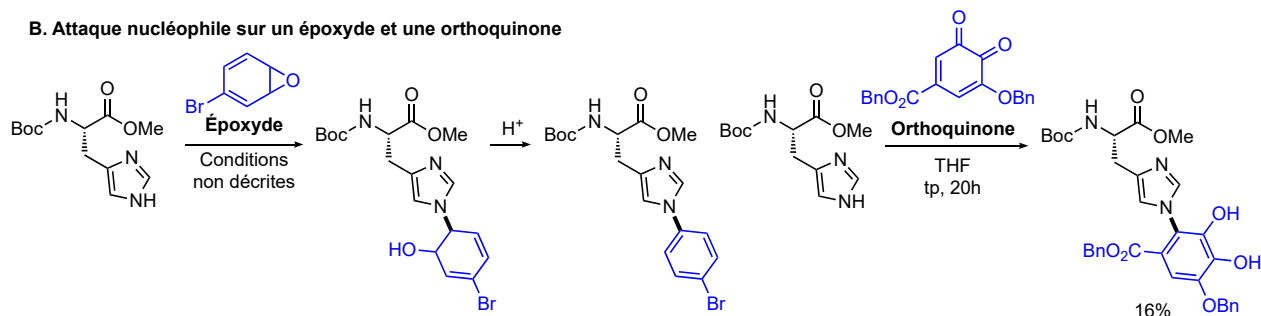
Pour ce qui est des méthodes de N-arylation de l'imidazole de l'histidine, il en existe un nombre plus conséquent et chronologiquement, les pionniers dans ce domaine exploitaient l'avantage du caractère nucléophile de l'azote N^τ pour réaliser différentes modifications. En effet, les premiers signes d'histidine arylée ont été rapportés dans les années 50 où une méthode de substitution nucléophile aromatique (S_NAr) donnait un mélange d'histidine N^{terminal}-arylée et N^τ-arylée en mettant en présence l'histidine avec du fluorodinitrobenzène.^[75, 76] Plus tard, Merrifield proposa de protéger d'abord l'azote terminal afin d'obtenir exclusivement l'arylation de l'imidazole (Figure 1-24A).^[77] La nécessité de la présence d'un groupement électro-attracteur ainsi qu'un halogène sur le cycle aromatique rend inévitablement la méthode très limitée en termes de diversité de groupements. Par la suite, dans le cadre d'études biochimiques, soit très loin de l'objectif de modification, l'histidine s'est retrouvée arylée par une attaque nucléophile, mais cette fois sur des composés « pro-aromatiques » tels que l'époxyde bromocyclohexadiène^[78] et l'orthoquinone (Figure 1-24B).^[79] En plus de l'inconvénient de devoir les préparer, la diversité des groupements aromatiques reste toujours très limitée. Il aura fallu attendre les travaux du groupe de Konopelski pour voir les prémices d'une diversité des groupements aromatiques installables sur l'histidine. En effet, en s'inspirant du couplage croisé organoplomb/cuivre développé par Barton,^[80, 81] ils ont réussi à installer des cycles aromatiques à la fois enrichis et appauvris en électrons et même de coupler l'imidazole de l'histidine avec la chaîne latérale d'une tyrosine protégée en ayant un rendement convenable (Figure 1-24C).^[82-84]

Figure 1-24 A – N^ε-Arylation par S_NAr ; B – N^ε-Arylation par attaque nucléophile sur un pro-aromatique ; C – N^ε-Arylation par couplage croisé de Barton

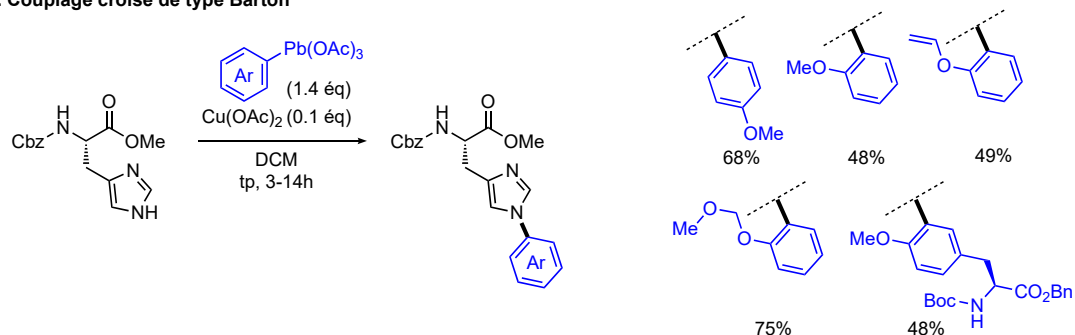
A. Substitution Nucléophile Aromatique



B. Attaque nucléophile sur un époxyde et une orthoquinone



C. Couplage croisé de type Barton

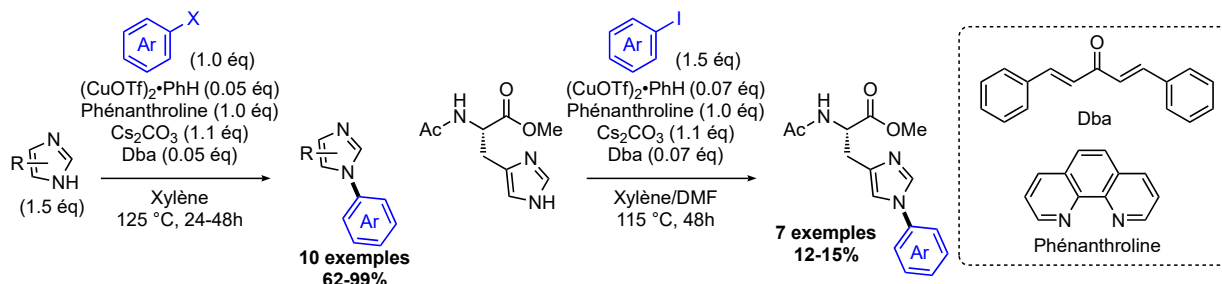


Conscient des problèmes de toxicité liés aux organoplombs et basé sur les travaux de Buchwald relatifs aux arylations de l'imidazole,^[85] Hanzlik réussit non sans peine à obtenir des nouveaux dérivés aromatiques de l'histidine en la mettant en présence d'halogénures d'aryle, d'une quantité catalytique de cuivre(I) avec de la phénanthroline et du dibenzylidèneacétone (Dba) comme ligands et du carbonate de césium en guise de base dans un mélange xylène/DMF à 115 °C (Figure 1-25A).^[86] Très proche des conditions optimales, il semblerait que l'activation de la liaison N–H de l'imidazole de l'histidine par simple chauffage n'était pas suffisante et qu'un chauffage par micro-onde était la clé, comme le démontre Jain à travers ses études. En effet, en y changeant le cuivre et le ligand et en chauffant simplement par micro-ondes, ces travaux remarquables impressionnent tant par la diversité des groupements aromatiques tolérés que par les rendements obtenus. Effectivement, toutes les positions (para, meta, ortho) des substituants sur le cycle

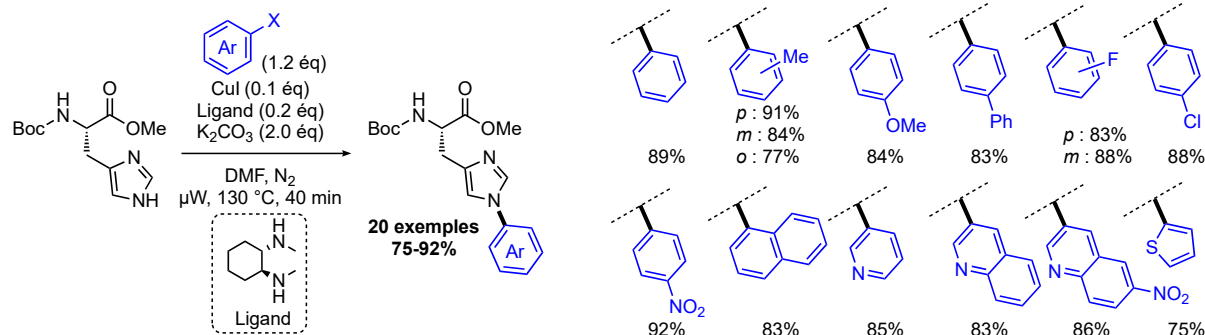
aromatique sont tolérées, qu'ils soient électrons donneurs ou accepteurs. De plus, les groupements hétéroaromatiques sont également applicables (Figure 1-25B).^[87]

Figure 1-25 A – Transposition des conditions d'arylation de l'imidazole dans le cadre de l'histidine ; B – N¹-Arylations de l'imidazole de l'histidine protégée par couplage de type Ullman-Goldberg chauffées par micro-onde

A. Arylations de type Ullman-Goldberg de l'imidazole par Buchwald et de l'histidine par Hanzlik

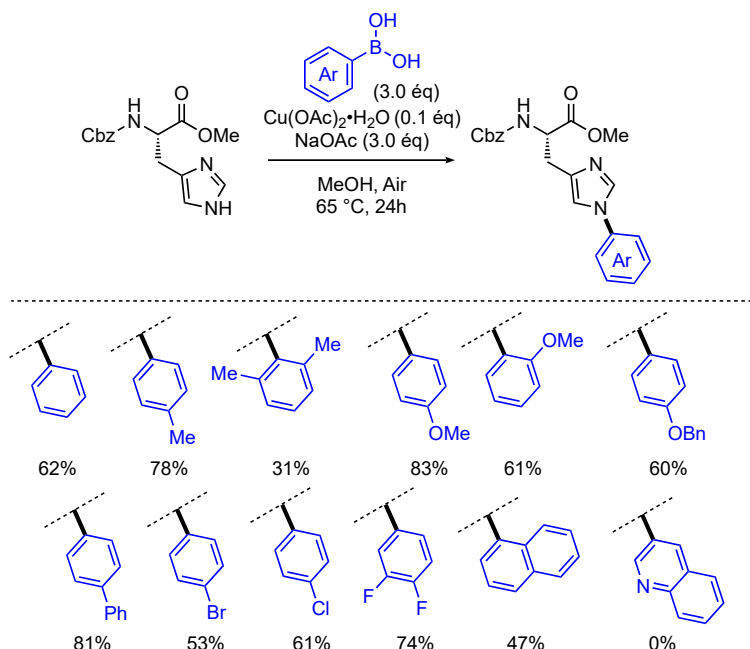


B. Couplages de type Ullman-Goldberg chauffés par micro-onde



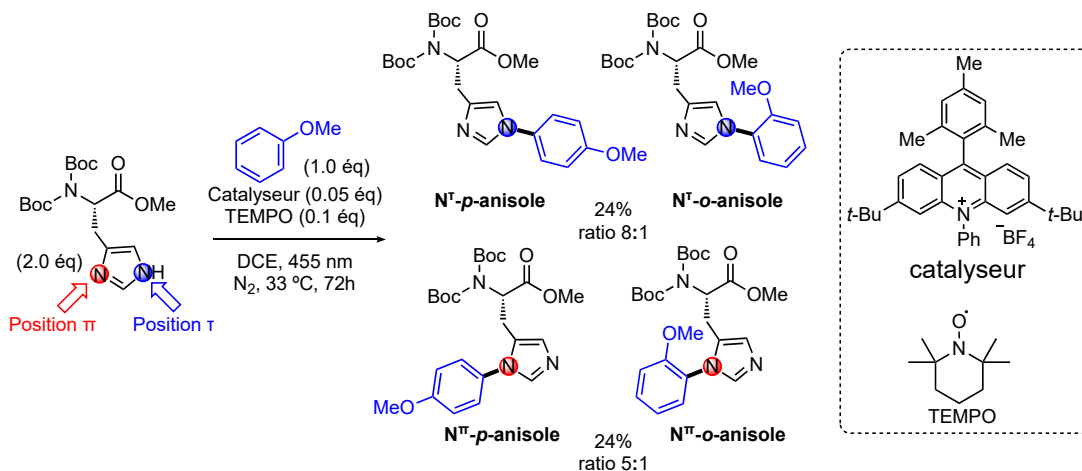
Une méthode alternative à celle par couplage de Ullman-Goldberg a été décrite par Campagne qui permet l'arylation de l'imidazole par un couplage croisé de Chan-Lam-Evans. Ces conditions ont l'avantage de se faire à une température plus douce et de se passer de solvant anhydre. En contrepartie, les cycles hétéroatomiques ne sont plus compatibles dans ces circonstances (Figure 1-26).^[88]

Figure 1-26 N^τ-Arylation de l'imidazole de l'histidine protégée par un couplage Chan-Lam-Evans



À ce jour, aucune méthode ne mentionne l'arylation régiosélective de la position N^τ de l'imidazole de l'histidine. Cependant, une méthode par photochimie proposée par Nicewicz peut apporter un semblant de réponse, mais malheureusement le manque de régiosélectivité rend cette dernière moins intéressante et reste encore sous explorée (Figure 1-27).^[89]

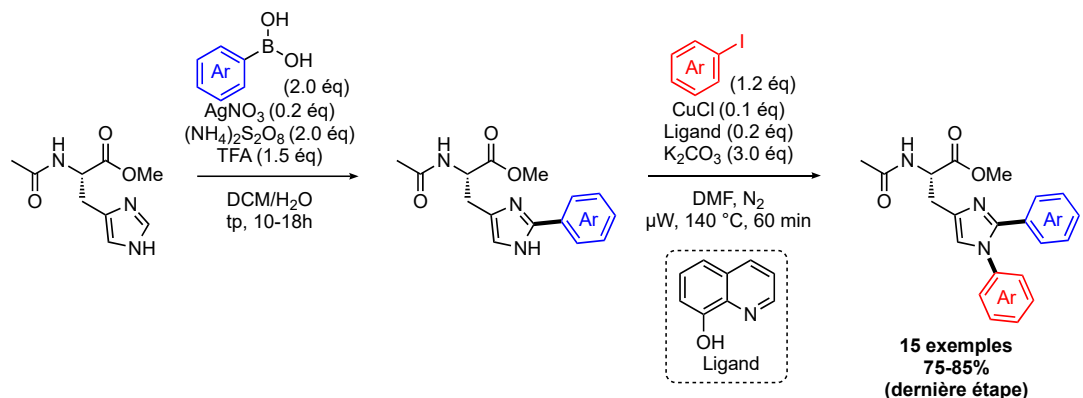
Figure 1-27 N-Arylations par réaction photochimique



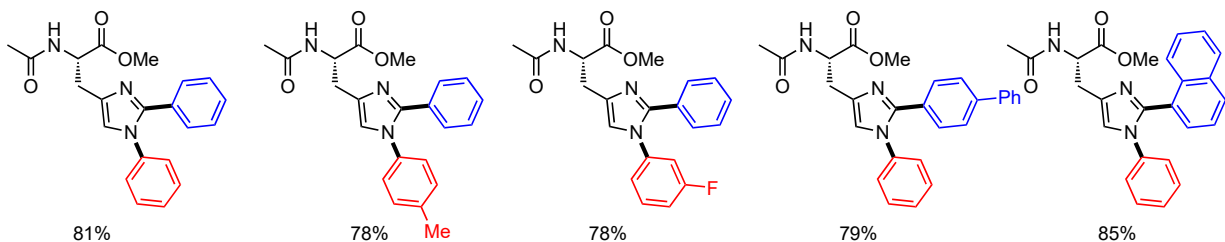
Il a été démontré qu'il était néanmoins possible de combiner deux méthodes d'arylations régiosélectives afin d'obtenir l'histidine doublement arylée à la fois à la position C² et N^τ. Pour ce faire, la position C² est

d'abord arylée avec les conditions de Minisci avant d'effectuer un couplage de type Ullman-Goldberg sur l'azote N^r de manière complètement régiosélective (Figure 1-28).^[90]

Figure 1-28 Double arylation de l'imidazole par réactions successives régiosélectives



Quelques exemples :



Ces derniers résultats datant de 2017 clôturent à ce jour les recherches autour de la modification de l'histidine et concluent par une remarquable preuve de la versatilité et de la diversité de sa chimie.

En parallèle à ces études, d'autres groupes ont proposé des méthodes de modification pour d'autres acides aminés, tels que le tryptophane ou encore la tyrosine. Ces études ne seront pas détaillées dans le cadre de cette thèse, mais il serait incongru de ne pas mentionner que notre groupe de recherche a participé à l'enrichissement méthodologique des modifications de ces deux acides aminés à travers l'utilisation d'organobismuthines.^[91, 92]

CHAPITRE 2

LES BISMUTHINES, UN NOUVEL OUTIL

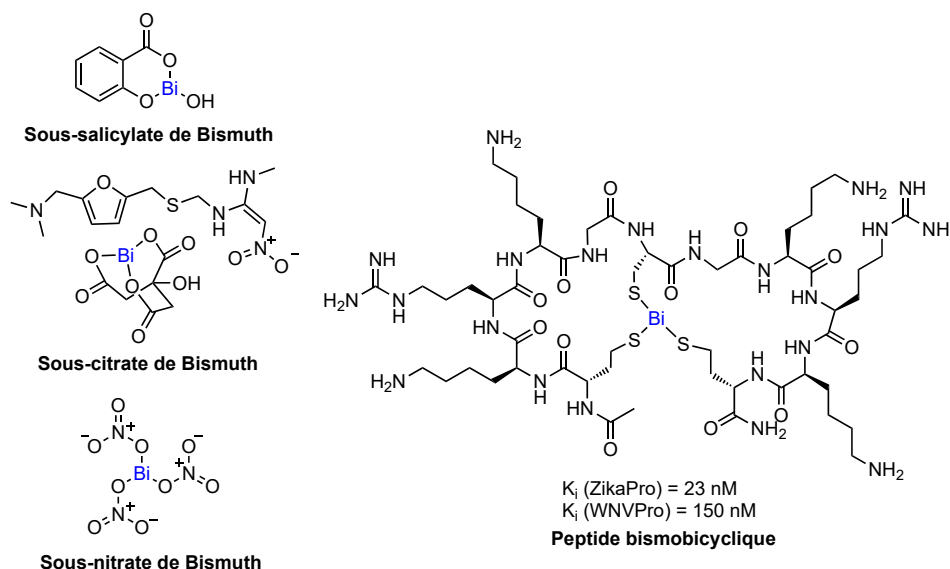
Dans ce chapitre il sera question tout d'abord d'appréhender les différentes facettes du bismuth avant de survoler ses multiples utilisations dans différents domaines d'application. En revanche, le domaine de la chimie de synthèse sera un peu plus détaillé et on verra comment le bismuth organique a su trouver sa place parmi les métaux de transition déjà fort employés dans ce secteur.

2.1 Les généralités

Identifié pour la première fois par Claude François Geoffroy en 1753, le bismuth est un métal du tableau périodique des éléments portant le numéro atomique 83 avec une configuration électronique $[\text{Xe}] 4f^{14}5d^{10}6s^26p^3$. Étant le dernier élément naturel de la famille des pnictogènes, soit la colonne 15 du tableau, il a longtemps été considéré comme le métal le plus lourd non radioactif. Toutefois, des chercheurs de l'institut d'astrophysique spatiale d'Orsay ont démontré que le bismuth émettait finalement des particules alpha lors de sa désintégration en thallium et en l'hélium avec une demi-vie de $1,9 \times 10^{19}$ années. Une demi-vie plus grande que l'âge de la Terre elle-même explique ainsi pourquoi il est considéré comme stable.^[93] À peu près deux fois plus abondant que l'or dans la croûte terrestre, le bismuth se trouve généralement sous la forme de minerai de bismuthinite (Bi_2S_3) et de bismite (Bi_2O_3). Son isolation élémentaire provient le souvent du débismuthage du plomb par des procédés pyrométallurgiques dont la Chine intervient comme le premier producteur au monde suivit du Laos et de la Corée du Sud.^[94]

Parce que le bismuth partage des similitudes physiques avec ses proches voisins, il a longtemps été confondu avec le plomb ou l'antimoine. Cependant contrairement à ces derniers, le bismuth montre une toxicité tout de même moindre et présente même des propriétés thérapeutiques.^[95] En effet, sous la forme de sous-salicylate dans le Pepto-Bismol®, de sous-citrate dans le Zantac® ou de sous-nitrate dans le Mammol®, ces principes actifs à base de bismuth sont utilisés comme un antiacide qui permet de soulager soit les brûlures et les ulcères de l'estomac, soit les démangeaisons causées par des piqûres ou de l'eczéma. De plus, usant des capacités chalcophile du bismuth (aime à se lier fortement au soufre), le groupe de Nitsche a récemment démontré qu'un peptide bismobicycliqué pouvait avoir un effet bénéfique contre les protéases du virus Zika (ZikaPro) et du Nil occidental (WNVPro) (Figure 2-1).^[96] Ses formidables propriétés doivent toutefois être nuancées, puisque lorsqu'accumulé dans l'organisme, le bismuth peut néanmoins provoquer des effets indésirables comme des néphropathies, des encéphalopathies, des ostéoporoses ou encore des hépatites.^[97]

Figure 2-1 Exemples de composés contenant du Bismuth aux propriétés thérapeutiques



Outre le domaine pharmaceutique, deux autres domaines se disputent également les ressources de bismuth à savoir le domaine de la chimie des matériaux et la chimie en synthèse organique. Dans le premier cas, c'est parce qu'il partage des propriétés physiques communes avec le plomb que le bismuth est devenu très rapidement son équivalent non toxique. Le plomb est ainsi remplacé par le bismuth dans les peintures, les écrans pararayons X ou encore dans les alliages métalliques qui en comportaient comme les tuyauteries ou les munitions de chasse. D'autres applications existent mais ne seront pas détaillées dans le cette thèse pour des raisons évidentes de concision. En revanche, dans le cadre de la chimie en synthèse organique, le bismuth et ses applications vont être vus un peu plus en détail.

2.2 Les organobismuthines, leurs synthèses et applications

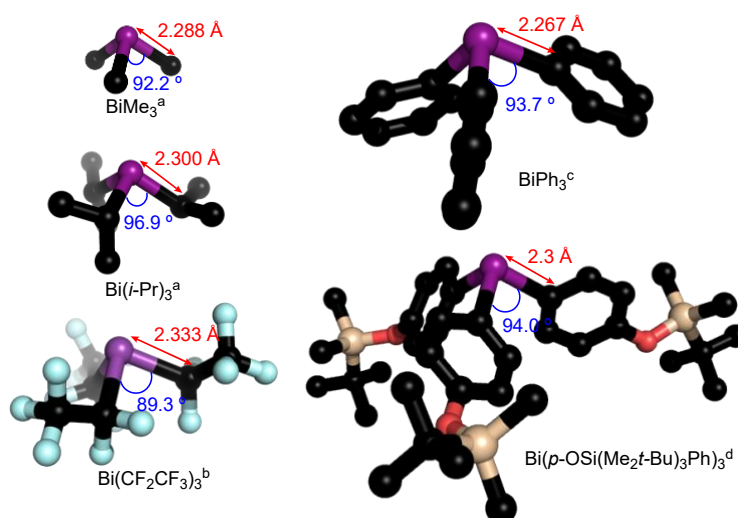
2.2.1 Synthèse des organobismuthines trivalents

On nomme par organobismuthines une espèce qui comporte au moins une liaison Bi-C. Ses formes stables se distinguent d'une part par les bismuths trivalents d'état d'oxydation +3, et d'une autre part par les pentavalents d'état d'oxydation +5. Les bismuths trivalents sont les plus documentés et se différencient selon s'ils sont trialkylés ou triarylés. Au-delà de la différence structurelle, les trialkylbismuthines sont liquides et pyrophoriques, alors que les triarylbismuthines sont solides et tout à fait stables à l'air. Ceci s'explique par la présence du doublet non-liant du bismuth fortement polarisé dans le cas des alkylbismuthines et qui est complètement diffus par résonance grâce aux groupements aromatiques dans le cas des arylbismuthines. Par des techniques de cryo-cristallisation et/ou cristallisation classique, les structures des alkyles et aryles bismuths ont pu être analysées en détail. Tous deux adoptent une géométrie

pyramidale trigonale avec des distances et des angles qui varient selon les groupements (Figure 2-2). On peut s'apercevoir que les angles de C–Bi–C sont nettement inférieurs à la valeur théorique de 109,5 ° établie selon la théorie de la répulsion des paires d'électrons de la couche de valence (RPECV). Ce phénomène particulier peut s'expliquer par les effets relativistes du bismuth qui sans rentrer dans les détails quantiques, permettent au doublet d'électrons libres de se localiser préférentiellement dans son orbitale 6s plutôt que dans l'orbitale hybridée sp^3 , ce qui a pour conséquence un angle de liaison réduit.^[98]

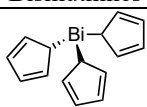
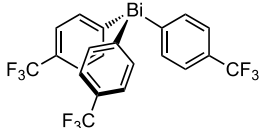
Figure 2-2 Structures cristallographiques de quelques organobismuthines trivalentes

Tiré de : ^a Schulz^[99] ; ^b Hoge^[100] ; ^c Bučinský^[Erreur! Référence de lien hypertexte non valide.] ; ^d Gagnon^[102]



Bien que moins faciles à isoler, les trialkylbismuthines seront les premières à être synthétisées. En effet, le professeur Löwig mentionnera en 1850 qu'en appliquant les mêmes conditions de synthèse des organoantimoine, il était possible d'obtenir le triéthylbismuth, sans plus de détails.^[103] Cette synthèse sera détaillée deux ans plus tard par Breed où il indiquera qu'en chauffant de l'idoéthane avec un alliage de bismuth et potassium il était possible d'obtenir le produit désiré.^[104] Fortement inspiré par cette méthode, en 1887 Michaelis et Polis rapporteront la première synthèse de triarylbismuthines. Pour ce faire, ils ont chauffé du bromobenzène avec un alliage de bismuth et sodium pour obtenir le triphénylbismuth attendu.^[105] À ce jour, la méthode la plus répandue est celle de l'utilisation d'un organomagnésien sur un sel de trihalogénobismuthine. Cette dernière datant du début du 20^{ième} siècle reste aujourd'hui la plus simple à mettre en place et permet à la fois d'obtenir des trialkyl-^[106, 107] et des triarylbismuths.^[108-110] Loin d'être la seule méthode disponible, d'autres groupes ont utilisé d'autres organométaux. Le Tableau 2-1 en liste quelques-uns sans être exhaustif.

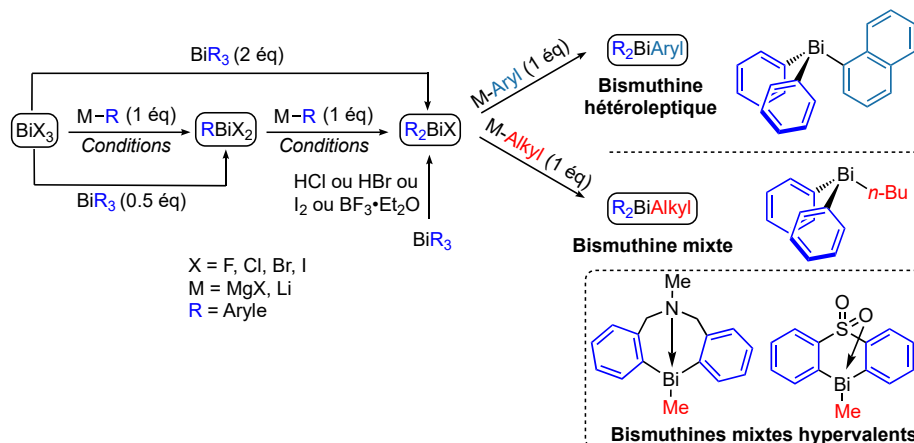
Tableau 2-1 Organométaux compatibles à la synthèse d'organobismuthines

$\text{BiX}_3 \xrightarrow[\text{Conditions}]{\text{M-R}} \text{BiR}_3 \quad \begin{matrix} \text{X} = \text{Cl, Br} \\ \text{M} = \text{Métal} \end{matrix}$					
Auteurs	Organométaux	Bismuthines	Auteurs	Organométaux	Bismuthines
Marquardt [111]	$\text{Me}_2\text{Zn}, \text{Et}_2\text{Zn}$	$\begin{matrix} \text{Me}-\text{Bi}-\text{Me} \\ \quad \\ \text{Me} \quad \text{Et} \end{matrix}$	Fisher et Schreiner [112]	NaCp	
Hilpert, Dimart et Grüttner [113]	Al_4C_3	$\begin{matrix} \text{Me}-\text{Bi}-\text{Me} \\ \\ \text{Me} \end{matrix}$	Dickson et West [114]	LiAlEt_4	$\begin{matrix} \text{Et}-\text{Bi}-\text{Et} \\ \\ \text{Et} \end{matrix}$
Hilpert et Grüttner [115]	HgPh_2	$\begin{matrix} \text{Ph}-\text{Bi}-\text{Ph} \\ \\ \text{Ph} \end{matrix}$	Naumann et Tyrre [116]	$\text{Cd}(\text{CF}_3)_2$	$\begin{matrix} \text{F}_3\text{C}-\text{Bi}-\text{CF}_3 \\ \\ \text{CF}_3 \end{matrix}$
Zakharkin et Okhlobystin [117]	AlR_3	$\begin{matrix} \text{Et}-\text{Bi}-\text{Et} & \text{Pr}-\text{Bi}-\text{Pr} \\ & \\ \text{Et} & \text{Pr} \\ i\text{-Bu}-\text{Bi}-i\text{-Bu} & \\ & \\ i\text{-Bu} & \end{matrix}$	Whitmire, Roesky et al. [118]	$\text{Li}(p\text{-CF}_3\text{Ph})$	

Lors des synthèses des organobismuthines rapportées précédemment, un composé d'halogénure de diorganobismuth est souvent décrit comme étant un sous-produit. Ce dernier issu de la conversion partielle du trihalogénobismuth, peut être obtenu plus systématiquement en ajoutant simplement un ou deux équivalents d'organolithien par rapport au sel de bismuth, comme le décrit Breunig.^[119] D'autre part, il est également possible d'obtenir ces espèces en faisant réagir du diéthyléthérate de trifluorure de bore, du bromure d'hydrogène ou encore du diiode avec du triarylbismuth conduisant respectivement au fluorure,^[120] au bromure,^[121] ou à l'iodure de diarylbismuth.^[122, 123] Une méthode par dismutation conduit également aux mêmes espèces et semble être la méthode la plus utilisée. Elle consiste en la simple mise en réaction du triarylbismuth avec son équivalent trihalogéné. La subtilité de cette dernière méthode est qu'une addition lente du sel de bismuth à une solution de triarylbismuth donnera bien l'espèce monohalogénée correspondante mais que l'addition d'ordre contraire conduira au dihalogénure d'arylbismuth.^[124] L'existence de ces espèces a permis l'émergence de nouveaux organobismuths trivalents dits hétéroleptiques. En effet, en ajoutant du naphthylmagnésien sur le bromure de diphenylobismuth,^[125] Challenger réussit la première synthèse d'un tel organobismuth et inspirera par la suite plusieurs groupes soit à travers les mêmes conditions,^[124, 126] soit à travers des turbo Grignard,^[127] soit à travers des organolithiens.^[128] En poussant le concept un peu plus loin, il est possible d'obtenir des organobismuths hétéroleptiques dits mixtes, ainsi nommés car le bismuth porte à la fois des groupements aromatiques et alkyliques. Ces espèces sont sensibles à l'air mais ne sont pas pyrophoriques comme leurs homologues trialkyliques.^[119, 129] Par ailleurs, l'utilisation d'une stratégie dans laquelle les deux cycles aromatiques sont liés par une fonction portant un caractère datif conduit à des organobismuthines trivalents hypervalents et

permet d'avoir des bismuthines tout à fait stables à l'air. Cette stratégie est encore peu explorée, seuls quelques exemples ayant été rapportés dans la littérature à ce jour (Figure 2-3).^[130-132] Ce phénomène d'hypervalence se caractérise par la faculté du bismuth à enfreindre la règle de l'octet et à accepter davantage d'électrons, ce qui mène parfois à des bismuths pentavalents.

Figure 2-3 Voies de synthèse des halogénures de bismuth et leurs transformations

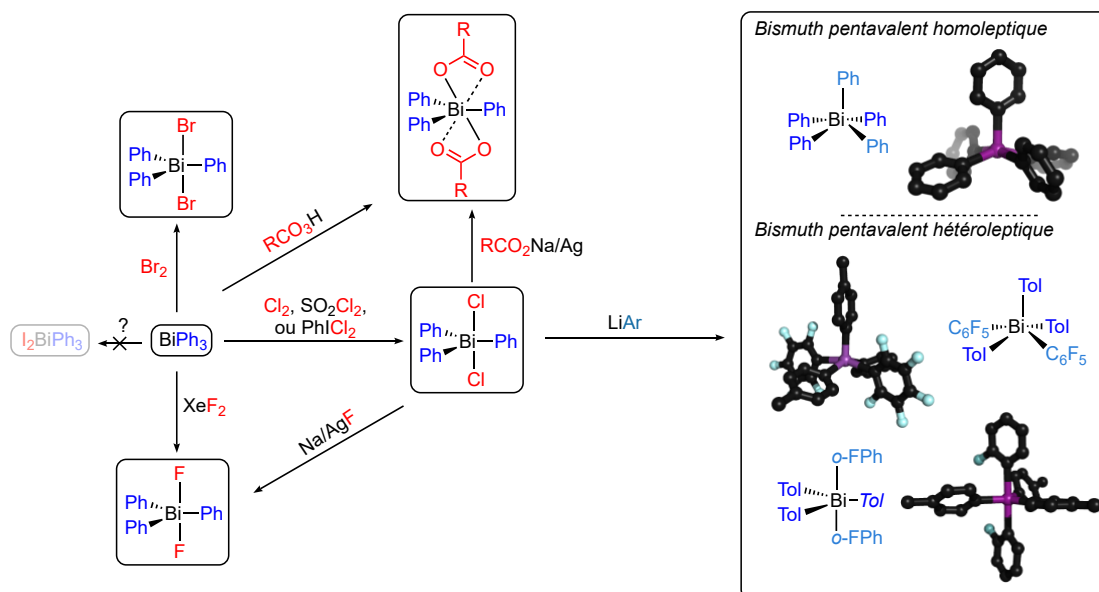


2.2.2 Synthèse des organobismuthines pentavalents

Les bismuths pentavalents présentent généralement une géométrie bipyramidale à base trigonale^[133] et comme leurs homologues trivalents, les dérivés alkyls sont sensibles à l'air et peu stables à température ambiante contrairement aux aromatiques correspondants. Pour ces derniers, il est possible de trouver dans la littérature autant de méthodes de synthèse que de dérivés mais elles ont en commun le fait qu'il est souvent nécessaire de passer par un intermédiaire trivalent. Les premières synthèses de tels bismuths ont été mentionnées par Michaelis et Polis en 1887, avec leurs triphényldihalogénobismuthines préparées en faisant simplement passer du chlore gazeux ou en ajoutant une solution de dibrome à une solution refroidie de triphénylbismuth pour obtenir respectivement l'espèce pentavalente dichlorée et/ou dibromée correspondante sous forme de précipités blancs.^[105] Cette méthode de synthèse sera imitée par de nombreux groupes^[110, 134] et l'inconvénient du dichlore gazeux sera vite détourné par l'utilisation plus pratique du chlorure de sulfuryle,^[124, 135, 136] et par du dichlorure d'iodobenzène dans les cas les plus spécifiques.^[137] Pour ce qui est des autres homologues halogénés, le fluorure correspondant a pu être obtenu à partir du triphényldichlorobismuth mis en présence de fluorure de sodium^[138] ou de fluorure d'argent,^[139] ou alors obtenu directement en oxydant le bismuth trivalent avec du difluorure de xénon.^[140-142] Malheureusement, malgré les tentatives, les bismuths pentavalents iodés restent encore absents de la littérature. Dans un autre registre, il est également possible d'obtenir des dicarboxylates de triarylbismuthines soit à partir du dichlorure d'arylbismuth(V) décrit plus haut mis en présence d'un carboxylate,^[143, 144] ou bien directement

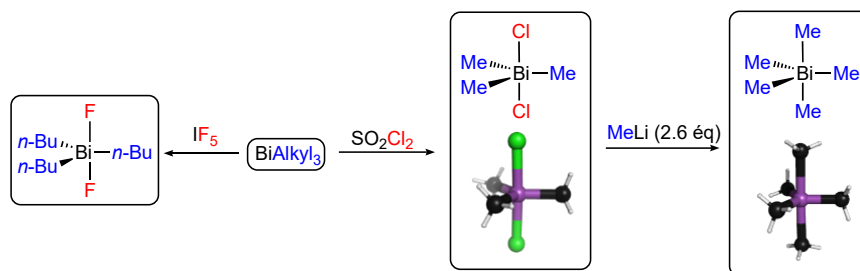
à partir de triarylbismuthines à travers une oxydation par un peracide.^[145-151] Les méthodes pour obtenir ces espèces sont diverses et variées, et ne seront pas présentées en détail dans cette thèse. Bien que très peu référencés dans la littérature, les bismuths pentavalents homo- et hétéroleptiques peuvent s'obtenir assez facilement en ajoutant un organolithien sur des dérivés halogénés correspondants. Wittig et Clauß seront les premiers à rapporter le pentaphénylbismuthine en 1952,^[152] qui inspirera par la suite les travaux de l'équipe de Seppelt avec leurs bismuthines pentavalentes hétéroleptiques. Curieusement, les pentaarylbismuthines adoptent généralement une structure plutôt pyramidale à base carrée et non plus bipyramidale trigonale comme ses homologues halogénés. Le tritolyl-*o*-difluorobismuthine endosse le rôle d'exception avec sa structure bipyramidale (Figure 2-4).^[142, 153]

Figure 2-4 Méthodes de synthèse des halogénures de triarylbismuthines et pentaarylbismuthines



Bien que très instables à température ambiante, il est également possible d'oxyder les trialkylbismuthines. En 1986, une analyse par ^{19}F -RMN de Maurer a permis de caractériser des traces du tributylodifluorobismuthine obtenu par oxydation de l'alkylbismuth correspondant avec du pentafluorure d'iode (IF_5).^[154] Plus tard, grâce à des analyses cristallographiques par Wallenhauer et de Seppelt, des preuves supplémentaires ont pu être apportées. En effet, ils ont réussi à synthétiser et à caractériser la première pentaméthylbismuthine en passant par un intermédiaire chloré de triméthylbismuthine.^[155] De manière assez surprenante, la structure géométrique du pentaméthylbismuth reste bipyramidale et n'adopte pas la structure pyramidale comme le pentaphénylbismuth (Figure 2-5).

Figure 2-5 Synthèses d'halogénures de trialkylbismuthines et du pentaméthylbismuthine

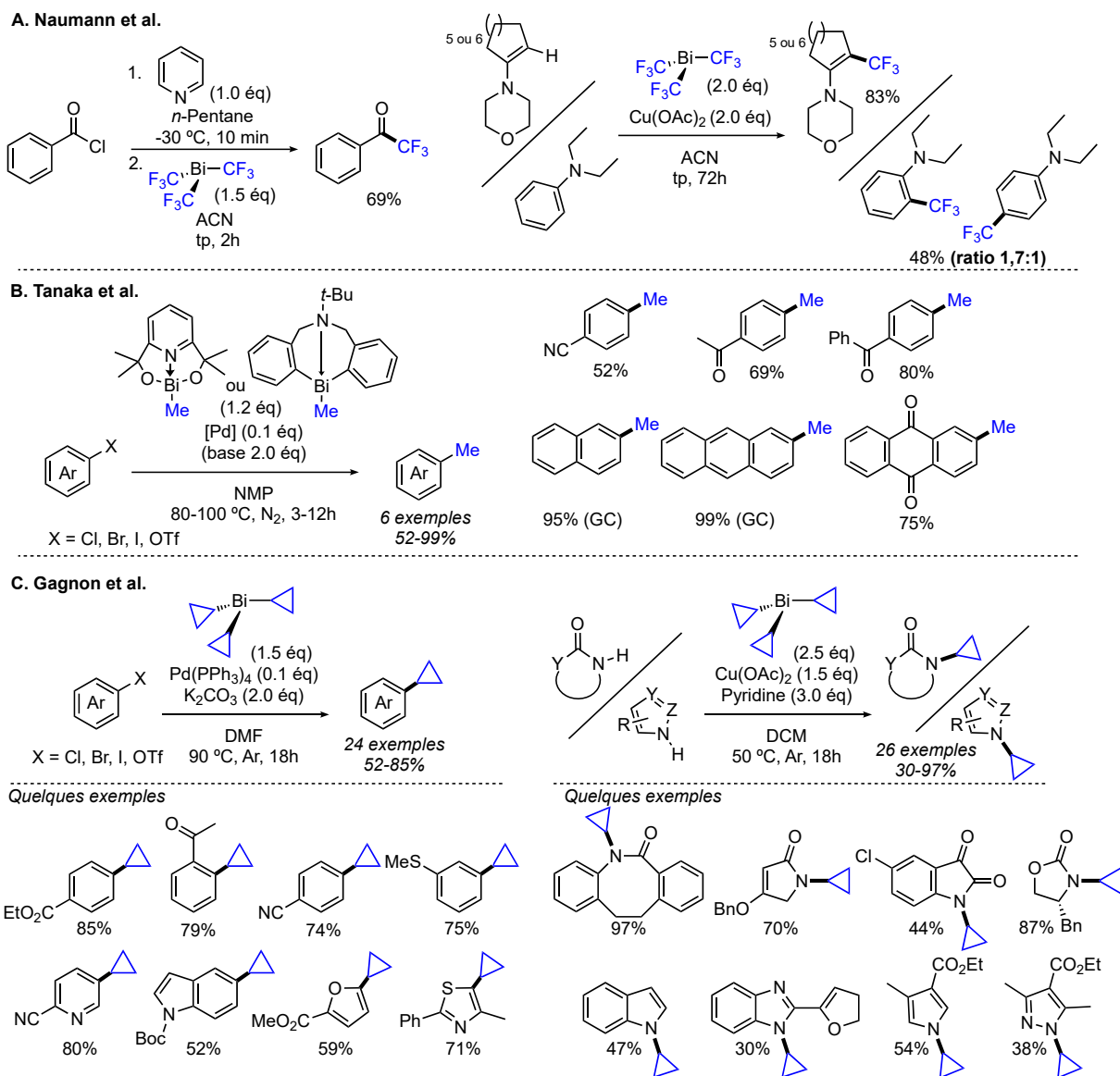


Les préparations des organobismuthines sont parfois des défis synthétiques et font souvent l'objet d'études cristallographiques. Outre l'élégance des clichés des rayons X, les organobismuthines proposent également une richesse réactionnelle appréciable.

2.2.3 Réaction d'alkylation par les alkylbismuthines

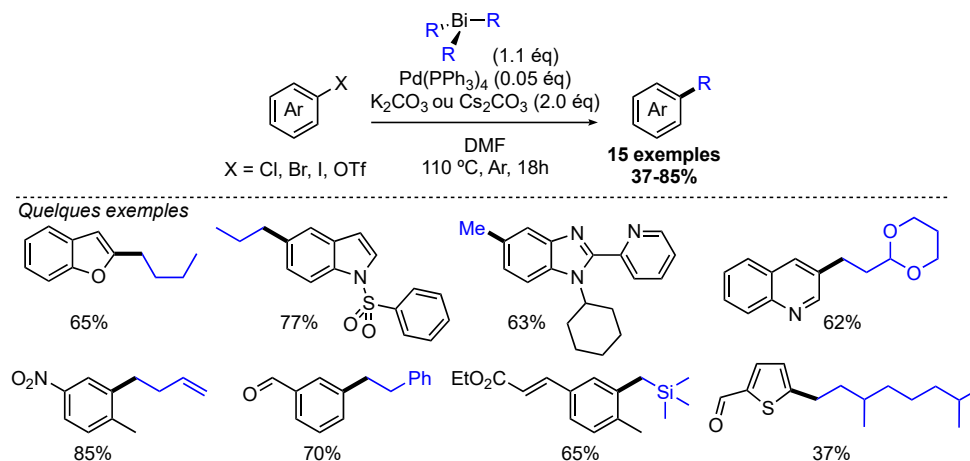
Certainement pour des raisons de stabilité, les réactions de transfert du groupement alkyle des bismuthines ne sont pas très communes. Toutefois, le groupe de Naumann réussit, sans l'aide d'un métal de transition à transférer le groupement trifluorométhyle du bismuth(III) correspondant en position α du chlorure de benzoyle pré-activé avec de la pyridine.^[156] Par la suite, ils ont découvert qu'en utilisant l'acétate de cuivre(II) il était possible de trifluorométhyliser à la fois une énamine et une aniline *N,N*-dialkylée. Sur cette dernière, un mélange de produits ortho et para est cependant obtenu avec un ratio 1,7 pour 1 (Figure 2-6A).^[157] Par la suite, l'équipe de Tanaka a développé des conditions d'alkylation catalysée au palladium en utilisant des organobismuthines hypervalents. Grâce à cette stratégie, les problèmes pyrophoriques relatifs aux alkylbismuths sont jugulés, mais vraisemblablement en raison de la réaction parasite potentielle d'élimination β -hydruure des palladiums, seuls les dérivés méthylés et insaturés du bismuth sont proposés. Leurs conditions permettent ainsi de méthyliser des pseudo-halogénures et halogénures d'aryle avec une quantité catalytique de palladium, avec ou sans base dans du *N*-méthyl-pyrrolidone (NMP) (Figure 2-6B).^[130, 158, 159] En s'inspirant de ces travaux, Gagnon a proposé d'appliquer les conditions avec son tricyclopropylbismuthine dont les résultats montrent une grande versatilité avec une tolérance aux hétérocycles jusque-là non vérifiée.^[160] Par ailleurs, en s'orientant avec les conditions d'arylation au cuivre de Barton–Finet–Chan^[161], il a démontré qu'il était également possible d'utiliser le tricyclopropylbismuth pour *N*-cyclopropyliser facilement des lactames, des imides, des indolidinones, des oxazolidinones, des azoles ou encore des indoles (Figure 2-6C).^[162]

Figure 2-6 A – Trifluorométhylation du chlorure de benzoyle, d'énamines et d'aniline ; B – Méthylation d'halogénure d'aryles ; C – Cyclopropylation d'halogénures d'aryles et N-cyclopropylation de lactames, d'azoles et d'indoles



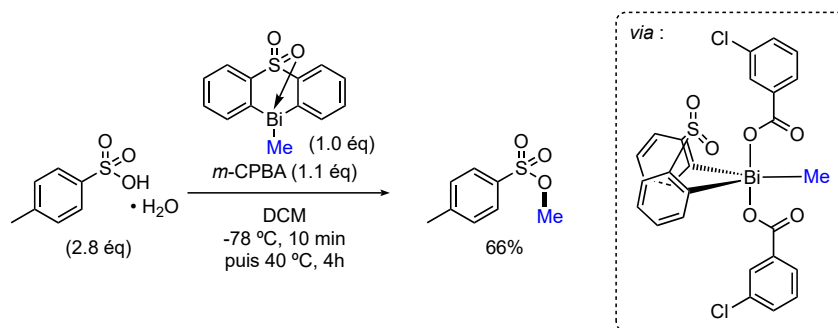
Pendant les études de cyclopropylation catalysée au palladium, l'équipe de Gagnon s'est aperçu avec surprise que leurs conditions permettaient de transférer le groupement éthyle du triéthylbismuthine avec un bon rendement de 81% sans pour autant être impacté par l'élimination β -hydrure tant redoutée. Ainsi, encouragés par ces résultats, ils ont par la suite soumis une variété d'alkylbismuths à différents halogénures d'aryles le tout en maintenant des rendements appréciables (Figure 2-7).^[163]

Figure 2-7 Alkylation d'halogénures d'aryles pallado-catalysées



Ces exemples concluent avec virtuosité les dernières réactions rapportées à ce jour d'alkylations par l'utilisation de trialkylbismuthines. Toutefois, bien que très anecdotique, la méthylation à travers un alkylbismuth pentavalent est possible. L'unique exemple disponible dans la littérature est mentionné par Mukaiyama et Sakurai avec la méthylation de l'acide para-toluènesulfonique à partir du méthylbismacyle hypervalent oxydé au préalable avec de l'acide *m*-chloroperbenzoïque (*m*-CPBA) (Figure 2-8).^[131]

Figure 2-8 Méthylation de l'acide p-toluènesulfonique par le dicarboxylate de méthylbismacyle



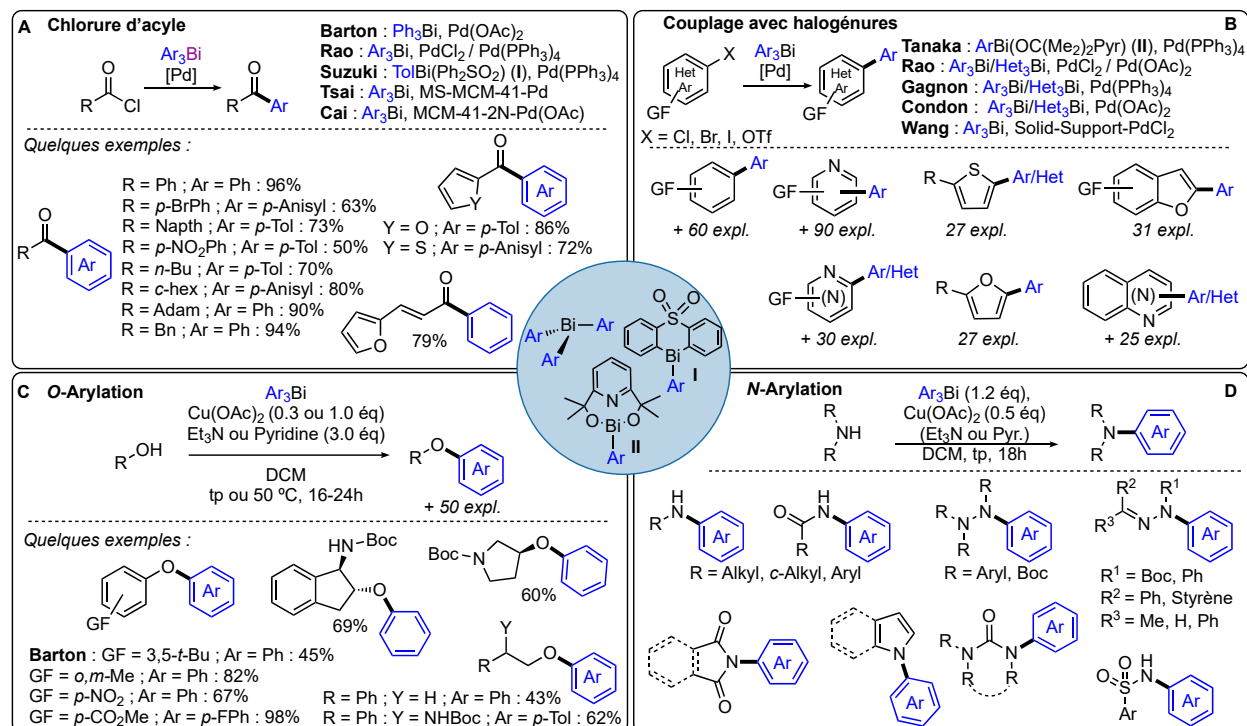
Avant d'embarquer dans les réactions d'arylation, il est nécessaire de mentionner que les méthodes d'arylation à partir d'organobismuthines sont légion dans la littérature et il serait inconvenant de mentionner l'ensemble des stratégies existantes. Ainsi, il sera présenté dans un premier temps une brève sélection des méthodes d'arylation à travers l'usage des arylbismuths trivalents avant de poursuivre dans un deuxième temps celles avec les pentavalents.

2.2.4 Réaction d'arylation par des arylbismuthines trivalentes

Les conditions de transfert d'alkyles vues précédemment peuvent s'appliquer dans le cadre des aryles de manière quasi systématique. En effet, les chlorures d'acyles peuvent se faire aryle à partir d'un arylbismuth trivalent avec une quantité catalytique de palladium. Les prémices de ces travaux ont été effectuées par Barton et ont inspiré un grand nombre de groupes.^[145] Notamment, l'équipe de Rao a non seulement su démontrer la grande robustesse de la méthode, mais également qu'une quantité substoechiométrique de triarylbismuth était suffisante puisque les trois aryles sont transférables.^[164-168] Suzuki est passé par l'usage d'une arylbismuthine hypervalente^[132] alors que les équipes de Tsai^[169] et Cai^[170] ont proposé des catalyseurs sur support solide contenant du palladium facilement recyclable (Figure 2-9A). Le couplage croisé catalysé au palladium avec des halogénures d'aryles reste remarquablement efficace. Le groupe de Tanaka a ouvert la voie en 1999 en couplant des aryltriflates avec le dialcoolate de méthylbismuth hypervalent.^[158] Par la suite, accompagné de Rao,^[171-174] ils ont participé activement au développement de la technique en couplant une variété importante de dérivés halogénés aromatiques tels que des thiophènes,^[175] des furanes^[176], des benzofuranes^[177] ou encore des pyridines.^[178] À ces travaux sont ajoutés les couplages avec des diazines, des benzopyridines et benzodiazines par les équipes de Condon^[179] et de Gagnon.^[180, 181] Pour conclure cette partie, des catalyseurs sur support solide ont également été développés par Wang et peuvent être ajoutés à cette liste déjà assez dense (Figure 2-9B).^[182] Les couplages croisés au cuivre donnent également accès avec grande facilité aux produits de *N*- et *O*-arylation. Les méthodes de cette dernière ont été beaucoup plus étudiées dans le cadre des bismuths pentavalents, mais il reste intéressant de mentionner que la première et unique *O*-arylation à partir d'un triarylbismuth a été observée par Barton en 1987. Pour ce faire, il a mis en réaction du 3,5-di-*t*-butylphénol avec du triphénylbismuth avec une quantité catalytique d'acétate de cuivre(II) et de la triéthylamine dans du dichlorométhane à température ambiante pendant 24 heures et a observé le produit *O*-phénylé avec un rendement de 25%. Ce rendement a pu être légèrement amélioré en ajoutant une quantité stœchiométrique de cuivre.^[183] Ce n'est que plus récemment que cette méthode a été explorée plus en détail par Gagnon et les résultats montrent qu'autant les phénols substitués que les phénols appauvris en électrons sont acceptés.^[91, 184] Dans la même lignée, ces mêmes conditions permettent également d'aryler des alcools primaires et secondaires avec des rendements sensiblement similaires (Figure 2-9C).^[185] En vue du caractère nucléophile accru des amines, les méthodes de *N*-arylations sont possibles et présentent sans trop de surprise, de bien meilleurs rendements. C'est à travers les études mentionnées plus tôt de 1987 que Barton a usé de l'avantage nucléophile des azotes pour effectuer ce type d'arylation.^[183] En utilisant une quantité catalytique de cuivre dans du dichlorométhane à température ambiante, il a réussi à aryle des azotes d'anilines, d'amines primaires et secondaires en terminant par des hydrazines et hydrazones. Hormis des changements mineurs tels que le nombre d'équivalents de cuivre ou l'ajout d'une base, ses conditions semblaient déjà être les conditions optimales puisqu'elles ont été largement utilisées. En effet, en passant par les arylations des pipéridines de

Banfi en 1994,^[186] des amides de Chan en 1996,^[161] des hydrazines de Kikugawa^[187] et de Ragnarsson en 2000,^[188] des imides de Hügel en 2006,^[189] des hydrazones de Mäeorg en 2007^[190] et des indoles et azoles de Gagnon en 2014, ces mêmes conditions ont toujours été préférées (Figure 2-9D).^[191]

Figure 2-9 A – Arylations pallado-catalysées de chlorures d'acyles ; B – Arylations pallado-catalysées d'halogénures d'aryles ; C – O-Arylations cupro-catalysées de phénols et d'alcools ; D – N-Arylations cupro-catalysées d'azotes



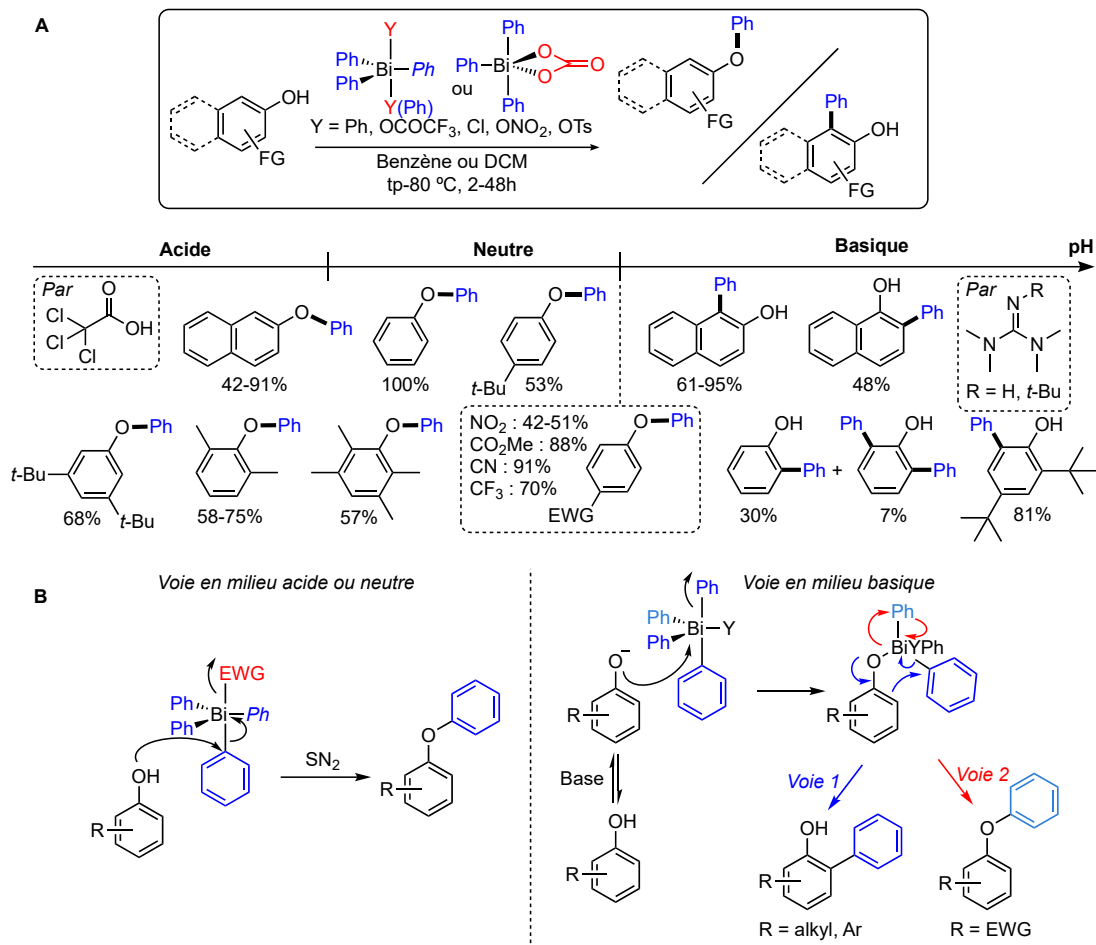
Cette sélection personnelle reflète en somme bien l'étendue du spectre des applications des triarylbismuthines en chimie de synthèse. Bien que déjà très dense, les réactions engageant des bismuthines trivalentes sont relativement moins nombreuses si comparées à celles qui passent par des homologues pentavalents. En effet, ces dernières ont eu le droit à une quinzaine d'années de plus de développement depuis leurs premières applications dans des réactions de transfert.

2.2.5 Réactions d'arylation par des arylbismuthines pentavalentes

Historiquement, la première réaction de transfert d'aryle a été découverte inopinément par Barton en 1980. En effet, en étudiant les capacités oxydantes des bismuths pentavalents, il s'est aperçu qu'en oxydant la quinine avec le carbonate de triphénylbismuth, il obtenait bien la quinone correspondante mais avec un groupement phényle additionnel en position alpha. Dans cette même publication, il a poursuivi ses recherches et réussi avec l'aide d'une base en sus, à aryle des stéroïdes carbonylés et un naphthol en position

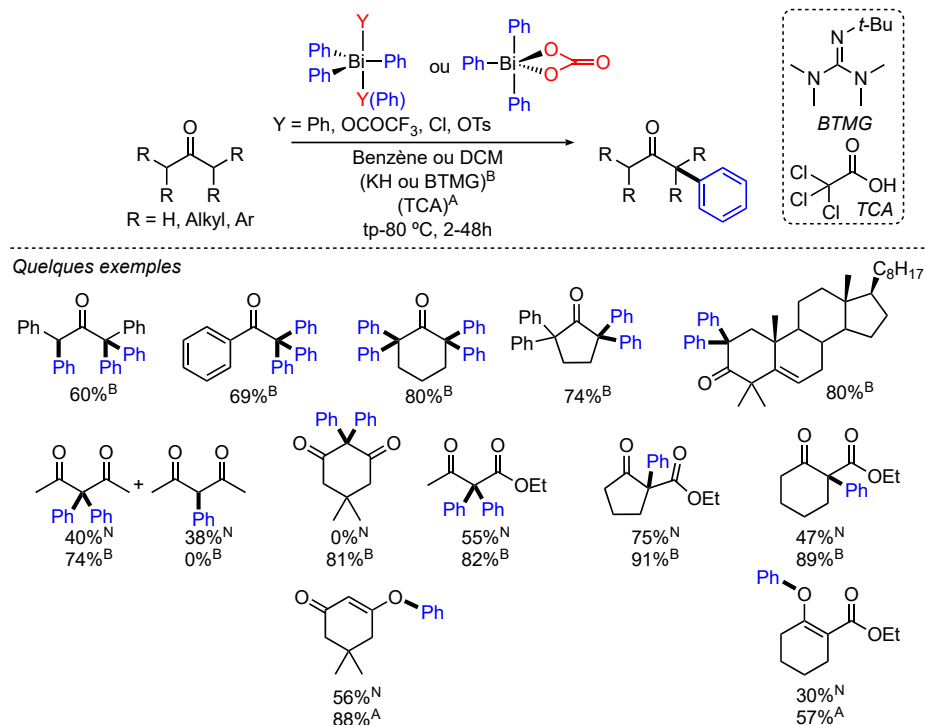
alpha du carbonyle et de l'alcool, respectivement.^[192] Fort de ces découvertes, il a conduit des études plus approfondies sur l'arylation de phénols et des carbonyles en utilisant une diversité de bismuths pentavalents. Ces premières études sur le phénol étaient d'une grande importance puisqu'elles permettent de mettre en lumière une régiosélectivité différente selon la nature du bismuth ainsi que le pH du milieu réactionnel. En effet, un bismuth de type triphénylé (Ph_3BiX_2) ou tétraphénylé pentavalent (Ph_4BiX) orientera plutôt vers une *O*-arylation en milieu acide ou neutre, et plutôt vers une *C^{ortho}*-arylation en milieu basique. D'un autre côté, un bismuth de type pentaphénylé (Ph_5Bi) n'étant pas stable, ne s'utilise uniquement qu'en milieu neutre et donnera systématiquement le produit d'*ortho*-arylation correspondant.^[124, 193, 194] Les phénols substitués avec un groupement électroattracteur en position *para* font office de singularités à cette règle puisque leurs arylations donneront invariablement le produit éthéré même à pH basique avec n'importe quel type de bismuth (Figure 2-10A). Cette particularité n'est pas très bien comprise à ce jour, mais l'isolation de l'intermédiaire phénolate de bismuth permet de confirmer que le mécanisme d'action ne passe pas par un mécanisme de type $\text{S}_{\text{N}}2$ comme proposé en milieu acide mais qu'il emprunte une voie alternative conduisant au produit de *O*-arylation (voir voie 2, Figure 2-10B).^[195]

Figure 2-10 A – Influence de la nature du bismuth pentavalent sur l'arylation régiosélective de dérivés phénols selon le pH ; B – Mécanismes proposés expliquant la régiosélectivité



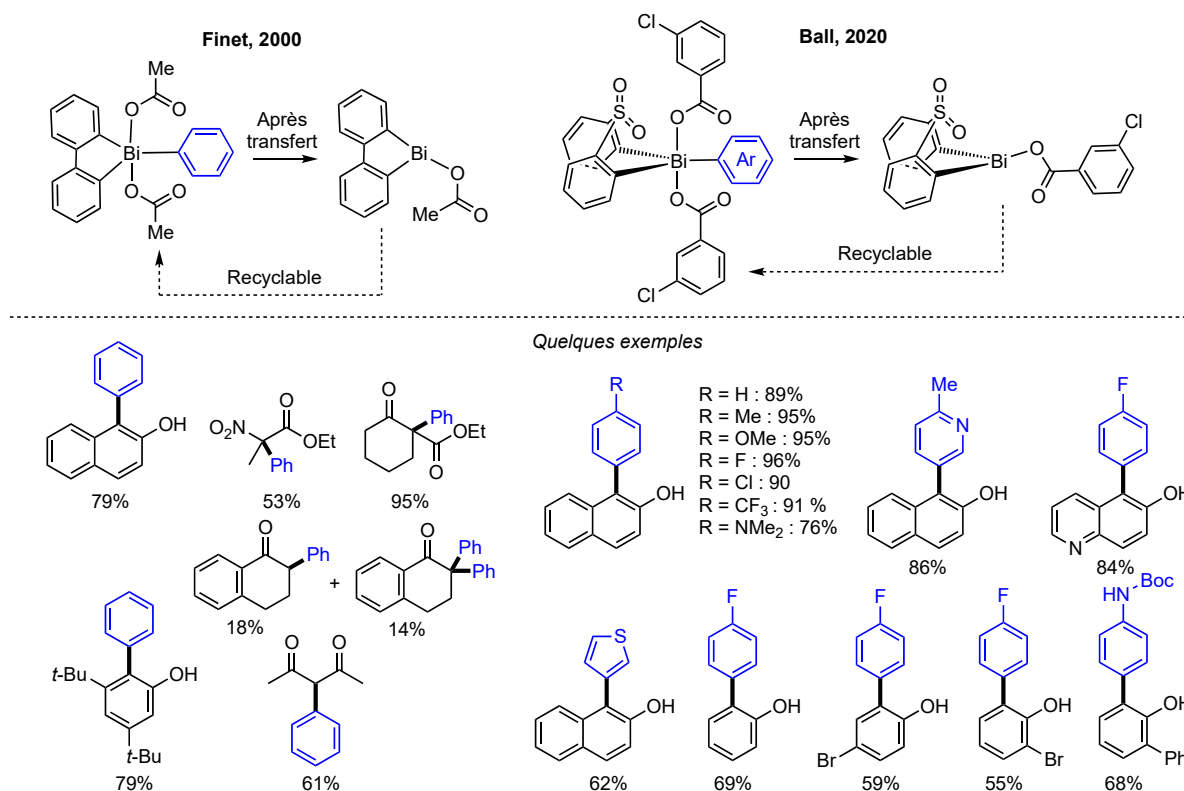
Dans le cadre des carbonyles, les résultats obtenus avec le phénol permettent d'anticiper avec plus ou moins de justesse la régiosélectivité de l'arylation. En effet dans un environnement basique, l'arylation des carbonyles s'effectue plus généralement en position alpha mais cette fois-ci, un environnement neutre semble suivre le même schéma et orienter également en alpha, toutefois avec des rendements nettement inférieurs. Des conditions acides ont été étudiées uniquement dans le cadre de diones et montrent la formation de quelques produits éthers (Figure 2-11).^[196]

Figure 2-11 α -Arylation de carbonyles par des bismuths pentavalents en milieu neutre et basique et *O*-arylation en milieu acide. Milieu : ^A = acide, ^N = neutre, ^B = basique



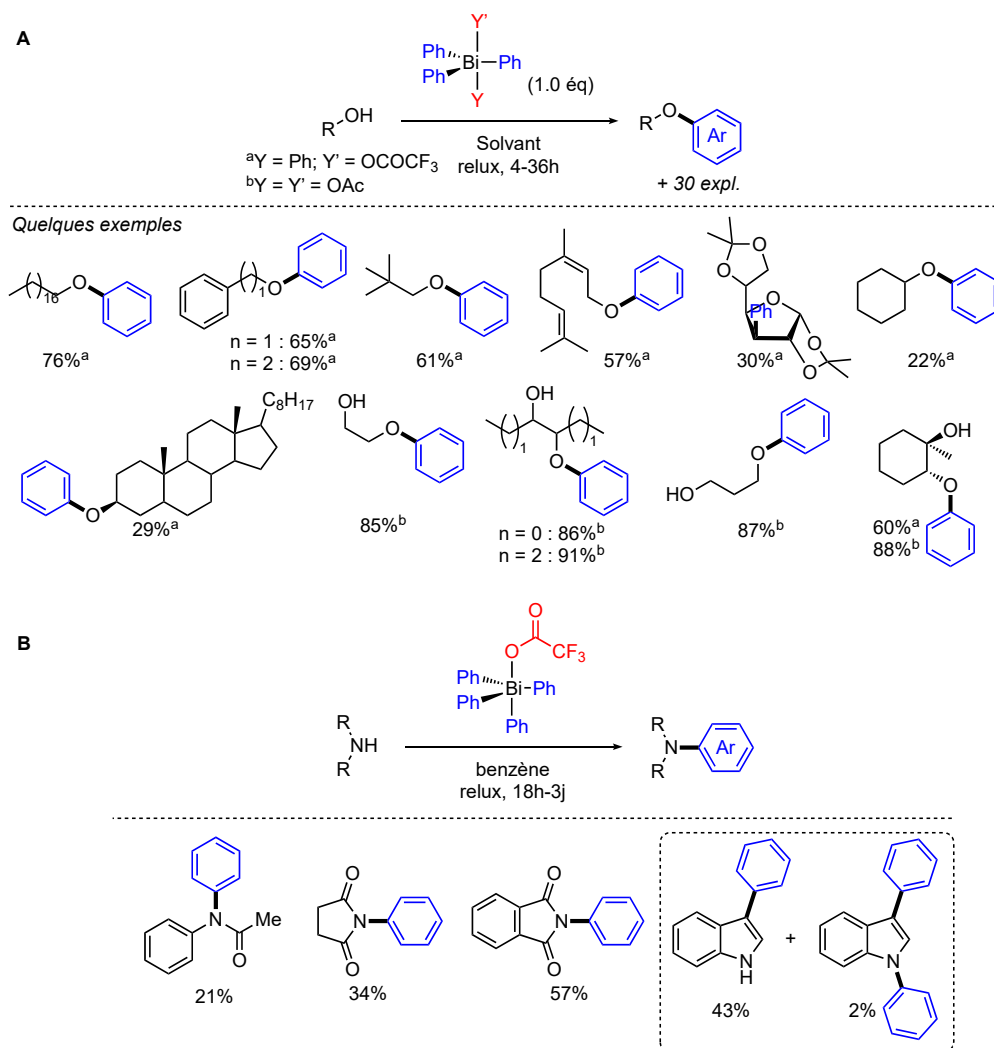
Le pouvoir d'insérer un groupement aryle régiosélectivement uniquement avec un bismuth pentavalent et surtout sans l'intervention d'un autre métal est un avantage incontestable de cette méthode. Cependant, la limitation de transférer uniquement un seul des groupements aromatiques parmi les trois (quatre ou cinq), apporte une certaine limitation à cette méthode en termes d'économie d'atomes. Cette faiblesse a pu être surmontée grâce aux travaux de Finet en 2000 qui, en utilisant des bismacycles pentavalents, a réalisé l'incroyable prouesse de maintenir la réactivité et la régiosélectivité des conditions déjà établies et d'en plus de pouvoir récupérer le sous-produit de bismacycle qui peut être réutilisé après de simples étapes de réactivation.^[197] Plus récemment, ces travaux ont été repris et remarquablement approfondis dans le cadre des phénols par Ball en 2020 (Figure 2-12).^[151]

Figure 2-12 C-Arylations régiosélectives par l'usage de bismacycles pentavalents



À l'image des bismuths trivalents, il est possible d'utiliser les bismuthines pentavalentes afin d'effectuer des réactions de *N*-[198-201] et *O*-arylation[202-205] cupro-catalysées. Mais la véritable force des bismuths pentavalents réside dans le fait de pouvoir effectuer ce genre de transfert sans l'intermédiaire d'un quelconque métal de transition. Les réactions avec le phénol en démontrent une partie, mais il est possible de trouver dans la littérature des arylations d'alcools plus simples et de diols, aisément en présence de bismuths pentavalents dans du solvant au reflux.[206-208] Les résultats obtenus montrent une efficacité de transfert raisonnable sur les alcools primaires, moyenne sur les secondaires et totalement nulle sur les tertiaires (Figure 2-13A). En revanche, dans le cadre des *N*-arylations, les résultats connus à ce jour sont beaucoup moins favorables à des conditions sans cuivre, mais peuvent toutefois être mentionnés. Dans ses travaux sur la *O*-arylation d'alcools, Barton a essayé ses conditions sur des amides, des imides et un indole.[196, 207] Les produits de *N*-arylations ont bel et bien été obtenus avec des rendements plutôt moyens, à l'exception de l'indole, où le produit de *C*³-arylation a été observé majoritairement (Figure 2-13B). Curieusement, la littérature récente ne fait mention d'aucune autre méthode d'arylation à la position *C*³ d'un indole à travers l'unique usage d'un organobismuth. Nonobstant, grâce à l'aide d'une catalyse au cuivre et un blocage de la position azotée, il est possible d'obtenir des produits de *C*³-arylation avec de bien meilleurs résultats.[209]

Figure 2-13 A – *O*-Arylation d'alcools et de diols à partir de bismuths pentavalents ; B – *N*-Arylation d'amides, d'imides et d'indoles à partir du trifluoroacétate tétraphénylbismuth.

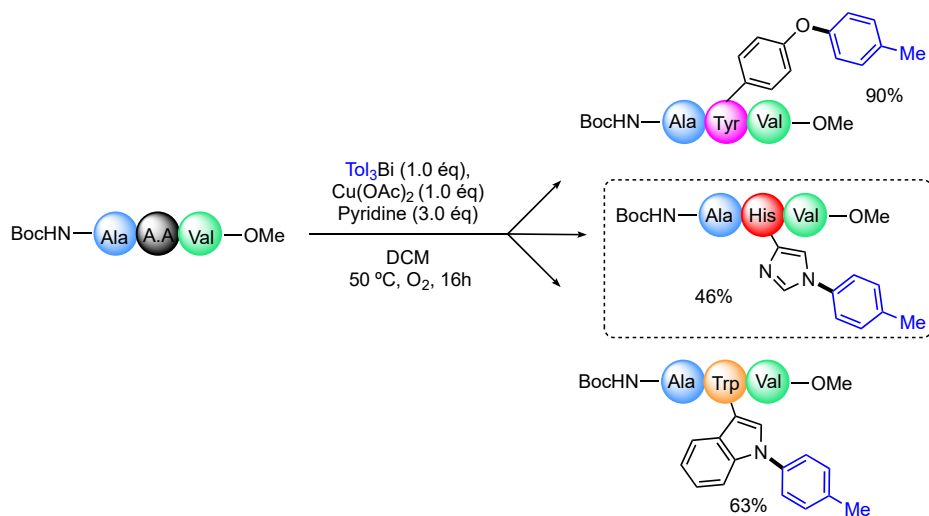


À travers ce chapitre, il a été possible d'appréhender les différentes facettes du bismuth à la fois en tant que métal mais surtout en tant que réactifs en chimie organique. Toutefois, à cette étape de lecture, il se peut qu'il soit déroutant de ne percevoir aucun lien concret entre l'histidine et le bismuth, mais comme mentionné à la fin du chapitre précédent, notre groupe de recherche a utilisé des bismuths trivalents afin de modifier sélectivement la chaîne latérale de différents acides aminés dans de petits peptides. Ces travaux marquent les premières phases de la genèse de cette thèse.

LE BISMUTH AU SERVICE DES MODIFICATIONS DE PEPTIDES

Les arylations de la chaîne latérale d'acides aminés par des bismuths trivalents ont été rapportées pour la première fois en 2020 par notre groupe de recherche. Les conditions développées permettent d'aryler efficacement l'oxygène du phénol de la tyrosine^[91] et l'azote de l'indole du tryptophane^[92] contenus dans de petits peptides. Au détour du développement de ces méthodes, les conditions d'arylation s'avéraient être également efficaces sur l'imidazole de l'histidine, une découverte fortuite sans précédent et aux applications potentiellement intéressantes (voir Figure 3-1). En effet, les techniques d'arylation vues dans le premier chapitre, bien que très fournies, ne présentent aucun cas d'arylation d'histidine au sein d'un peptide. Ce genre de modification post-synthétique est non seulement très recherchée en chimie médicinale où une modification structurale de peptides peut avoir des effets thérapeutiques intéressants, mais la modification sélective de l'histidine est également un outil extrêmement convoité en biochimie avec les différentes étiquettes bio-marquantes aux vertus diverses. Mais avant toutes ambitions d'applications concrètes, il est nécessaire de passer par la recherche fondamentale dont cette thèse va essayer d'y apporter une petite pierre et ainsi enrichir cet immense édifice.

Figure 3-1. Résultat d'arylation d'un tripeptide contenant l'histidine durant les études précédentes



CHAPITRE 3

COPPER-PROMOTED N-ARYLATION OF THE IMIDAZOLE SIDE CHAIN OF PROTECTED HISTIDINE BY USING TRIARYLBISMUTH REAGENTS

3.1 Introduction

Ce chapitre présentera les résultats issus du développement d'une nouvelle méthode d'arylation de l'imidazole de l'histidine sous forme protégée ou peptidique utilisant des triarylbismuthines. En passant par des étapes d'optimisation et des études établissant les limites de la méthode, il a été possible de démontrer que non seulement l'arylation de l'histidine était robuste en tolérant une grande variété de dérivés aromatiques, mais également que la méthode présente une grande spécificité à l'histidine comparée à ses homologues nucléophiles.

3.2 Article issu de ces travaux

Chan, H.-C., Bueno, B., Le Roch, A. and Gagnon, A. (2021). Copper-Promoted N-Arylation of the Imidazole Side Chain of Protected Histidine by Using Triarylbismuth Reagents. *Chemistry – A European Journal*, 27(53), 13330-13336. <https://doi.org/10.1002/chem.202102186>

Copper-Promoted N-Arylation of the Imidazole Side Chain of Protected Histidine by Using Triarylbismuth Reagents

Hwai-Chien Chan,^[a] Bianca Bueno,^[a] Adrien Le Roch,^[a] and Alexandre Gagnon^{*[a]}

[a] Université du Québec à Montréal
Département de chimie
C.P. 8888, Succursale Centre-Ville
Montréal, Québec, Canada, H3C 3P8
E-mail: gagnon.alexandre@uqam.ca

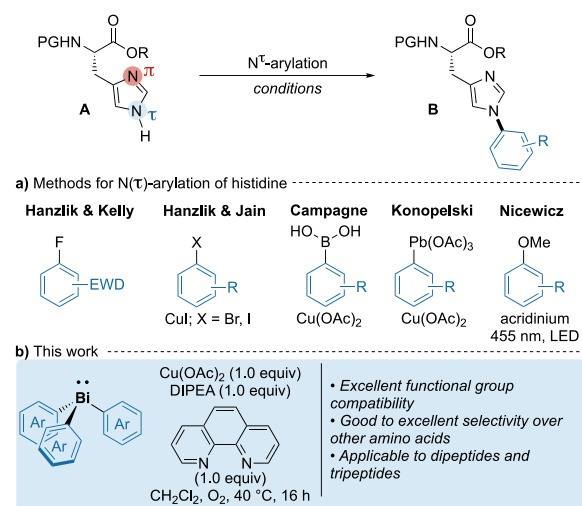
This paper is dedicated to our colleague and dear friend Prof. Victor Snieckus for his exceptional lifetime contribution to organic Chemistry

Abstract: The N-arylation of the side chain of histidine by using triarylbismuthines is reported. The reaction is promoted by copper(II) acetate in dichloromethane at 40 °C under oxygen in the presence of diisopropylethylamine and 1,10-phenanthroline and allows the transfer of aryl groups with substituents at any position of the aromatic ring. The reaction shows excellent functional group tolerance and is applicable to dipeptides where the histidine is located at the N terminus. A histidine-guided backbone N–H arylation was observed in dipeptides where the histidine occupies the C terminus.

Bismuth is the heaviest stable non-radioactive element of the periodic table.^[1] Contrary to its neighboring elements, this heavy metal shows relatively low toxicity, as exemplified by medicinal ingredients that contain inorganic bismuth salts.^[2] Due to its borderline behavior as a metal and a classic pnictogen, bismuth compounds have served both as Lewis acids and bases.^[3]

Triarylbismuthines are versatile organometallic reagents that have been used in a wide range of transition metalcatalyzed reactions.^[4] Our group reported a portfolio of coppercatalyzed methods for the arylation of NH-containing

heterocycles,^[5] phenols^[6] and amino alcohols^[7] that involve triarylbismuthines. Recently, we transposed some of these protocols to the arylation of the side chain of tryptophan^[8] and tyrosine.^[9] In the course of our studies, we found that the side chain of histidine was considerably reactive under those conditions, resulting in the N-arylation of the imidazole moiety.^[9] This observation was in agreement with reports from Finet and more recently Jadhav showing that imidazole can be smoothly arylated by triarylbismuthines in the presence of copper salts.^[10] Motivated by these results, we decided to further investigate this reaction. Various methods have been reported for the N-arylation of the side chain of histidine, usually in their N,O-diprotected form. While the arylation can in principle occur at the proximal N(π) or distal N(τ) atom of the imidazole ring in **A**, the vast majority of these methods result in the arylation of the sterically more accessible N(τ) position to deliver product **B** (Scheme 1).^[11]



Scheme 1. Methods for the N(τ)-arylation of the side chain of histidine.

The first procedures leading to the arylation of the side chain of histidine relied on an S_NAr reaction with aryl fluorides, as shown first by Hanzlik^[12] and then by Kelly.^[13] However, like most S_NAr reactions, these methods require very high temperatures and/or the presence of electron-withdrawing groups on the aryl substrate. Hanzlik and Jain developed two copper(I)-catalyzed protocols to arylate the side chain of histidine using aryl bromides and iodides.^[14] While their methods show excellent scope, high temperatures are required. Using a Chan-Evans-Lam approach, Campagne reported an efficient procedure to arylate the imidazole side chain of histidine.^[15] Although the reaction operates under air at a moderate temperature, limited functional groups were introduced on the aryl moiety. Histidine has also been arylated using aryl lead triacetates, as shown by Konopelski, but the toxicity associated with lead reagents considerably limits the use of the method.^[16] Recently, Nicewicz performed the arylation of histidine according to a

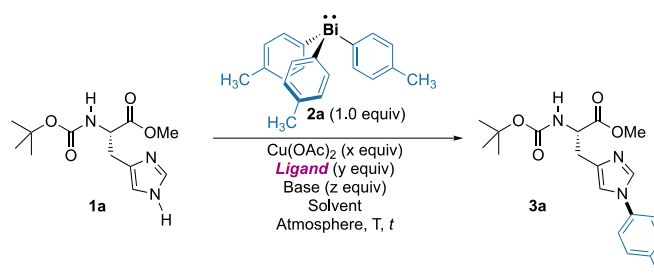
photoredox protocol that involves an aryl methyl ether under irradiation at 455 nm in the presence of an acridinium photosensitizer.^[17] However, a mixture of arylation products at N(π) and N(τ) connected to the ortho and para position of anisole was obtained.

There is an interest for general methods that lead to the modification of amino acids, not only in medicinal chemistry where peptides are used as lead compounds, but also in chemical biology where arylated amino acids have been found to play a role in biochemical processes.^[18] Even though procedures exist to arylate the side chain of histidine, additional methods that operate under smooth conditions while offering excellent functional group compatibility are still highly desirable. In this context, we would like to report herein our investigation into the copper(II)-promoted arylation of histidine using triarylbismuth reagents.

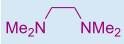
We commenced our work by optimizing the conditions for the arylation of the imidazole side chain of Boc-protected histidine methyl ester **1a** with tri(4-tolyl)bismuth **2a** (Table 1). Using the conditions that we reported for the arylation of the side chain of tyrosine,^[9] we obtained the desired product in 81% yield (entry 1). After testing a few organic bases, we identified N,N-diisopropylethylamine (DIPEA) as a suitable replacement for the undesirable pyridine and found that the reaction could be performed at 40 °C with only 1.0 equivalent of DIPEA (entries 2 to 4). With the objective of improving the yield, we added ligands **L1–L5** and found that 1,10-phenanthroline **L4** and bathophenanthroline **L5** provided the best improvement in the yield of the reaction (entries 5 to 9). Using **L4** as the ligand, we verified the necessity of performing the reaction under oxygen and found a considerable drop in the yield when replacing it by air or argon (entry 10 and 11). Conducting the reaction in other solvents led to the identification of THF and acetonitrile as suitable replacements for dichloromethane (entries 12 to 14). A complete loss of reactivity was observed upon performing the reaction at room temperature, demonstrating the importance of heat (entry 15). Minimal impact on the yield was observed upon reducing the catalyst loading to 0.7 equivalent or lowering the reaction time to 4 h (entry 16 and 17).

We then investigated the scope of the reaction using various triarylbismuth reagents (Scheme 2). To maximize the yields, we used conditions from entry 8 in Table 1. Our results show that the reaction tolerates substituents at any position of the aromatic ring (**3a–d**), is unaffected by electronic effects (**3e–i**), is compatible with a broad range of functional groups (**3j–q**), allows the transfer of a naphthyl unit (**3r**) and functions with Cbz and Fmoc protecting groups (**3s, t**). We performed some entries in acetonitrile (yields indicated in brackets) and observed that the transfer proceeded efficiently with the electronically poor 4-fluorophenyl group (**3f**) but not with the electronically rich 4-methoxyphenyl (**3e**) or sterically encumbered 2-tolyl (**3c**). Also, while the efficiency was preserved with Cbz-protected histidine (**3s**), a dramatic drop was observed with the Fmoc counterpart (**3t**).

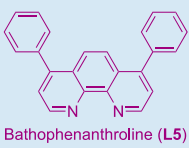
Table 1. Optimization of the reaction conditions for the copper-promoted N-arylation of the imidazole side chain of N-Boc histidine methyl ester **1a** using tri(4-tolyl)bismuth **2a**.^[a]

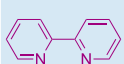


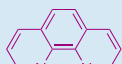
Ligands


TMEDA (L1)


DMEDA (L2)

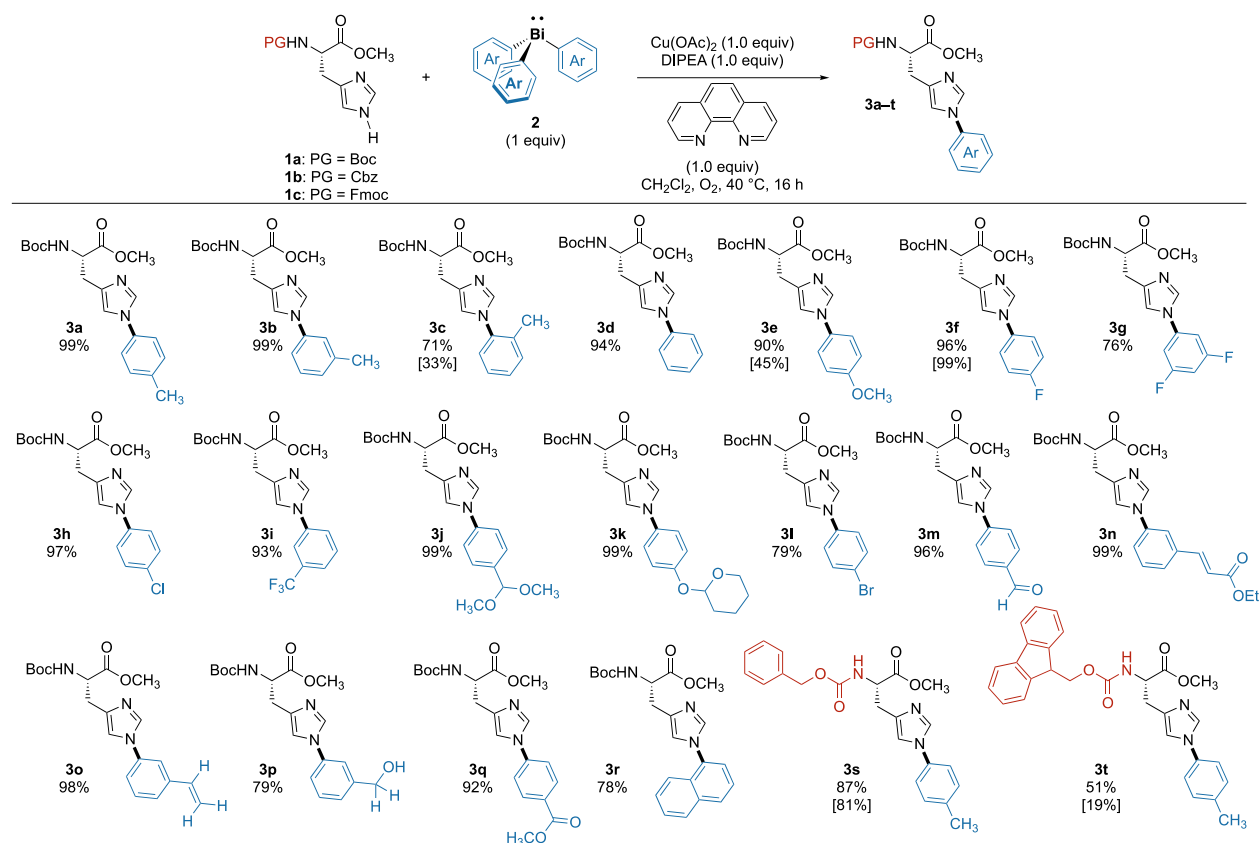

Bathophenanthroline (L5)


Bipy (L3)


1,10-Phenanthroline (L4)

Entry	Cu(OAc) ₂ (x equiv)	Ligand (y equiv)	Base (z equiv)	Solvent	Atmosphere	T (°C)	t (h)	Yield 3a (%) ^[b]
1	1.0 equiv	–	Pyridine (3.0 equiv)	CH ₂ Cl ₂	O ₂	50	16	81
2	1.0 equiv	–	Et ₃ N (3.0 equiv)	CH ₂ Cl ₂	O ₂	50	16	72
3	1.0 equiv	–	DIPEA (3.0 equiv)	CH ₂ Cl ₂	O ₂	50	16	82
4	1.0 equiv	–	DIPEA (1.0 equiv)	CH ₂ Cl ₂	O ₂	40	16	84
5	1.0 equiv	L1 (1.0 equiv)	DIPEA (1.0 equiv)	CH ₂ Cl ₂	O ₂	40	16	37
6	1.0 equiv	L2 (1.0 equiv)	DIPEA (1.0 equiv)	CH ₂ Cl ₂	O ₂	40	16	72
7	1.0 equiv	L3 (1.0 equiv)	DIPEA (1.0 equiv)	CH ₂ Cl ₂	O ₂	40	16	85
8	1.0 equiv	L4 (1.0 equiv)	DIPEA (1.0 equiv)	CH ₂ Cl ₂	O ₂	40	16	99
9	1.0 equiv	L5 (1.0 equiv)	DIPEA (1.0 equiv)	CH ₂ Cl ₂	O ₂	40	16	99
10	1.0 equiv	L4 (1.0 equiv)	DIPEA (1.0 equiv)	CH ₂ Cl ₂	Air	40	16	88
11	1.0 equiv	L4 (1.0 equiv)	DIPEA (1.0 equiv)	CH ₂ Cl ₂	Argon	40	16	42
12	1.0 equiv	L4 (1.0 equiv)	DIPEA (1.0 equiv)	DMF	O ₂	40	16	87
13	1.0 equiv	L4 (1.0 equiv)	DIPEA (1.0 equiv)	THF	O ₂	40	16	99
14	1.0 equiv	L4 (1.0 equiv)	DIPEA (1.0 equiv)	MeCN	O ₂	40	16	98
15	1.0 equiv	L4 (1.0 equiv)	DIPEA (1.0 equiv)	MeCN	O ₂	r.t.	16	0
16	0.7 equiv	L4 (0.7 equiv)	DIPEA (1.0 equiv)	MeCN	O ₂	40	16	93
17	1.0 equiv	L4 (1.0 equiv)	DIPEA (1.0 equiv)	MeCN	O ₂	40	4	93

^[a] Reaction performed in a sealed tube placed in an oil bath at the indicated temperature and under the specified atmosphere. ^[b] Yield of isolated pure product. DMF = N,N-dimethylformamide. THF = tetrahydrofuran.



Scheme 2. Scope of the N-arylation of the side chain of N-protected histidine methyl esters **1a–c**. Yields are for isolated pure products; yields in brackets are for reactions performed in MeCN instead of CH₂Cl₂.

Before transposing our protocol to histidine-containing peptides, we sought to evaluate the relative reactivity of amino acids possessing a nucleophilic side chain under our newly optimized conditions (Figure 1).

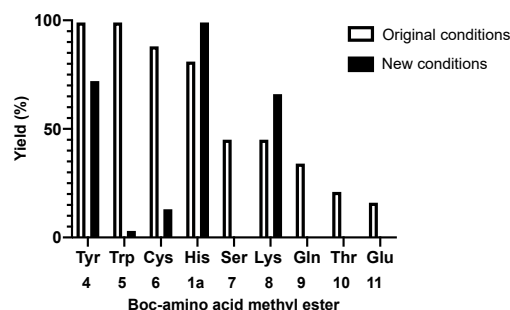
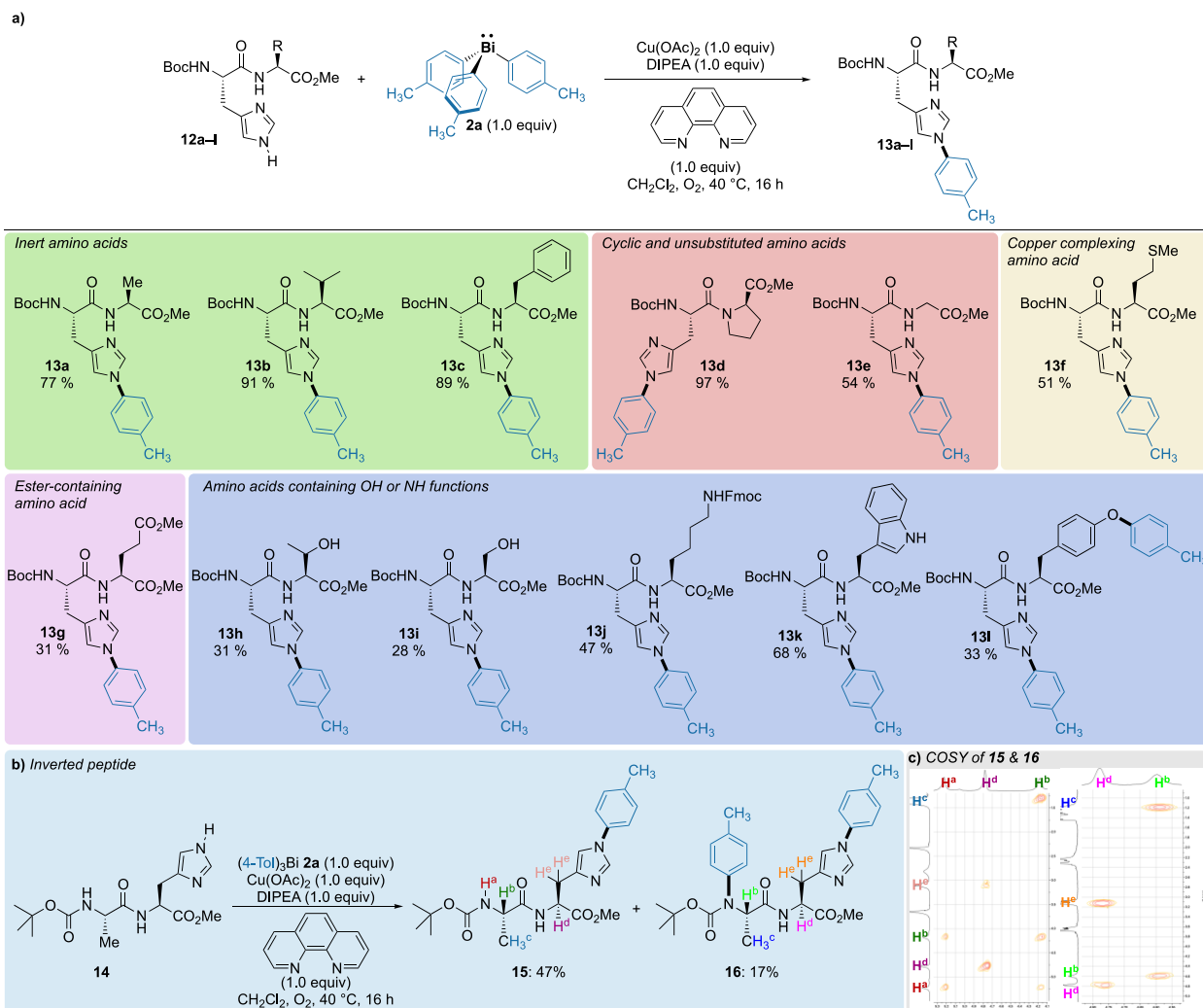


Figure 1. Arylation of amino acids possessing a nucleophilic side chain. Original conditions: amino acid N-Boc methyl ester (1.0 equiv), (p-Tol)₃Bi (1.0 equiv), Cu(OAc)₂ (1.0 equiv), pyridine (3.0 equiv), CH₂Cl₂, O₂, 50 °C, 16 h.^[9] New conditions: amino acid N-Boc methyl ester (1.0 equiv), (p-Tol)₃Bi (1.0 equiv), Cu(OAc)₂ (1.0 equiv), DIPEA (1.0 equiv), 1,10-phen (1.0 equiv), CH₂Cl₂, O₂, 40 °C, 16 h. Numbers refer to the N-Boc methyl ester amino acids substrates.

Under our original conditions, N-Boc tyrosine methyl ester **4** and N-Boc tryptophan methyl ester **5** were almost equally reactive, followed by N-Boc cysteine methyl ester **6** and N-Boc histidine methyl ester **1a** (Figure 1, white bars).^[9] This

lack of differentiation in reactivity was expected to lead to issues of chemoselectivity during the arylation of histidine-containing peptides. Under our new conditions (Figure 1, black bars), a dramatic change in the order of reactivity between the amino acids was observed: N-Boc histidine methyl ester **1a** was now the most reactive amino acid, followed by N-Boc tyrosine methyl ester **4** and N-Boc lysine methyl ester **8**. N-Boc tryptophan methyl ester **5** and N-Boc cysteine methyl ester **6** were found to be poorly reactive under these new conditions: indeed, N-Boc tryptophan methyl ester **5** gave the arylated product in only 12% yield with 88% recovered starting material while N-Boc cysteine methyl ester **6** gave the desired product in 19% along with 59% recovered starting material. N-Boc serine methyl ester **7**, N-Boc glutamine methyl ester **9**, N-Boc threonine methyl ester **10** and N-Boc glutamic acid mono methyl ester **11** were all recovered intact. While it is difficult to explain this drastic change in ranking of amino acids between the original and the new conditions, it must be hypothesized that the lower reaction temperature combined with the presence of the 1,10-phenanthroline ligand are necessarily responsible for this phenomenon. Further mechanistic and theoretical studies will be needed to better understand the difference in reactivity under both sets of conditions.



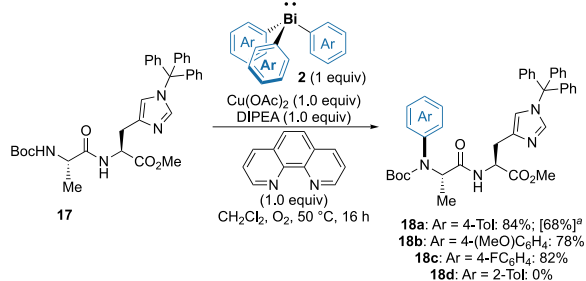
Scheme 3. a) N-Arylation of histidine-containing dipeptides **12a–l**. b) Double histidine and backbone N-arylation of “inverted” Boc-Ala-His-OMe. Yields of isolated pure products. c) COSY spectra of product **15** and **16** (full spectra in Supporting Information).

Having established the order of reactivity of protected amino acids under our arylating conditions, we explored the chemoselectivity in the context of histidine-containing dipeptides where histidine occupies the N-terminal position (Scheme 3a). Thus, dipeptides **12a–l** were prepared by conventional methods (see the Supporting Information for details) and were reacted with tri(4-tolyl)bismuth **2a** under our optimized conditions. In the event, good to excellent yields were obtained with dipeptides where histidine is connected to an inert or cyclic amino acid (**13a–d**). Lower yields were obtained when the histidine is connected to an unsubstituted glycine residue (**13e**), a copper-complexing methionine (**13f**) or an ester-containing amino acid (**13g**). Using bathophenanthroline instead of phenanthroline resulted in similar yields for **13a** (78%) and **13f** (54%), motivating us to pursue with the simpler phenanthroline ligand. The arylation of Boc-His-Thr-OMe and Boc-His-Ser-OMe gave the products of histidine arylation **13h** and **13i** in low yields, even though N-Boc threonine and serine methyl esters were found to be unreactive under these conditions in our amino acids study (Figure 1c, f). The results suggest that the arylation of amino acids in peptides greatly depends on

the immediate environment. However, as anticipated from our amino acid studies, Boc-His-Trp-OMe provided exclusively product **13k** from arylation on the histidine residue while Boc-His-Tyr-OMe led to the diarylated product **13l**.

To verify if the position of the histidine in the dipeptide has an impact on the outcome of the reaction, Boc-Ala-His-OMe **14** was submitted to our arylation reaction (Scheme 3b). Firstly, we found that the product of imidazole N-arylation **15** was formed in lower yield than in the complementary dipeptide Boc-His-Ala-OMe **13a**, suggesting that the position of histidine in the dipeptide affects the yield of the reaction. Secondly, product **16** where two aryl groups are transferred was isolated in 17% yield. 2D NMR analysis showed that the second aryl group had been introduced on the N–H backbone of alanine, thus the amino acid preceding histidine (so-called n–1 residue; Scheme 3c). This type of N–H backbone arylation has previously been reported by Ball in the copper- and nickel-catalyzed arylation of peptides using boronic acids.^[19] We sought to gain some understanding about this copper-catalyzed histidinedirected backbone N–H arylation involving triarylbi-bismuth reagents.

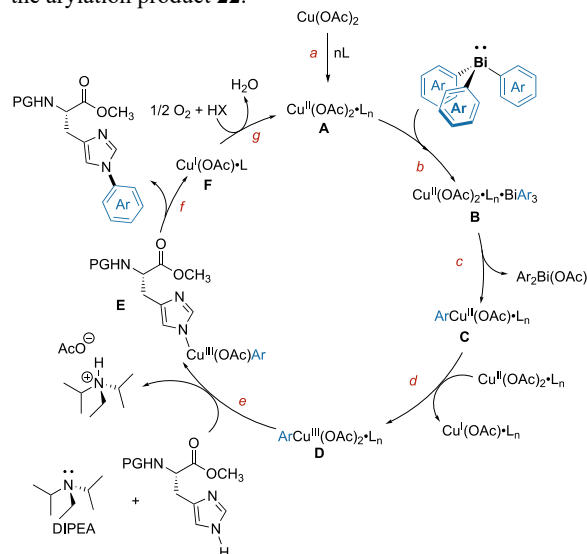
To prevent the arylation of the histidine, we prepared dipeptide **17** where the imidazole ring is protected with a trityl group (Scheme 4). When submitted to our arylation conditions, peptide **17** provided the corresponding product of N–H arylation **18a** in a modest 68% yield. This yield was increased to 84% simply by raising the temperature to 50 °C. Rapid investigation of the scope of this transformation indicated that aryl groups with electron donating (**18b**) and withdrawing (**18c**) substituents could be efficiently transferred using this protocol. However, we found that the presence of a substituent in ortho position was not accepted, possibly due to exacerbated steric congestion generated by the tert-butyloxycarbonyl protecting group (**18d**).



Scheme 4. Histidine-directed N–H backbone arylation of Boc-Ala-His(Nt-Trt)-OMe **17**. Yield of isolated pure products. ^[a] Performed at 40 °C instead of 50 °C.

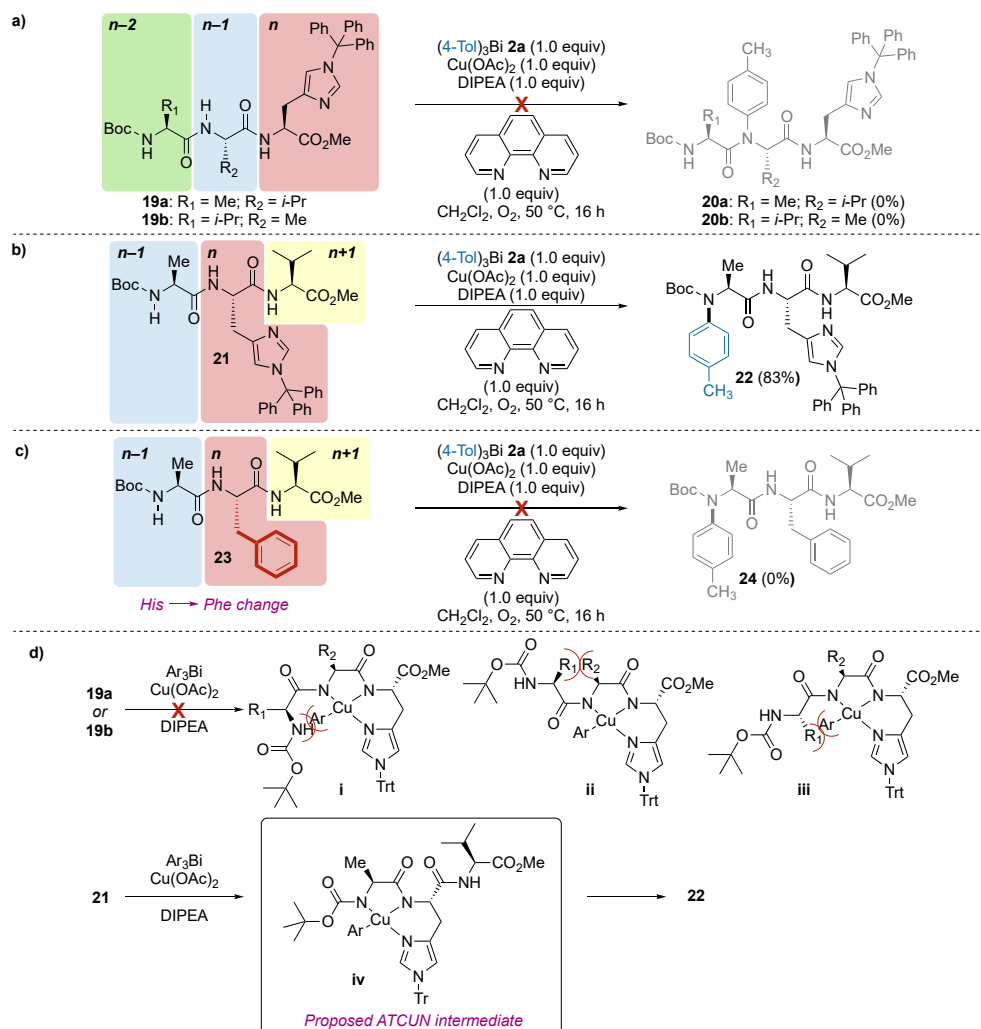
To determine if the reaction is guided by the histidine residue or simply proceeds on the terminal NHBoc moiety, we submitted Boc-Ala-Val-His(Trt)-OMe **19a** to our conditions where a histidine-directed process should lead to the arylation of the central n-1 valine residue (Scheme 5a). In the event, no reaction was observed and the starting material was recovered intact. To make sure that the reaction was not inhibited by the sterically encumbered isopropyl residue at R₂, we submitted the “reverse” tripeptide Boc-Val-Ala-His(Trt)-OMe **19b** to our conditions but again, no reaction was observed. The histidine was then moved to the central position, as shown in Boc-Ala-His(Trt)-Val-OMe **21**, and in this case, the product of arylation at the n-1 position **22** was isolated in 83% yield (Scheme 5b). Finally, to confirm our hypothesis that the imidazole ring acts as a directing group, we prepared tripeptide **23** where the histidine is replaced by a phenyl alanine (Scheme 5c). As anticipated, product of arylation **24** could not be detected and the starting peptide **23** was recovered in 83% yield. Altogether, these results suggest that: 1) the arylation is guided by the histidine; 2) the arylation proceeds at the n-1 position, that is, one residue before the histidine; 3) the arylation occurs only if the n-1 position is also the NHBoc terminus. These results can be best explained by an ATCUN-like motif (amino terminal Cu and Ni) where the copper center is chelated by the imidazole moiety and the peptide backbone (Scheme 5d).^[20] In the case of **19a** or **19b**, formation of the putative copper(III) species would lead to a severe steric clash between the NHBoc and the aryl group (complex i) thus preventing the arylation process. Moreover, the complex would not be able to avoid this steric hindrance

since rotation around the C–N or C–C bond would lead to conformers with steric repulsion between R₁ and R₂ (species ii) or R₁ and the Ar group (species iii). On the contrary, in the case of **21**, formation of an ATCUN-like intermediate iv where the copper center is simultaneously chelated by the imidazole, the internal amide, the terminal NBoc and the aryl group would be less hindered and thus allowed, leading to the arylation product **22**.



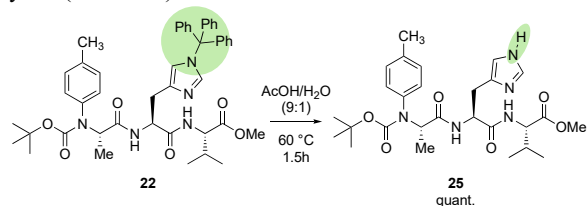
Scheme 6. Proposed mechanism for the N-arylation of the imidazole side chain of histidine. L=1,10-phenanthroline.

The proposed mechanism for the arylation of the imidazole side chain of histidine begins by the complexation of cupric acetate by phenanthroline to give species **A** (Scheme 6, step a). Complexation of the triarylbiarylborane to this species would then generate species **B** (step b) on which transmetalation of the aryl group from Bi to Cu^{II} would occur (step c) to give species **C** concomitantly with diarylbismuth acetate. Disproportion between the copper(II) complex **C** and copper(II) diacetate would provide the copper(III) intermediate **D** along with copper(I) acetate (step d). Complexation of the imidazole ring of histidine to copper and deprotonation by DIPEA would then lead to the key intermediate **E** where a copper(III) center is simultaneously ligated to the imidazole ring, the aryl group and an acetate (step e). Reductive elimination would provide the desired N-arylated product along with cuprous acetate (species **F**, step f). Oxidation of this copper(I) species by oxygen would regenerate the initial copper(II) catalyst. This mechanism is based on the mechanism proposed for the Chan-Evans-Lam reaction and involves a 2:1 ratio of Cu(OAc)₂:Ar₃Bi.^[21] Based on our previous results, we exclude a mechanism where copper diacetate oxidizes the triarylbiarylborane species to its pentavalent triarylbiarylborane diacetate counterpart.^[22] The mechanism for the arylation of the NH backbone of peptides would presumably be similar except that it would proceed through the ATCUN complex iv presented in Scheme 5.



Scheme 5. a) Attempted backbone NH arylation of tripeptides where histidine occupies the C-terminal position. b) Successful backbone NH-arylation of tripeptides where histidine occupies the central position. c) Attempted backbone NH-arylation of tripeptide where the histidine is replaced by a nonchelating phenyl alanine residue. d) Proposed ATCUN models to explain the difference in reactivity between tripeptides **19a,b** and **21**. Yields are of isolated pure products.

Finally, we verified that the trityl group could be chemoselectively removed under mild conditions, a process which was achieved by stirring **22** in a 9:1 mixture of acetic acid and water at 60 °C for 1.5 h to afford **25** in quantitative yield (Scheme 7).



Scheme 7. Selective trityl removal in arylated peptide **22**.

In conclusion, we have developed a protocol to arylate the imidazole side chain of histidine by using triarylbismuthines. The reaction is promoted by cupric acetate, uses 1,10-phenanthroline as a ligand and DIPEA as the base, and operates in dichloromethane at 40 °C under an oxygen atmosphere. The reaction is insensitive to electronic

or steric effects, shows excellent functional group compatibility, and can be performed with Boc, Cbz or Fmoc protecting groups. The protocol was applied to histidine-containing dipeptides where histidine is at the N terminus. Chemoselective arylation on the histidine residue was obtained in most cases, except when histidine was connected to a tyrosine. Dipeptides where the histidine is at the C terminus resulted in imidazole and N-H backbone arylation. Protection of the imidazole with a trityl group led to exclusive backbone arylation. The arylation occurs at the n-1 position when this position is also the N-terminal position. Chemoselective removal of the trityl group was accomplished under mild conditions.

Acknowledgements

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Scientific Advancement grant. We would like to thank Prof. Steve Bourgault for useful discussions and advice.

Conflict of Interest

The authors declare no conflict of interest.

Keywords: copper catalysis • histidine • imidazole • N-arylation • organobismuthines

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3.3 Informations supplémentaires

Les informations supplémentaires concernant les protocoles expérimentaux, les caractérisations et les spectres RMN ^1H et ^{13}C des composés synthétisés sont disponible en Annexe A et sur <https://chemistry-europe.onlinelibrary.wiley.com/doi/10.1002/chem.202102186>.

3.4 Contributions des auteurs à l'article

L'auteur de la thèse, et premier auteur de l'article, a réalisé une petite partie de la table d'optimisation des conditions (Table 1), la préparation de la moitié des triarylbismuths pour le schéma 2 et le reste des réactions pour les schémas 3 à 7 ainsi que pour la figure 1. Ceci inclut l'ensemble de la synthèse des dipeptides et tripeptides. L'auteur a également rédigé l'ensemble de la partie expérimentale, le traitement des spectres RMN et a contribué à l'écriture de l'article, la recherche bibliographique affiliée et la préparation des figures.

La seconde auteure, Bianca Bueno, a contribué à la majorité des réactions d'optimisation de la table 1 ainsi que l'intégralité des réactions pour le schéma 2. Elle a également fait la relecture et la correction de la partie expérimentale.

Le troisième auteur, Adrien Le Roch, a contribué à l'initiation du projet avec des réactions démontrant la compatibilité des conditions à l'arylation de l'histidine.

L'auteur de correspondance, le Professeur Alexandre Gagnon, a effectué la rédaction du corps de l'article ainsi que le processus de soumission de celui-ci et a aidé l'auteur principal dans la relecture de la partie expérimentale.

CHAPITRE 4

ON THE COPPER-PROMOTED BACKBONE ARYLATION OF THE HISTIDINE-CONTAINING PEPTIDES USING TRIARYLBISMUTHINES

4.1 Introduction

Suites aux développements des réactions d'arylation de l'imidazole de l'histidine, il a été possible de démontrer que la position du résidu au sein du peptide avait une importance sur la régiospécificité lors du transfert de l'aryle. En effet, un acide aminé précédant l'histidine au sein d'un di- ou tripeptide possède un azote alpha (N^{terminal}) suffisamment actif pour se faire aryle. Une observation très peu documentée dans la littérature et qui a été jugée intéressante à investiguer davantage.

4.2 Article issu de ces travaux

Chan, H.-C. and Gagnon, A. (2022). On the Copper-Promoted Backbone Arylation of Histidine-Containing Peptides using Triarylismuthines. *Synthesis*. <https://doi.org/10.1055/a-1786-6578>

On the Copper-Promoted Backbone Arylation of Histidine-Containing Peptides using Triarylismuthines

Hwai-Chien Chan^[a] and Alexandre Gagnon^{*[a]}

[a] Université du Québec à Montréal
Département de chimie
C.P. 8888, Succursale Centre-Ville
Montréal, Québec, Canada, H3C 3P8
E-mail: gagnon.alexandre@uqam.ca

This paper is dedicated to the memory Dr. Michael Boš for his remarkable contribution to the pharmaceutical industry in the Montreal area and his friendship.

Abstract We report herein our detailed investigation on the histidine-directed backbone arylation of histidine-containing peptides using triarylismuth reagents. The reaction proceeds on the backbone NH of the amino acid that precedes the histidine, the so-called $n-1$ position. The protocol is applicable to dipeptides where the histidine is located at the C-terminus and to tripeptides where the histidine occupies the central position. The transformation is promoted by copper(II) acetate in the presence of phenanthroline (Phen) and diisopropylethylamine in dichloromethane at 50 °C

under oxygen. An excellent scope was observed for the triarylismuthines. In all cases, the imidazole ring of the histidine is protected with a trityl group to prevent the arylation of the side chain. An ATCUN-like model is proposed to explain the observed results.

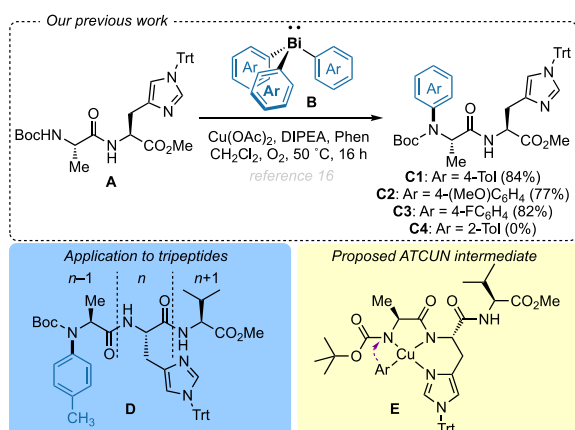
Key words triarylismuthines, histidine, peptides, backbone arylation, copper catalysis

Organobismuth compounds are versatile organometallic species that can be easily accessed from

inexpensive and low-toxic bismuth trichloride.¹ Originally pioneered by Barton, Finet, and Dodonov in the 1980s,² these species have recently seen a remarkable renaissance with the work of Burford,³ Dóstał,⁴ Lichtenberg,⁵ L. T. Ball,⁶ and Cornella,⁷ who greatly contributed to demonstrating their unique properties and expanding their synthetic utility. Our own interest in organobismuth chemistry resulted in the development of a series of metal-catalyzed reactions which led to the construction of C–C,⁸ C–N,⁹ C–O,¹⁰ and C–S bonds.¹¹

Amino acids and peptides, long considered as poor choices for drug discovery, are now gaining traction in medicinal chemistry due to their non-planarity and their ability to form strong and selective interactions with various biological targets.¹² As a consequence, multiple methods to modify amino acids and peptides have appeared in the literature over the past decade.¹³ In addition to giving access to potential drug candidates, many of these methods also allow the chemoselective installation of small organic groups that help modulate the biophysical properties of the peptide or investigate the role played by specific residues in the peptide's activity.

Recently, we published three papers on the arylation of the side chain of tryptophan,¹⁴ tyrosine,¹⁵ and histidine¹⁶ using triarylbiimidines. In the course of our studies on the arylation of histidine-containing peptides, we found that the histidine residue was capable of directing the arylation on the backbone of a dipeptide at the so-called *n*–1 position (Scheme 1, A → C). To prevent the arylation of the side chain of the histidine, the imidazole ring was protected with a trityl group.



Scheme 1. Our work on the copper-catalyzed histidine-directed backbone arylation of peptides using triarylbiimidines (Phen = phenanthroline)

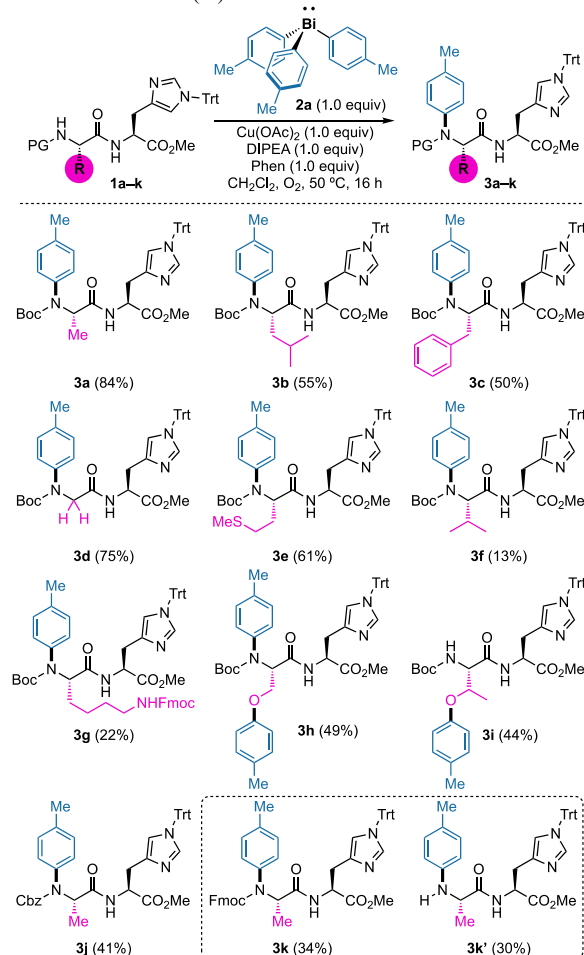
Our result was in line with that of Z. T. Ball, who observed similar results during the nickel/copper-catalyzed arylation of histidine-containing peptides using arylboronic acids.¹⁷ Ball's method was remarkably powerful due to its ability to selectively arylate large proteins at the position preceding the histidine.

As a preliminary investigation on the copper-catalyzed histidine-directed backbone arylation of peptides using triarylbiimidines, we found that the reaction was not affected by electronic effects on the aryl ring (C1–3), but was completely inhibited by substituents in the ortho position (C4) (see Scheme 1). Using Boc-Ala-His(Trt)-Val-OMe to test this manifold, we also discovered that the backbone arylation

proceeded only when the *n*–1 residue was located at the N-terminus of the peptide, delivering the arylated product D. To explain these results, the ATCUN-like (amino terminal copper and nickel) model E was proposed (Scheme 1).¹⁸

Hoping that our method could be complementary to Ball's method, where the amino acid which is arylated does not have to be at the terminal position,¹⁷ and to other methods which indiscriminately arylate the N terminus of peptides,¹⁹ we decided to undertake a thorough investigation of this intriguing copper-promoted histidine-directed backbone arylation of peptides.

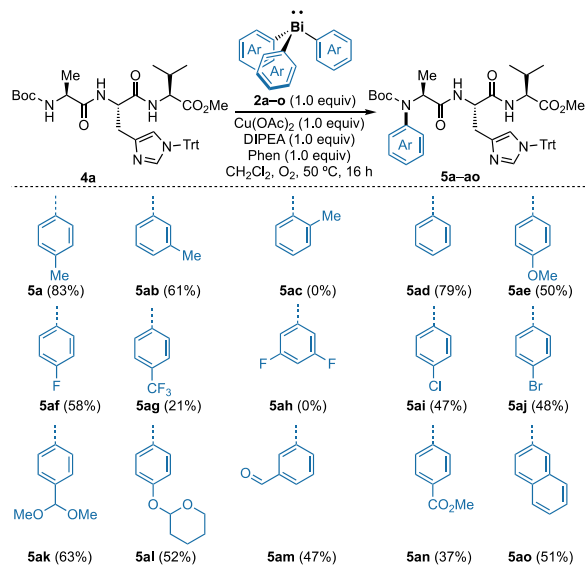
Having not demonstrated the scope of the *n*–1 amino acid in the reaction A → C above, we began by arylating dipeptides **1a–k** using tris(4-tolyl)bismuth (**2a**) under our previously reported conditions (Scheme 2).¹⁶ In the event, fair to good yields were obtained in cases where the N-terminal amino acid was an alanine (**3a**), a leucine (**3b**), a phenylalanine (**3c**), a glycine (**3d**), or a methionine (**3e**). However, a low yield was obtained for N-Boc-Val-His(Trt)-OMe (**3f**), presumably due to severe steric congestion between the aryl, the tert-butyloxycarbonyl, and the isopropyl group. Chemoselective backbone arylation occurred in dipeptide **3g**, which contains an Fmoc-protected lysine, albeit in low yield. Submitting Boc-Ser-His(Trt)-OMe to our conditions provided the diarylation product **3h** in modest yield. To our surprise, a reversal of chemoselectivity was observed for Boc-Thr-His(Trt)-OMe, where preferential arylation of the OH moiety on the threonine side chain occurred (**3i**).



Scheme 2. Scope of the N-terminal amino acid in the histidine-directed backbone arylation of histidine-containing dipeptides

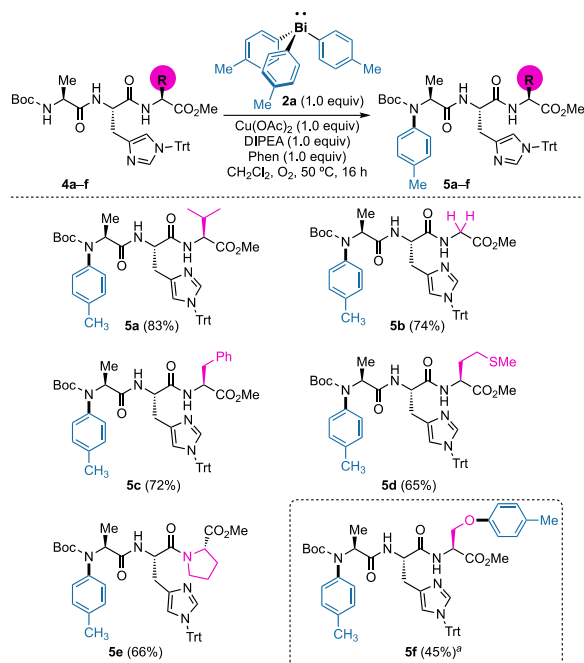
The backbone arylation proceeded less efficiently on peptides protected at the N-terminal with a benzyloxy carbonyl (compare **3j** with **3a**) or a fluorenylmethoxycarbonyl (compare **3k** with **3a**) group (Scheme 2). Unexpectedly, in the case of Fmoc-Ala-His(Trt)-OMe, we also isolated a considerable amount of the unprotected arylated product **3k'**. These two examples indicate that while the arylation can be performed with Cbz-, Fmoc-, and Boc-protected peptides, it is more efficient with the latter.

Having already explored the scope of the aryl group with dipeptides (cf. products **C1–4**), we decided to study the arylation of Boc-Ala-His(Trt)-Val-OMe (**4a**) using various triarylbi-muthines (Scheme 3). As predicted from our studies on dipeptide A, we found that the reaction operates reasonably well with *para*- and *meta*-substituted or unsubstituted aryl groups, but not with *ortho*-substituted ones (**5a–ad**). Electron-rich and -deficient aryl groups were transferred in comparable yields (**5ae–af**). However, a low yield was observed with *p*-(trifluoromethyl)phenyl (**5ag**), while no product could be isolated with 3,5-difluorophenyl (**5ah**). The process tolerates halogens as well as various functional groups (**5ai–an**), offering points of entry for further diversification. A naphthyl group was also transferred in modest yield (**5ao**).



Scheme 3. Scope of the triarylbi-muthines in the arylation of Boc-Ala-His(Trt)-Val-OMe

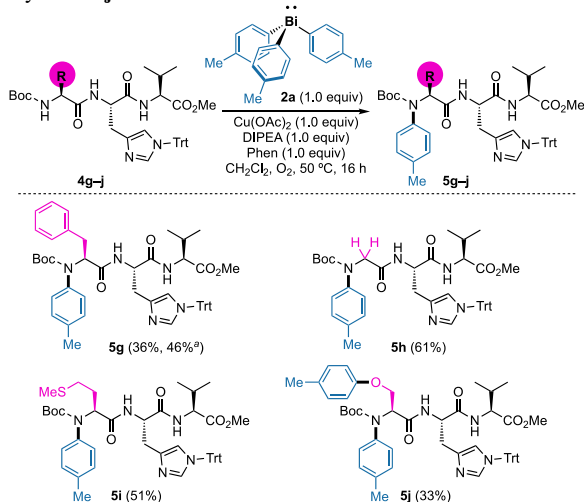
We then moved our focus to the C-terminal *n+1* residue of tripeptides **4a–f** (Scheme 4), with the assumption that this position, as it is remote from the arylation site, should not have a dramatic impact on the efficiency of the process, unless it has a reactive side chain. As anticipated, tripeptides bearing a glycine (**5b**), a phenylalanine (**5c**), a methionine (**5d**), or a proline (**5e**) at the *n+1* position gave similar yields as the original valine-containing tripeptide (**5a**). As expected from compound **3h**, the presence of a serine at the *n+1* position resulted in the formation of the double arylation product **5f**. This product was accompanied by a small amount of mono-arylated product on the serine residue, suggesting that the diarylation proceeds first on the serine and then on the terminal NH.



Scheme 4. Scope of the C-terminal *n+1* amino acid in the backbone N–H arylation of tripeptides. ^a Accompanied by 6% of the mono-arylated product on serine.

Using tripeptides **4g–j**, we then evaluated the impact of the amino acid at the *n–1* position on the arylation process (Scheme 5). As anticipated, because the arylation proceeds on the NH backbone at that location, we observed a much greater influence from this residue than the one at the *n+1* position. For instance, a simple, although slightly more encumbered phenylalanine led to a lower yield of the desired product than an alanine (**5g** vs **5a**). The yield could be improved to 46% by using 1.5 equivalents of each reagent.

Arylation of Boc-Gly-His(Trt)-Val-OMe and Boc-Met-His(Trt)-Val-OMe afforded the respective products **5h** and **5i** in slightly higher yields than **5g**, but still lower than **5a** (Scheme 5). As predicted from **3h** and **5f**, arylation of Boc-Ser-His(Trt)-Val-OMe **4j** exclusively provided the product of double arylation **5j**.

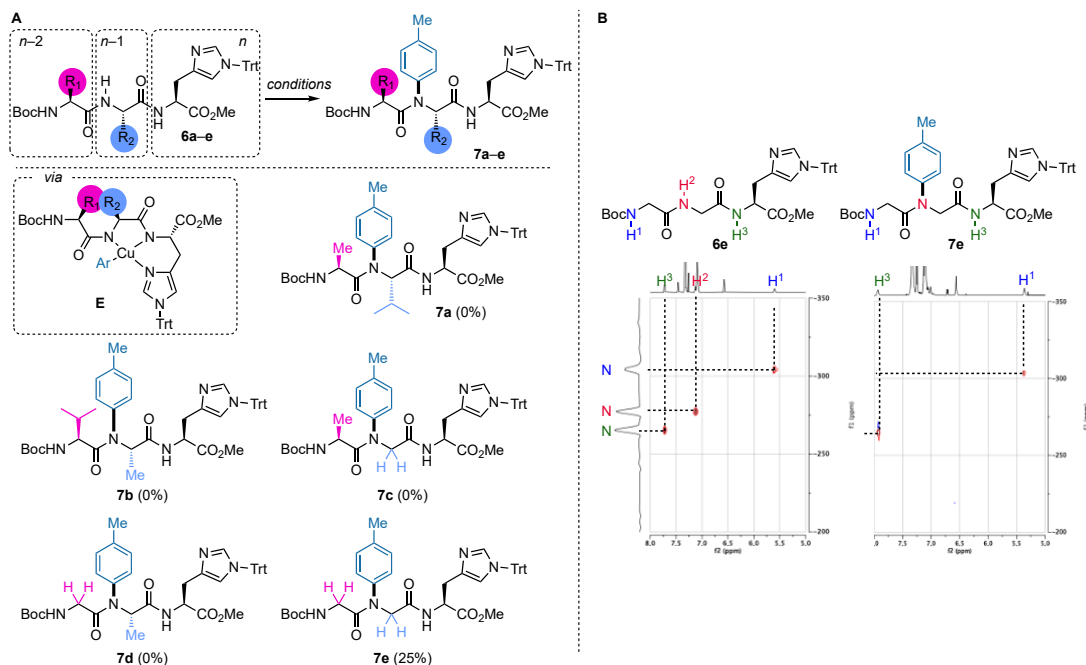


Scheme 5. Scope of the N-terminal *n–1* amino acid in the backbone N–H arylation of tripeptides. ^a Reaction performed with **2a** (1.5 equiv), Cu(OAc)₂, DIPEA, Phen.

In our previous paper,¹⁶ we observed that tripeptides **6a,b**, where the histidine is at the C-terminal *n* position, failed to deliver the desired product of backbone arylation **7a,b** at *n-1* (Scheme 6). To explain these results, we proposed the ATCUN model **E**, where steric congestion between the side chain residues R¹ and R² of the amino acids at the *n-1* and *n-2* positions prevents the formation of the copper(III) intermediate, thus shutting down the arylation process. This led us to conclude that the histidine-directed backbone arylation proceeds on the amino acid preceding the histidine *only* if this residue is *also* at the N-terminal position.

To test this hypothesis, we submitted tripeptides **6c** and **6d**, where a glycine at *n-1* or *n-2* is expected to reduce the congestion and therefore permit the arylation reaction. In the event, the unreacted peptides were recovered intact (Scheme 6), suggesting that our model is either incorrect or that the release

of steric congestion is still not sufficient to allow the formation of the presumed intermediate **E**. Therefore, we then tested tripeptide Boc-Gly-Gly-His(Trt)-OMe (**6e**) under our conditions and obtained the desired backbone arylation product **7e** in 13% yield. The yield could be increased to 25% by using 1.5 equivalents of tritolylbismuth, copper acetate, DIPEA, and phenanthroline. Comparison of the HSQC ¹H-¹⁵N NMR spectra between **6e** and **7e** showed the disappearance of the hydrogen on the central glycine, thus confirming that the arylation proceeded at the *n-1* position, that is, left from the histidine (Scheme 6B). Despite the low yields, these results appear to support our model **E**, leading us to slightly revise our original conclusion: the histidine-directed backbone arylation occurs at the amino acid preceding the histidine if this residue is located at the N-terminal position or if it is part of a Gly-Gly-His triad.



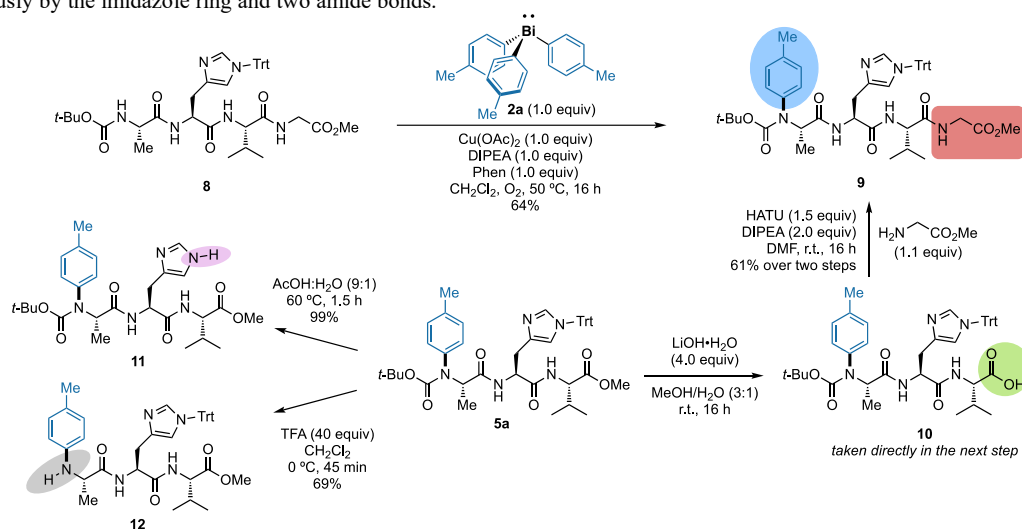
Scheme 6. (A) Studies aimed at testing the ATCUN model **E** for the histidine-directed backbone arylation of tripeptides where the histidine occupies the C-terminal position. Reagents and conditions: Tol₃Bi (1.5 equiv), Cu(OAc)₂ (1.5 equiv), DIPEA (1.5 equiv), Phen (1.5 equiv), CH₂Cl₂, O₂, 50 °C, 16h. (B) HSQC ¹H-¹⁵N spectra of **6e** and **7e** (full spectra in Supporting Information, SI).

To explore the limit of our method, we submitted tetrapeptide Boc-Ala-His(Trt)-Val-Gly-OMe (**8**) to our conditions and obtained the corresponding product **9** in 64% yield, indicating that the process is poorly affected by elongation of the peptide on the C-terminus (Scheme 7). However, we believe that the limit of our procedure has been reached due to solubility issues associated with higher peptides in dichloromethane. Previous attempts at changing the solvent resulted in drastic erosion in the yield of the reaction. To further increase the versatility of our method, we developed conditions to selectively remove each protecting group in **5a** (Scheme 7). The ester could easily be saponified using lithium hydroxide in a 3:1 mixture of methanol and water, resulting in **10**, which was directly coupled with glycine methyl ester by using HATU, to generate **9** in 61% yield over two steps. The ¹H NMR spectrum of **9** obtained via the coupling of **10** was identical to that obtained through the arylation of **8**, confirming that the tolyl group was installed on the alanine residue on **8**. The trityl group was selectively removed by treating **5a** with acetic acid in water at 60 °C for 1.5 hours, providing **11** in quantitative yield. Selective

removal of the tert-butyloxycarbonyl group could be accomplished by treating **5a** with trifluoroacetic acid in dichloromethane at 0 °C for 45 minutes, affording **12** in 69% yield.

In conclusion, we developed a histidine-directed backbone arylation of peptides using triarylbismuthines. The reaction is catalyzed by cupric acetate and operates in dichloromethane at 50 °C under oxygen in the presence of phenanthroline as the copper ligand and diisopropylethylamine as the base. The reaction proceeds on the backbone of dipeptides and tripeptides at the amino acid preceding the histidine only when it occupies the N-terminal position. The arylation of the backbone of tripeptides where the histidine is located at the C-terminus is possible only with the Gly-Gly-His triad. In all cases, the imidazole of the histidine is protected with a trityl group to prevent the arylation of the side chain. Good scope was observed for the triarylbismuth, except for ortho-substituted ones. The protocol was applied to one tetrapeptide. Conditions were developed for the selective removal of each protecting group. The reaction is presumed to proceed via an ATCUN-like

intermediate, where the copper metal is complexed simultaneously by the imidazole ring and two amide bonds.



Scheme 7. Histidine-directed backbone arylation of tetrapeptide **8** and further derivatization of tripeptide **5a**

Conflict of Interest

The authors declare no conflict of interest.

Funding Information

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Supporting Information

Supporting information for this article is available online at <https://doi.org/10.1055/a-1786-6578>.

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4.3 Informations supplémentaires

Les informations supplémentaires concernant les protocoles expérimentaux, les caractérisations et les spectres RMN ^1H et ^{13}C des composés synthétisés sont disponible en Annexe B

4.4 Contributions des auteurs à l'article

L'auteur de la thèse, et premier auteur de l'article, a réalisé l'ensemble des réactions. L'auteur a également rédigé la partie expérimentale, le traitement des spectres RMN et a contribué à l'écriture de l'article, la recherche bibliographique affiliée et la préparation des figures.

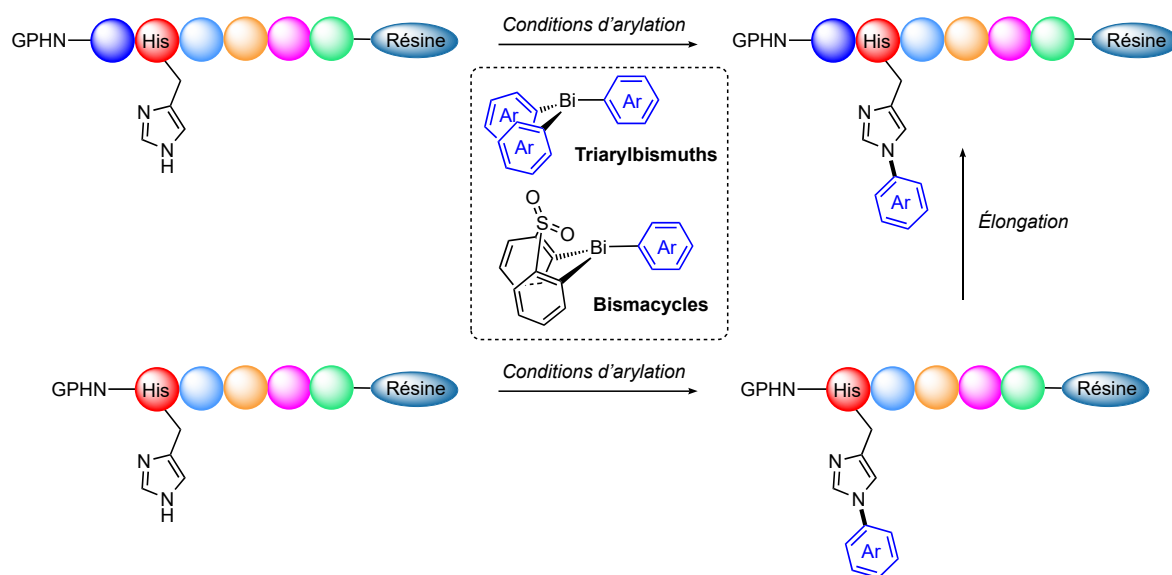
L'auteur de correspondance, le Professeur Alexandre Gagnon, a effectué la rédaction du corps de l'article ainsi que le processus de soumission de celui-ci et a aidé l'auteur principal dans la relecture de la partie expérimentale.

CONCLUSION

À travers cette thèse il a été possible de faire un peu plus connaissance avec l'histidine en y apprenant comment elle était produite et métabolisée ainsi que ses implications dans les divers processus biologiques. Par la suite, il a été possible de voir que son groupement imidazole pouvait être amené à des modifications naturelles et artificielles. Ces dernières ont été détaillées et présentent un champ large de possibilité avec les travaux remarquables à la fois d'alkylation et d'arylation. D'un autre côté, le bismuth a été présenté comme un métal relativement récent dans le domaine de la chimie de synthèse et mérite une attention toute particulière au vu du fort potentiel qu'il présente. Cette thèse a tenté de le prouver en ajoutant un outil supplémentaire à l'arsenal des modifications post-synthétiques de peptides et de l'histidine elle-même. La méthode développée permet d'aryler l'imidazole de l'histidine avec une compatibilité avec des groupements aromatiques enrichis ou appauvris en électrons et avec des substituants à toutes les positions. De plus, il a été démontré que l'histidine présentait une capacité à diriger l'arylation sur l'azote du résidu placé juste avant elle.

La durée d'une thèse étant évidemment trop courte, il n'a pas été possible d'explorer toutes les facettes des organobismuths et des études supplémentaires peuvent encore être effectuées. En effet, étudier le comportement des organobismuthines dans le cadre de peptides plus longs est une perspective intéressante. La synthèse sur support solide des peptides faciliterait grandement la préparation de tels peptides et l'étape d'arylation pourrait éventuellement s'effectuer entre deux phases d'élongation. Une autre limite assez sensible dans l'utilisation des triarylbismuths est qu'un seul des groupements aryle parmi les trois n'est transférable. Cette limitation pourrait être éventuellement contournée en passant par une stratégie employant des bismacycles hypervalents (voir figure 4-1).

Figure 4-1. Perspectives sur l'étude du comportement des organobismuthines sur de longs peptides synthétisés sur support solide



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Annexe A

Informations supplémentaires

*Copper-Promoted N-Arylation of the Imidazole Side Chain of Protected Histidine by Using
Triarylbismuth Reagents*

SUPPORTING INFORMATION

Copper-Promoted *N*-Arylation of the Imidazole Side Chain of Protected

Histidine using Triarylbiomuth Reagents

Hwai-Chien Chan, Bianca Bueno, Adrien Le Roch, Alexandre Gagnon*

Département de chimie
Université du Québec à Montréal
C.P. 8888, Succ. Centre-Ville, Montréal, Québec, Canada, H3C 3P8
gagnon.alexandre@uqam.ca

1. General information	A2
2. Triarylbiomuthines (2) used in this work	A3
3. General procedure for the optimization of the reaction conditions for the coupling between Boc-His-OMe (1a) and tri(4-tolyl)biomuth (2a)	A4
4. General procedure for the coupling between <i>N</i> -protected histidine methyl ester (1a) and triarylbiomuthines (2)	A4
5. General procedure for the coupling between <i>N</i> -protected amino acid methyl ester and tri(4-tolyl)biomuthines (2a)	A14
6. Procedures for the preparation of dipeptides	A16
a. Procedure for synthesis of Boc-His-A.A.-OMe (12a-f and 12j-l)	A16
b. Procedure for synthesis of intermediate Boc-His(Trt)-Glu(OMe)-OMe (12g') ; Boc-His(Trt)-Thr-OMe (12h') and Boc-His(Trt)-Ser-OMe (12i')	A21
c. Procedure for synthesis of Boc-His-Glu(OMe)-OMe (12g); Boc-His-Thr-OMe (12h) and Boc-His-Ser-OMe (12i)	A23
d. Procedure for synthesis Boc-Ala-His-OMe (14)	A24
e. Procedure for synthesis Boc-Ala-His(trt)-OMe (17)	A25
7. Procedure for the preparation of aryl dipeptides	A26
a. General procedure for synthesis of Boc-His(4-tolyl)-A.A.-OMe (13a-l)	A26
b. Procedure for synthesis of Boc-Ala-His(4-tolyl)-OMe (15) and Boc-N(4-tolyl)-Ala-His(4-tolyl)-OMe (16)	A33
c. General procedure for synthesis of Boc-N(Aryl)-Ala-His(trt)-OMe (18a-c)	A34
8. Procedure for the preparation of tripeptides	A36
a. General procedure for synthesis of Boc-A.A ₁ -A.A ₂ -His(Trt)-OMe (19)	A36

b.	<i>Procedure for synthesis of Boc-Ala-His(Trt)-Val-OMe (21)</i>	A38
c.	<i>Procedure for synthesis of Boc-Ala-Phe-Val-OMe (23)</i>	A39
9.	Procedure for the preparation of aryl tripeptides	A40
a.	<i>Procedure for synthesis of Boc-N(Tolyl)-Ala-His(trt)-Val-OMe (22)</i>	A40
b.	<i>Procedure for synthesis of Boc-N(Tolyl)-Ala-His-Val-OMe (25)</i>	A41

1. General information

Unless otherwise indicated, all reactions were run under ambient atmosphere in non-flame dried glassware. For reactions performed under oxygen, 99.6% extra dry oxygen was used. Unless otherwise stated, commercial reagents were used without further purification. Triarylbismuthines were prepared according to Hébert et al.ⁱ Anhydrous solvents were obtained using a MBRAUN (model MB-SPS 800) encapsulated solvent purification system. The evolution of reactions was monitored by analytical thin-layer chromatography using silica gel 60 F254 precoated plates. Flash chromatography was performed employing 220 mesh silica (Silicycle) using the indicated solvent system according to standard techniques. Nuclear magnetic resonance spectra ¹H were recorded on a BrukerAvance-III 300MHz spectrometer or a BrukerAvance-III 600MHz and ¹³C were recorded on a BrukerAvance-III 75MHz spectrometer or a BrukerAvance-III 150MHz. Chemical shifts for ¹H-NMR spectra are recorded in parts per million from tetramethylsilane with the solvent resonance as the internal standard (chloroform, δ 7.26 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, p = quintuplet, m = multiplet, dd = doublet of doublet, dt = doublet of triplet, dq = doublet of quartet, tt = triplet of triplet, td = triplet of doublet, s(br) = broad singlet), coupling constant *J* in Hz and integration. Chemical shifts for ¹³C-NMR spectra are recorded in parts per million from tetramethylsilane using the central peak of deuteriochloroform (77.16 ppm) as the internal standard. All ¹³C-NMR spectra were obtained with complete proton decoupling. IR spectra were recorded on a Thermo Scientific Nicolet 6700 PT-IR from thin films and are reported in reciprocal centimeters (cm⁻¹). Melting points were measured using Büchi M560 melting point apparatus in open capillaries.

ⁱ Hébert, M.; Petiot, P.; Benoit, E.; Dansereau, J.; Ahmad, T.; Le Roch, A.; Ottenwaelde, X.; Gagnon, A. *J. Org. Chem.* **2016**, *81*, 5401–5416

2. Triarylbiomuthines (2) used in this work

The organobismuthines used in this publication are illustrated in **Figure S1**. The procedures for synthesis of these organobismuthines can be found in the following references: ^a P. Petiot, A. Gagnon, *Eur. J. Org. Chem.*, **2013**, 5282; ^b P. Petiot, J. Dansereau, A. Gagnon, *RSC Adv.*, **2014**, 4, 22255; ^c D. H. R. Barton, N. Y. Bhatnagar, J.-P. Finet and W. B. Motherwell, *Tetrahedron*, **1986**, 42, 3111; ^d R. Ding, C.-S. Ge, Y.-J. Chen, D. Wanga, C.-J. Li, *Tetrahedron Lett.*, **2002**, 43, 7789; ^e M. H. bert, P. Petiot, E. Benoit, J. Dansereau, T. Ahmad, A. Le Roch, X. Ottenwaelder, A. Gagnon, *J. Org. Chem.*, **2016**, 81, 5401; ^f X. Marset, J. Torregrosa-Crespo, R. M. Martínez-Espinosa, G. Guillena, D. J. Ramón, *Green Chem.*, **2019**, 21, 4127.

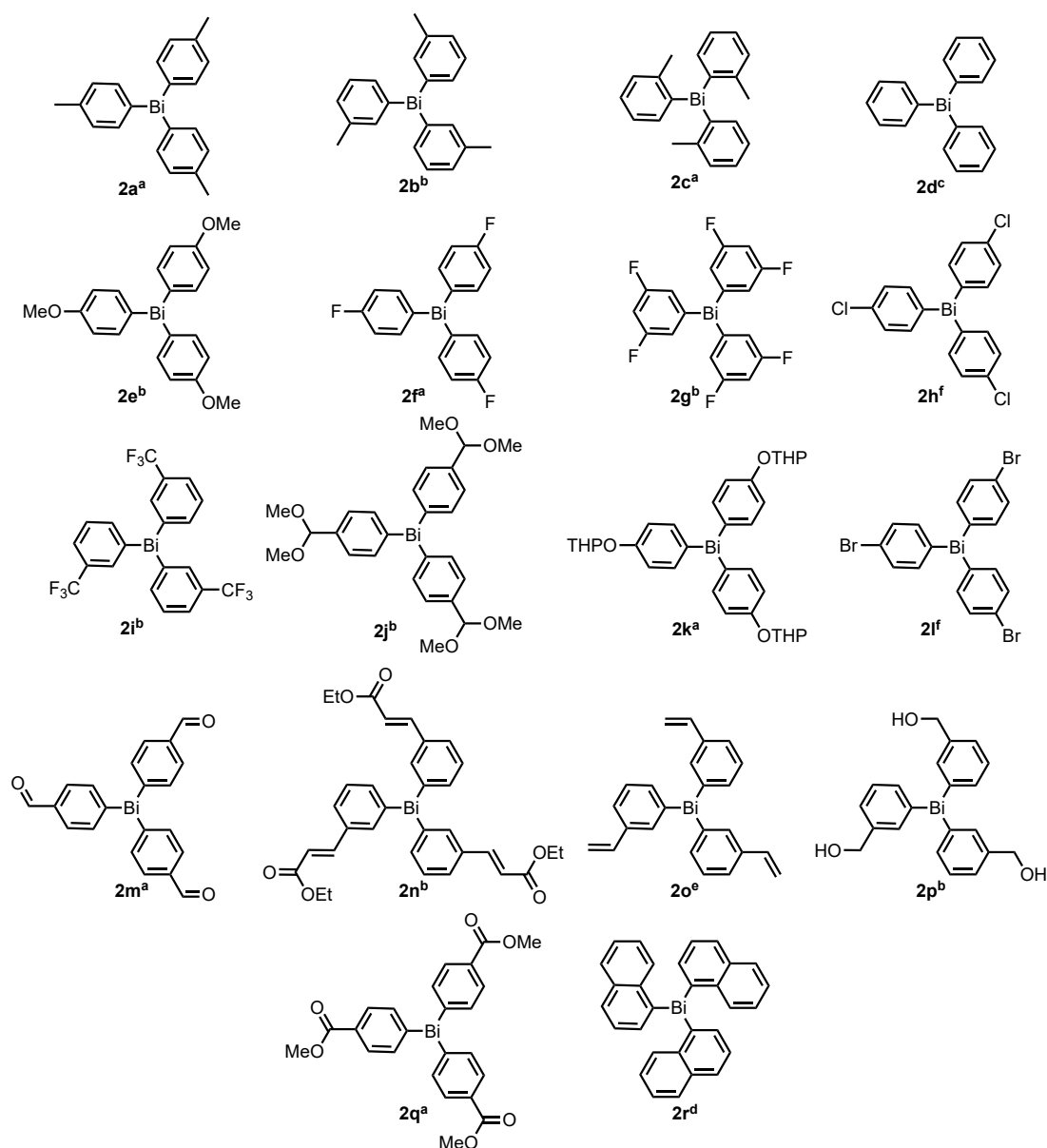
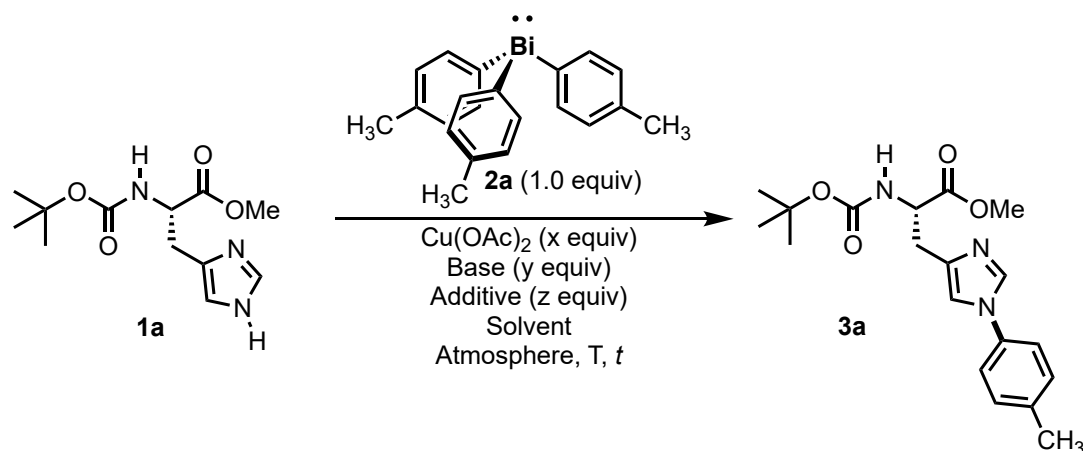


Figure S1. Functionalized organobismuthines used in this publication

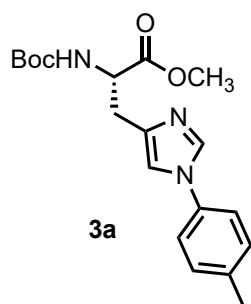
3. General procedure for the optimization of the reaction conditions for the coupling between Boc-His-OMe (**1a**) and tri(4-tolyl)bismuth (**2a**)



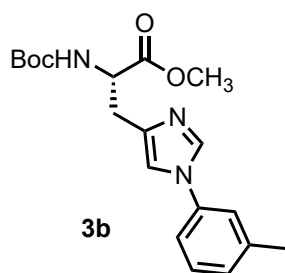
Manuscript, Table 1: In a sealed tube, the base (y equiv) was added to a mixture of the *N*-protected *L*-histidine methyl ester **1a** (1.0 equiv), tri(4-tolyl)bismuthine **2a** (1.0 equiv), copper(II) acetate (x equiv) and additive (z equiv) in CH_2Cl_2 (0.1 M). The mixture was stirred at the corresponding temperature under the corresponding atmosphere for 4 or 16 hours. The solvent was evaporated under reduce pressure and the residue was purified on silica gel chromatography system (2% MeOH/ CH_2Cl_2) to afford **3a** as a yellow thick oil. See section 4 for complete characterization of **3a**.

4. General procedure for the coupling between *N*-protected histidine methyl ester (**1a**) and triarylbismuthines (**2**)

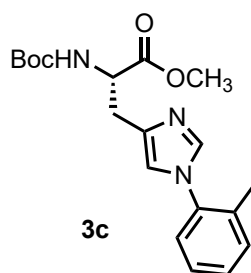
Manuscript, Scheme 2: In a sealed tube, *N,N*-diisopropylethylamine (25.5 μL , 0.15 mmol, 1.0 equiv) was added to the *N*-protected *L*-histidine methyl ester **1a** (0.15 mmol, 1.0 equiv), the corresponding triarylbismuthine **2** (0.15 mmol, 1.0 equiv), copper(II) acetate (27 mg, 0.15 mmol, 1.0 equiv) and phenanthroline (27 mg, 0.15 mmol, 1.0 equiv) in CH_2Cl_2 (0.1 M). Oxygen was bubbled in the reaction mixture before stirred in an oil bath set at 40 $^\circ\text{C}$ for 16 hours. The solvent was evaporated under reduce pressure and the residue was purified on silica gel chromatography using the corresponding eluent system to afford desired pure product **3a–r**.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(4-tolyl)-*L*-histidinate (3a)

The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40.0 mg) and tri(4-tolyl)bismuthine **2a** (72 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3a** as a yellow thick oil (53 mg, 99%): *R*_f = 0.46 (5% MeOH/CH₂Cl₂); ¹H-NMR (300 MHz, CDCl₃) δ 7.73 (s(br), 1H), 7.28 – 7.17 (m, 4H), 7.04 (s(br), 1H), 5.90 (d, *J* = 6.5 Hz, 1H), 4.59 (s(br), 1H), 3.70 (s, 3H), 3.29 – 2.89 (m, 2H), 2.37 (s, 3H), 1.42 (s, 9H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.7, 155.7, 137.6, 134.9, 130.5, 121.3, 79.8, 53.6, 52.3, 30.4, 28.4, 21.0; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3350, 3129, 2973, 2928, 1741, 1703, 1521, 1483, 1365, 1251, 1159, 1051, 1019, 813; HRMS (ESI) calcd. for [C₁₉H₂₅N₃O₄ + H]⁺: 360.1918, found 360.1922.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(3-tolyl)-*L*-histidinate (3b)

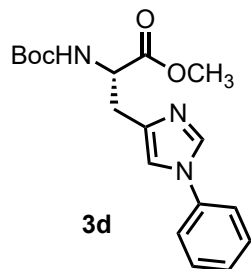
The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(3-tolyl)bismuthine **2b** (72 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3b** as a yellow thick oil (53 mg, 99%): *R*_f = 0.44 (5% MeOH/CH₂Cl₂); ¹H-NMR (600 MHz, CDCl₃) δ 7.82 (s(br), 1H), 7.33 (t, *J* = 7.7 Hz, 1H), 7.17 – 7.10 (m, 3H), 7.08 (s(br), 1H), 5.89 (d, *J* = 7.2 Hz, 1H), 4.60 (s, 1H), 3.71 (s, 3H), 3.23 – 3.02 (m, 2H), 2.40 (s, 3H), 1.42 (s, 9H); ¹³C-NMR (150 MHz, CDCl₃) δ 172.6, 155.7, 140.2, 137.1, 129.8, 128.4, 122.0, 118.4, 79.8, 53.5, 52.4, 30.3, 28.4, 21.5; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3241, 3127, 3106, 2970, 2950, 2926, 1743, 1697, 1614, 1594, 1538, 1506, 1436, 1363, 1158, 1069, 1048, 1030, 985, 854, 786, 691; HRMS (ESI) calcd. for [C₁₉H₂₅N₃O₄ + H]⁺: 360.1918, found 360.1923.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(2-tolyl)-*L*-histidinate (3c)

The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(2-tolyl)bismuthine **2c** (72 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3c** as a yellow oil (38 mg, 71%): *R*_f = 0.51 (3% MeOH/CH₂Cl₂); ¹H-NMR (300 MHz, CDCl₃) δ 7.53 (s(br), 1H), 7.38 – 7.28 (m, 2H), 7.28 – 7.24 (m, 1H), 7.18 (d, *J* = 7.6 Hz, 1H), 6.84 (s(br), 1H), 5.93 (d, *J* = 7.8 Hz, 1H), 4.65 – 4.55 (m, 1H), 3.71 (s, 3H), 3.23 – 3.01 (m, 2H), 2.15 (s, 3H), 1.43 (s, 9H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.6, 155.7, 136.5, 133.9, 131.5,

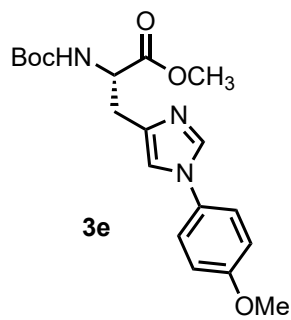
129.1, 127.0, 126.5, 79.8, 53.7, 52.3, 30.3, 28.5, 17.7; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹):** 3357, 2974, 2930, 1744, 1708, 1501, 1436, 1391, 1365, 1163, 1022, 992, 969, 761, 719; **HRMS (ESI)** calcd. for [C₁₉H₂₅N₃O₄ + H]⁺: 360.1918, found 360.1929.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(phenyl)-*L*-histidinate (3d**)**

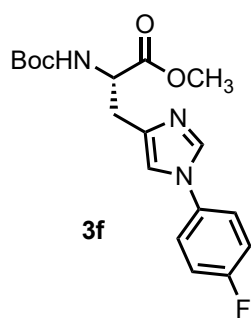


The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and triphenylbismuthine **2d** (66 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3d** as a yellow oil (48 mg, 94%): *R*_f = 0.46 (5% MeOH/CH₂Cl₂); **¹H-NMR (600 MHz, CDCl₃)** δ 7.86 (s(br), 1H), 7.49 – 7.40 (m, 2H), 7.36 – 7.29 (m, 3H), 7.09 (s(br), 1H), 5.89 (d, *J* = 7.5 Hz, 1H), 4.59 (s(br), 1H), 3.70 (s, 3H), 3.21 – 3.03 (m, 2H), 1.41 (s, 9H); **¹³C-NMR (150 MHz, CDCl₃)** δ 172.5, 155.7, 138.3, 137.1, 135.2, 130.0, 127.7, 121.3, 116.1, 79.8, 53.5, 52.4, 30.2, 28.4; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹):** 3382, 3012, 2971, 2951, 1741, 1698, 1600, 1500, 1435, 1391, 1365, 1157, 1055, 753, 692; **HRMS (ESI)** calcd. for [C₁₈H₂₃N₃O₄ + H]⁺: 346.1761, found 346.1775.

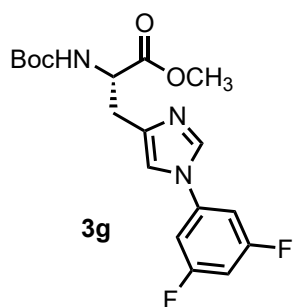
Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(4-methoxyphenyl)-*L*-histidinate (3e**)**



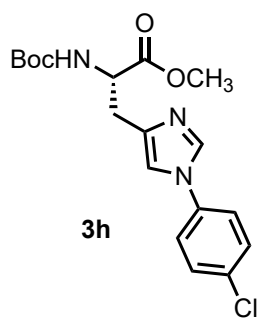
The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(4-methoxyphenyl)bismuthine **2e** (79 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3e** as a yellow oil (50 mg, 90%): *R*_f = 0.34 (5% MeOH/CH₂Cl₂); **¹H-NMR (600 MHz, CDCl₃)** δ 7.77 (s(br), 1H), 7.24 (d, *J* = 8.8 Hz, 2H), 7.00 (s(br), 1H), 6.95 (d, *J* = 8.8 Hz, 2H), 5.90 (d, *J* = 7.2 Hz, 1H), 4.58 (s, 1H), 3.81 (s, 3H), 3.70 (s, 3H), 3.19 – 3.11 (m, 1H), 3.11 – 3.04 (m, 1H), 1.41 (s, 9H); **¹³C-NMR (150 MHz, CDCl₃)** δ 172.5, 159.1, 155.7, 137.9, 135.3, 130.4, 123.1, 116.6, 115.0, 79.8, 55.7, 53.5, 52.3, 30.2, 28.4; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹):** 3373, 3263, 3134, 2975, 2841, 1741, 1693, 1606, 1518, 1436, 1390, 1250, 1160, 1126, 1069, 1028, 983, 970, 835, 756; **HRMS (ESI)** calcd. for [C₁₉H₂₅N₃O₅ + H]⁺: 376.1867, found 376.1879.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(4-fluorophenyl)-*L*-histidinate (3f**)**

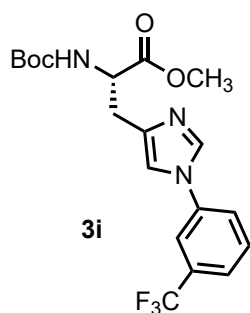
The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(4-fluorophenyl)bismuthine **2f** (74 mg). The crude material was purified on silica gel (3% MeOH/CH₂Cl₂) to afford **3f** as a yellow oil (52 mg, 96%): *R*_f = 0.33 (5% MeOH/CH₂Cl₂); ¹H-NMR (600 MHz, CDCl₃) δ 7.85 (s(br), 1H), 7.36 – 7.28 (m, 2H), 7.14 (t, *J* = 8.4 Hz, 2H), 7.04 (s(br), 1H), 5.87 (d, *J* = 7.2 Hz, 1H), 4.59 (s, 1H), 3.71 (s, 3H), 3.21 – 3.03 (m, 2H), 1.40 (s, 9H); ¹³C-NMR (150 MHz, CDCl₃) δ 172.5, 161.8 (d, *J* = 246.7 Hz), 155.7, 133.4, 123.4 (d, *J* = 8.4 Hz), 116.9 (d, *J* = 23.0 Hz), 79.8, 53.4, 52.4, 30.2, 29.7, 28.4; ¹⁹F-NMR (282 MHz, CDCl₃) δ -113.7; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3125, 2980, 2932, 2850, 1744, 1701, 1515, 1437, 1391, 1366, 1226, 1159, 1058, 1022, 839, 818, 753, 665; HRMS (ESI) calcd. for [C₁₈H₂₂FN₃O₄ + H]⁺: 364.1667, found 364.1683.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(3,5-difluorophenyl)-*L*-histidinate (3g**)**

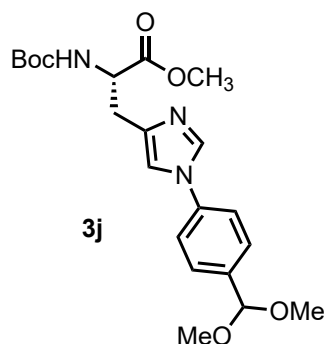
The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tris(3,5-difluorophenyl)bismuthine **2g** (82 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3g** as a yellow solid (43 mg, 76%): *R*_f = 0.77 (5% MeOH/CH₂Cl₂); *m.p.* = 129 – 131 °C; ¹H-NMR (300 MHz, CDCl₃) δ 7.84 (s, 1H), 7.07 (s(br), 1H), 6.98 – 6.86 (m, 2H), 6.80 (tt, *J* = 8.7, 2.2 Hz, 1H), 5.78 (d, *J* = 7.5 Hz, 1H), 4.60 (s(br), 1H), 3.72 (s, 3H), 3.34 – 2.81 (m, 2H), 1.42 (s, 9H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.5, 165.3 (d, *J* = 14.1 Hz), 162.0 (d, *J* = 14.3 Hz), 155.7, 104.613 (d, *J* = 28.6 Hz), 104.612 (d, *J* = 9.1 Hz), 103.3, 103.0, 102.6, 79.9, 53.3, 52.4, 30.4, 28.4; ¹⁹F-NMR (282 MHz, CDCl₃) δ -106.3; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3253, 3134, 3110, 2959, 2931, 2851, 1742, 1692, 1626, 1605, 1540, 1506, 1477, 1435, 1364, 1285, 1260, 1220, 1162, 1127, 1067, 994, 851, 836, 758, 674; HRMS (ESI) calcd. for [C₁₈H₂₁F₂N₃O₄ + H]⁺: 382.1573, found 382.1580.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(4-chlorophenyl)-*L*-histidinate (3h)

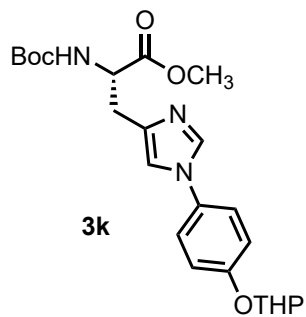
The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(4-chlorophenyl)bismuthine **2h** (81 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3h** as a thick yellow oil (55 mg, 97%): *R*_f = 0.76 (5% MeOH/CH₂Cl₂); ¹H-NMR (300 MHz, CDCl₃) δ 7.78 (s, 1H), 7.41 (dt, *J* = 9.3, 2.4 Hz, 2H), 7.27 (dt, *J* = 9.4, 2.4 Hz, 2H), 7.04 (s(br), 1H), 5.84 (d, *J* = 7.5 Hz, 1H), 4.58 (s(br), 1H), 3.70 (s, 3H), 3.23 – 2.98 (m, 2H), 1.40 (s, 9H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.6, 155.7, 135.7, 133.3, 130.1, 122.5, 79.8, 53.4, 52.4, 30.4, 28.4; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3375, 2977, 1743, 1705, 1504, 1436, 1392, 1365, 1306, 1250, 1210, 1161, 1096, 1014, 991, 968, 829, 754; HRMS (ESI) calcd. for [C₁₈H₂₂ClN₃O₄ + H]⁺: 380.1372, found 380.1376.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(3-(trifluoromethyl)phenyl)-*L*-histidinate (3i)

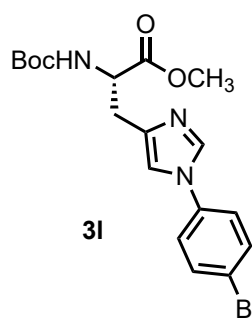
The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(3-(trifluoromethyl)phenyl)bismuthine **2i** (96 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3i** as a yellow oil (57 mg, 93%): *R*_f = 0.7 (5% MeOH/CH₂Cl₂); ¹H-NMR (600 MHz, CDCl₃) δ 7.99 (s, 1H), 7.65 – 7.53 (m, 4H), 7.15 (s, 1H), 5.83 (d, *J* = 7.4 Hz, 1H), 4.60 (s, 1H), 3.71 (s, 3H), 3.20 – 3.06 (m, 2H), 1.40 (s, 9H); ¹³C-NMR (150 MHz, CDCl₃) δ 172.4, 155.7, 139.0, 137.5, 135.2, 132.7 (q, *J* = 131.7 Hz), 130.9, 124.5, 124.4 (q, *J* = 14.4 Hz), 123.4 (q, *J* = 1084.5 Hz), 118.20 (q, *J* = 14.7 Hz), 115.9, 79.9, 53.3, 52.4, 30.2, 28.4; ¹⁹F-NMR (282 MHz, CDCl₃) δ -62.88; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 2977, 2934, 1744, 1705, 1619, 1599, 1505, 1437, 1392, 1366, 1322, 1166, 1125, 1098, 1068, 1021, 803, 756, 697; HRMS (ESI) calcd. for [C₁₉H₂₂F₃N₃O₄ + H]⁺: 414.1635, found 414.1648.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(4-(dimethoxymethyl)phenyl)-*L*-histidinate (3j)

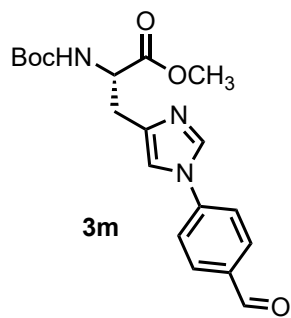
The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(4-(dimethoxymethyl)phenyl)bismuthine **2j** (98 mg). The crude material was purified on basified silica gel (2% MeOH/CH₂Cl₂) to afford **3j** as a yellow oil (62 mg, 99%): *R*_f = 0.45 (5% MeOH/CH₂Cl₂); ¹H-NMR (600 MHz, CDCl₃) δ 7.79 (s, 1H), 7.53 (d, *J* = 8.4 Hz, 2H), 7.32 (d, *J* = 8.5 Hz, 2H), 7.06 (s, 1H), 5.86 (d, *J* = 8.0 Hz, 1H), 5.40 (s, 1H), 4.58 (X of ABX, *J* = 12.7, 5.2 Hz, 1H), 3.70 (s, 3H), 3.31 (s, 6H), 3.13 (A of ABX, *J* = 14.8, 5.7 Hz, 1H), 3.08 (B of ABX, *J* = 14.7, 4.9 Hz, 1H), 1.40 (s, 9H); ¹³C-NMR (150 MHz, CDCl₃) δ 172.5, 155.7, 137.7, 137.1, 135.2, 128.5, 120.9, 115.9, 102.3, 79.8, 53.5, 52.8, 52.3, 30.3, 28.4, 8.7; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 2937, 2831, 1744, 1705, 1613, 1522, 1489, 1436, 1366, 1206, 1614, 1101, 1054, 985, 969, 818, 753; HRMS (ESI) calcd. for [C₂₁H₂₉N₃O₆ + H]⁺: 420.2129, found 420.2138.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(4-((tetrahydro-2H-pyran-2-yl)oxy)phenyl)-*L*-histidinate (3k)

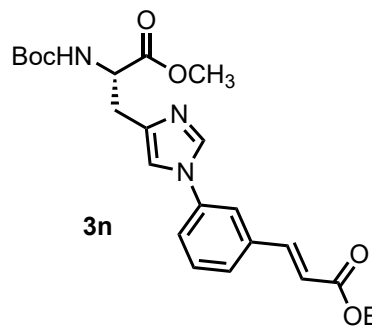
The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(4-((tetrahydro-2H-pyran-2-yl)oxy)phenyl)bismuthine **2k** (110 mg). The crude material was purified on basified silica gel (2% MeOH/CH₂Cl₂) to afford **3k** as a yellow oil (66 mg, 99%): *R*_f = 0.47 (5% MeOH/CH₂Cl₂); ¹H-NMR (600 MHz, CDCl₃) δ 7.69 (s(br), 1H), 7.21 (dt, *J* = 9.7, 2.5 Hz, 2H), 7.10 (dt, *J* = 9.7, 2.6 Hz, 2H), 6.99 (s(br), 1H), 5.90 (d, *J* = 7.4 Hz, 1H), 5.41 (t, *J* = 3.1 Hz, 1H), 4.57 (s, 1H), 3.85 (ddd, *J* = 12.0, 9.2, 2.1 Hz, 1H), 3.69 (s, 3H), 3.62 – 3.56 (m, 1H), 3.19 – 2.98 (m, 2H), 2.02 – 1.92 (m, 1H), 1.88 – 1.81 (m, 2H), 1.73 – 1.63 (m, 2H), 1.62 – 1.54 (m, 1H), 1.41 (s, 9H); ¹³C-NMR (150 MHz, CDCl₃) δ 172.6, 156.5, 155.7, 131.2, 122.8, 117.6, 96.6, 79.7, 62.1, 53.5, 52.3, 30.3, 29.7, 28.4, 25.1, 18.7; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 2946, 2874, 1747, 1709, 1515, 1456, 1436, 1365, 1237, 1201, 1166, 1109, 1074, 1035, 958, 919, 835, 752; HRMS (ESI) calcd. for [C₂₃H₃₁N₃O₆ + H]⁺: 446.2286, found 446.2293.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(4-bromophenyl)-*L*-histidinate (3l**)**

The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(4-bromophenyl)bismuthine **2l** (101 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3l** as a thick yellow oil (50 mg, 79%): *R*_f = 0.67 (5% MeOH/CH₂Cl₂); ¹H-NMR (600 MHz, CDCl₃) δ 7.88 (s, 1H), 7.58 (d, *J* = 8.6 Hz, 2H), 7.24 (d, *J* = 8.7 Hz, 2H), 7.06 (s, 1H), 5.84 (d, *J* = 7.7 Hz, 1H), 4.66–4.51 (m, 1H), 3.71 (s, 3H), 3.15 (A of ABX, *J* = 14.6, 4.8 Hz, 1H), 3.10 (B of ABX, *J* = 14.7, 4.1 Hz, 1H), 1.41 (s, 9H); ¹³C-NMR (150 MHz, CDCl₃) δ 172.5, 155.7, 136.1, 133.2, 122.8, 121.2, 79.9, 53.4, 52.4, 30.2, 28.4; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 2979, 1744, 1701, 1500, 1436, 1365, 1161, 1068, 1010, 966, 826, 752; HRMS (ESI) calcd. for [C₁₈H₂₂BrN₃O₄ + H]⁺: 424.0867, found 424.0876.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(4-formylphenyl)-*L*-histidinate (3m**)**

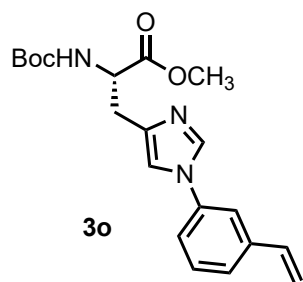
The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(4-formylphenyl)bismuthine **2m** (78 mg). The crude material was purified on silica gel (3% MeOH/CH₂Cl₂) to afford **3m** as a yellow oil (53 mg, 96%): *R*_f = 0.37 (5% MeOH/CH₂Cl₂); ¹H-NMR (300 MHz, CDCl₃) δ 10.00 (s, 1H), 8.02–7.92 (m, 3H), 7.52 (d, *J* = 8.5 Hz, 2H), 7.18 (s, 1H), 5.81 (d, *J* = 8.0 Hz, 1H), 4.70–4.50 (m, 1H), 3.70 (s, 3H), 3.20–2.98 (m, 2H), 1.39 (s, 9H); ¹³C-NMR (75 MHz, CDCl₃) δ 190.6, 172.5, 155.6, 141.4, 135.1, 131.7, 120.9, 79.9, 53.3, 52.4, 30.4, 28.4; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3358, 3125, 2980, 2849, 2742, 1744, 1698, 1605, 1519, 1486, 1436, 1392, 1366, 1305, 1206, 1164, 1059, 1023, 966, 835, 752; HRMS (ESI) calcd. for [C₁₉H₂₃N₃O₅ + H]⁺: 374.1711, found 374.1724.

Ethyl (*S,E*)-3-(3-(4-(2-((*tert*-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)-1*H*-imidazol-1-yl)phenyl)acrylate (3n**)**

The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and triethyl 3,3',3''-(bismuthanetriyltris(benzene-3,1-diyl))(2*E*,2'*E*,2''*E*)-triacrylate **2n** (109 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3n** as a yellow oil (65 mg, 99%): *R*_f = 0.45 (5% MeOH/CH₂Cl₂); ¹H-NMR (600 MHz, CDCl₃)

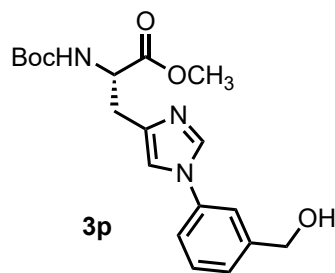
δ 7.90 (s(br), 1H), 7.65 (d, J = 16.0 Hz, 1H), 7.52 – 7.44 (m, 3H), 7.35 (d, J = 7.1 Hz, 1H), 7.13 (s(br), 1H), 6.47 (d, J = 16.0 Hz, 1H), 5.86 (d, J = 6.8 Hz, 1H), 4.60 (s, 1H), 4.25 (q, J = 7.1 Hz, 2H), 3.71 (s, 3H), 3.24 – 3.00 (m, 2H), 1.41 (s, 9H), 1.32 (t, J = 7.1 Hz, 3H); **$^{13}\text{C-NMR}$ (150 MHz, CDCl_3)** δ 172.5, 166.5, 155.7, 142.8, 137.7, 136.5, 130.6, 127.0, 122.7, 120.5, 120.4, 79.8, 60.8, 53.4, 52.4, 30.3, 28.4, 14.3; **IR (ATR)/ $\bar{\nu}$ (cm^{-1})**: 2980, 1705, 1640, 1589, 1499, 1366, 1307, 1162, 1027, 983, 860, 753; **HRMS (ESI)** calcd. for $[\text{C}_{23}\text{H}_{29}\text{N}_3\text{O}_6 + \text{H}]^+$: 444.2129, found 444.2131.

Methyl N^α -(*tert*-butoxycarbonyl)- N^ϵ -(3-vinylphenyl)-*L*-histidinate (**3o**)



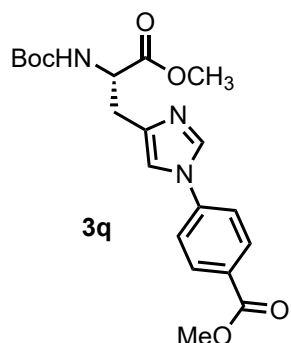
The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(3-vinylphenyl)bismuthine **2o** (77 mg). The crude material was purified on silica gel (2% MeOH/ CH_2Cl_2) to afford **3o** as a yellow oil (54 mg, 98%): R_f = 0.57 (5% MeOH/ CH_2Cl_2); **$^1\text{H-NMR}$ (600 MHz, CDCl_3)** δ 7.87 (s(br), 1H), 7.41 – 7.34 (m, 2H), 7.33 (s, 1H), 7.20 (d, J = 7.5 Hz, 1H), 6.70 (dd, J = 17.6, 10.9 Hz, 1H), 5.91 (d, J = 6.5 Hz, 1H), 5.79 (d, J = 17.5 Hz, 1H), 5.33 (d, J = 10.9 Hz, 1H), 4.59 (s(br), 1H), 3.70 (s, 3H), 3.19 – 2.99 (m, 2H), 1.40 (s, 9H); **$^{13}\text{C-NMR}$ (150 MHz, CDCl_3)** δ 172.5, 155.6, 139.6, 137.5, 135.6, 130.1, 125.4, 120.5, 119.0, 115.9, 79.7, 53.4, 52.3, 30.2, 28.3; **IR (ATR)/ $\bar{\nu}$ (cm^{-1})**: 2979, 1744, 1705, 1607, 1585, 1498, 1436, 1392, 1366, 1162, 1055, 1023, 991, 915, 751; **HRMS (ESI)** calcd. for $[\text{C}_{20}\text{H}_{25}\text{N}_3\text{O}_4 + \text{H}]^+$: 372.1918, found 372.1930.

Methyl N^α -(*tert*-butoxycarbonyl)- N^ϵ -(3-(hydroxymethyl)phenyl)-*L*-histidinate (**3p**)



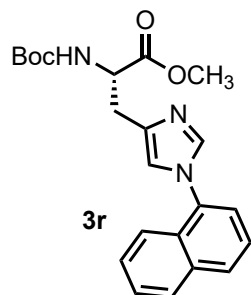
The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(3-(hydroxymethyl)phenyl)bismuthine **2p** (79 mg). The crude material was purified on silica gel (3% MeOH/ CH_2Cl_2) to afford **3p** as a yellow oil (44 mg, 79%): R_f = 0.45 (10% MeOH/ CH_2Cl_2); **$^1\text{H-NMR}$ (300 MHz, CDCl_3)** δ 7.72 (s(br), 1H), 7.46 – 7.28 (m, 3H), 7.18 (d, J = 7.8 Hz, 1H), 7.06 (s, 1H), 5.83 (d, J = 8.0 Hz, 1H), 4.73 (s, 2H), 4.61 – 4.49 (m, 1H), 4.30 (s, 1H), 3.70 (s, 3H), 3.16 – 2.97 (m, 2H), 1.40 (s, 9H); **$^{13}\text{C-NMR}$ (75 MHz, CDCl_3)** δ 172.6, 155.7, 143.9, 137.2, 135.2, 130.0, 125.8, 119.9, 119.4, 116.0, 79.9, 64.0, 53.6, 52.4, 29.8, 28.4; **IR (ATR)/ $\bar{\nu}$ (cm^{-1})**: 3310, 2920, 2850, 1741, 1699, 1499, 1456, 1436, 1365, 1249, 1214, 1160, 1052, 1022, 785; **HRMS (ESI)** calcd. for $[\text{C}_{19}\text{H}_{25}\text{N}_3\text{O}_5 + \text{H}]^+$: 376.1867, found 376.1884.

Methyl (S)-4-(4-(2-((*tert*-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)-1H-imidazol-1-yl)benzoate (3q)



The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and trimethyl 4,4',4''-bismuthanetriyltribenzoate **2q** (91 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3q** as a white solid (55 mg, 92%): *R_f* = 0.34 (5% MeOH/CH₂Cl₂); *m.p.* = 50–52 °C; ¹H-NMR (300 MHz, CDCl₃) δ 8.11 (dt, *J* = 8.9, 2.0 Hz, 2H), 7.84 (s, 1H), 7.40 (dt, *J* = 9.0, 2.0 Hz, 2H), 7.12 (s, 1H), 5.83 (d, *J* = 8.0 Hz, 1H), 4.69–4.47 (m, 1H), 3.91 (s, 3H), 3.70 (s, 3H), 3.20–3.01 (m, 2H), 1.40 (s, 9H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.5, 166.0, 155.6, 140.5, 135.1, 131.6, 129.0, 120.3, 115.4, 79.8, 53.4, 52.4, 30.4, 29.7, 28.4; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3341, 2919, 2843, 1708, 1609, 1520, 1485, 1435, 1365, 1281, 1210, 1162, 1111, 1056, 1017, 966, 855, 769, 694; HRMS (ESI) calcd. for [C₂₀H₂₅N₃O₆ + H]⁺: 404.1816, found 404.1820.

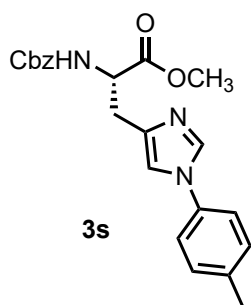
Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(naphthalen-1-yl)-*L*-histidinate (3r)



The general procedure was followed starting from (*L*)-Boc-His-OMe **1a** (40 mg) and tri(naphthalen-1-yl)bismuthine **2r** (88 mg). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3r** as a yellow oil (46 mg, 78%): *R_f* = 0.76 (5% MeOH/CH₂Cl₂); ¹H-NMR (300 MHz, CDCl₃) δ 8.02–7.88 (m, 2H), 7.71 (s, 1H), 7.61–7.47 (m, 4H), 7.42 (d, *J* = 6.5 Hz, 1H), 7.04 (s, 1H), 6.00 (d, *J* = 7.8 Hz, 1H), 4.66 (s, 1H), 3.74 (s, 3H), 3.32–3.07 (m, 2H), 1.44 (s, 9H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.6, 155.7, 134.3, 133.9, 129.5, 129.5, 128.4, 127.8, 127.1, 125.3, 123.7, 122.3, 79.8, 53.8, 52.4, 30.4, 28.5; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3413, 3345, 2981, 2950, 1741, 1706, 1599, 1488, 1435, 1365, 1161, 1055, 1022, 940, 864, 803, 773, 753, 644; HRMS (ESI) calcd. for [C₂₂H₂₅N₃O₄ + H]⁺: 396.1918, found 396.1920.

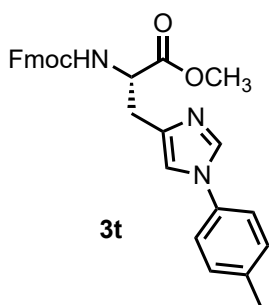
Methyl *N*^α-((benzyloxy)carbonyl)-*N*^ε-(4-tolyl)-*L*-histidinate (3s)

In a sealed tube, *N,N*-diisopropylethylamine (22.5 μL, 0.13 mmol, 1.0 equiv) was added to the (*L*)-Cbz-His-OMe **1b** (40 mg, 0.13 mmol, 1.0 equiv), tri(4-tolyl)bismuthine **2a** (64 mg, 0.13 mmol, 1.0 equiv), copper(II) acetate (24 mg, 0.13 mmol, 1.0 equiv) and phenanthroline (24 mg, 0.13 mmol, 1.0 equiv) in CH₂Cl₂ (0.1 M). The mixture was purged with oxygen before stirred in an oil bath set at 40 °C for 16 hours under oxygen. The solvent was evaporated under



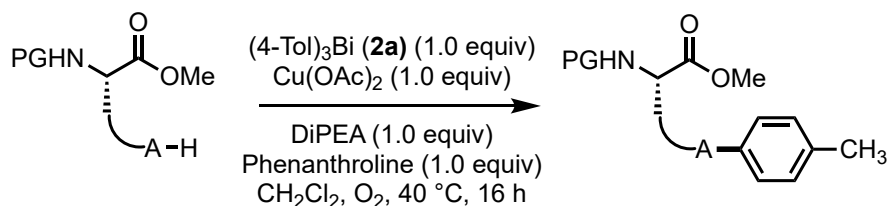
reduce pressure and the crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **3s** as a yellow oil (45 mg, 87%): R_f = 0.26 (5% MeOH/CH₂Cl₂); **¹H-NMR (300 MHz, CDCl₃)** δ 7.72 (s, 1H), 7.39 – 7.29 (m, 5H), 7.27 – 7.14 (m, 4H), 7.07 – 6.91 (m, 1H), 6.31 (d, J = 6.7 Hz, 1H), 5.12 (d, J = 2.4 Hz, 2H), 4.67 (s, 1H), 3.72 (s, 3H), 3.29 – 3.00 (m, 2H), 2.38 (s, 3H); **¹³C-NMR (150 MHz, CDCl₃)** δ 172.3, 156.2, 137.6, 136.6, 134.9, 130.5, 129.5, 128.5, 128.2, 128.1, 121.3, 116.2, 66.9, 54.0, 52.4, 30.2, 21.0; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3324, 3034, 2945, 1713, 1518, 1487, 1454, 1435, 1307, 1247, 1207, 1175, 1027, 969, 815, 739, 697; **HRMS (ESI)** calcd. for [C₂₂H₂₃N₃O₄ + H]⁺: 394.1761, found 394.1771.

Methyl N^α-(((9H-fluoren-9-yl)methoxy)carbonyl)-N^ε-(4-tolyl)-L-histidinate (**3t**)



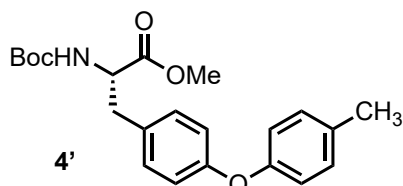
In a sealed tube, *N,N*-diisopropylethylamine (17.5 μ L, 0.10 mmol, 1.0 equiv) was added to the (*L*)-Fmoc-His-OMe **1c** (40 mg, 0.10 mmol, 1.0 equiv), tri(4-tolyl)bismuthine **2a** (49 mg, 0.10 mmol, 1.0 equiv), copper(II) acetate (19 mg, 0.10 mmol, 1.0 equiv) and phenanthroline (18 mg, 0.10 mmol, 1.0 equiv) in CH₂Cl₂ (0.1 M). The mixture was purged with oxygen before stirred in an oil bath set at 40 °C for 16 hours under oxygen. The solvent was evaporated under reduce pressure and the crude material was purified on silica gel (1% MeOH/CH₂Cl₂) to afford **3t** as an orange oil (25 mg, 51%): R_f = 0.35 (5% MeOH/CH₂Cl₂); **¹H-NMR (300 MHz, CDCl₃)** δ 10.89 (s(br), 1H), 8.55 (s, 1H), 7.72 (d, J = 7.5 Hz, 2H), 7.58 (t, J = 6.5 Hz, 2H), 7.43 – 7.16 (m, 8H), 6.66 – 6.51 (m, 1H), 4.66 (s, 1H), 4.45 – 4.24 (m, 2H), 4.16 (t, J = 7.0 Hz, 1H), 3.80 (s, 3H), 3.53 – 3.39 (m, 1H), 3.39 – 3.23 (m, 1H), 2.42 (s, 3H); **¹³C-NMR (150 MHz, CDCl₃)** δ 171.0, 156.5, 143.9, 143.8, 141.4, 140.8, 133.1, 132.6, 131.1, 127.9, 127.2, 125.4, 122.0, 120.0, 118.5, 67.4, 53.7, 53.2, 47.1, 27.8, 21.2; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3297, 3137, 2950, 2919, 1669, 1541, 1513, 1437, 1177, 1130, 1079, 1045, 816, 797, 759, 740, 719; **HRMS (ESI)** calcd. for [C₂₉H₂₇N₃O₄ + H]⁺: 482.2074, found 482.2086.

5. General procedure for the coupling between *N*-protected amino acid methyl ester and tri(4-tolyl)bismuthines (**2a**)



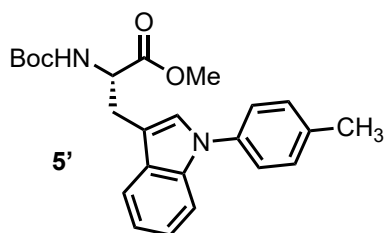
Manuscript, Figure 1: In a sealed tube, *N,N*-diisopropylethylamine (1.0 equiv) was added to the *N*-protected (*L*)-amino acid methyl ester (40 mg, 1.0 equiv), tri(4-tolyl)bismuthine **2a** (1.0 equiv), copper(II) acetate (1.0 equiv) and phenanthroline (1.0 equiv) in CH₂Cl₂ (0.1 M). Oxygen was bubbled in the reaction mixture before stirred in an oil bath set at 40 °C for 16 hours. The solvent was evaporated under reduce pressure and the residue was purified on silica gel chromatography using the corresponding eluent system to afford desired pure product.

Methyl (S)-2-((*tert*-butoxycarbonyl)amino)-3-(4-(*p*-tolylloxy)phenyl)propanoate (**4'**)



The general procedure was followed starting from (*L*)-Boc-Tyr-OMe (40 mg, 0.14 mmol), tri(4-tolyl)bismuthine **2a** (65 mg, 0.14 mmol), *N,N*-diisopropylethylamine (23.0 μ L, 0.14 mmol), copper(II) acetate (25 mg, 0.14 mmol) and phenanthroline (25 mg, 0.14 mmol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **4'** as a colorless oil (38 mg, 73%). *R*_f = 0.53 (30% EtOAc/Hexane); This compound has been previously reported ⁱⁱ : ¹H-NMR (600 MHz, CDCl₃) δ 7.13 (d, *J* = 8.3 Hz, 2H), 7.06 (d, *J* = 8.3 Hz, 2H), 6.91 (d, *J* = 2.4 Hz, 2H), 6.89 (d, *J* = 2.4 Hz, 2H), 5.03 (d, *J* = 7.9 Hz, 1H), 4.57 (X of ABX, *J* = 13.3, 6.1 Hz, 1H), 3.72 (s, 3H), 3.09 (A of ABX, *J* = 13.8, 5.5 Hz, 1H), 3.01 (B of ABX, *J* = 13.9, 6.1 Hz, 1H), 2.34 (s, 3H), 1.43 (s, 9H).

Methyl *N* ^{α} -(*tert*-butoxycarbonyl)-1-(*p*-tolyl)-*L*-tryptophanate (**5'**)

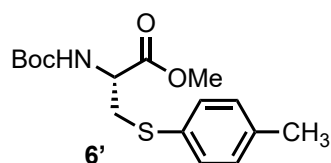


The general procedure was followed starting from (*L*)-Boc-Trp-OMe (40 mg, 0.13 mmol), tri(4-tolyl)bismuthine **2a** (61 mg, 0.13 mmol), *N,N*-diisopropylethylamine (21.5 μ L, 0.13 mmol), copper(II) acetate (23 mg, 0.13 mmol) and

ⁱⁱ Le Roch, A.; Hébert, M.; Gagnon, A. *Eur. J. Org. Chem.* **2020**, 33, 5363–5367

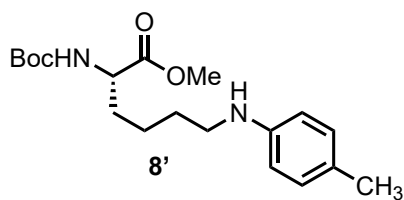
phenanthroline (23 mg, 0.13 mmol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **5'** as a white solid (6 mg, 12%). The compound contained impurities. R_f = 0.30 (EtOAc:Hexane 2:8); This compound has been previously reportedⁱⁱⁱ: **¹H-NMR (300 MHz, CDCl₃)** δ 7.59 (d, J = 7.3 Hz, 1H), 7.49 (d, J = 7.5 Hz, 1H), 7.39 – 7.26 (m, 4H), 7.24 – 7.09 (m, 3H), 5.11 (d, J = 7.0 Hz, 1H), 4.74 – 4.65 (m, 1H), 3.71 (s, 3H), 3.41 – 3.25 (m, 2H), 2.43 (s, 3H), 1.43 (s, 9H).

Methyl *N*-(*tert*-butoxycarbonyl)-*S*-(*p*-tolyl)-*L*-cysteinate (**6'**)



The general procedure was followed starting from (*L*)-Boc-Cys-OMe (40 mg, 0.17 mmol), tri(4-tolyl)bismuthine **2a** (82 mg, 0.17 mmol), *N,N*-diisopropylethylamine (29.0 μ L, 0.17 mmol), copper(II) acetate (31 mg, 0.17 mmol) and phenanthroline (31 mg, 0.17 mmol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **6'** as a yellow oil (11 mg, 19%). The compound contained impurities R_f = 0.30 (1% MeOH/CH₂Cl₂); This compound has been previously reportedⁱⁱ: **¹H-NMR (300 MHz, CDCl₃)** δ 7.31 (dt, J = 8.5, 2.2 Hz, 2H), 7.10 (d, J = 7.9 Hz, 2H), 5.45 – 5.19 (m, 1H), 4.60 – 4.45 (m, 1H), 3.54 (s, 3H), 3.32 (d, J = 4.7 Hz, 2H), 2.31 (s, 3H), 1.41 (s, 9H).

Methyl *N*²-(*tert*-butoxycarbonyl)-*N*⁶-(*p*-tolyl)-*L*-lysinate (**8'**)



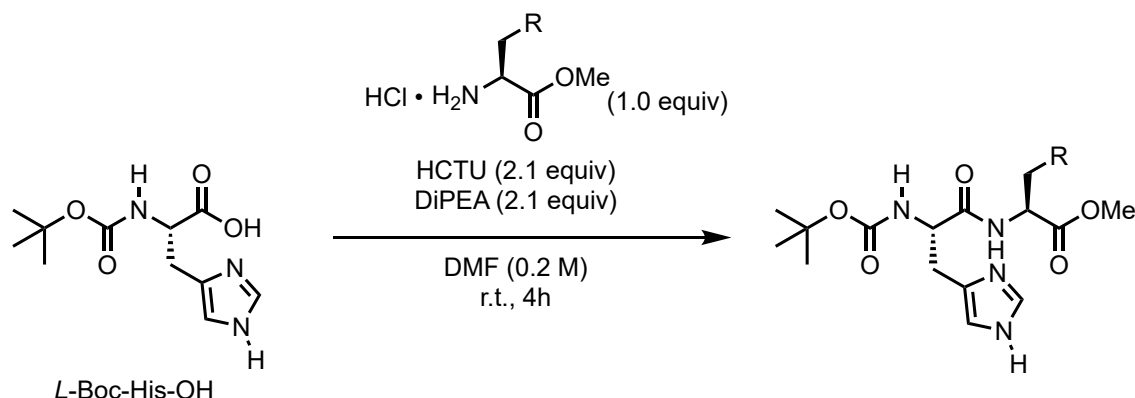
The general procedure was followed starting from (*L*)-Boc-Lys-OMe•HCl (40 mg, 0.14 mmol), tri(4-tolyl)bismuthine **2a** (65 mg, 0.14 mmol), *N,N*-diisopropylethylamine (23.0 μ L, 0.14 mmol), copper(II) acetate (25 mg, 0.14 mmol) and phenanthroline (25 mg, 0.14 mmol). The crude material was purified on silica gel (1% MeOH/CH₂Cl₂) to afford **8'** as a yellow oil (31 mg, 66%). R_f = 0.74 (5% MeOH/CH₂Cl₂); This compound has been previously reported^{iv}: **¹H-NMR (300 MHz, CDCl₃)** δ 6.98 (d, J = 8.0 Hz, 2H), 6.53 (dt, J = 9.1, 2.6 Hz, 2H), 5.05 (d, J = 7.9 Hz, 1H), 4.32 (dd, J = 12.4, 6.9 Hz, 1H), 3.73 (s, 3H), 3.09 (t, J = 6.9 Hz, 2H), 2.23 (s, 3H), 1.90 – 1.75 (m, 1H), 1.71 – 1.55 (m, 3H), 1.53 – 1.33 (m, 11H).

ⁱⁱⁱ Le Roch, A.; Chan, H-C.; Gagnon, A. *Eur. J. Org. Chem.* **2020**, 36, 5815–5819

^{iv} Sun, H-B.; Gong, L.; Tian, Y-B.; Wu, J-G.; Zhang, X.; Liu, J.; Fu, Z.; Niu, D. *Angew. Chem. Int. Ed.* **2018**, 57, 1–6

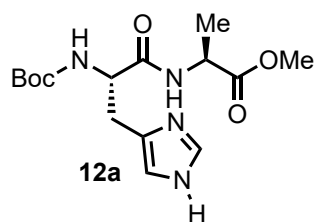
6. Procedures for the preparation of dipeptides

a. Procedure for synthesis of Boc-His-A.A.-OMe (12a–f and 12j–l)



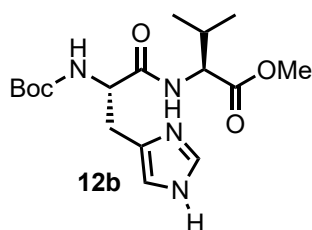
Manuscript, Scheme 3a): In a 50 mL flask, DIPEA (280 μL , 1.64 mmol, 2.1 equiv) was added to L-Boc-His-OH (200 mg, 783 μmol , 1.0 equiv), L-H-A.A.-OMe $\cdot\text{HCl}$ (783 μmol , 1.0 equiv) and HCTU (681 mg, 1.65 mmol, 2.1 equiv) in DMF (4 mL, 0.2 M). The mixture was stirred for 4h at room temperature before being extracted twice with dichloromethane. The combined organic phases were successively washed with sat. aq. NaHCO_3 (2x20 mL), brine (2x20 mL), dried over Na_2SO_4 , filtered and concentrated under reduce pressure. The resulting crude product was purified by column chromatography using the indicated eluent system to afford the desired pure product.

Methyl (*tert*-butoxycarbonyl)-L-histidyl-L-alaninate (12a)



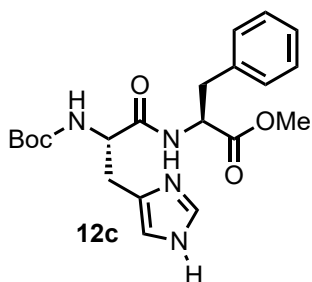
The general procedure was followed starting from L-H-Ala-OMe $\cdot\text{HCl}$ (109 mg, 783 μmol). The crude material was purified on silica gel (10% MeOH/ CH_2Cl_2) to afford **12a** as a white solid (109 mg, 41%). R_f = 0.28 (10% MeOH/DCM); **m.p.** = 94 – 96 $^\circ\text{C}$; **$^1\text{H-NMR}$ (600**

MHz, CDCl_3) δ 7.74 (s, 1H), 7.60 – 7.32 (m, 2H), 6.79 (s, 1H), 5.99 – 5.86 (d, J = 5.5 Hz, 1H), 4.51 – 4.35 (m, 2H), 3.68 (s, 3H), 3.09 – 2.98 (m, 2H), 1.38 (s, 9H), 1.31 (d, J = 7.2 Hz, 3H); **$^{13}\text{C-NMR}$ (150 MHz, CDCl_3)** δ 173.5, 171.8, 155.8, 135.2, 131.6, 119.5, 80.2, 54.2, 52.6, 48.3, 29.7, 28.4, 17.7; **IR (ATR)/ $\bar{\nu}$ (cm^{-1})**: 3288, 3195, 2959, 2919, 1753, 1705, 1683, 1662, 1540, 1494, 1361, 1254, 1196, 1154, 1019, 853, 761, 617; **HRMS (ESI)** calcd. for $[\text{C}_{15}\text{H}_{24}\text{N}_4\text{O}_5 + \text{H}]^+$: 341.1820, found 341.1817.

Methyl (*tert*-butoxycarbonyl)-*L*-histidyl-*L*-valinate (12b)

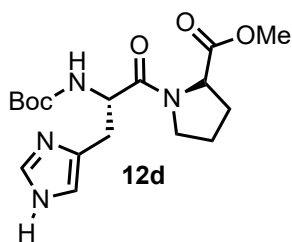
The general procedure was followed starting from *L*-H-Val-OMe•HCl (131 mg, 783 μ mol). The crude material was purified on silica gel (10% MeOH/CH₂Cl₂) to afford **12b** as a white solid (225 mg, 78%). *R_f* = 0.32 (10% MeOH/DCM); *m.p.* = 82 – 84 °C; ¹H-NMR (300

MHz, CDCl₃) δ 8.44 (s, 1H), 7.60 (s, 1H), 7.43 (d, *J* = 6.6 Hz, 1H), 6.85 (s, 1H), 5.94 (d, *J* = 5.9 Hz, 1H), 4.54 – 4.30 (m, 2H), 3.70 (s, 3H), 3.13 (A of ABX, *J* = 15.0, 4.4 Hz, 1H), 3.01 (B of ABX, *J* = 14.9, 6.3 Hz, 1H), 2.20 – 2.02 (m, 1H), 1.43 (s, 9H), 0.82 (dd, *J* = 6.8, 2.6 Hz, 6H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.0, 155.8, 135.0, 132.7, 117.8, 80.0, 57.5, 54.5, 52.0, 30.8, 28.2, 18.8, 17.6; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3306, 2963, 1738, 1659, 1515, 1436, 1366, 1250, 1210, 1162, 1019, 843, 760; HRMS (ESI) calcd. for [C₁₇H₂₈N₄O₅ + H]⁺: 369.2133, found 369.2131.

Methyl (*tert*-butoxycarbonyl)-*L*-histidyl-*L*-phenylalaninate (12c)

The general procedure was followed starting from *L*-H-Phe-OMe•HCl (169 mg, 783 μ mol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **12c** as a white solid (322 mg, 99%). *R_f* = 0.62 (10% MeOH/DCM); *m.p.* = 85 – 87 °C; ¹H-NMR (300

MHz, CDCl₃) δ 10.35 (s, 1H), 7.53 (s, 2H), 7.24 – 7.11 (m, 3H), 6.98 (d, *J* = 6.4 Hz, 2H), 6.76 (s, 1H), 6.03 (d, *J* = 7.2 Hz, 1H), 4.72 (dd, *J* = 12.9, 6.3 Hz, 1H), 4.38 (s(br), 1H), 3.60 (s, 3H), 3.09 – 2.88 (m, 4H), 1.36 (s, 9H); ¹³C-NMR (75 MHz, CDCl₃) δ 171.8, 155.7, 135.9, 135.1, 132.5, 129.1, 128.5, 127.0, 118.1, 80.1, 54.4, 53.5, 52.3, 37.7, 29.2, 28.2; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3280, 2978, 1739, 1658, 1496, 1455, 1435, 1366, 1248, 1216, 1163, 1020, 843, 746, 700; HRMS (ESI) calcd. for [C₂₁H₂₈N₄O₅ + H]⁺: 417.2133, found 417.2146.

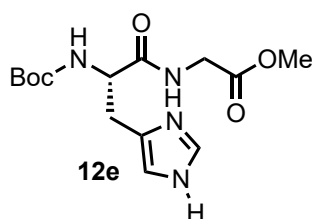
Methyl (*tert*-butoxycarbonyl)-*L*-histidyl-*L*-prolinate (12d)

The general procedure was followed starting from *L*-H-Pro-OMe•HCl (101 mg, 783 μ mol). The crude material was purified on silica gel (4% MeOH/CH₂Cl₂) to afford **12d** as a white solid (163 mg, 57%). *R_f* = 0.14 (5% MeOH/DCM); *m.p.* = 59 – 62 °C; ¹H-NMR (300

MHz, CDCl₃) δ 8.53 (s(br), 1H), 7.68 (s, 1H), 6.94 (s, 1H), 5.43 (d, *J* = 7.8 Hz, 1H), 4.66 – 4.56 (m, 1H), 4.55 – 4.47 (m, 1H), 3.79 (s, 3H), 3.67 – 3.56 (m, 1H), 3.25 – 3.15 (m, 1H), 3.08 (d, *J* = 5.6 Hz, 2H), 2.32 – 2.17 (m, 1H), 2.02 – 1.88 (m, 3H), 1.41 (s, 9H); ¹³C-

NMR (150 MHz, CDCl₃) δ 173.4, 170.6, 155.3, 135.3, 128.1, 122.6, 79.9, 59.1, 52.7, 52.0, 47.1, 38.6, 29.0, 28.3, 25.0; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3275, 2977, 1739, 1704, 1635, 1506, 1436, 1365, 1248, 1161, 1046, 1019, 841, 751; **HRMS (ESI)** calcd. for [C₁₇H₂₆N₄O₅ + H]⁺: 367.1976, found 367.1983.

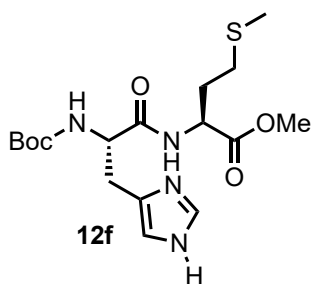
Methyl (tert-butoxycarbonyl)-L-histidylglycinate (12e)



The general procedure was followed starting from L-H-Gly-OMe•HCl (99 mg, 0.78 mmol). The crude material was purified on silica gel (5% MeOH/CH₂Cl₂) to afford **12e** as a white solid (54 mg, 21%). **R_f** = 0.38 (10% MeOH/DCM); **m.p.** = 86 – 88 °C; **¹H-NMR (600**

MHz, CDCl₃) δ 9.31 (s(br), 1H), 7.88 (s(br), 1H), 7.51 (s, 1H), 6.80 (s, 1H), 5.91 (s, 1H), 4.43 (s, 1H), 4.03 – 3.90 (m, 2H), 3.69 (s, 3H), 3.16 – 3.06 (m, 1H), 3.06 – 2.99 (m, 1H), 1.38 (s, 9H); **presence of conformers** **¹³C-NMR (151 MHz, CDCl₃)** δ 172.6, 170.6, 155.8, 135.4, 131.4, 119.8, 80.3, 54.2, 53.3, 52.4, 41.3, 38.7, 29.4, 28.4, 18.3; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3276, 2920, 2850, 1747, 1674, 1520, 1436, 1366, 1248, 1211, 1162, 1022; **HRMS (ESI)** calcd. for [C₁₄H₂₂N₄O₅ + H]⁺: 327.1663, found 327.1680.

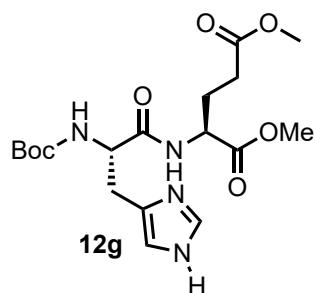
Methyl (tert-butoxycarbonyl)-L-histidyl-L-methioninate (12f)



The general procedure was followed starting from L-H-Met-OMe•HCl (156 mg, 783 μ mol). The crude material was purified on silica gel (10% MeOH/CH₂Cl₂) to afford **12f** as a white solid (228 mg, 73%). **R_f** = 0.23 (10% MeOH/DCM); **m.p.** = 62 – 64 °C; **¹H-NMR (300**

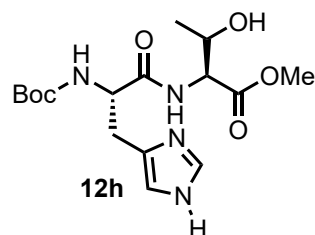
MHz, CDCl₃) δ 8.06 (s, 1H), 7.69 (d, *J* = 6.6 Hz, 1H), 7.51 (s, 1H), 6.75 (s, 1H), 6.06 (d, *J* = 7.5 Hz, 1H), 4.59 – 4.48 (m, 1H), 4.40 – 4.28 (m, 1H), 3.60 (s, 3H), 2.97 (d, *J* = 5.1 Hz, 2H), 2.30 (t, *J* = 7.6 Hz, 2H), 2.07 – 1.95 (m, 1H), 1.93 (s, 3H), 1.91 – 1.76 (m, 1H), 1.31 (s, 9H); **presence of conformers** **¹³C-NMR (150 MHz, CDCl₃)** δ 172.3, 172.0, 155.7, 135.0, 131.9, 118.5, 80.2, 54.2, 52.6, 51.6, 31.1, 29.8, 29.1, 28.3, 15.2; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3288, 2921, 2852, 1739, 1661, 1506, 1436, 1366, 1273, 1348, 1162, 1019, 841, 778; **HRMS (ESI)** calcd. for [C₁₇H₂₈N₄O₅S + H]⁺: 401.1853, found 401.1869.

Dimethyl *N*^α-(*tert*-butoxycarbonyl)-*L*-histidyl-*L*-glutamate (12g)



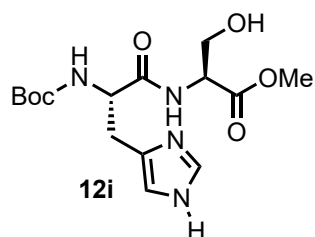
The procedure is described below, see section **6c** for complete characterization of **12g**.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*L*-histidyl-*L*-allothreoninate (12h)



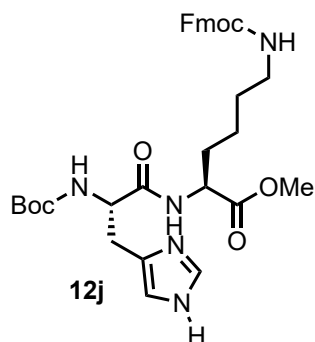
The procedure is described below, see section **6c** for complete characterization of **12h**.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*L*-histidyl-*L*-serinate (12i)



The procedure is described below, see section **6c** for complete characterization of **12i**.

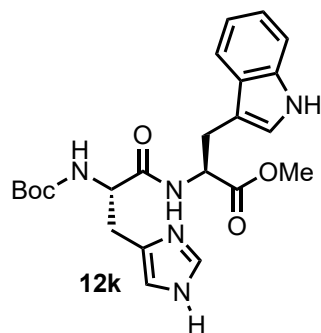
Methyl *N*⁶-(((9*H*-fluoren-9-yl)methoxy)carbonyl)-*N*²-(*tert*-butoxycarbonyl)-*L*-histidyl)-*L*-lysinate (12j)



The general procedure was followed starting from *L*-H-Lys(Fmoc)-OMe•HCl (328 mg, 783 μmol). The crude material was purified on silica gel (3% MeOH/CH₂Cl₂) to afford **12j** as a white solid (454 mg, 94%). *R*_f = 0.46 (10% MeOH/DCM); *m.p.* = 76 – 78 °C; ¹H-NMR (600 MHz, CDCl₃) δ 10.35 (s(br), 1H), 7.71 (d, *J* = 7.7 Hz, 3H), 7.55 (d, *J* = 7.4 Hz, 2H), 7.35 (t, *J* = 7.5 Hz, 2H), 7.29 – 7.22 (m, 3H), 6.81 (s, 1H), 6.41 (s(br), 1H), 5.53 (t, *J* = 5.8 Hz, 1H), 4.46 (s(br), 2H), 4.35 (d, *J* = 7.0 Hz, 2H), 4.16 (t, *J* = 7.0 Hz, 1H), 3.64 (s, 3H), 3.19 – 3.11 (m, 1H), 3.08 – 2.95 (m, 3H), 1.78 – 1.67 (m, 1H), 1.60 – 1.51 (m, 1H), 1.40 (s, 9H), 1.37 – 1.29 (m, 3H), 1.03 – 0.89 (m, 1H); ¹³C-NMR (150 MHz, CDCl₃) δ 172.5, 171.5, 165.8, 157.1, 155.7, 143.9, 143.9, 141.3, 134.9, 127.7, 127.1, 125.1, 120.0, 80.1, 66.7, 54.5, 52.4, 52.2, 47.2, 38.6, 31.5, 29.7, 29.1, 28.3, 21.9; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3408, 3288,

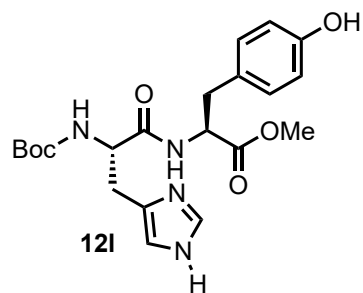
2936, 1694, 1515, 1449, 1366, 1248, 1161, 1021, 841, 760, 740; **HRMS (ESI)** calcd. for $[C_{33}H_{41}N_5O_7 + H]^+$: 620.3079, found 620.3097.

Methyl (*tert*-butoxycarbonyl)-*L*-histidyl-*L*-tryptophanate (**12k**)



The general procedure was followed starting from *L*-H-Trp-OMe•HCl (200 mg, 783 μ mol). The crude material was purified on silica gel (4% MeOH/CH₂Cl₂) to afford **12k** as a white solid (249 mg, 70%). *R_f* = 0.33 (10% MeOH/DCM); *m.p.* = 97 – 99 °C; **¹H-NMR (600 MHz, CDCl₃)** δ 10.20 (s, 1H), 9.40 (s, 1H), 7.44 (s, 1H), 7.41 (d, *J* = 7.1 Hz, 1H), 7.33 (s, 1H), 7.28 (d, *J* = 8.0 Hz, 1H), 7.10 (t, *J* = 7.5 Hz, 1H), 7.03 (t, *J* = 7.4 Hz, 1H), 6.78 (s, 1H), 6.67 (s, 1H), 6.20 (d, *J* = 5.6 Hz, 1H), 4.79 (s, 1H), 4.45 (s, 1H), 3.55 (s, 3H), 3.20 (s, 2H), 2.99 (s, 2H), 1.36 (s, 9H); **¹³C-NMR (150 MHz, CDCl₃)** δ 172.3, 172.0, 155.8, 136.3, 135.1, 132.4, 127.3, 123.5, 121.8, 119.2, 118.1, 117.9, 111.6, 108.8, 80.3, 54.7, 53.1, 52.4, 29.2, 28.2, 27.4; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3289, 2972, 1660, 1506, 1456, 1435, 1366, 1245, 1161, 1010, 845, 741; **HRMS (ESI)** calcd. for $[C_{23}H_{29}N_5O_5 + H]^+$: 456.2242, found 456.2243.

Methyl (*tert*-butoxycarbonyl)-*L*-histidyl-*L*-tyrosinate (**12l**)

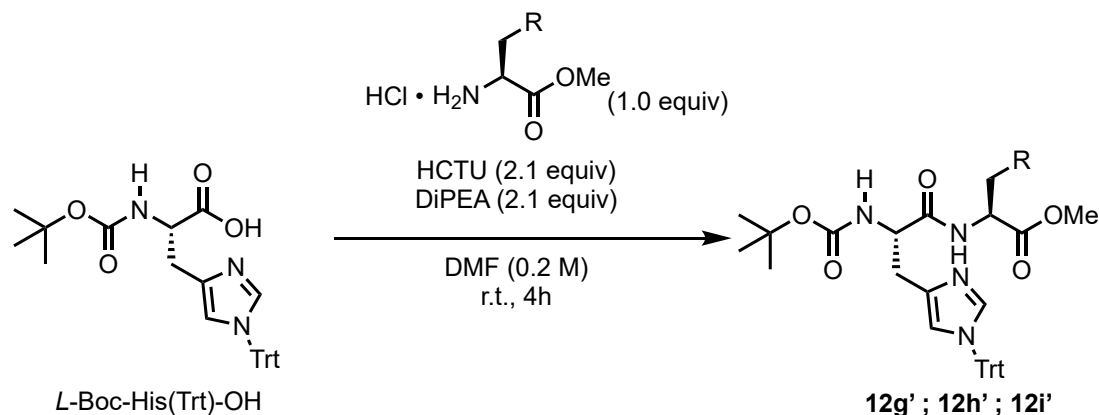


The general procedure was followed starting from *L*-H-Tyr-OMe•HCl (182 mg, 783 μ mol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **12l** as a white solid (276 mg, 81%). This compound has been previously reported.ⁱⁱ

R_f = 0.24 (10% MeOH/DCM); *m.p.* = 93 – 95 °C; **¹H-NMR (300 MHz, CDCl₃)** δ 8.25 (s(br), 1H), 7.47 (s, 1H), 7.24 – 7.12 (m, 1H), 6.77 (d, *J* = 8.4 Hz, 2H), 6.73 – 6.57 (m, 3H), 5.96 (s(br), 1H), 4.81 – 4.63 (m, 1H), 4.38 (s(br), 1H), 3.67 (s, 3H), 3.07 – 2.82 (m, 4H), 1.38 (s, 9H); **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3284, 2972, 1661, 1515, 1437, 1366, 1248, 1162, 1020, 828, 753; **HRMS (ESI)** calcd. for $[C_{21}H_{28}N_4O_6 + H]^+$: 433.2082, found 433.2094.

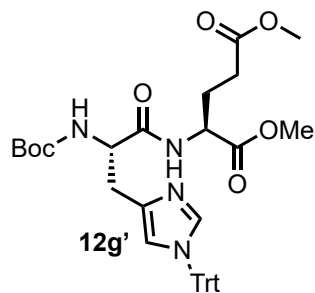
ⁱⁱ Le Roch, A.; Hébert, M.; Gagnon, A. *Eur. J. Org. Chem.* **2020**, 33, 5363–5367

b. Procedure for synthesis of intermediate Boc-His(Trt)-Glu(OMe)-OMe (12g') ; Boc-His(Trt)-Thr-OMe (12h') and Boc-His(Trt)-Ser-OMe (12i')



In a 50 mL flask, DIPEA (215 μL , 1.27 mmol, 2.1 equiv) was added to *L*-Boc-His(Trt)-OH (300 mg, 603 μmol , 1.0 equiv), *L*-H-A.A.-OMe $\cdot\text{HCl}$ (603 μmol , 1.0 equiv) and HCTU (524 mg, 1.27 mmol, 2.1 equiv) in DMF (3 mL, 0.2 M). The mixture was stirred for 4h at room temperature before being extracted twice with dichloromethane. The combined organic phases were successively washed with sat. aq. NaHCO_3 (2x20 mL), brine (2x20 mL), dried over Na_2SO_4 , filtered and concentrated under reduce pressure. The resulting crude product was purified by column chromatography using the indicated eluent system to afford the desired pure product (**12g'** ; **12h'** and **12i'**).

Dimethyl *N* $^\alpha$ -(*tert*-butoxycarbonyl)-*N* $^\epsilon$ -trityl-*L*-histidyl-*L*-glutamate (12g'**)**

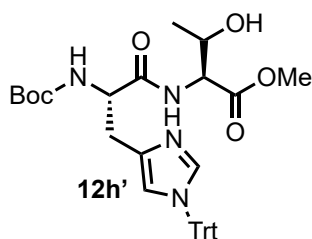


In a 50 mL flask, DIPEA (145 μL , 844 μmol , 2.1 equiv) was added to *L*-Boc-His(Trt)-OH (200 mg, 402 μmol , 1.0 equiv), *L*-Glu(OMe)-OMe $\cdot\text{HCl}$ (85 mg, 0.40 mmol, 1.0 equiv) and HCTU (349 mg, 844 μmol , 2.1 equiv) in DMF (2 mL, 0.2 M). The crude material was purified on silica gel (2% MeOH/ CH_2Cl_2) to afford **12g'** as a white solid (207 mg, 79%). R_f = 0.33 (5% MeOH/ DCM); **m.p.** = 68 – 70 $^\circ\text{C}$;

$^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 7.58 (d, J = 6.6 Hz, 1H), 7.34 (d, J = 1.2 Hz, 1H), 7.31 – 7.22 (m, 9H), 7.10 – 7.01 (m, 6H), 6.62 (s, 1H), 6.39 – 6.26 (m, 1H), 4.57 – 4.44 (m, 1H), 4.44 – 4.33 (m, 1H), 3.60 (s, 3H), 3.52 (s, 3H), 3.01 (A of ABX, J = 14.9, 4.6 Hz, 1H), 2.89 (B of ABX, J = 14.8, 6.1 Hz, 1H), 2.37 – 2.20 (m, 2H), 2.20 – 2.03 (m, 1H), 1.95 – 1.80 (m, 1H), 1.38 (s, 9H); **$^{13}\text{C-NMR}$**

(75 MHz, CDCl₃) δ 172.9, 171.8, 171.7, 155.6, 142.2, 138.4, 136.6, 129.7, 128.0, 128.0, 119.5, 79.6, 75.3, 54.4, 52.3, 51.6, 51.5, 30.0, 29.8, 28.3, 27.3; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3221, 2945, 1737, 1712, 1660, 1493, 1445, 1365, 1242, 1204, 1164, 1131, 1038, 1016, 1001, 871, 841, 748, 701, 659, 638; HRMS (ESI) calcd. for [C₃₇H₄₂N₄O₇ + H]⁺: 655.3126, found 655.3128.

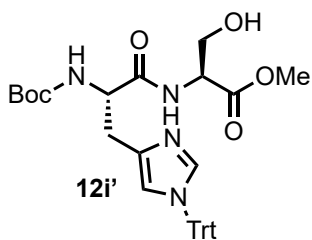
Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-trityl-*L*-histidyl-*L*-allothreoninate (12h')



The general procedure was followed starting from *L*-H-Thr-OMe•HCl (102 mg, 603 μ mol). The crude material was purified on silica gel (5% MeOH/CH₂Cl₂) to afford **12h'** as a white solid (225 mg, 61%). *R*_f = 0.55 (5% MeOH/DCM); *m.p.* = 105 – 107 °C; ¹H-NMR (300 MHz, CDCl₃) δ 7.46 – 7.37 (m, 2H), 7.37 – 7.28 (m, 9H), 7.15 – 7.05

(m, 6H), 6.66 (d, *J* = 1.0 Hz, 1H), 5.83 (d, *J* = 5.5 Hz, 1H), 4.68 – 4.34 (m, 3H), 4.30 – 4.13 (m, 1H), 3.63 (s, 3H), 3.22 – 2.91 (m, 2H), 1.41 (s, 9H), 1.16 (d, *J* = 6.4 Hz, 3H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.3, 171.3, 155.6, 142.2, 138.8, 136.2, 129.8, 128.3, 128.2, 120.2, 80.0, 75.6, 68.4, 58.2, 55.3, 52.4, 30.7, 28.4, 20.2; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3288, 2959, 1744, 1674, 1493, 1445, 1366, 1159, 1019, 1101, 841, 748, 701, 660, 638; HRMS (ESI) calcd. for [C₃₅H₄₀N₄O₆ + H]⁺: 613.3021, found 613.3034.

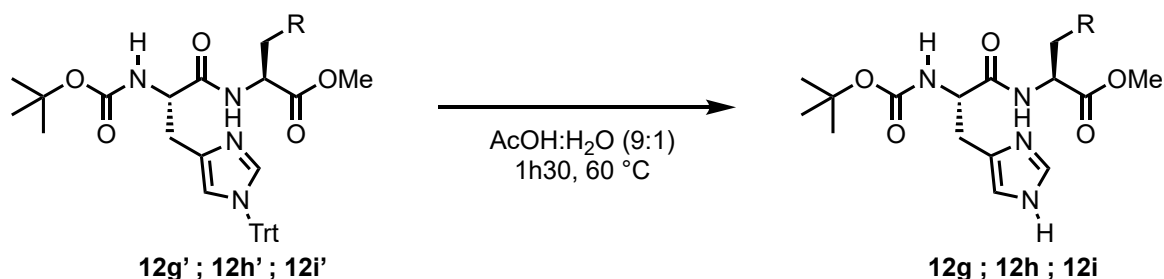
Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-trityl-*L*-histidyl-*L*-serinate (12i')



The general procedure was followed starting from *L*-H-Ser-OMe•HCl (94 mg, 0.60 mmol). The crude material was purified on silica gel (5% MeOH/CH₂Cl₂) to afford **12i'** as a white solid (221 mg, 61%). *R*_f = 0.47 (5% MeOH/DCM); *m.p.* = 93 – 95 °C; ¹H-NMR (300 MHz, CDCl₃) δ 7.44 (s, 1H), 7.37 – 7.29 (m, 10H), 7.15 – 7.05 (m, 6H),

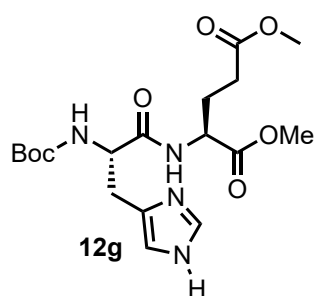
6.66 (s, 1H), 5.28 (s, 1H), 4.70 – 4.30 (m, 3H), 4.17 (A of ABX, *J* = 11.7, 3.4 Hz, 1H), 3.83 (B of ABX, *J* = 11.7, 3.7 Hz, 1H), 3.74 (s, 3H), 3.29 (A' of A'B'X', *J* = 15.0, 3.4 Hz, 1H), 2.92 (B' of A'B'X', *J* = 15.0, 6.3 Hz, 1H), 1.44 (s, 9H); ¹³C-NMR (75 MHz, CDCl₃) δ 171.7, 171.0, 155.4, 142.0, 139.2, 135.4, 129.8, 128.4, 128.3, 121.1, 80.4, 75.9, 61.6, 55.6, 53.6, 52.7, 30.2, 28.4; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3275, 2963, 2928, 1743, 1674, 1493, 1445, 1365, 1238, 1207, 1162, 1131, 1020, 1001, 841, 747, 700, 660, 638; HRMS (ESI) calcd. for [C₃₄H₃₈N₄O₆ + H]⁺: 599.2864, found 599.2875.

c. Procedure for synthesis of Boc-His-Glu(OMe)-OMe (**12g**); Boc-His-Thr-OMe (**12h**) and Boc-His-Ser-OMe (**12i**)

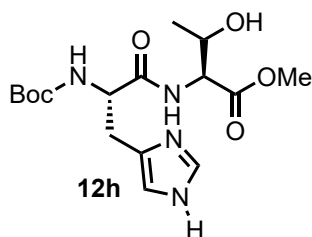


In a flask, *L*-Boc-His(Trt)-A.A-OMe in AcOH:H₂O (9:1, 0.35 M) was stirred for 1h30 at 60 °C. After cooling down at room temperature, the solvent was co-evaporated with cyclohexane under reduce pressure. The resulting crude product was purified by column chromatography using the indicated eluent system to afford the desired pure product (**12g ; 12h and 12i**).

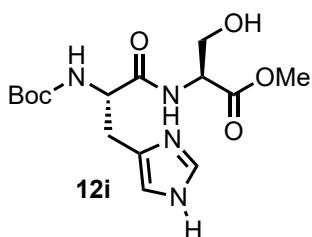
Dimethyl *N*^α-(*tert*-butoxycarbonyl)-*L*-histidyl-*L*-glutamate (12g**)**



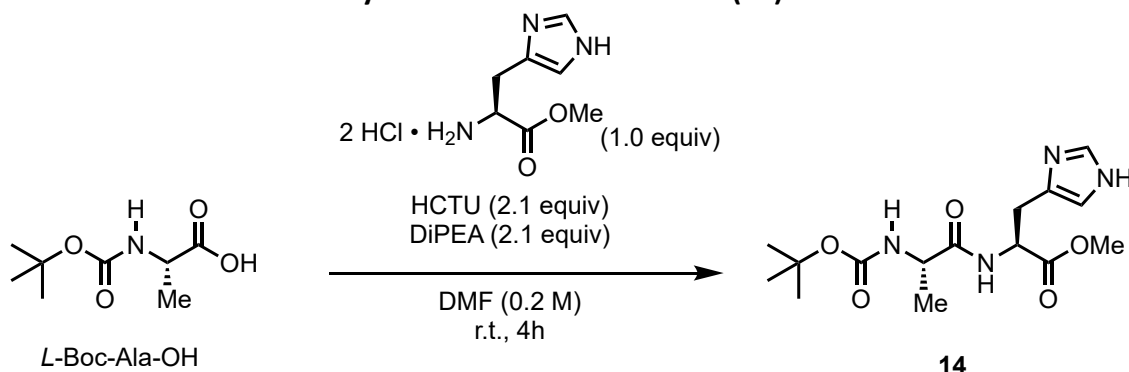
The general procedure was followed starting from *L*-Boc-His-Glu(OMe)-OMe **12g'** (130 mg, 200 μmol). The crude material was purified on silica gel (7% MeOH/CH₂Cl₂) to afford **12g** as a white solid (64 mg, 78%). *R*_f = 0.28 (10% MeOH/DCM); *m.p.* = 47 – 49 °C; ¹H-NMR (600 MHz, CDCl₃) δ 10.50 (s, 1H), 7.67 (s(br), 1H), 7.63 (s, 1H), 6.82 (s, 1H), 6.00 (d, *J* = 5.6 Hz, 1H), 4.53 – 4.46 (m, 1H), 4.40 (s(br), 1H), 3.66 (s, 3H), 3.60 (s, 3H), 3.04 (s, 2H), 2.30 – 2.19 (m, 2H), 2.16 – 2.08 (m, 1H), 1.95 – 1.85 (m, 1H), 1.37 (s, 9H); ¹³C-NMR (150 MHz, CDCl₃) δ 173.2, 172.1, 172.0, 155.7, 135.1, 132.0, 118.5, 80.2, 54.4, 52.6, 51.8, 51.8, 29.8, 29.2, 28.3, 26.9; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3300, 2955, 1733, 1666, 1515, 1436, 1366, 1249, 1208, 1162, 1018, 985, 844, 755, 616; HRMS (ESI) calcd. for [C₁₈H₂₈N₄O₇ + H]⁺: 413.2031, found 413.2048.

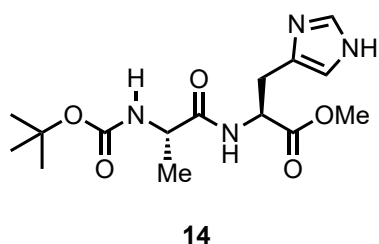
Methyl *N*^α-(*tert*-butoxycarbonyl)-*L*-histidyl-*L*-allothreoninate (12h**)**

The general procedure was followed starting from *L*-Boc-His-Thr-OMe **12h'** (183 mg, 300 μmol). The crude material was purified on silica gel (7% MeOH/CH₂Cl₂) to afford **12h** as a white solid (89 mg, 81%). *R*_f = 0.19 (5% MeOH/DCM); *m.p.* = 90 – 92 °C; ¹H-NMR (300 MHz, CDCl₃) δ 8.65 (s(br), 2H), 7.68 – 7.48 (m, 2H), 6.84 (s, 1H), 5.93 (d, *J* = 7.2 Hz, 1H), 4.56 – 4.37 (m, 2H), 4.32 – 4.19 (m, 1H), 3.63 (s, 3H), 3.04 (s(br), 2H), 1.36 (s, 9H), 1.14 (d, *J* = 6.1 Hz, 3H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.6, 171.5, 155.7, 135.3, 131.6, 118.9, 80.2, 67.8, 58.4, 54.4, 52.5, 29.6, 28.3, 20.0; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3270, 2979, 1661, 1515, 1435, 1366, 1250, 1161, 1084, 1018, 846, 743; HRMS (ESI) calcd. for [C₁₆H₂₆N₄O₆ + H]⁺: 371.1925, found 371.1927.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*L*-histidyl-*L*-serinate (12i**)**

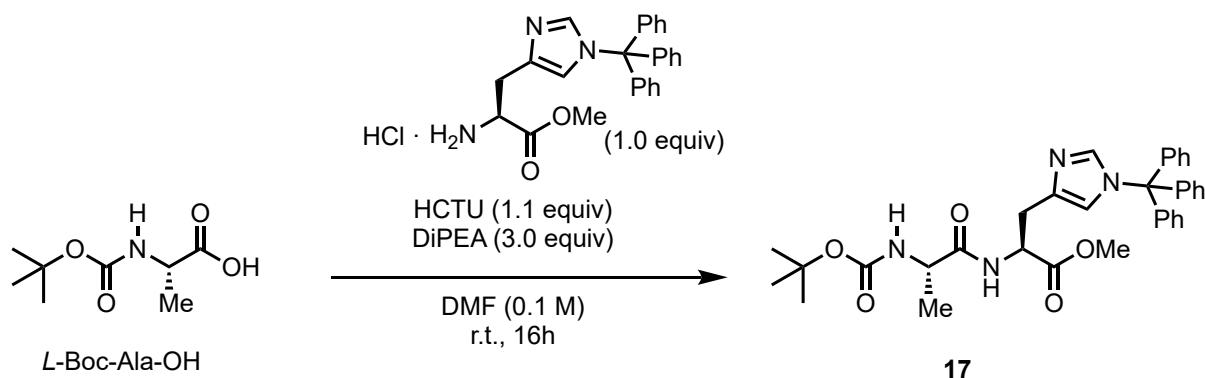
The general procedure was followed starting from *L*-Boc-His-Ser-OMe **12i'** (195 mg, 326 μmol). The crude material was purified on silica gel (7% MeOH/CH₂Cl₂) to afford **12i** as a white solid (86 mg, 74%). *m.p.* = 90 – 92 °C; ¹H-NMR (300 MHz, CDCl₃) δ 8.16 (s(br), 2H), 7.72 (s(br), 1H), 7.57 (s, 1H), 6.83 (s, 1H), 5.88 – 5.75 (m, 1H), 4.57 (s(br), 1H), 4.47 – 4.30 (m, 1H), 3.90 (s(br), 2H), 3.67 (s, 3H), 3.12 – 2.95 (m, 2H), 1.36 (s, 9H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.3, 171.1, 155.7, 135.5, 131.9, 118.5, 80.4, 62.0, 55.2, 54.4, 52.7, 29.7, 28.3; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3279, 2979, 1740, 1662, 1509, 1366, 1245, 1161, 1051, 1020, 847, 758; HRMS (ESI) calcd. for [C₁₅H₂₄N₄O₆ + H]⁺: 357.1769, found 357.1769.

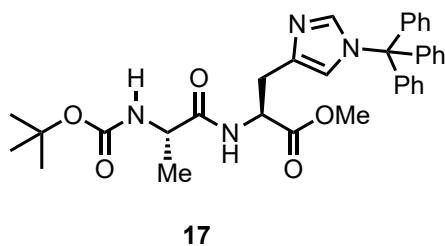
d. Procedure for synthesis Boc-Ala-His-OMe (14**)**

Methyl (tert-butoxycarbonyl)-L-alanyl-L-histidinate (14)

Manuscript, Scheme 3b) : In a 50 mL flask, DIPEA (190 μ L, 1.11 mmol, 2.1 equiv) was added to *L*-Boc-Ala-OH (100 mg, 529 μ mol, 1.0 equiv), *L*-H-His-OMe $\cdot$ 2HCl (128 mg, 529 μ mol, 1.0 equiv) and HCTU (459 mg, 1.11 mmol, 2.1 equiv) in DMF (2.6 mL, 0.2 M). The mixture was stirred for 4h at room

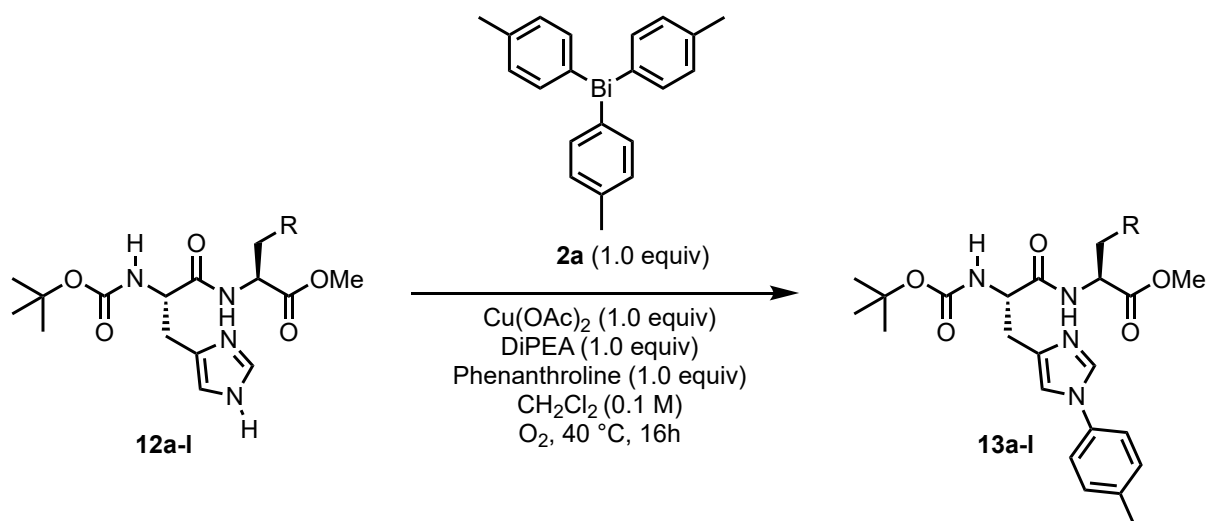
temperature before being extracted twice with dichloromethane. The combined organic phases were successively washed with sat. aq. NaHCO₃ (2x20 mL), brine (2x20 mL), dried over Na₂SO₄, filtered and concentrated under reduce pressure. The resulting crude product was purified on silica gel (5% MeOH/CH₂Cl₂) to afford **14** as a white solid (97mg, 54%). **R_f** = 0.26 (10% MeOH/DCM); **m.p.** = 145 – 147 °C; **¹H-NMR (300 MHz, CDCl₃)** δ 8.44 (s(br), 1H), 7.59 (d, *J* = 6.7 Hz, 1H), 7.50 (s, 1H), 6.74 (s, 1H), 5.59 (d, *J* = 6.4 Hz, 1H), 4.80 – 4.67 (m, 1H), 4.22 – 4.02 (m, 1H), 3.64 (s, 3H), 3.17 – 3.03 (m, 2H), 1.38 (s, 9H), 1.30 (d, *J* = 7.0 Hz, 3H); **¹³C-NMR (75 MHz, CDCl₃)** δ 173.1, 171.5, 155.9, 135.4, 129.0, 119.3, 80.2, 52.8, 52.5, 50.5, 29.7, 28.3, 18.2; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3381, 3248, 2972, 1728, 1694, 1645, 1553, 1505, 1435, 1337, 1291, 1231, 1162, 1067, 1005, 947, 758, 620; **HRMS (ESI)** calcd. for [C₁₅H₂₄N₄O₅ + H]⁺: 341.1820, found 341.1818.

e. Procedure for synthesis Boc-Ala-His(trt)-OMe (17)

Methyl *N*^α-((*tert*-butoxycarbonyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidinate (17**)**

Manuscript, Scheme 4: In a 50 mL flask, DIPEA (570 μ L, 3.35 mmol, 3.0 equiv) was added to *L*-Boc-Ala-OH (211 mg, 1.12 mmol, 1.0 equiv), *L*-H-His(trt)-OMe•HCl (500 mg, 1.12 mmol, 1.0 equiv) and HCTU (508 mg, 1.23 mmol, 1.1 equiv) in DMF (11 mL, 0.1 M). The mixture was stirred

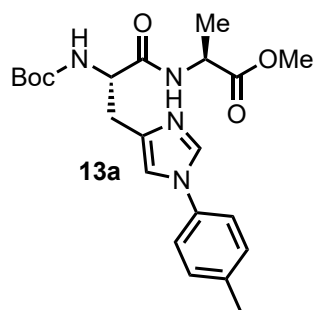
for 16h at room temperature before being extracted twice with ethyl acetate. The combined organic phases were successively washed with sat. aq. NaHCO₃ (2x30 mL), brine (2x30 mL), dried over Na₂SO₄, filtered and concentrated under reduce pressure. The resulting crude product was purified on silica gel (30% EtOAc/Hexane) to afford **17** as a white solid (606 mg, 93%). *R*_f = 0.42 (5% MeOH/DCM); *m.p.* = 74 – 76 °C; ¹H-NMR (300 MHz, CDCl₃) δ 7.60 (d, *J* = 5.2, 1H), 7.37 (d, *J* = 1.1 Hz, 1H), 7.35 – 7.27 (m, 9H), 7.15 – 6.98 (m, 6H), 6.54 (s, 1H), 5.33 (s, 1H), 4.77 (dt, *J* = 8.0, 4.7 Hz, 1H), 4.29 – 4.13 (m, 1H), 3.58 (s, 3H), 3.09 – 2.97 (m, 2H), 1.39 (s, 9H), 1.34 (d, *J* = 7.0 Hz, 3H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.4, 171.3, 155.0, 142.1, 138.5, 136.2, 129.6, 128.0, 119.5, 79.3, 75.2, 52.3, 52.0, 49.9, 29.6, 28.2, 18.9; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3284, 2977, 1744, 1711, 1673, 1492, 1445, 1365, 1243, 1163, 1133, 1033, 1001, 870, 848, 748, 700, 660, 638; HRMS (ESI) calcd. for [C₃₄H₃₈N₄O₅ + H]⁺: 583.2915, found 583.2943.

7. Procedure for the preparation of aryl dipeptides**a. General procedure for synthesis of Boc-His(4-tolyl)-A.A.-OMe (**13a-l**)**

Manuscript, Scheme 3a : In a sealed tube, *N,N*-diisopropylethylamine (20.5 μ L, 0.12 mmol,

1.0 equiv) was added to the *N*-protected *L*-histidine *L*-amio acid methyl ester **12a–l** (0.12 mmol, 1.0 equiv), tri(4-tolyl)bismuthine **2a** (58 mg, 0.12 mmol, 1.0 equiv), copper(II) acetate (22 mg, 0.12 mmol, 1.0 equiv) and phenanthroline (22 mg, 0.12 mmol, 1.0 equiv) in CH₂Cl₂ (0.1 M). Oxygen was bubbled in the reaction mixture before stirred in an oil bath set at 40 °C for 16 hours. The solvent was evaporated under reduce pressure and the residue was purified on silica gel chromatography using the corresponding eluent system to afford desired pure product **13a–l**.

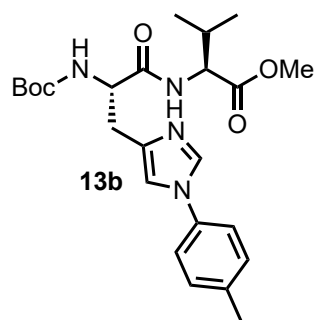
Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(*p*-tolyl)-*L*-histidyl-*L*-alaninate (**13a**)



The general procedure was followed starting from Boc-(*L*)His-(*L*)Ala-OMe **12a** (41 mg, 0.12 mmol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **13a** as a white solid (40 mg, 77%). *R*_f = 0.26 (5% MeOH/CH₂Cl₂) ; *m.p.* = 62 – 64 °C; ¹H-NMR (600 MHz, CDCl₃) δ 7.80 (s, 1H), 7.51 (s(br), 1H), 7.24 – 7.19 (m, 4H), 7.08 (s, 1H), 6.21 (s, 1H), 4.55 – 4.43 (m, 2H), 3.66 (s, 3H),

3.19 – 3.10 (m, 1H), 3.01 (dd, *J* = 14.4, 4.9 Hz, 1H), 2.37 (s, 3H), 1.42 (s, 9H), 1.26 (d, *J* = 7.1 Hz, 3H); ¹³C-NMR (150 MHz, CDCl₃) δ 173.1, 171.3, 155.8, 138.6, 137.7, 134.9, 134.7, 130.5, 121.2, 116.5, 79.9, 54.5, 52.4, 48.1, 29.8, 28.4, 21.0, 18.2; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3310, 2922, 1744, 1683, 1653, 1520, 1453, 1366, 1244, 1207, 1163, 1149, 816; HRMS (ESI) calcd. for [C₂₂H₃₀N₄O₅ + H]⁺: 431.2289, found 431.2308.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(*p*-tolyl)-*L*-histidyl-*L*-valinate (**13b**)

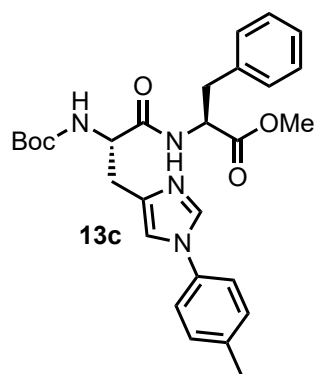


The general procedure was followed starting from Boc-(*L*)His-(*L*)Val-OMe **12b** (40 mg, 0.11 mmol), tri(4-tolyl)bismuthine **2a** (52 mg, 0.11 mmol), *N,N*-diisopropylethylamine (18.5 μL, 0.11 mmol), copper(II) acetate (20 mg, 0.11 mmol) and phenanthroline (20 mg, 0.11 mmol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **13b** as a white solid (45 mg, 91%). *R*_f = 0.38

(5% MeOH/CH₂Cl₂); *m.p.* = 55 – 57 °C; ¹H-NMR (300 MHz, CDCl₃) δ 7.75 (s, 1H), 7.35 (s(br), 1H), 7.25 – 7.14 (m, 4H), 7.06 (s, 1H), 6.47 (d, *J* = 4.2 Hz, 1H), 4.48 (s(br), 1H), 4.45 – 4.35 (m, 1H),

3.64 (s, 3H), 3.26 – 3.11 (m, 1H), 3.06 – 2.91 (m, 1H), 2.36 (s, 3H), 2.10 – 1.94 (m, 1H), 1.44 (s, 9H), 0.73 (d, $J = 6.6$ Hz, 6H); $^{13}\text{C-NMR}$ (75 MHz, CDCl_3) δ 171.0, 170.6, 154.8, 137.9, 136.6, 133.9, 133.7, 129.5, 120.2, 115.6, 79.0, 56.3, 51.1, 30.2, 28.8, 27.4, 20.0, 17.8, 16.6; **IR (ATR)/ $\bar{\nu}$ (cm^{-1}):** 3297, 2963, 2923, 1739, 1667, 1519, 1487, 1435, 1366, 1248, 1205, 1163, 1019, 845, 816, 624; **HRMS (ESI)** calcd. for $[\text{C}_{24}\text{H}_{34}\text{N}_4\text{O}_5 + \text{H}]^+$: 459.2602, found 459.2611.

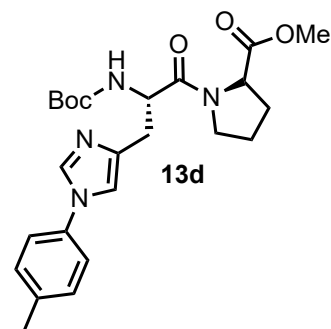
Methyl N^α -(*tert*-butoxycarbonyl)- N^ϵ -(*p*-tolyl)-*L*-histidyl-*L*-phenylalaninate (**13c**)



The general procedure was followed starting from Boc-(*L*)His-(*L*)Phe-OMe **12c** (50 mg, 0.12 mmol). The crude material was purified on silica gel (2% MeOH/ CH_2Cl_2) to afford **13c** as a yellow solid (54 mg, 89%); $R_f = 0.44$ (5% MeOH/ CH_2Cl_2); **m.p.** = 66 – 68 °C; $^1\text{H-NMR}$ (600 MHz, CDCl_3) δ 7.88 (s, 1H), 7.60 (s, 1H), 7.31 (s, 1H), 7.26 – 7.17 (m, 5H), 7.14 (s(br), 1H), 7.06 – 7.00 (m, 2H), 6.28 (s, 1H), 4.82 (s, 1H), 4.66 – 4.37 (m, 2H), 3.65 (s, 3H), 3.18 (s(br), 1H), 3.13 –

3.00 (m, 3H), 2.41 (s, 3H), 1.44 (s, 9H); $^{13}\text{C-NMR}$ (150 MHz, CDCl_3) δ 171.6, 171.4, 155.7, 138.4, 137.8, 136.1, 134.7, 134.6, 130.5, 129.3, 128.5, 126.9, 121.3, 116.5, 79.9, 54.5, 53.5, 52.1, 38.0, 29.7, 28.3, 21.0; **IR (ATR)/ $\bar{\nu}$ (cm^{-1}):** 3288, 2922, 1741, 1682, 1647, 1520, 1495, 1455, 1365, 1247, 1214, 1165, 1047, 1020, 816, 670; **HRMS (ESI)** calcd. for $[\text{C}_{28}\text{H}_{34}\text{N}_4\text{O}_5 + \text{H}]^+$: 507.2602, found 507.2611.

Methyl N^α -(*tert*-butoxycarbonyl)- N^ϵ -(*p*-tolyl)-*L*-histidyl-*L*-prolinate (**13d**)

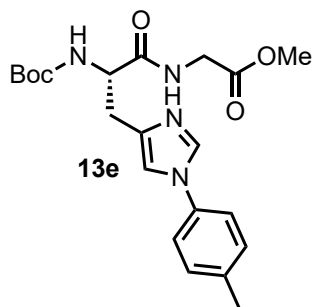


The general procedure was followed starting from Boc-(*L*)His-(*L*)Pro-OMe **12d** (44 mg, 0.12 mmol). The crude material was purified on silica gel (2% MeOH/ CH_2Cl_2) to afford **13d** as a yellow solid (53 mg, 97%). $R_f = 0.37$ (5% MeOH/ CH_2Cl_2); **m.p.** = 68 – 70 °C; **presence of conformers**: $^1\text{H-NMR}$ (600 MHz, CDCl_3) δ 7.85 (s, 1H), 7.31 – 7.26 (m, 2H), 7.24 – 7.18 (m, 3H), 5.60 – 5.44 (m, 1H), 4.80

– 4.67 (m, 1H), 4.48 (d, $J = 4.6$ Hz, 1H), 3.78 – 3.70 (m, 1H), 3.65 (s, 3H), 3.58 – 3.51 (m, 1H), 3.14 – 3.03 (m, 1H), 2.99 – 2.87 (m, 1H), 2.35 (s, 3H), 2.20 – 2.10 (m, 1H), 1.93 (s, 3H), 1.35 (s, 9H); $^{13}\text{C-NMR}$ (150 MHz, CDCl_3) δ 172.6, 170.8, 155.5, 137.4, 134.9, 130.5, 130.4, 126.9, 121.1, 117.0, 79.6, 58.9, 54.3, 52.2, 47.0, 42.5, 29.1, 28.3, 25.0, 21.0; **IR (ATR)/ $\bar{\nu}$ (cm^{-1}):** 3275, 2968,

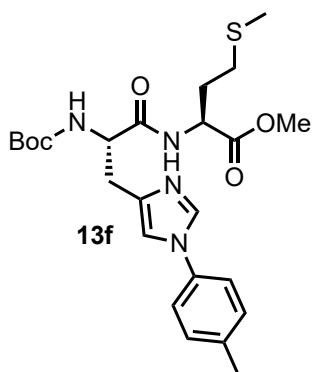
1741, 1704, 1643, 1520, 1488, 1435, 1365, 1163, 1018, 843, 817; **HRMS (ESI)** calcd. for $[C_{24}H_{32}N_4O_5 + H]^+$: 457.2446, found 457.2467.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^τ-(*p*-tolyl)-*L*-histidylglycinate (13e**)**

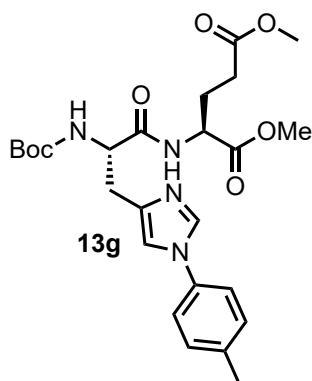


The general procedure was followed starting from Boc-(*L*)His-Gly-OMe **12e** (39 mg, 0.12 mmol). The crude material was purified on silica gel (2% MeOH/ CH_2Cl_2) to afford **13e** as a white solid (27 mg, 54%). R_f = 0.23 (5% MeOH/ CH_2Cl_2); **m.p.** = 52 – 54 °C; **¹H-NMR (600 MHz, $CDCl_3$)** δ 7.79 (s(br), 1H), 7.54 (s(br), 1H), 7.24 (s, 4H), 7.10 (s(br), 1H), 6.27 (s, 1H), 4.52 (s(br), 1H), 4.07 – 3.92 (m, 2H), 3.67 (s, 3H), 3.19 (s(br), 1H), 3.04 (s(br), 1H), 2.38 (s, 3H), 1.44 (s, 9H); **¹³C-NMR (150 MHz, $CDCl_3$)** δ 172.2, 170.2, 155.9, 137.6, 134.9, 130.5, 121.4, 116.5, 80.1, 54.6, 52.3, 41.3, 29.8, 28.5, 21.1; **IR (ATR)/ $\bar{\nu}$ (cm^{-1})**: 3292, 2972, 2923, 1748, 1668, 1519, 1487, 1365, 1246, 1204, 1163, 1020, 815; **HRMS (ESI)** calcd. for $[C_{21}H_{28}N_4O_5 + H]^+$: 417.2133, found 417.2120.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^τ-(*p*-tolyl)-*L*-histidyl-*L*-methioninate (13f**)**

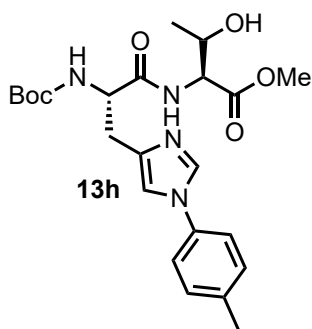


The general procedure was followed starting from Boc-(*L*)His-(*L*)Met-OMe **12f** (48 mg, 0.12 mmol). The crude material was purified on silica gel (2% MeOH/ CH_2Cl_2) to afford **13f** as a white solid (30 mg, 51%). R_f = 0.38 (5% MeOH/ CH_2Cl_2); **m.p.** = 53 – 55 °C; **¹H-NMR (300 MHz, $CDCl_3$)** δ 7.81 (s, 1H), 7.53 (s(br), 1H), 7.24 (s, 4H), 7.09 (s, 1H), 6.36 (s(br), 1H), 4.66 – 4.57 (m, 1H), 4.49 (s(br), 1H), 3.67 (s, 3H), 3.24 – 3.16 (m, 1H), 3.00 (dd, J = 14.4, 5.2 Hz, 1H), 2.38 (s, 3H), 2.34 – 2.23 (m, 2H), 2.11 – 2.01 (m, 1H), 1.96 (s, 3H), 1.93 – 1.83 (m, 1H), 1.45 (s, 9H); **¹³C-NMR (150 MHz, $CDCl_3$)** δ 172.1, 171.6, 155.8, 138.6, 137.8, 134.9, 134.7, 130.5, 121.3, 116.6, 80.1, 54.6, 52.5, 51.6, 31.7, 29.8, 29.7, 28.4, 21.1, 15.4; **IR (ATR)/ $\bar{\nu}$ (cm^{-1})**: 3296, 2920, 1740, 1667, 1520, 1367, 1164, 1023, 847, 817; **HRMS (ESI)** calcd. for $[C_{24}H_{34}N_4O_5S + H]^+$: 491.2323, found 491.2341.

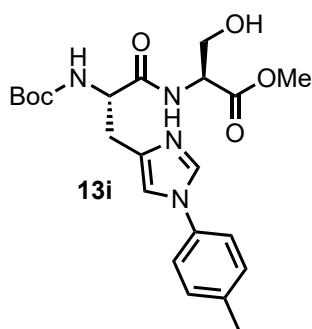
Dimethyl *N*^α-(*tert*-butoxycarbonyl)-*N*^τ-(*p*-tolyl)-*L*-histidyl-*L*-glutamate (13g**)**

The general procedure was followed starting from Boc-(*L*)His-(*L*)Glu(OMe)-OMe **12g** (64 mg, 0.16 mmol), tri(4-tolyl)bismuthine **2a** (75 mg, 0.16 mmol), *N,N*-diisopropylethylamine (26.5 μ L, 0.16 mmol), copper(II) acetate (28 mg, 0.16 mmol) and phenanthroline (28 mg, 0.16 mmol). The crude material was purified on silica gel (2% MeOH/ CH_2Cl_2) to afford **13g** as a white solid (24 mg, 31%). R_f = 0.57 (10% MeOH/ CH_2Cl_2); **m.p.** = 46 – 48 °C; $^1\text{H-NMR}$ (300 MHz, CDCl_3) δ

7.84 (s, 1H), 7.46 (s(br), 1H), 7.24 (s, 4H), 7.10 (s, 1H), 6.33 (s(br), 1H), 4.60 – 4.43 (m, 2H), 3.67 (s, 3H), 3.53 (s, 3H), 3.25 – 3.11 (m, 1H), 3.06 – 2.95 (m, 1H), 2.38 (s, 3H), 2.24 – 2.04 (m, 3H), 1.96 – 1.80 (m, 1H), 1.44 (s, 9H); $^{13}\text{C-NMR}$ (150 MHz, CDCl_3) δ 177.9, 173.1, 172.0, 171.7, 155.8, 138.0, 134.8, 134.6, 130.6, 121.4, 116.8, 80.1, 54.6, 52.5, 51.7, 51.7, 29.8, 29.8, 28.4, 27.4, 21.1; **IR (ATR)/ $\bar{\nu}$ (cm^{-1})**: 3333, 2953, 1708, 1520, 1437, 1367, 1249, 1207, 1166, 1022, 848, 819; **HRMS (ESI)** calcd. for $[\text{C}_{25}\text{H}_{34}\text{N}_4\text{O}_7 + \text{H}]^+$: 503.2500, found 503.2511.

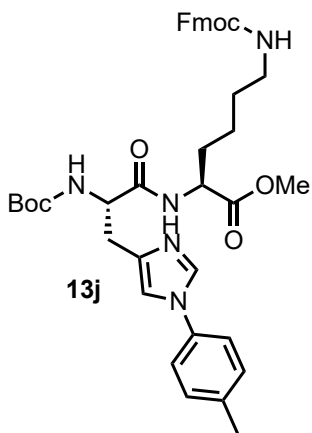
Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^τ-(*p*-tolyl)-*L*-histidyl-*L*-allothreoninate (13h**)**

The general procedure was followed starting from Boc-(*L*)His-(*L*)Thr-OMe **12h** (44 mg, 0.12 mmol). The crude material was purified on silica gel (2% MeOH/ CH_2Cl_2) to afford **13h** as a white solid (17 mg, 31%). R_f = 0.35 (5% MeOH/ CH_2Cl_2); **m.p.** = 82 – 84 °C ; $^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 7.82 (s, 1H), 7.51 – 7.37 (m, 1H), 7.24 (s, 4H), 7.10 (s, 1H), 6.05 (s(br), 1H), 4.64 – 4.46 (m, 2H), 4.28 – 4.15 (m, 1H), 3.69 (s, 3H), 3.21 (A of ABX, J = 14.7, 5.0 Hz, 1H), 3.04 (B of ABX, J = 14.8, 5.9 Hz, 1H), 2.38 (s, 3H), 1.43 (s, 9H), 1.09 (d, J = 6.4 Hz, 3H); $^{13}\text{C-NMR}$ (75 MHz, CDCl_3) δ 172.2, 171.2, 155.7, 137.9, 135.3, 135.2, 134.6, 130.5, 121.3, 117.0, 80.2, 68.5, 58.0, 54.6, 52.5, 30.5, 28.4, 21.1, 19.9; **IR (ATR)/ $\bar{\nu}$ (cm^{-1})**: 3301, 2972, 2919, 1689, 1646, 1520, 1435, 1366, 1252, 1164, 1015, 845, 814, 747; **HRMS (ESI)** calcd. for $[\text{C}_{23}\text{H}_{32}\text{N}_4\text{O}_6 + \text{H}]^+$: 461.2395, found 461.2416.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(*p*-tolyl)-*L*-histidyl-*L*-serinate (13i**)**

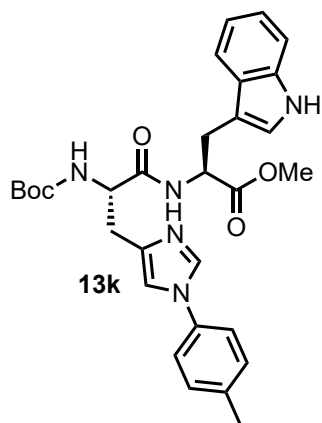
The general procedure was followed starting from Boc-(*L*)His-(*L*)Ser-OMe **12i** (43 mg, 0.12 mmol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **13i** as a white solid (15 mg, 28%). *R*_f = 0.26 (10% MeOH/CH₂Cl₂); *m.p.* = 65 – 67 °C; ¹H-NMR (300 MHz, CDCl₃) δ 7.88 (s, 1H), 7.47 (s, 1H), 7.25 (s, 4H), 7.13 (s, 1H), 5.59 (s, 1H), 4.66 – 4.42 (m, 2H), 4.13 (A of ABX, *J* = 11.7,

3.3 Hz, 1H), 3.85 (B of ABX, *J* = 11.6, 3.8 Hz, 1H), 3.75 (s, 3H), 3.43 – 3.28 (m, 1H), 3.01 (dd, *J* = 14.5, 5.7 Hz, 1H), 2.39 (s, 3H), 1.44 (s, 9H); ¹³C-NMR (150 MHz, CDCl₃) δ 171.6, 170.9, 155.5, 138.1, 137.0, 135.6, 134.5, 130.6, 121.5, 117.6, 80.4, 62.0, 55.6, 54.0, 52.6, 29.8, 28.4, 21.1; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3297, 2932, 1741, 1667, 1520, 1367, 1247, 1164, 1073, 1023, 817; HRMS (ESI) calcd. for [C₂₂H₃₀N₄O₆ + H]⁺: 447.2238, found 447.2246.

Methyl *N*⁶-(((9H-fluoren-9-yl)methoxy)carbonyl)-*N*²-(*N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(*p*-tolyl)-*L*-histidyl)-*L*-lysinate (13j**)**

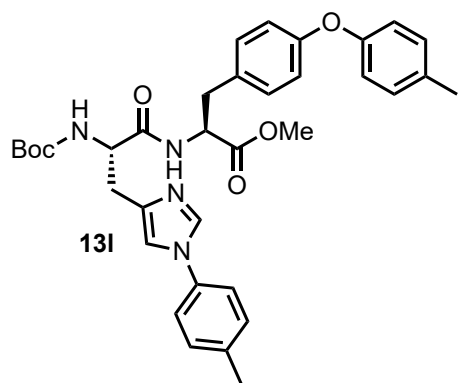
The general procedure was followed starting from Boc-(*L*)His-(*L*)Lys(Fmoc)-OMe **12j** (60 mg, 0.97 mmol), tri(4-tolyl)bismuthine **2a** (47 mg, 0.97 mmol), *N,N*-diisopropylethylamine (16.5 μL, 0.97 mmol), copper(II) acetate (18 mg, 0.97 mmol) and phenanthroline (18 mg, 0.97 mmol). The crude material was purified on silica gel (% MeOH/CH₂Cl₂) to afford **13j** as a white solid (32 mg, 47%). *R*_f = 0.24 (5% MeOH/CH₂Cl₂); ¹H-NMR (300 MHz, CDCl₃) δ 8.35 (s, 1H), 8.25 – 8.13 (m, 1H), 7.75 (d, *J* = 7.4 Hz, 2H), 7.61 (d, *J* = 7.1 Hz, 2H), 7.45 –

7.25 (m, 8H), 7.17 (s, 1H), 6.05 – 5.89 (m, 1H), 5.23 – 5.08 (m, 1H), 4.73 (s, 1H), 4.49 – 4.39 (m, 1H), 4.35 (d, *J* = 7.1 Hz, 2H), 4.21 (t, *J* = 6.8 Hz, 1H), 3.68 (s, 3H), 3.59 – 3.44 (m, 1H), 3.29 – 3.14 (m, 2H), 3.07 – 2.91 (m, 1H), 2.43 (s, 3H), 1.94 – 1.71 (m, 2H), 1.63 – 1.37 (m, 4H), 1.32 (s, 9H); IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3321, 2922, 2852, 1720, 1688, 1650, 1520, 1450, 1256, 1167, 1019, 814, 758, 730; HRMS (ESI) calcd. for [C₄₀H₄₇N₅O₇ + H]⁺: 710.3548, found 710.3569.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*N*^ε-(*p*-tolyl)-*L*-histidyl-*L*-tryptophanate (13k**)**

The general procedure was followed starting from Boc-(*L*)His-(*L*)Trp-OMe **12k** (55 mg, 0.12 mmol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **13k** as a white solid (45 mg, 68%). *R*_f = 0.24 (5% MeOH/CH₂Cl₂); *m.p.* = 96 – 98 °C; ¹H-NMR (300 MHz, CDCl₃) δ 8.18 (s, 1H), 7.61 (s, 1H), 7.45 (d, *J* = 7.8 Hz, 2H), 7.29 (d, *J* = 8.0 Hz, 1H), 7.23 – 7.15 (m, 2H), 7.15 – 7.08 (m, 3H), 7.08 – 7.02 (m, 2H), 6.95 (s, 1H), 6.12 (s(br), 1H), 4.84 (dd, *J* = 12.7, 5.7 Hz, 1H), 4.52 (s(br), 1H), 3.57 (s, 3H), 3.27 (dd, *J* = 14.7,

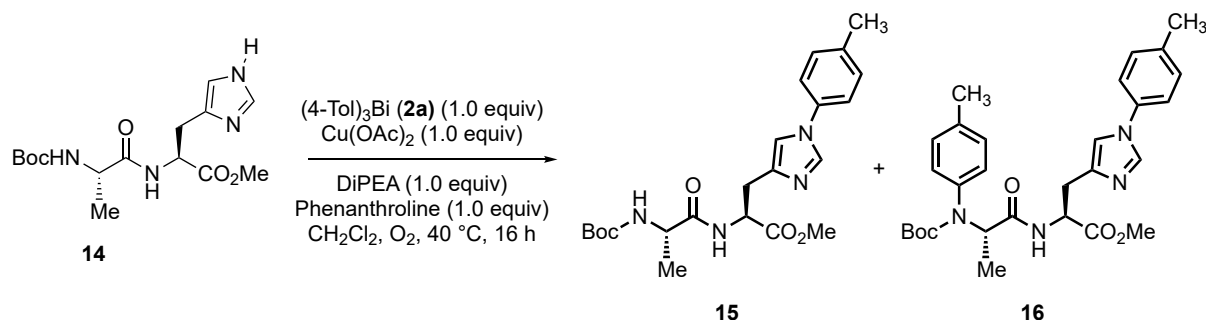
5.5 Hz, 1H), 3.16 (dd, *J* = 14.7, 5.0 Hz, 2H), 3.01 (dd, *J* = 14.5, 5.9 Hz, 1H), 2.37 (s, 3H), 1.39 (s, 9H); ¹³C-NMR (150 MHz, CDCl₃) δ 172.1, 171.5, 155.8, 139.0, 137.7, 136.2, 134.7, 130.5, 127.2, 123.1, 122.2, 121.3, 119.7, 118.8, 116.5, 111.3, 80.0, 54.8, 53.0, 52.3, 30.6, 28.4, 27.9, 21.1; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3299, 2928, 1663, 1519, 1437, 1367, 1249, 1163, 1022, 846, 743; HRMS (ESI) calcd. for [C₃₀H₃₅N₅O₅ + H]⁺: 546.2711, found 546.2709.

Methyl (*S*)-2-((*S*)-2-((*tert*-butoxycarbonyl)amino)-3-(1-(*p*-tolyl)-1*H*-imidazol-4-yl)propanamido)-3-(4-(*p*-tolylloxy)phenyl)propanoate (13l**)**

The general procedure was followed starting from Boc-(*L*)His-(*L*)Tyr-OMe **12l** (52 mg, 0.12 mmol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **13l** as a yellow solid (24 mg, 33%). *R*_f = 0.33 (5% MeOH/CH₂Cl₂); *m.p.* = 60 – 62 °C; ¹H-NMR (600 MHz, CDCl₃) δ 7.64 (s(br), 1H), 7.45 (s(br), 1H), 7.14 – 7.06 (m, 4H), 7.01 (d, *J* = 8.2 Hz, 3H), 6.82 (s(br), 2H), 6.76 (d, *J* =

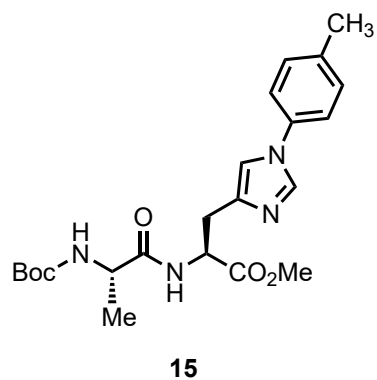
8.4 Hz, 2H), 6.70 (d, *J* = 7.4 Hz, 2H), 6.20 (s(br), 1H), 4.67 (s(br), 1H), 4.39 (s(br), 1H), 3.53 (s, 3H), 3.07 (s(br), 1H), 2.90 (s(br), 3H), 2.27 (s, 3H), 2.22 (s, 3H), 1.33 (s, 9H); ¹³C-NMR (150 MHz, CDCl₃) δ 171.7, 156.9, 155.8, 154.7, 137.7, 134.8, 133.0, 130.5, 130.3, 121.3, 119.2, 119.2, 118.4, 116.4, 80.0, 53.7, 52.3, 38.7, 37.4, 28.4, 21.1, 20.8, 14.2; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3279, 2977, 1741, 1705, 1640, 1500, 1436, 1365, 1238, 1163, 1045, 1017, 843, 750; HRMS (ESI) calcd. for [C₃₅H₄₀N₄O₆ + H]⁺: 613.3021, found 613.3037.

b. Procedure for synthesis of Boc-Ala-His(4-tolyl)-OMe (15) and Boc-N(4-tolyl)-Ala-His(4-tolyl)-OMe (16)



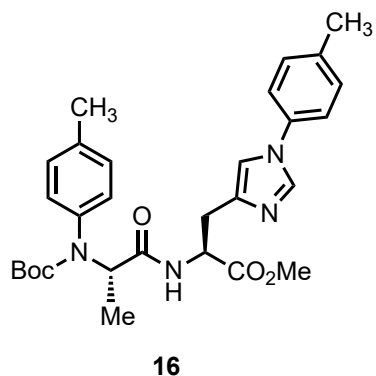
Manuscript, Scheme 3b): In a sealed tube, *N,N*-diisopropylethylamine (25.0 μ L, 0.15 mmol, 1.0 equiv) was added to the methyl (*tert*-butoxycarbonyl)-*L*-alanyl-*L*-histidinate **14** (50 mg, 0.15 mmol, 1.0 equiv), tri(4-tolyl)bismuthine **2a** (71 mg, 0.15 mmol, 1.0 equiv), copper(II) acetate (27 mg, 0.15 mmol, 1.0 equiv) and phenanthroline (27 mg, 0.15 mmol, 1.0 equiv) in CH₂Cl₂ (1.50 mL, 0.1 M). Oxygen was bubbled in the reaction mixture before stirred in an oil bath set at 40 °C for 16 hours. The solvent was evaporated under reduce pressure and the residue was purified on silica gel chromatography using 2% MeOH/CH₂Cl₂ to afford **15** and **16** reported below.

Methyl *N* ^{α} -((*tert*-butoxycarbonyl)-*L*-alanyl)-*N* ^{ϵ} -(*p*-tolyl)-*L*-histidinate (15)

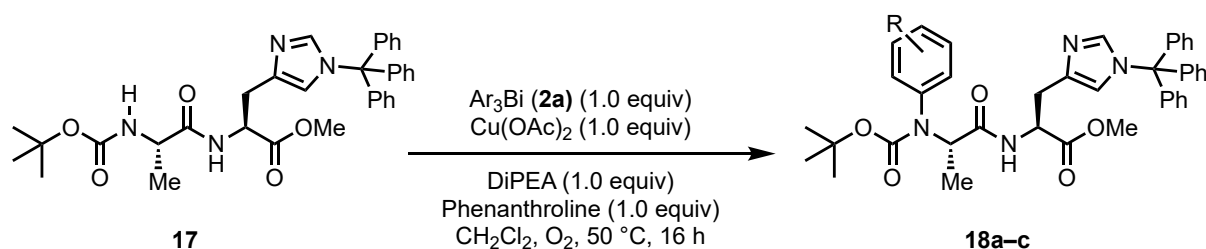


Yellow solid (30 mg, 47%). *R*_f = 0.54 (10% MeOH/CH₂Cl₂); *m.p.* = 54 – 56 °C; ¹H-NMR (300 MHz, CDCl₃) δ 7.74 (s, 1H), 7.52 (s(br), 1H), 7.24 (s, 4H), 7.08 (s, 1H), 5.26 (s, 1H), 4.90 – 4.76 (m, 1H), 4.30 – 4.14 (m, 1H), 3.70 (s, 3H), 3.25 – 3.04 (m, 2H), 2.38 (s, 3H), 1.41 (s, 9H), 1.37 (d, *J* = 7.0 Hz, 3H); **presence of conformers**: ¹³C-NMR (150 MHz, CDCl₃) δ 172.8, 171.7, 155.5, 137.8, 135.2, 134.7, 130.5, 121.4, 116.6, 79.9, 55.3, 52.5, 52.4,

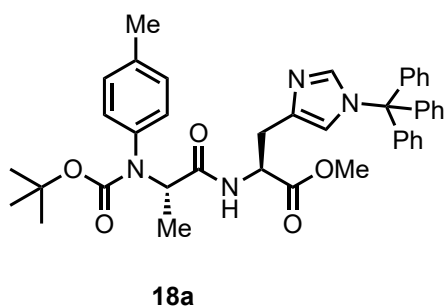
50.3, 43.3, 29.6, 28.4, 21.1, 18.9, 12.5; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3292, 2977, 2923, 1742, 1668, 1519, 1489, 1439, 1365, 1247, 1213, 1163, 1068, 1020, 842, 817, 754; HRMS (ESI) calcd. for [C₂₂H₃₀N₄O₅ + H]⁺: 431.2289, found 431.2290.

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl)-*N*^ε-(*p*-tolyl)-*L*-histidinate (16)

Yellow solid (13 mg, 17%). R_f = 0.62 (10% MeOH/CH₂Cl₂); **m.p.** = 60 – 62 °C; **¹H-NMR (300 MHz, CDCl₃)** δ 7.85 (s(br), 1H), 7.73 (s, 1H), 7.25 – 7.14 (m, 4H), 7.14 – 7.00 (m, 5H), 4.89 – 4.78 (m, 1H), 4.72 – 4.61 (m, 1H), 3.74 (s, 3H), 3.21 – 3.14 (m, 2H), 2.38 (s, 3H), 2.29 (s, 3H), 1.33 (s, 9H), 1.28 (d, J = 7.2 Hz, 3H); **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3346, 2977, 2923, 1743, 1681, 1515, 1367, 1251, 1210, 1162, 1022, 843; **HRMS (ESI)** calcd. for [C₂₉H₃₆N₄O₅ + H]⁺: 521.2759, found 521.2781.

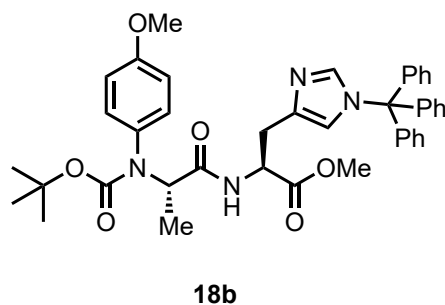
c. General procedure for synthesis of Boc-*N*(Aryl)-Ala-His(trt)-OMe (18a–c)

Manuscript, Scheme 4: In a sealed tube, *N,N*-diisopropylethylamine (17.5 μ L, 0.10 mmol, 1.0 equiv) was added to the methyl *N*^α-((*tert*-butoxycarbonyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidinate **17** (60 mg, 0.10 mmol, 1.0 equiv), triarylbi-muthine (0.10 mmol, 1.0 equiv), copper(II) acetate (19 mg, 0.10 mmol, 1.0 equiv) and phenanthroline (19 mg, 0.10 mmol, 1.0 equiv) in CH₂Cl₂ (1.00 mL, 0.1 M). Oxygen was bubbled in the reaction mixture before stirred in an oil bath set at 50 °C for 16 hours. The solvent was evaporated under reduce pressure and the residue was purified on silica gel chromatography using the corresponding eluent system to afford the desired pure product **18a–c**.

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidinate (18a**)**

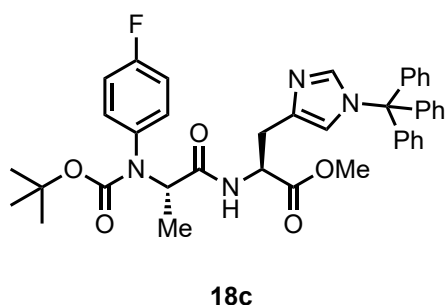
The general procedure was followed starting from tri(4-tolyl)bismuthine **2a** (50 mg, 0.10 mmol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **18a** as a white solid (58 mg, 84%). *R*_f = 0.47 (5% MeOH/CH₂Cl₂); *m.p.* = 64 – 66 °C; ¹H-NMR (300 MHz, CDCl₃) δ 8.07 (s, 1H), 7.38 – 7.28 (m, 10H), 7.17 (d, *J* = 8.1

Hz, 2H), 7.12 – 7.00 (m, 8H), 6.58 (s, 1H), 4.85 – 4.75 (m, 1H), 4.67 – 4.57 (m, 1H), 3.60 (s, 3H), 3.12 (A of ABX, *J* = 14.6, 5.1 Hz, 1H), 3.00 (B of ABX, *J* = 14.6, 4.8 Hz, 1H), 2.29 (s, 3H), 1.36 (s, 9H), 1.30 (d, *J* = 7.1 Hz, 3H); ¹³C-NMR (150 MHz, CDCl₃) δ 172.3, 171.9, 154.9, 142.3, 138.7, 137.9, 136.4, 129.8, 129.3, 128.4, 128.1, 119.6, 80.8, 75.4, 52.9, 52.1, 38.7, 29.8, 28.4, 21.1, 15.8; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3288, 2972, 2928, 1746, 1687, 1514, 1494, 1445, 1366, 1323, 1159, 1132, 748, 701, 660, 637; HRMS (ESI) calcd. for [C₄₁H₄₄N₄O₅ + H]⁺: 673.3385, found 673.3418.

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(4-methoxyphenyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidinate (18b**)**

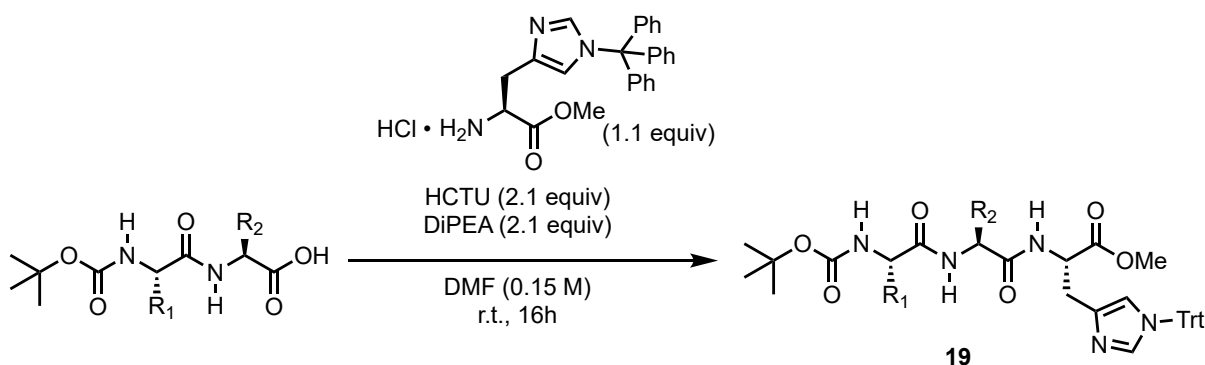
The general procedure was followed starting from tri(4-methoxyphenyl)bismuthine (55 mg, 0.10 mmol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **18b** as a white solid (55 mg, 78%). *R*_f = 0.47 (5% MeOH/CH₂Cl₂); *m.p.* = 61 – 63 °C; ¹H-NMR (600 MHz, CDCl₃) δ 8.01 (s(br), 1H), 7.36 (s, 1H),

7.33 – 7.27 (m, 9H), 7.21 (s(br), 2H), 7.12 – 7.04 (m, 6H), 6.77 (d, *J* = 8.8 Hz, 2H), 6.59 (s, 1H), 4.82 – 4.77 (m, 1H), 4.65 (s(br), 1H), 3.75 (s, 3H), 3.60 (s, 3H), 3.12 (A of ABX, *J* = 14.6, 5.2 Hz, 1H), 3.00 (B of ABX, *J* = 14.6, 4.6 Hz, 1H), 1.35 (s, 9H), 1.29 – 1.23 (m, 3H); ¹³C-NMR (150 MHz, CDCl₃) δ 172.4, 171.9, 158.3, 155.3, 142.4, 138.7, 136.5, 133.3, 130.0, 129.9, 128.2, 119.7, 113.9, 80.7, 75.5, 55.5, 52.9, 52.2, 38.7, 29.8, 28.4, 15.9; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3275, 3048, 2919, 1722, 1667, 1493, 1445, 1238, 1207, 1034, 741, 700, 660; HRMS (ESI) calcd. for [C₄₁H₄₄N₄O₆ + H]⁺: 689.3334, found 689.3347.

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(4-fluorophenyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidinate (18c**)**

The general procedure was followed starting from tri(4-fluorophenyl)bismuthine (51 mg, 0.10 mmol). The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **18c** as a colorless oil (57 mg, 82%). *R*_f = 0.34 (5% MeOH/CH₂Cl₂); ¹H-NMR (600 MHz, CDCl₃) δ 8.07 (s(br), 1H), 7.37 – 7.28 (m, 12H), 7.13 – 7.04 (m, 6H), 6.93 (t, *J* =

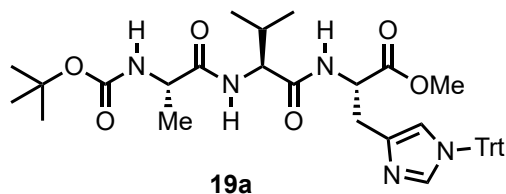
8.6 Hz, 2H), 6.58 (s, 1H), 4.80 (X of ABX, *J* = 12.0, 5.0 Hz, 1H), 4.66 (s(br), 1H), 3.59 (s, 3H), 3.11 (A of ABX, *J* = 14.7, 5.1 Hz, 1H), 2.99 (B of ABX, *J* = 14.6, 4.4 Hz, 1H), 1.35 (s, 9H), 1.28 (d, *J* = 6.1 Hz, 3H); ¹³C-NMR (150 MHz, CDCl₃) δ 172.1, 171.8, 161.4 (d, *J* = 244.5 Hz), 154.91, 142.33, 138.72, 136.53, 130.7 (d, *J* = 8,31 Hz), 129.85, 128.21, 128.18, 119.67, 115.4 (d, *J* = 22.3 Hz), 81.05, 75.47, 52.92, 52.16, 38.72, 29.72, 28.35, 15.94; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3319, 2980, 2930, 1747, 1689, 1508, 1445, 1367, 1319, 1215, 1153, 1132, 846, 828, 747, 700, 661, 638; HRMS (ESI) calcd. for [C₄₀H₄₁FN₄O₅ + H]⁺: 677.3134, found 677.3133.

8. Procedure for the preparation of tripeptides**a. General procedure for synthesis of Boc-A.A₁-A.A₂-His(Trt)-OMe (**19**)**

Manuscript, Scheme 5a): In a 50 mL flask, DIPEA (186 μ L, 1.09 mmol, 2.1 equiv) was added to Boc-A.A₁-A.A₂-OH (150 mg, 520 μ mol, 1.0 equiv), *L*-H-His(Trt)-OMe•HCl (256 mg, 572 μ mol, 1.1 equiv) and HCTU (452 mg, 1.09 mmol, 2.1 equiv) in DMF (3.5 mL, 0.15 M). The mixture was stirred for 16h at room temperature before being extracted twice with dichloromethane. The combined organic phases were successively washed with sat. aq. NaHCO₃ (2x20 mL), brine (2x20 mL), dried over Na₂SO₄, filtered and concentrated under reduce pressure. The

resulting crude product was purified by column chromatography using the indicated eluent system to afford the desired pure product (**19a-b**).

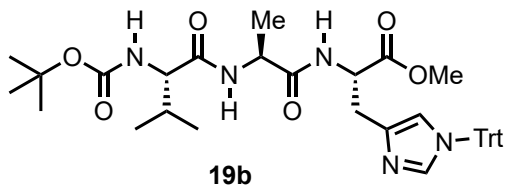
Methyl *N*^α-(*tert*-butoxycarbonyl)-*L*-alanyl-*L*-valyl-*N*^ε-trityl-*L*-histidinate (19a**)**



The general procedure was followed. The crude material was purified on silica gel (1% MeOH/CH₂Cl₂) to afford **19a** as a white solid (259 mg, 73%): **R_f** = 0.34 (5% MeOH/CH₂Cl₂); **m.p.**: 87 – 89 °C; **¹H-NMR**

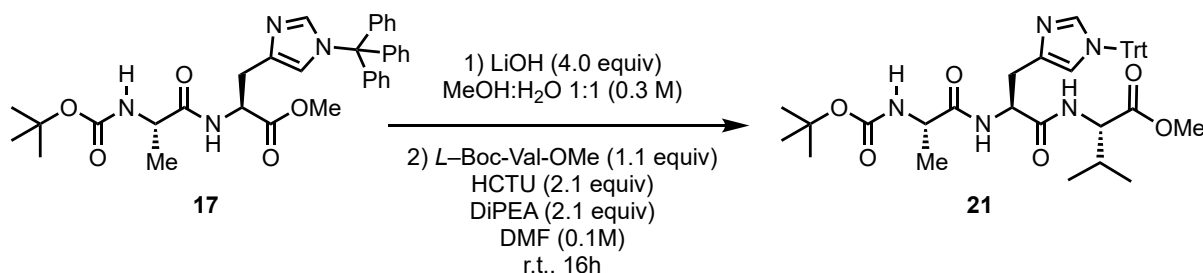
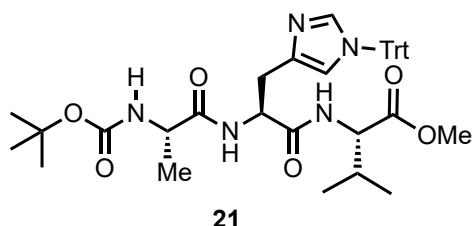
(300 MHz, CDCl₃) δ 7.82 (d, *J* = 7.4 Hz, 1H), 7.43 (s, 1H), 7.35 – 7.25 (m, 9H), 7.15 – 7.09 (m, 1H), 7.09 – 7.01 (m, 6H), 6.58 (s, 1H), 5.44 (s, 1H), 4.79 – 4.63 (m, 1H), 4.31 (dd, *J* = 8.2, 5.7 Hz, 1H), 4.23 – 4.09 (m, 1H), 3.53 (s, 3H), 3.01 (A of ABX, *J* = 14.6, 5.1 Hz, 1H), 2.94 (B of ABX, *J* = 14.9, 5.0 Hz, 1H), 2.20 – 2.05 (m, 1H), 1.38 (s, 9H), 1.26 (d, *J* = 7.0 Hz, 3H), 0.90 (t, *J* = 6.1 Hz, 6H); **presence of conformers** **¹³C-NMR** (75 MHz, CDCl₃) δ 172.9, 171.1, 170.8, 155.5, 141.8, 138.3, 135.5, 129.6, 128.1, 128.1, 119.7, 79.8, 75.6, 58.2, 55.4, 52.5, 52.0, 50.0, 43.4, 38.5, 31.1, 29.1, 28.2, 18.8, 17.9, 17.6, 12.7; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3292, 2972, 1650, 1493, 1445, 1162, 838, 748, 701, 660; **HRMS (ESI)** calcd. for [C₃₉H₄₇N₅O₆ + H]⁺: 682.3599, found 682.3609.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*L*-valyl-*L*-alanyl-*N*^ε-trityl-*L*-histidinate (19b**)**



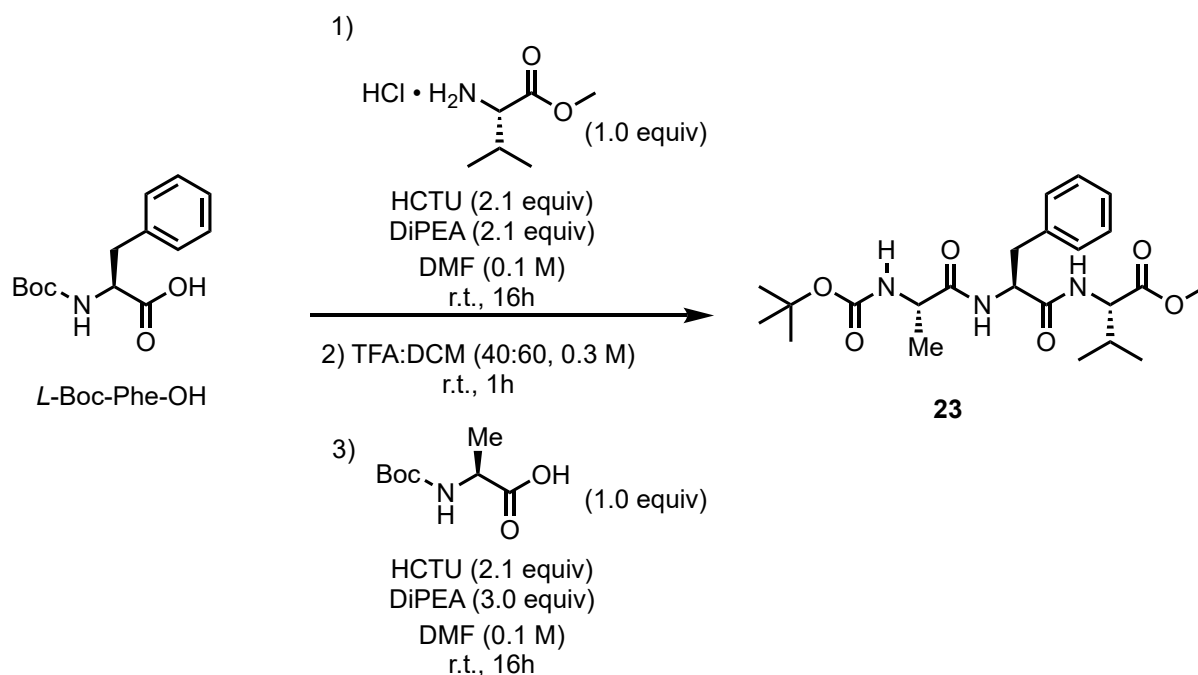
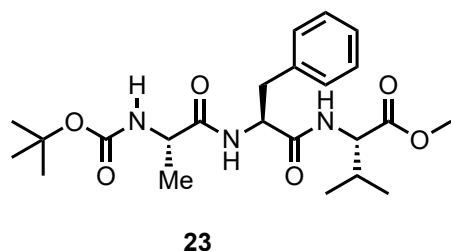
The general procedure was followed. The crude material was purified on silica gel (2% MeOH/CH₂Cl₂) to afford **19b** as a white solid (248 mg, 70%): **R_f** = 0.36 (5% MeOH/CH₂Cl₂); **m.p.**: 92 – 94 °C; **¹H-NMR**

(300 MHz, CDCl₃) δ 7.70 (d, *J* = 7.8 Hz, 1H), 7.45 (s, 1H), 7.34 – 7.27 (m, 9H), 7.13 – 7.00 (m, 7H), 6.56 (d, *J* = 1.0 Hz, 1H), 5.25 (d, *J* = 8.9 Hz, 1H), 4.80 – 4.70 (m, 1H), 4.48 (p, *J* = 6.9 Hz, 1H), 4.01 – 3.91 (m, 1H), 3.55 (s, 3H), 3.04 (A of ABX, *J* = 14.9, 5.1 Hz, 1H), 2.94 (B of ABX, *J* = 10.2, 5.6 Hz, 1H), 2.14 – 1.99 (m, 1H), 1.39 (s, 9H), 1.36 (d, *J* = 7.1 Hz, 3H), 0.85 (dd, *J* = 13.5, 6.8 Hz, 6H); **presence of conformers** **¹³C-NMR** (75 MHz, CDCl₃) δ 172.0, 171.4, 171.2, 155.9, 141.9, 138.4, 135.6, 129.7, 128.2, 128.2, 119.9, 79.7, 75.7, 59.6, 55.6, 52.6, 52.2, 49.0, 43.6, 38.6, 31.2, 29.2, 28.3, 19.3, 18.6, 17.5, 12.8; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3275, 2972, 1745, 1651, 1493, 1445, 1366, 1160, 1132, 838, 748, 701, 660; **HRMS (ESI)** calcd. for [C₃₉H₄₇N₅O₆ + H]⁺: 682.3599, found 682.3606.

b. Procedure for synthesis of Boc-Ala-His(Trt)-Val-OMe (21)**Methyl *N*^α-((*tert*-butoxycarbonyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (21)**

Manuscript, Scheme 5b): In a 25 mL flask, lithium hydroxide (26 mg, 1.1 mmol, 4.0 equiv) was added to a solution of methyl *N*^α-((*tert*-butoxycarbonyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (**17**) (160 mg, 281 μmol, 1.0 equiv)

in methanol:water 1:1 (1 mL). The mixture was stirred for 16h at room temperature before concentrated to water under reduce pressure. The residue was lyophilized to obtain Boc-Ala-His(Trt)-O⁻Li⁺ as white solid, used without further purification. In a 50 mL flask, DIPEA (100 μL, 590 μmol, 2.1 equiv) was added to Boc-Ala-His(Trt)-O⁻Li⁺, L-H-Val-OMe•HCl (52 mg, 0.31 mmol, 1.1 equiv) and HCTU (244 mg, 590 μmol, 2.1 equiv) in DMF (2.8 mL, 0.1 M). The mixture was stirred for 16h at room temperature before being extracted twice with dichloromethane. The combined organic phases were successively washed with sat. aq. NaHCO₃ (2x20 mL), brine (2x20 mL), dried over Na₂SO₄, filtered and concentrated under reduce pressure. The resulting crude product was purified by column chromatography (2% MeOH/CH₂Cl₂) to afford **21** as a white solid (89 mg, 46%): *R*_f = 0.42 (5% MeOH/CH₂Cl₂); *m.p.*: 79 – 81 °C; ¹H-NMR (300 MHz, CDCl₃) δ 8.46 (d, *J* = 6.1 Hz, 1H), 7.52 (d, *J* = 8.2 Hz, 1H), 7.38 – 7.27 (m, 10H), 7.13 – 7.00 (m, 6H), 6.66 (s, 1H), 5.22 (d, *J* = 6.6 Hz, 1H), 4.78 – 4.65 (m, 1H), 4.39 (dd, *J* = 8.4, 5.3 Hz, 1H), 4.24 – 4.12 (m, 1H), 3.64 (s, 3H), 3.11 (A of ABX, *J* = 15.1, 4.3 Hz, 1H), 2.89 (B of ABX, *J* = 15.1, 5.9 Hz, 1H), 2.14 – 2.01 (m, 1H), 1.38 (d, *J* = 7.2 Hz, 3H), 1.35 (s, 9H), 0.80 (d, *J* = 6.8 Hz, 6H); ¹³C-NMR (75 MHz, CDCl₃) δ ¹³C NMR (75 MHz, CDCl₃) δ 172.8, 171.9, 171.0, 155.4, 142.2, 138.2, 136.9, 129.8, 128.2, 128.1, 119.7, 79.8, 75.5, 57.6, 53.3, 52.0, 50.7, 30.9, 29.3, 28.4, 19.1, 18.0; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3275, 2970, 1740, 1659, 1493, 1445, 1366, 1245, 1205, 1159, 1001, 841, 748, 701, 659, 638; HRMS (ESI) calcd. for [C₃₉H₄₇N₅O₆ + H]⁺: 682.3599, found 682.3594.

c. Procedure for synthesis of Boc-Ala-Phe-Val-OMe (**23**)Methyl (*tert*-butoxycarbonyl)-*L*-alanyl-*L*-phenylalanyl-*L*-valinate (**23**)

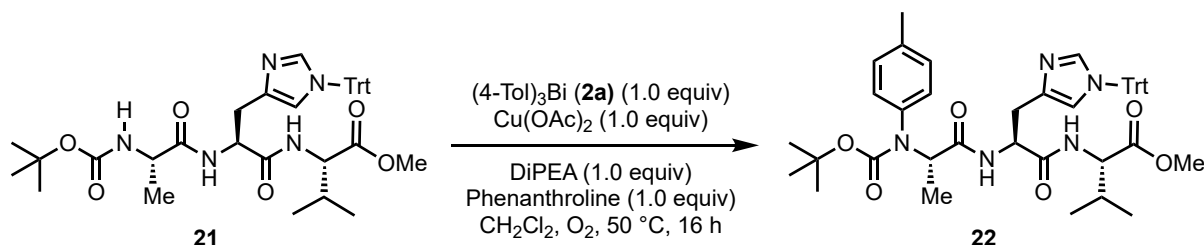
Manuscript, Scheme 5c): In a 50 mL flask, DIPEA (270 μL , 1.58 mmol, 2.1 equiv) was added to *L*-Boc-Phe-OH (200 mg, 754 μmol , 1.0 equiv), H-Val-OMe $\cdot$ HCl (126 mg, 754 μmol , 1.0 equiv), and HCTU (655 mg, 1.58 mmol, 2.1 equiv) in DMF (7.5 mL, 0.1 M). The mixture was stirred

for 16h at room temperature before concentrated under reduce pressure. The residue was diluted in dichloromethane and successively washed with sat. aq. NaHCO_3 (2x20 mL), brine (2x20 mL), dried over Na_2SO_4 , filtered before concentrated under reduce pressure. The thick yellow oil residue was diluted in TFA: CH_2Cl_2 (40:60 ; 0.3 M) and stirred at room temperature for 1h. The trifluoroacetic acid was completely removed by several co-evaporation with dichloromethane under reduce pressure. To the residue was added Boc-Ala-OH (143 mg, 754 μmol , 1.0 equiv) and HCTU (655 mg, 1.58 mmol, 2.1 equiv) in DMF (7.5 mL, 0.1 M) followed by DIPEA (385 μL , 2.26 mmol, 3.0 equiv). The mixture was stirred for 16h at room temperature before concentrated under reduce pressure. The residue was diluted in dichloromethane and successively washed with sat. aq. NaHCO_3 (2x20 mL), brine (2x20 mL), dried over Na_2SO_4 , filtered before concentrated under reduce pressure. The resulting crude product was purified by column chromatography (3% MeOH/ CH_2Cl_2) to afford **23** as a thick yellow oil (312 mg, 92%):

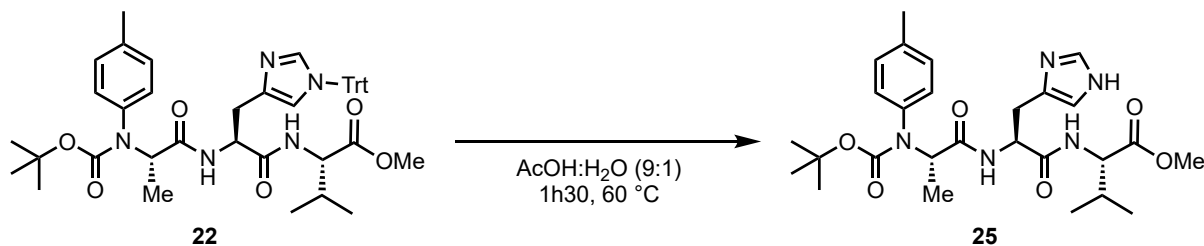
R_f = 0.41 (10% MeOH/CH₂Cl₂); ¹H-NMR (300 MHz, CDCl₃) δ 7.31 – 7.14 (m, 4H), 6.87 (d, J = 7.9 Hz, 1H), 6.60 (d, J = 7.5 Hz, 1H), 5.07 (d, J = 7.2 Hz, 1H), 4.71 (q, J = 7.2 Hz, 1H), 4.41 (dd, J = 8.6, 5.3 Hz, 1H), 4.21 – 4.07 (m, 1H), 3.67 (s, 3H), 3.06 (d, J = 7.2 Hz, 2H), 2.12 – 1.99 (m, 2H), 1.40 (s, 9H), 1.28 (d, J = 7.1 Hz, 3H), 0.82 (dd, J = 10.3, 6.9 Hz, 6H); ¹³C-NMR (75 MHz, CDCl₃) δ 173.0, 171.8, 170.9, 155.4, 136.5, 129.3, 128.4, 126.8, 79.8, 57.4, 54.3, 52.0, 50.4, 38.6, 31.1, 28.3, 18.8, 17.8; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3287, 2969, 1741, 1712, 1646, 1499, 1454, 1366, 1246, 1206, 1164, 1049, 1023, 742, 698; HRMS (ESI) calcd. for [C₂₃H₃₅N₃O₆ + H]⁺: 450.2599, found 450.2610.

9. Procedure for the preparation of aryl tripeptides

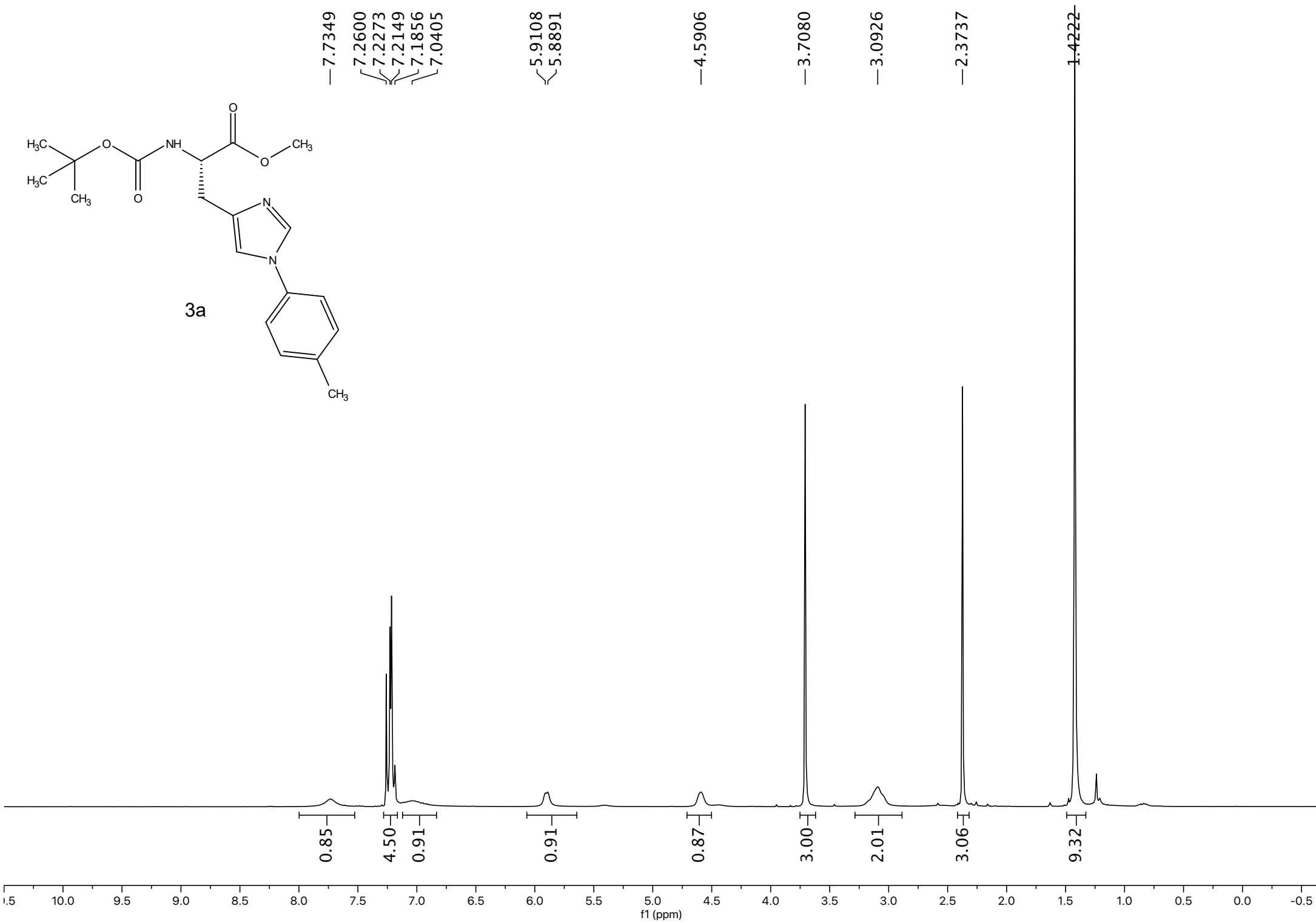
a. Procedure for synthesis of Boc-*N*(Tolyl)-Ala-His(trt)-Val-OMe (**22**)

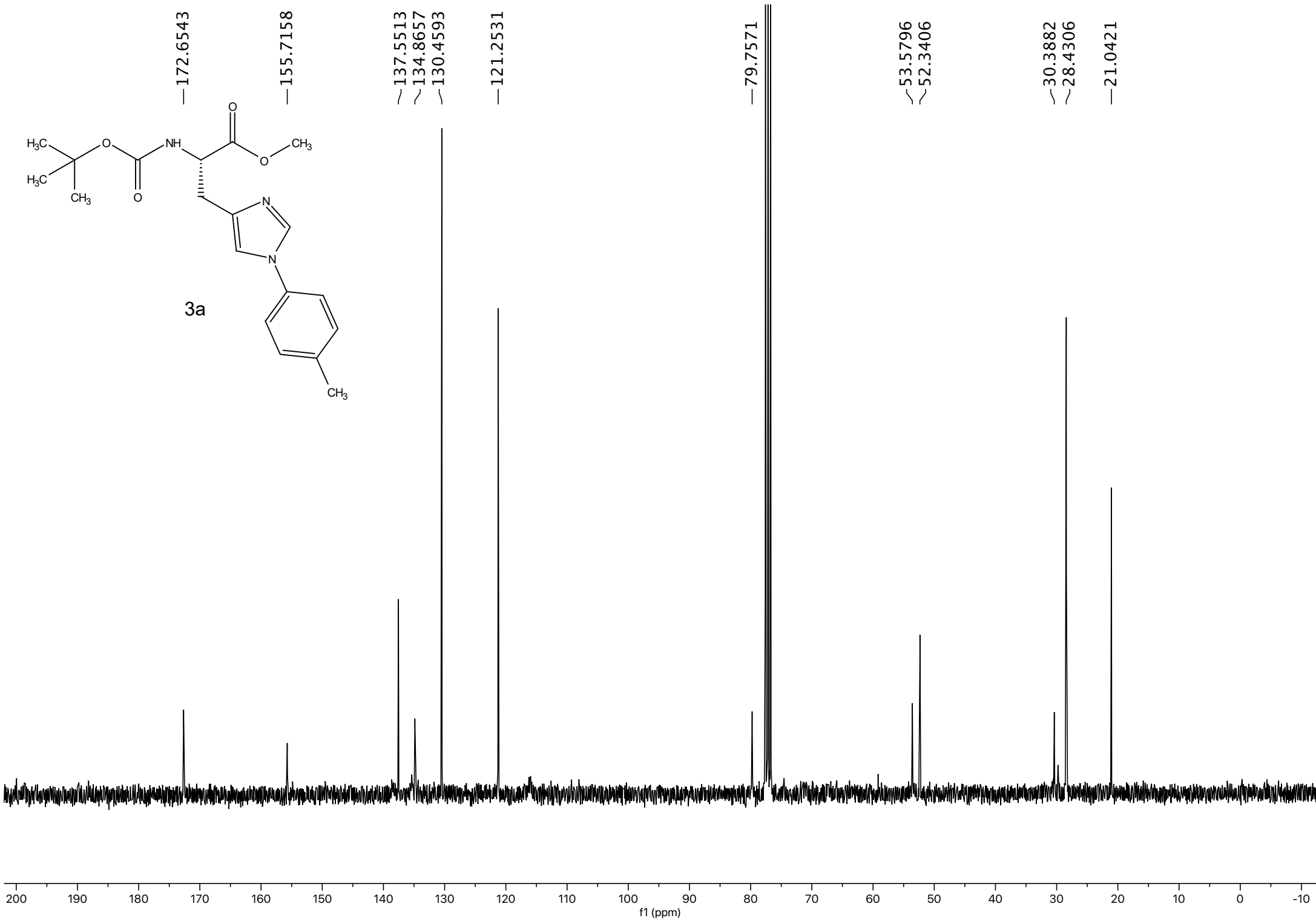


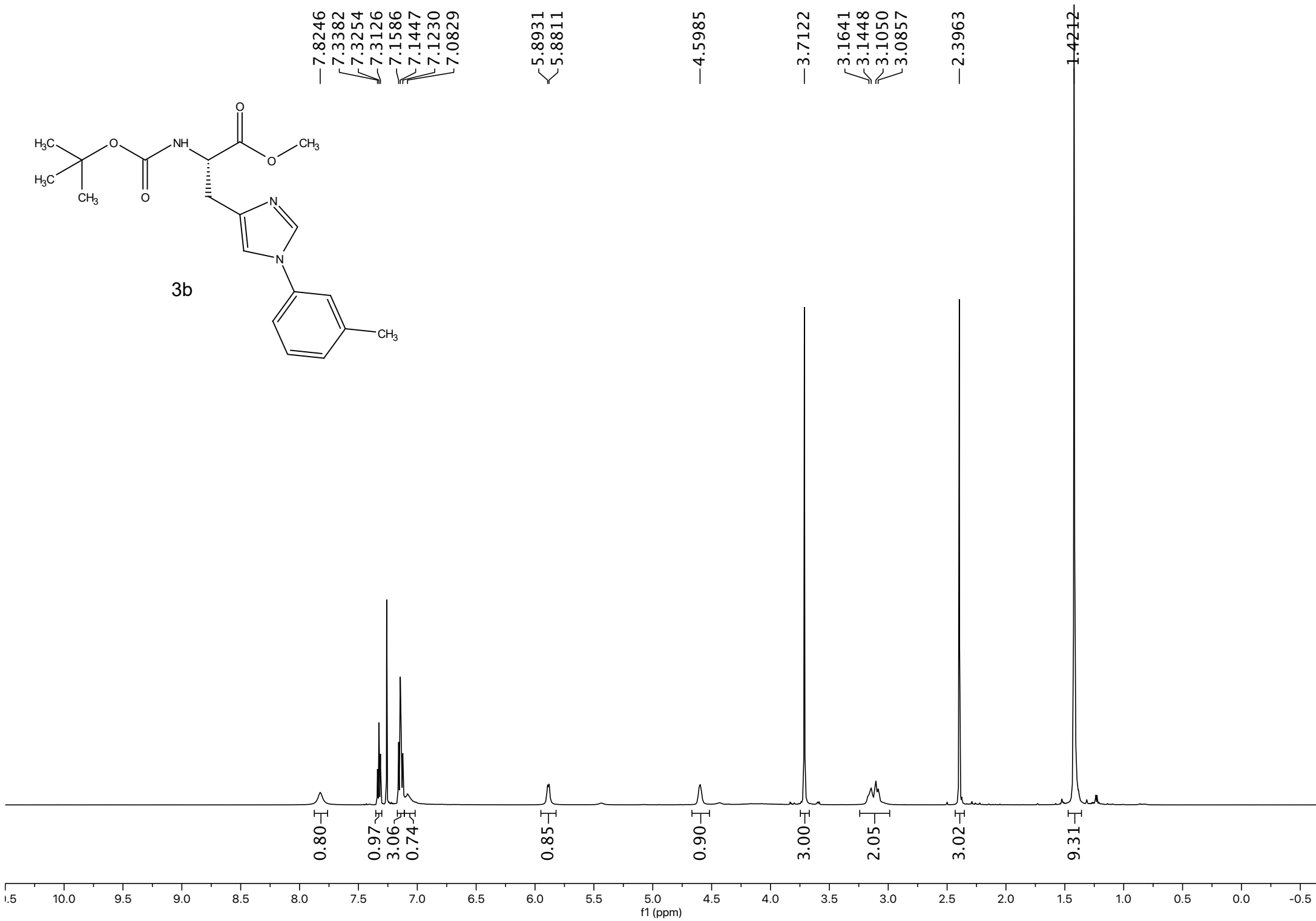
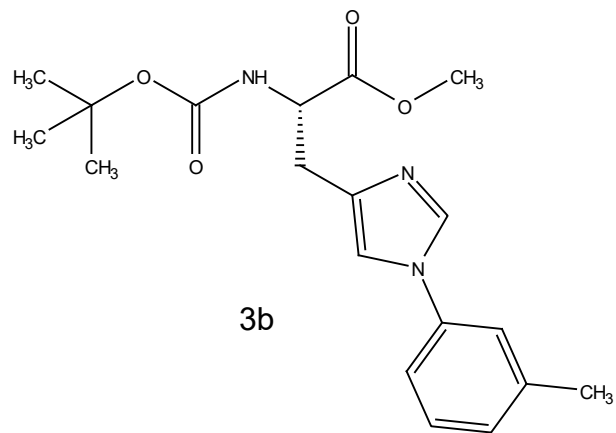
Manuscript, Scheme 5b: In a sealed tube, *N,N*-diisopropylethylamine (25.0 μ L, 0.15 mmol, 1.0 equiv) was added to methyl α -((tert-butoxycarbonyl)-L-alanyl)-*N* ϵ -trityl-L-histidyl-L-valinate **21** (100 mg, 0.15 mmol, 1.0 equiv), tri(4-tolyl)bismuthine **2a** (71 mg, 0.15 mmol, 1.0 equiv), copper(II) acetate (27 mg, 0.15 mmol, 1.0 equiv) and phenanthroline (27 mg, 0.15 μ mol, 1.0 equiv) in CH₂Cl₂ (0.1 M). The reaction mixture was purged with oxygen before stirred in an oil bath set at 50 °C for 16 hours. The solvent was evaporated under reduce pressure and the residue was purified on silica gel chromatography (2% MeOH/CH₂Cl₂) to afford **22** as a white solid (94 mg, 83%): R_f = 0.46 (5% MeOH/CH₂Cl₂); m.p.: 134 – 136 °C; ¹H-NMR (300 MHz, CDCl₃) δ 8.35 (d, J = 7.2 Hz, 1H), 8.08 (s, 1H), 7.74 (d, J = 5.9 Hz, 1H), 7.41 – 7.31 (m, 10H), 7.10 (dd, J = 5.9, 1.7 Hz, 8H), 7.04 (d, J = 8.2 Hz, 1H), 6.87 (s, 1H), 5.27 – 5.11 (m, 1H), 4.50 – 4.29 (m, 2H), 3.67 (s, 3H), 3.54 (d, J = 15.1 Hz, 1H), 2.96 – 2.81 (m, 1H), 2.27 (s, 3H), 2.26 – 2.20 (m, 1H), 1.31 (d, J = 6.6 Hz, 3H), 1.22 (s, 9H), 1.02 (dd, J = 15.7, 6.8 Hz, 6H); ¹³C-NMR (75 MHz, CDCl₃) δ 172.1, 172.0, 170.7, 154.4, 140.0, 138.5, 136.6, 134.3, 132.2, 129.8, 129.4, 129.2, 128.9, 128.1, 120.9, 80.6, 78.7, 58.7, 52.8, 52.0, 30.6, 30.2, 28.3, 21.1, 19.3, 18.5, 15.7; IR (ATR)/ $\bar{\nu}$ (cm⁻¹): 3155, 2970, 1740, 1668, 1514, 1495, 1447, 1367, 1155, 1129, 1019, 1001, 839, 747, 701, 662, 640; HRMS (ESI) calcd. for [C₄₆H₅₃N₅O₆ + H]⁺: 772.4069, found 772.4058.

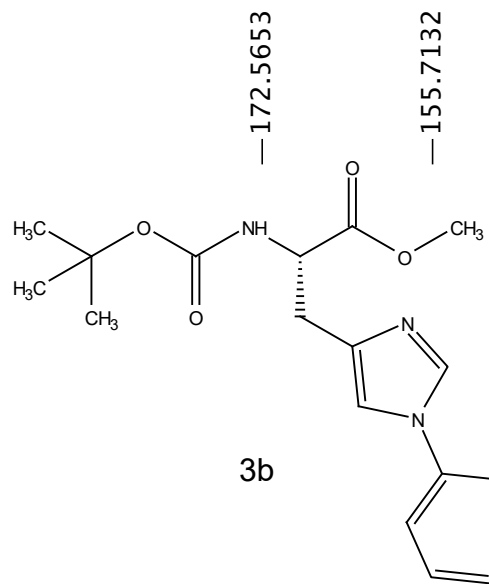
b. Procedure for synthesis of Boc-*N*(Tolyl)-Ala-His-Val-OMe (25)

Manuscript, Scheme 7: In a flask, Boc-*N*(Tolyl)-Ala-His(trt)-Val-OMe **22** (50 mg, 0.65 mmol) in AcOH:H₂O (9:1, 0.35 M) was stirred for 1h30 at 60 °C. After cooling down at room temperature, the solvent was co-evaporated with cyclohexane under reduce pressure. The white solid residue was purified on silica gel chromatography (5% MeOH/CH₂Cl₂) to afford **25** as a white solid (35 mg, 99%): **R_f** = 0.46 (6% MeOH/CH₂Cl₂); **m.p.**: 154 – 157 °C; **¹H-NMR (600 MHz, CDCl₃)** δ 7.86 (s(br), 1H), 7.62 (s(br), 1H), 7.56 (s, 1H), 7.14 – 7.08 (m, 4H), 6.83 (s, 1H), 4.80 – 4.75 (m, 1H), 4.57 – 4.51 (m, 1H), 4.39 (dd, *J* = 8.3, 5.2 Hz, 1H), 3.71 (s, 3H), 3.16 (dd, *J* = 15.1, 4.7 Hz, 1H), 3.09 (dd, *J* = 15.2, 5.9 Hz, 1H), 2.33 (s, 3H), 2.18 – 2.11 (m, 1H), 1.36 (s, 9H), 1.30 (d, *J* = 6.9 Hz, 3H), 0.86 (dd, *J* = 6.8, 3.5 Hz, 6H); **¹³C-NMR (150 MHz, CDCl₃)** δ 172.5, 172.2, 171.3, 155.2, 137.6, 136.8, 134.9, 129.4, 128.2, 81.1, 64.2, 57.8, 53.1, 52.2, 30.6, 28.8, 28.3, 28.2, 21.1, 19.0, 17.8, 15.5; **IR (ATR)/ $\bar{\nu}$ (cm⁻¹)**: 3287, 2971, 1741, 1661, 1514, 1455, 1391, 1367, 1323, 1255, 1158, 1107, 1019, 855, 832, 755, 622; **HRMS (ESI)** calcd. for [C₂₇H₃₉N₅O₆ + H]⁺: 530.2973, found 530.2976.









3b

—172.5653

—155.7132

—140.2022

—137.1220

~129.7915

~128.4346

—122.0057

—118.4123

—79.7816

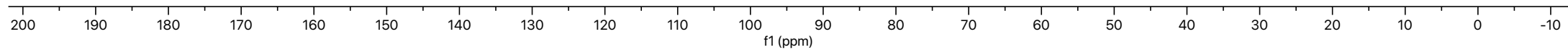
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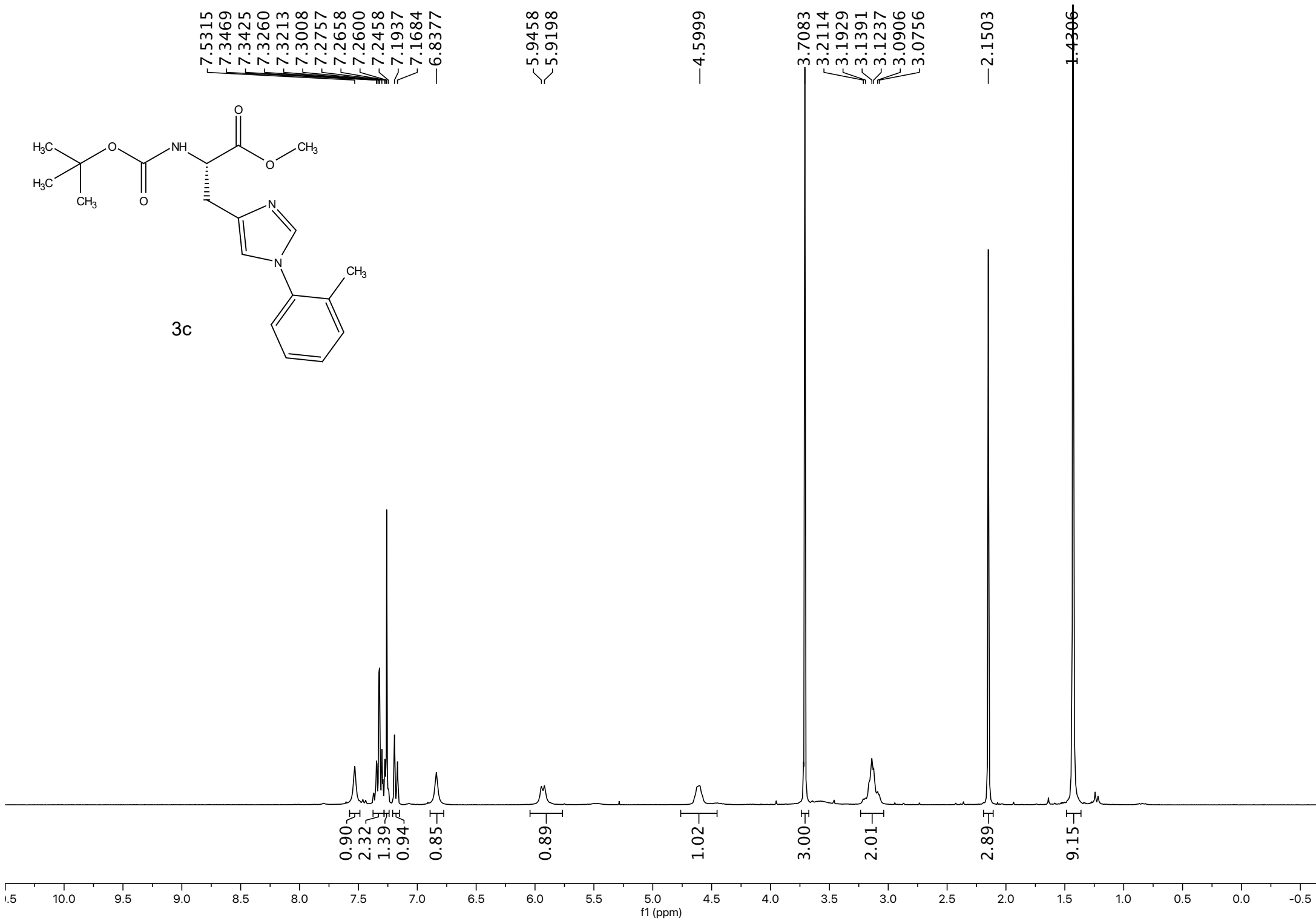
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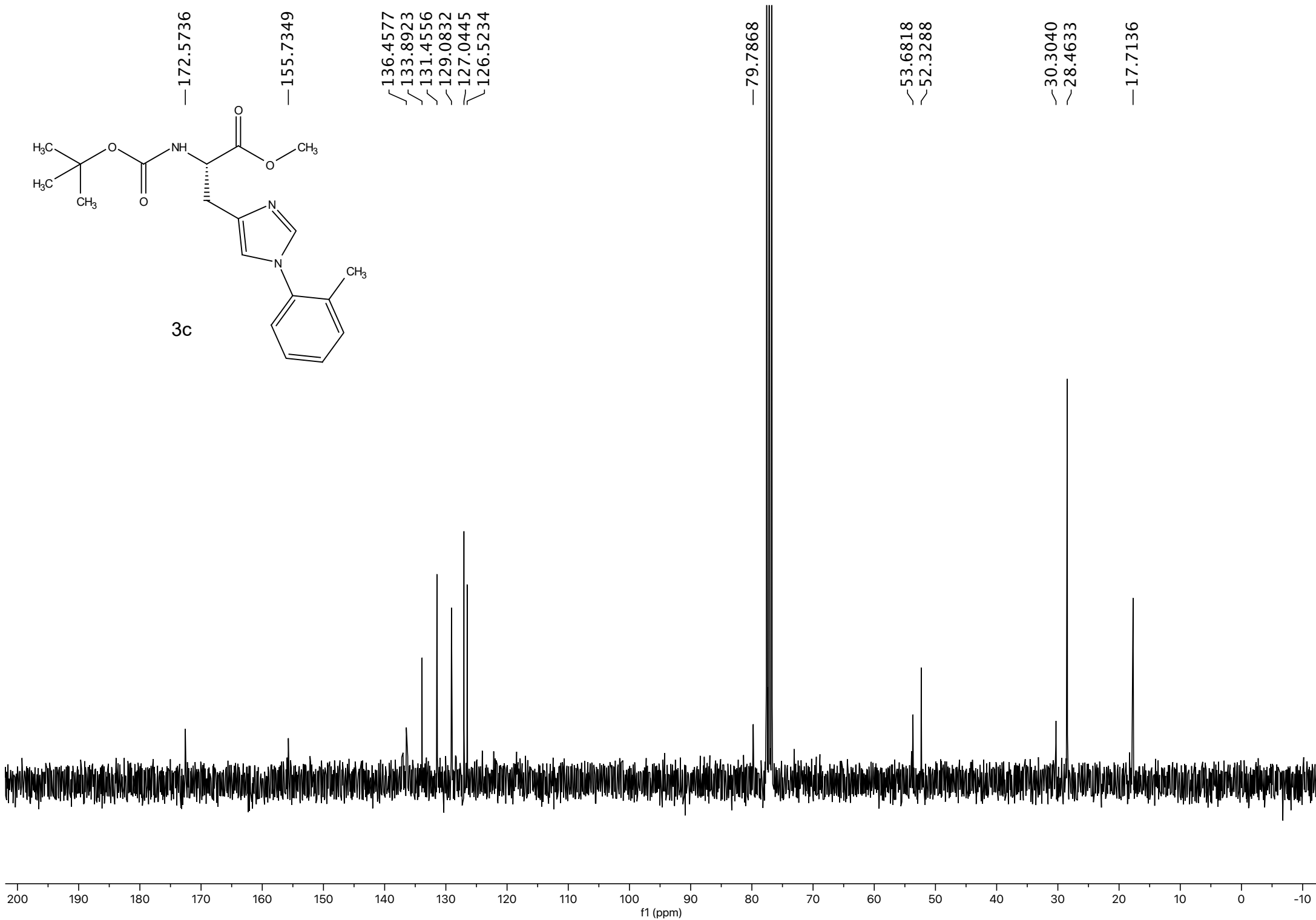
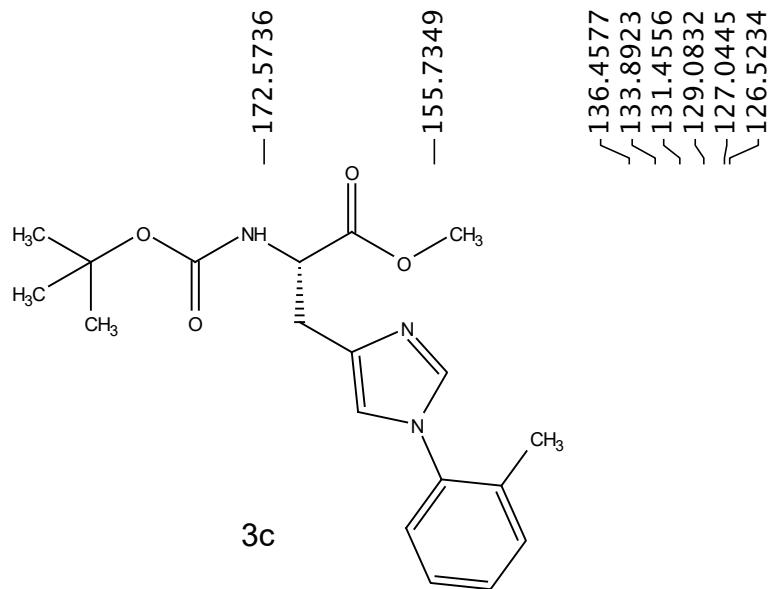
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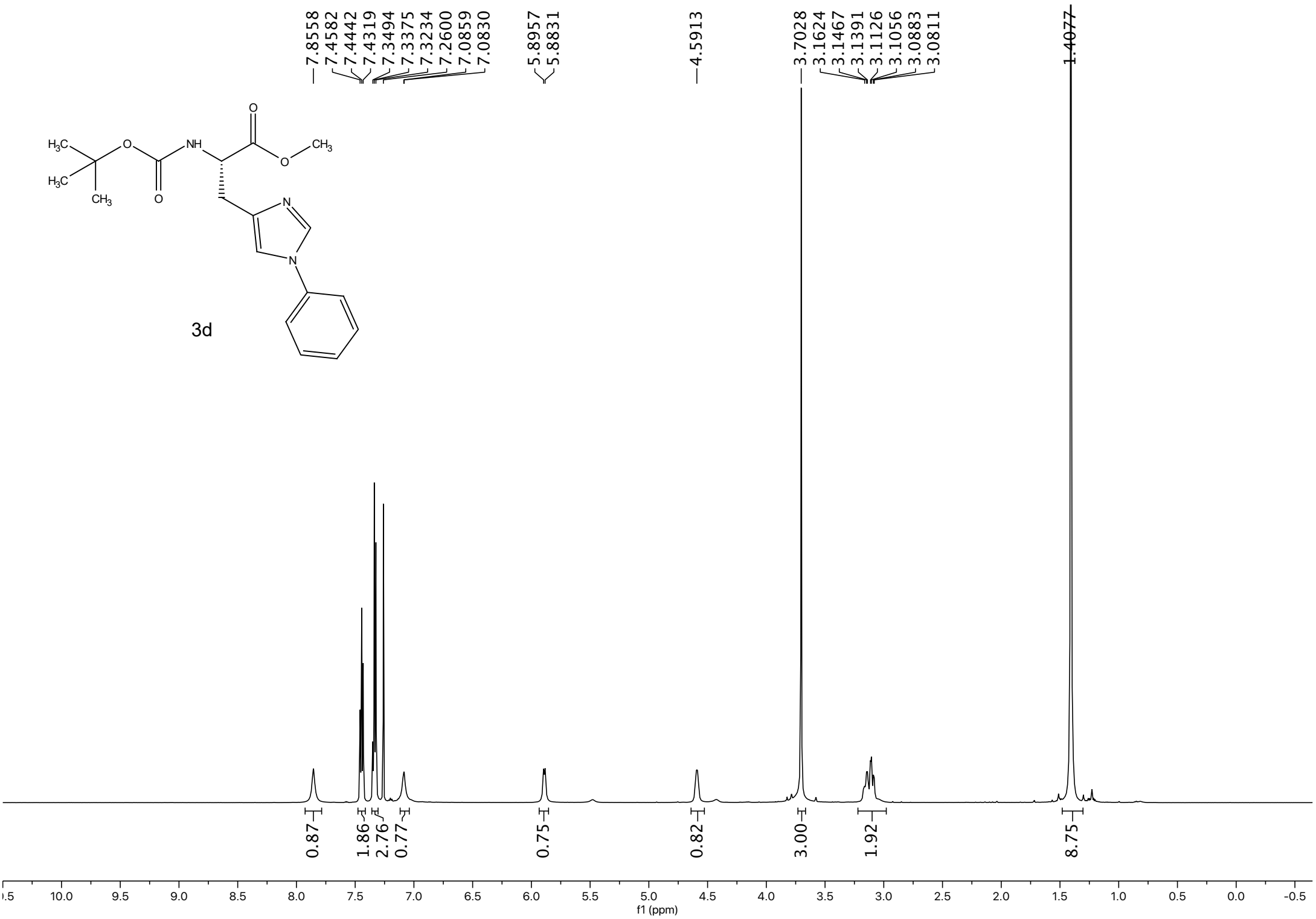
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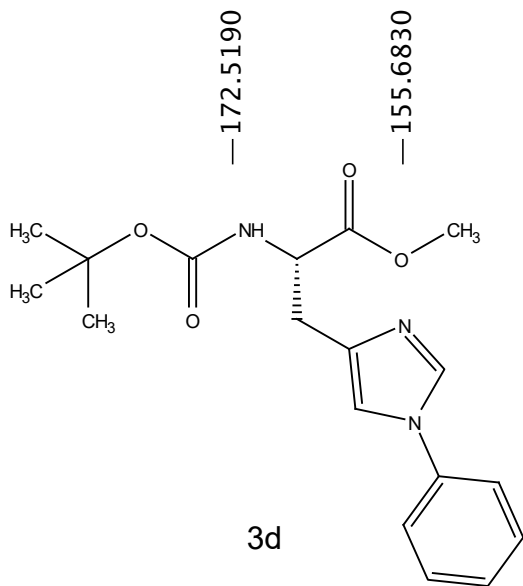
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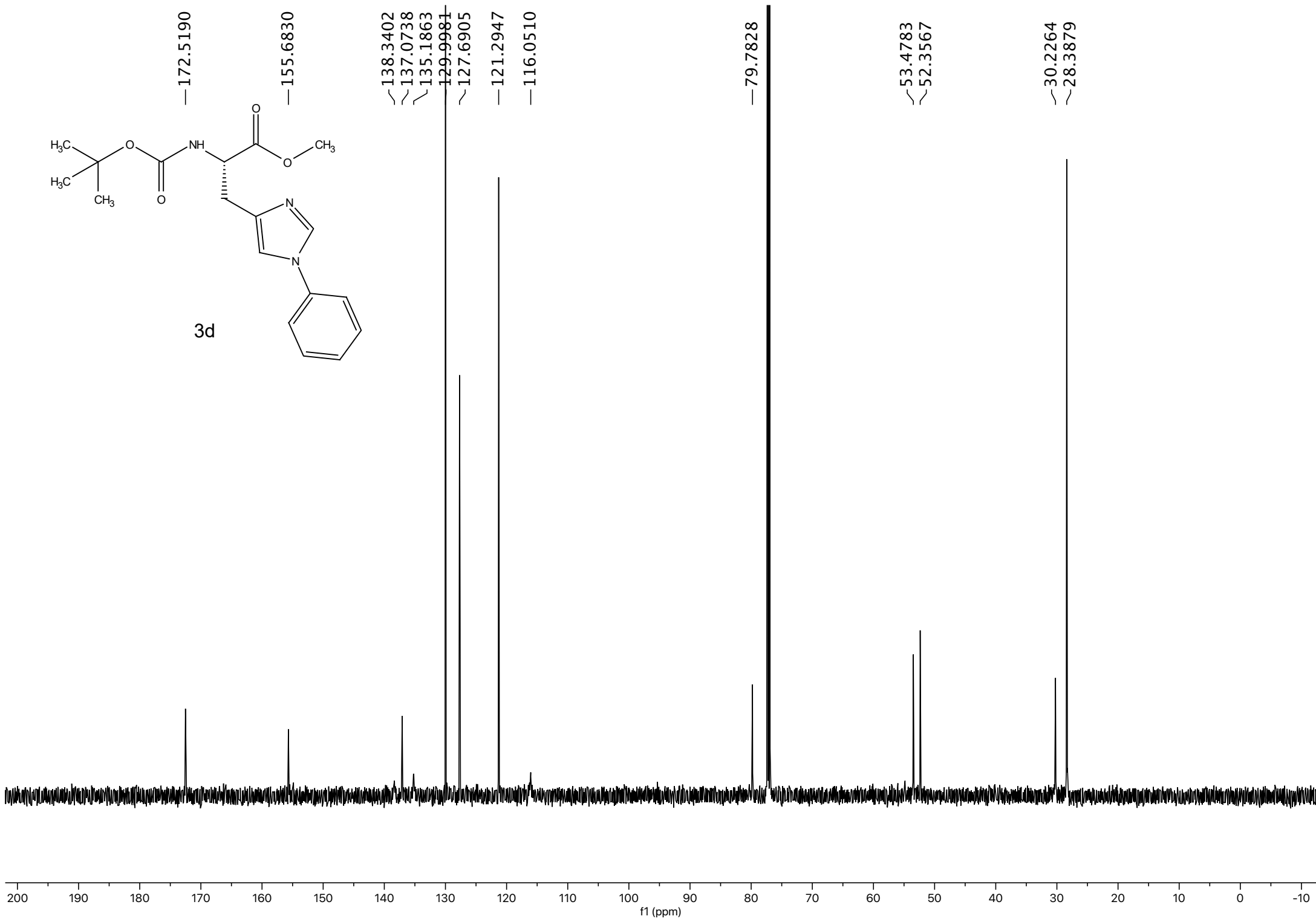




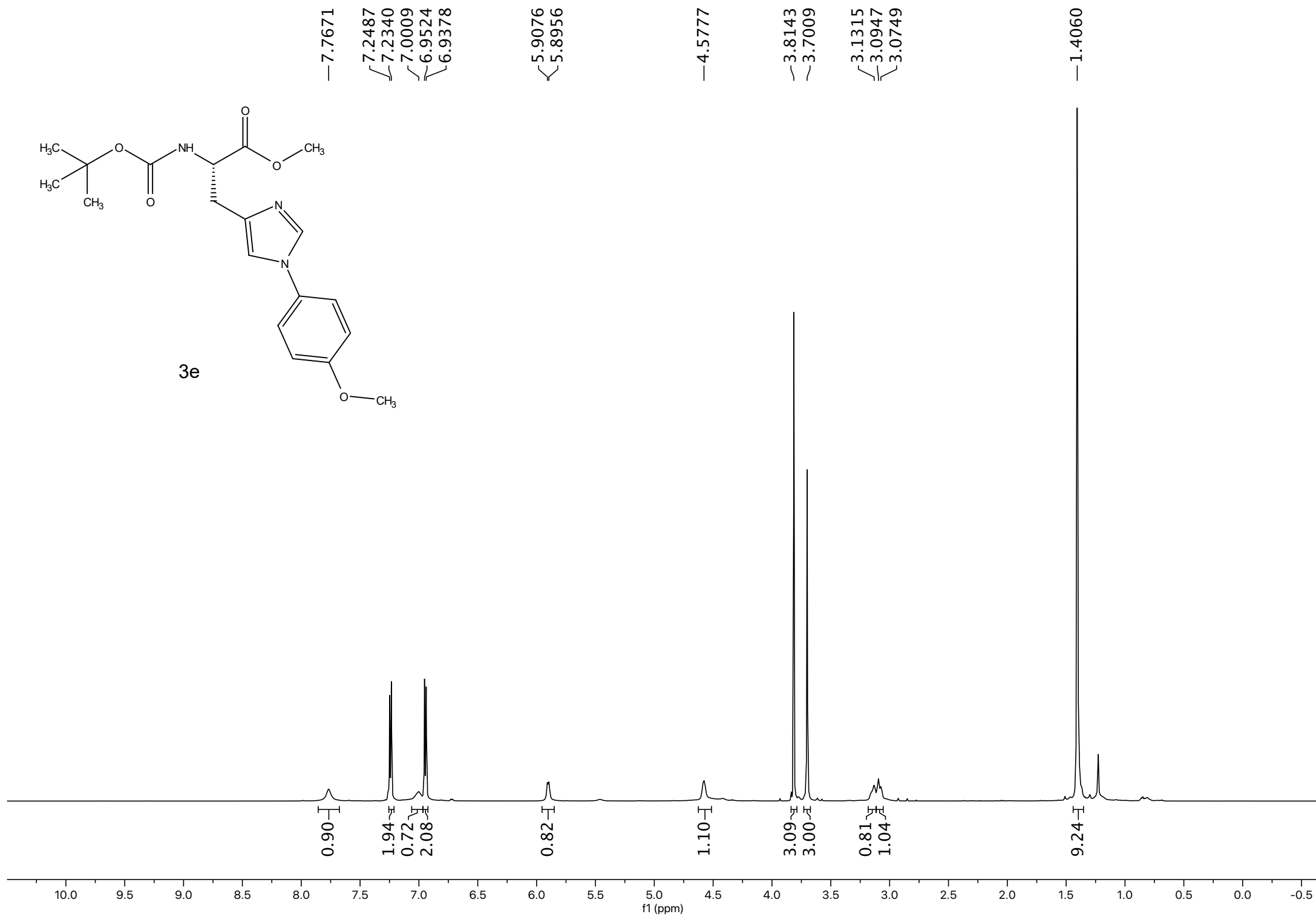
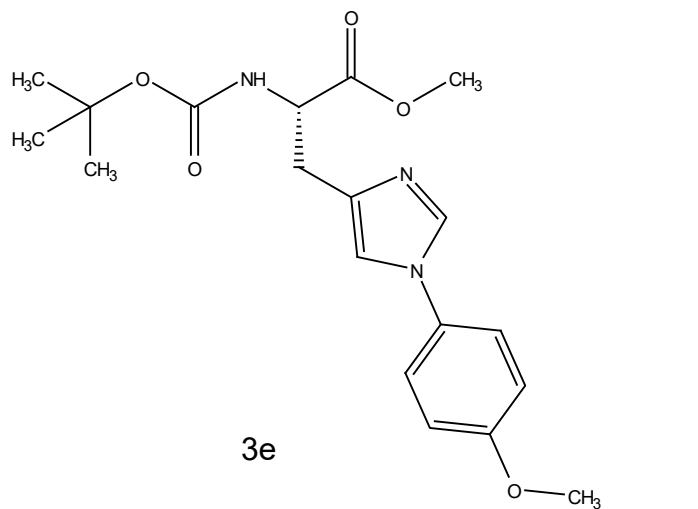


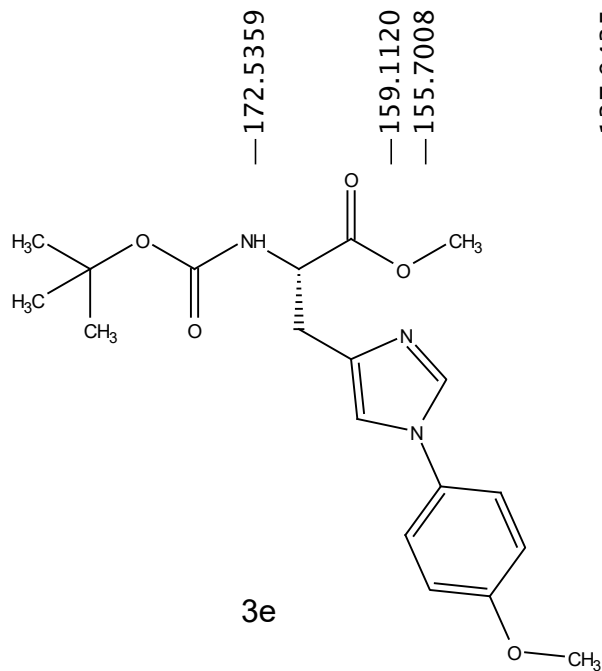


3d



3e

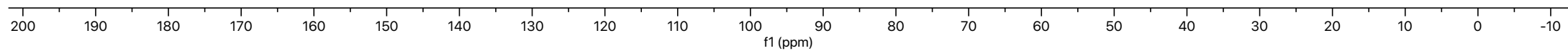


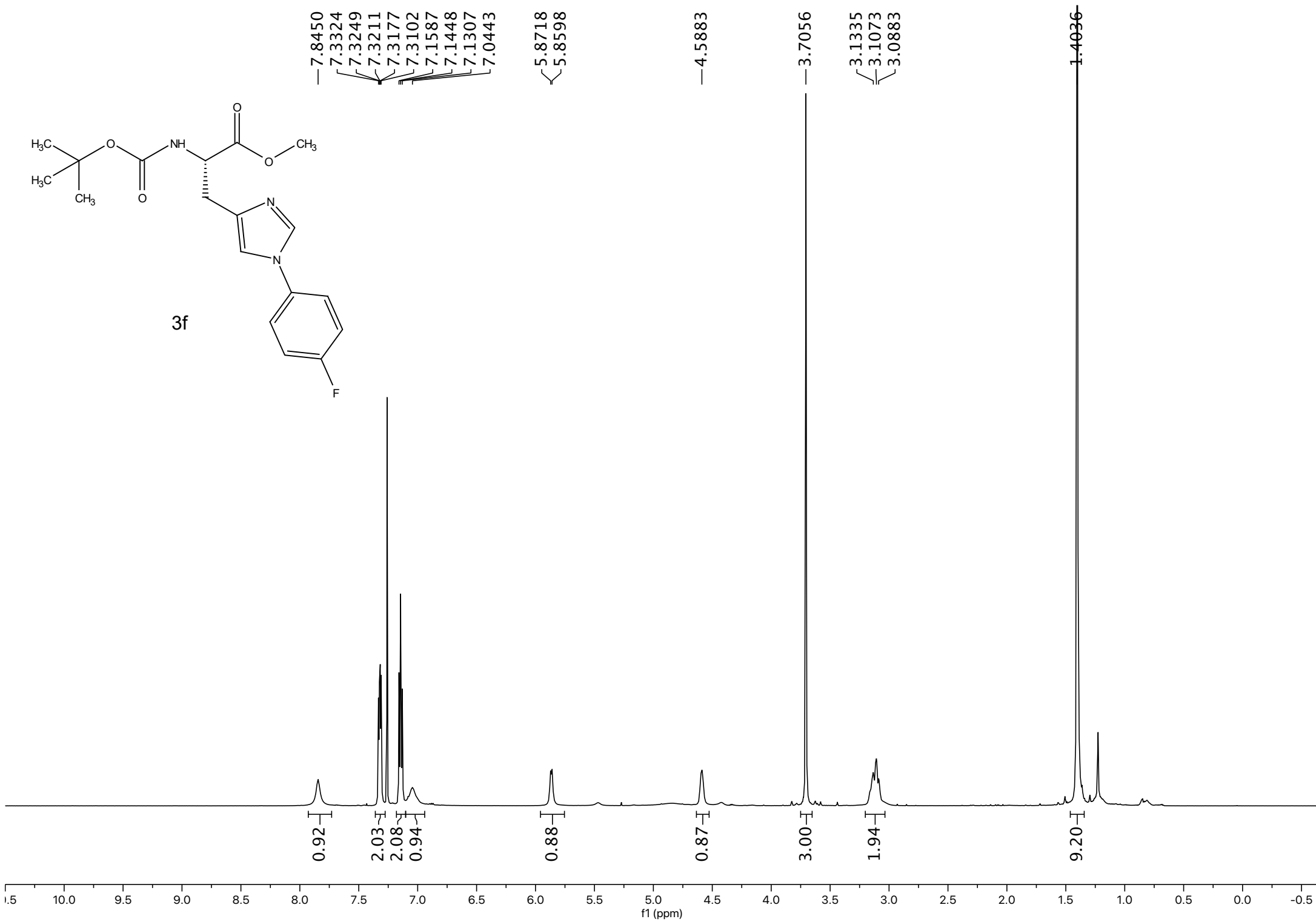


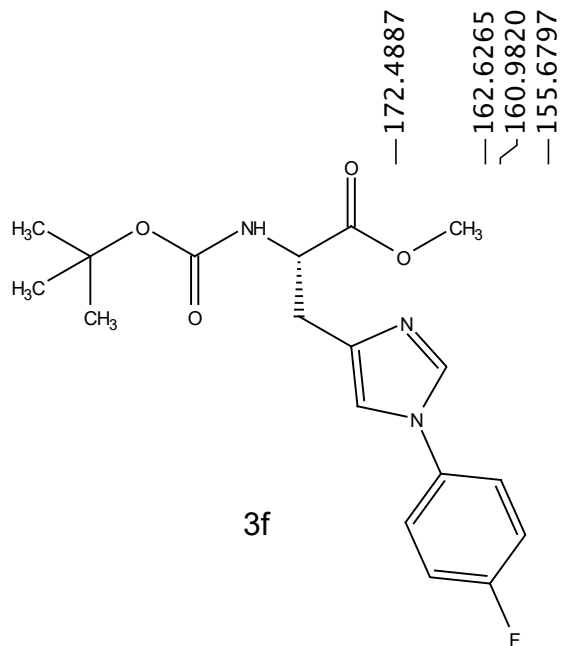
3e

Chemical shifts (ppm) labeled above the spectrum:

- 172.5359
- 159.1120
- 155.7008
- 137.9435
- 135.3287
- 130.4255
- 123.0500
- 116.5852
- 115.0273
- 79.7495
- 55.6745
- 53.5120
- 52.3357
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- 28.3964

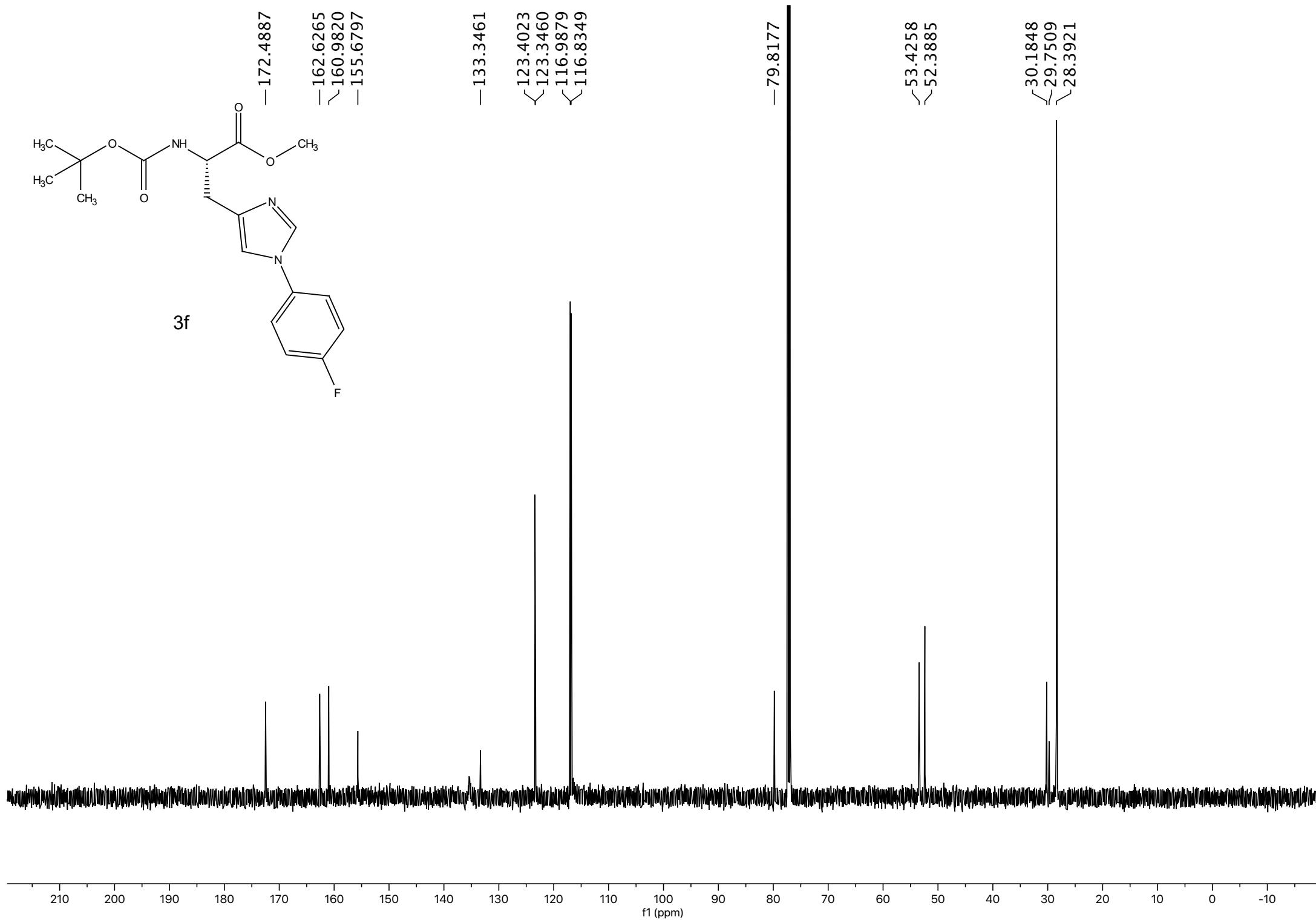


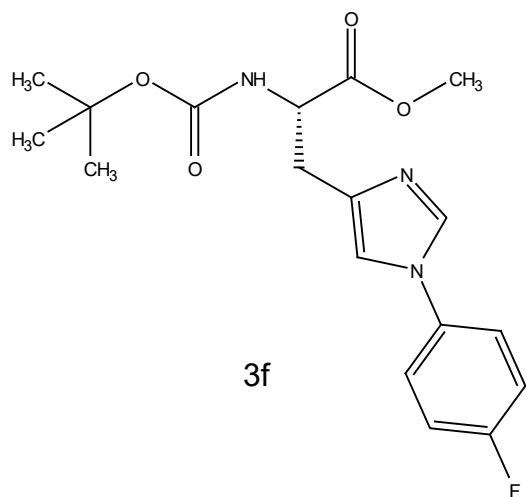




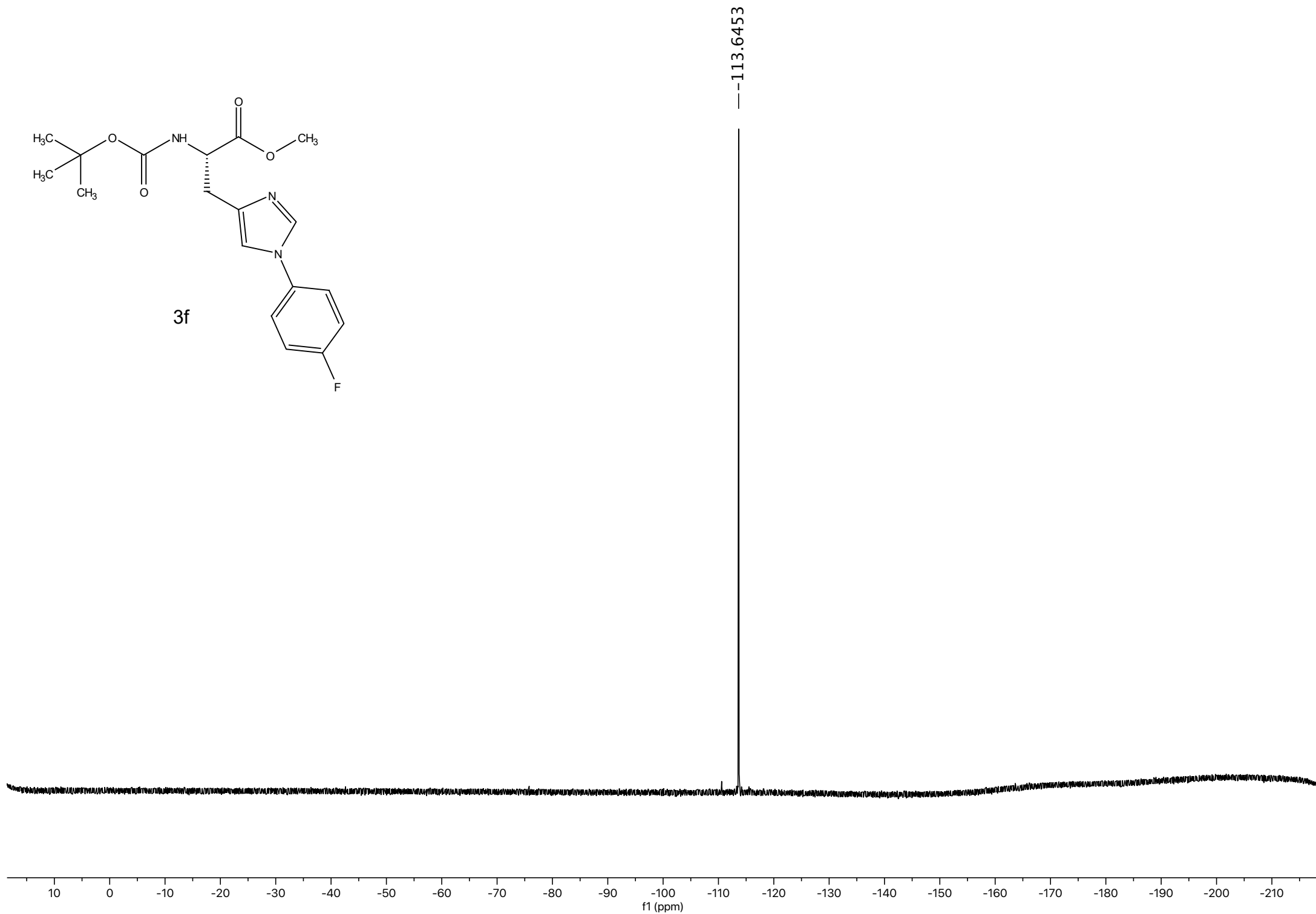
3f

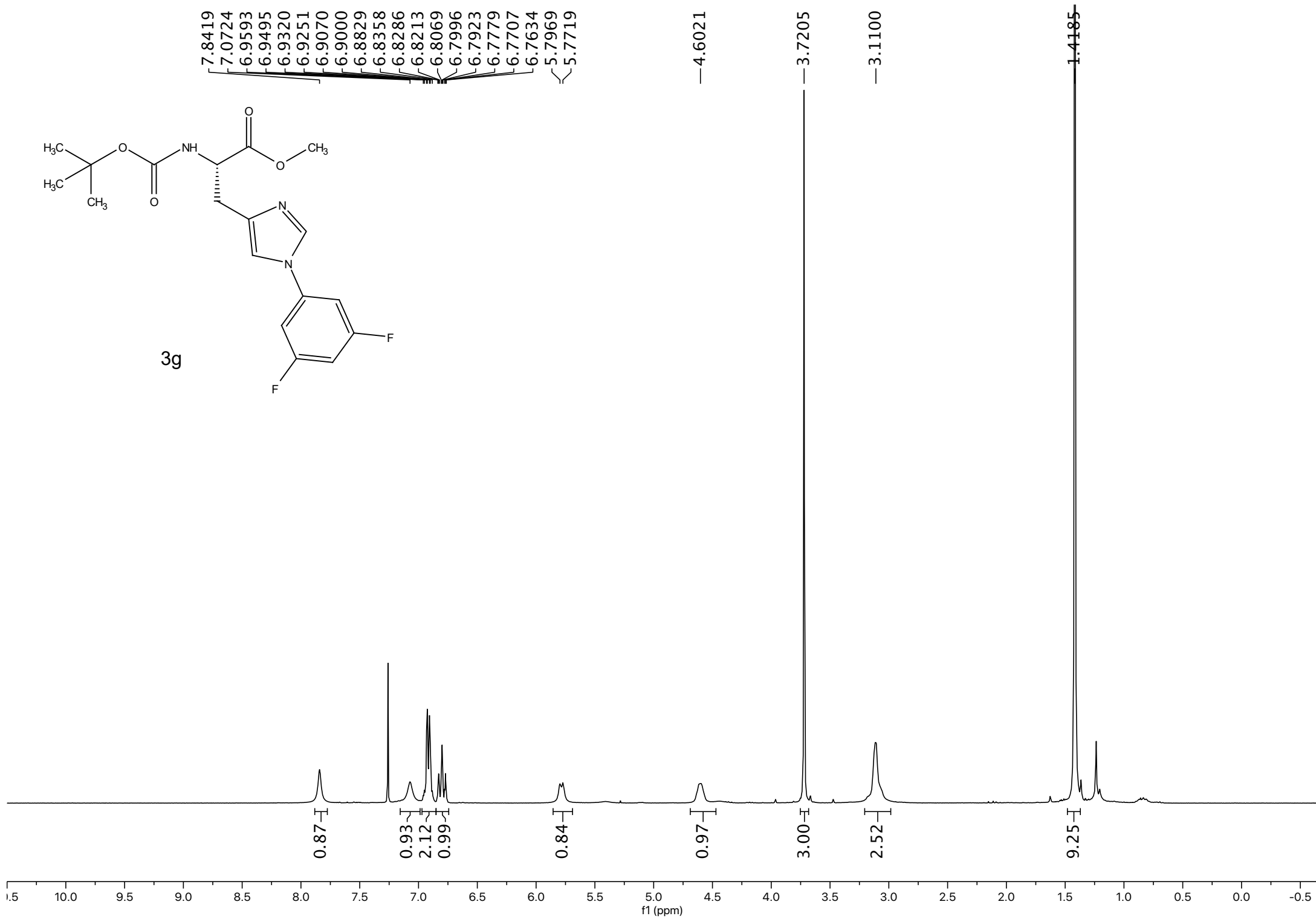
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30.1848
29.7509
28.3921



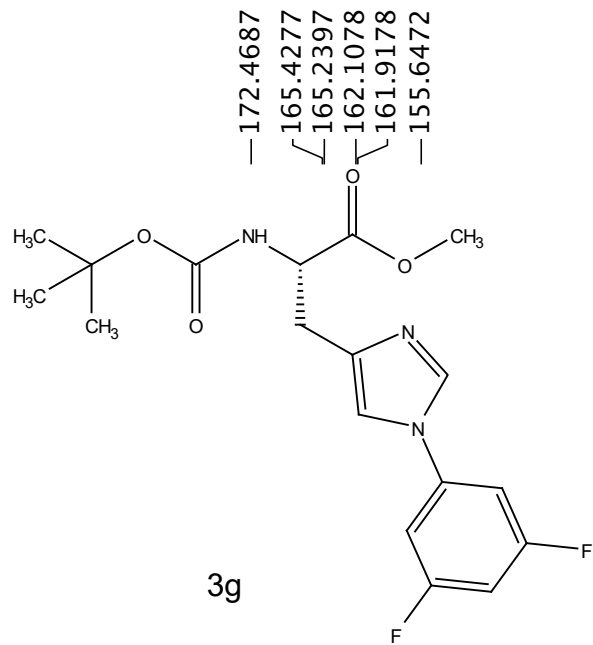


3f





3g



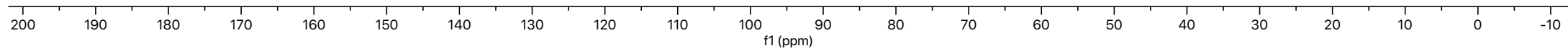
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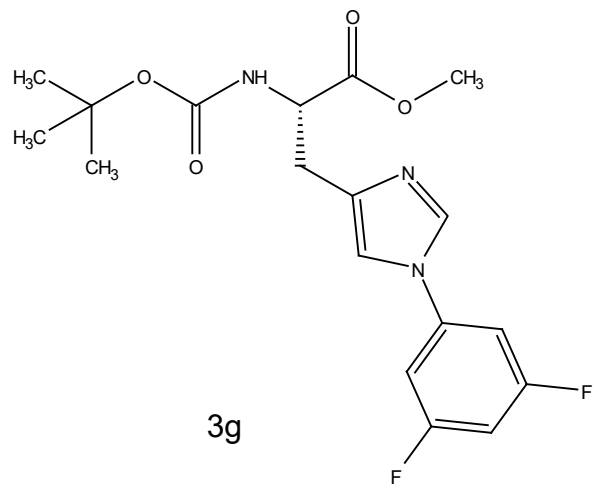
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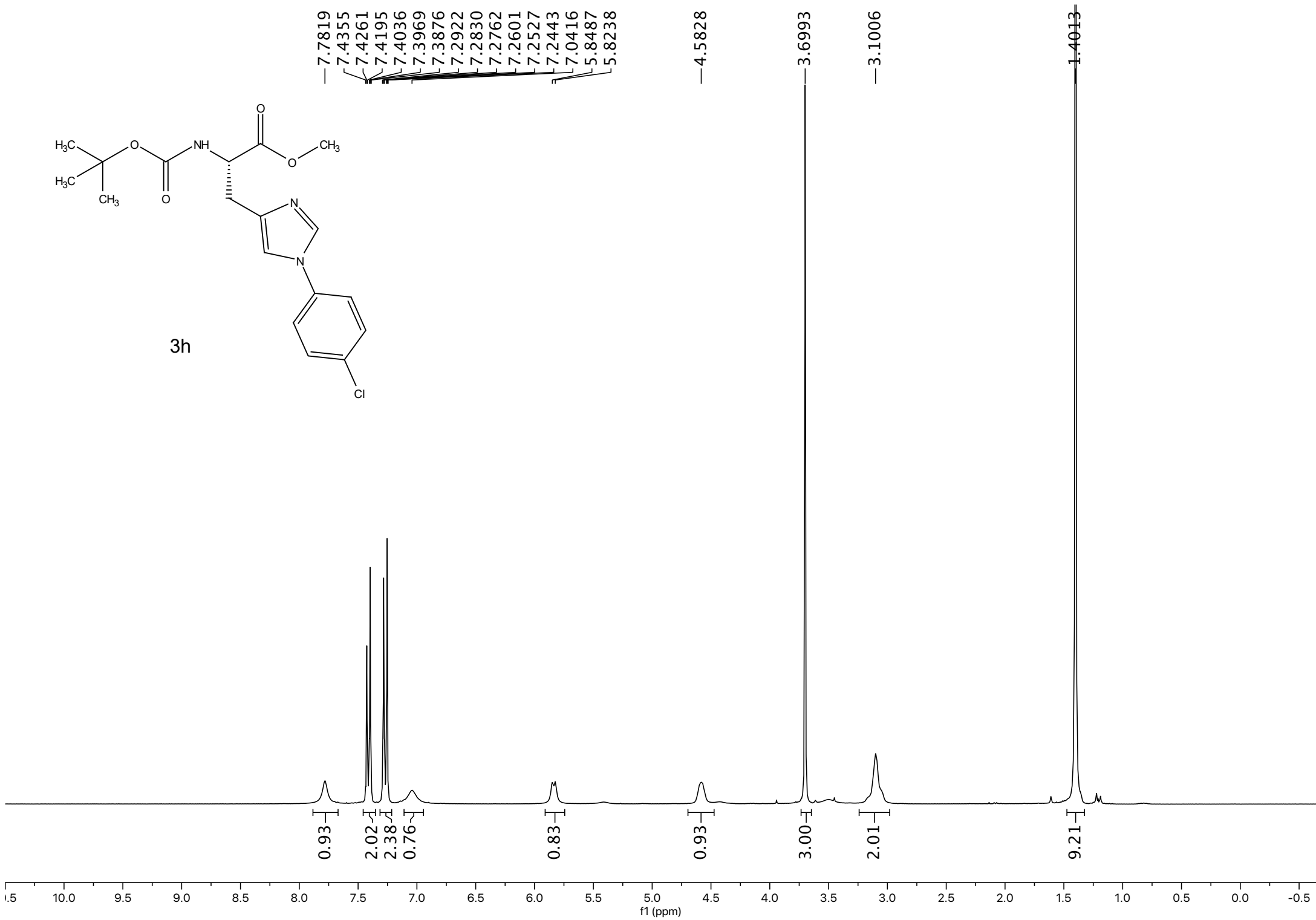
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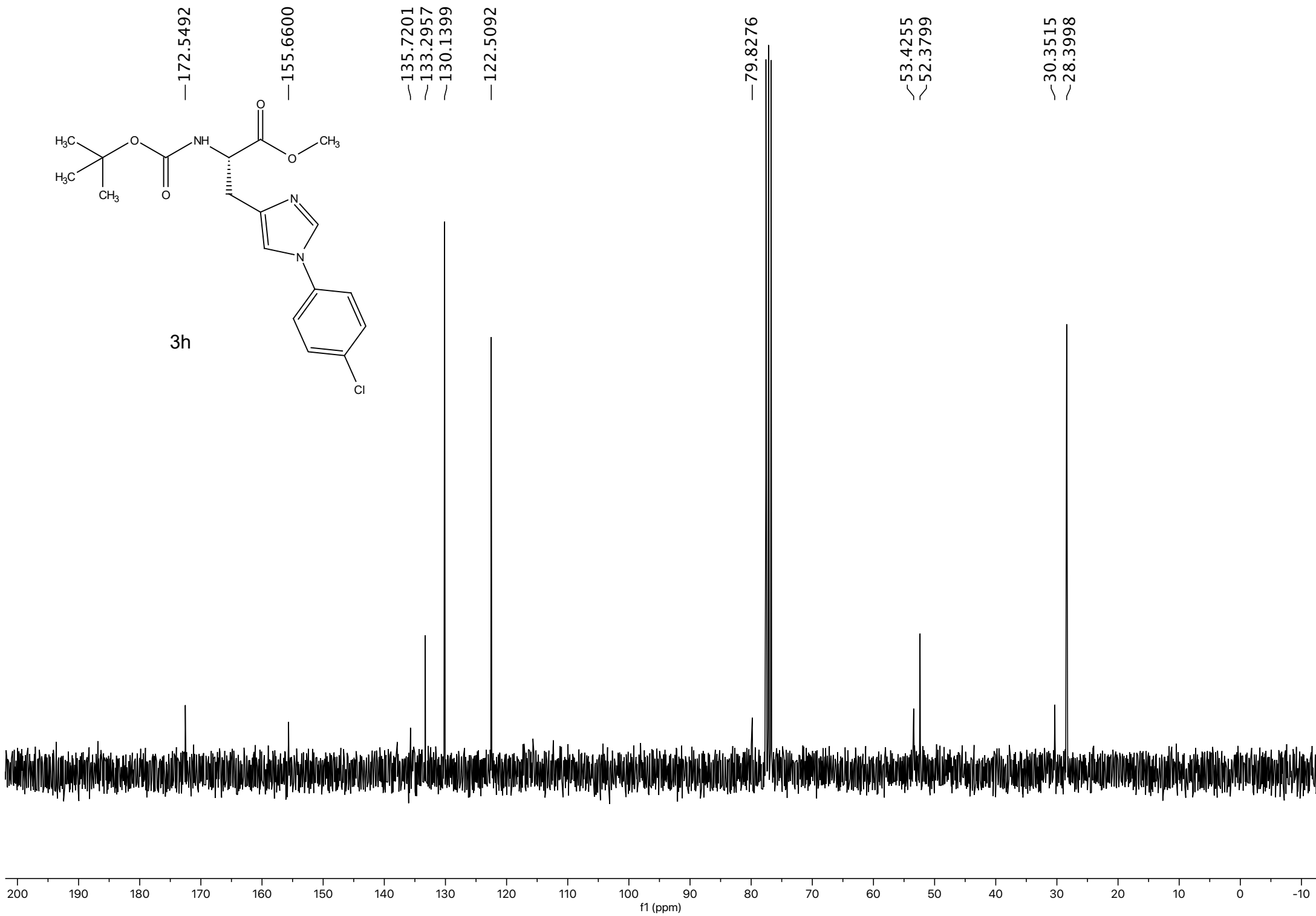
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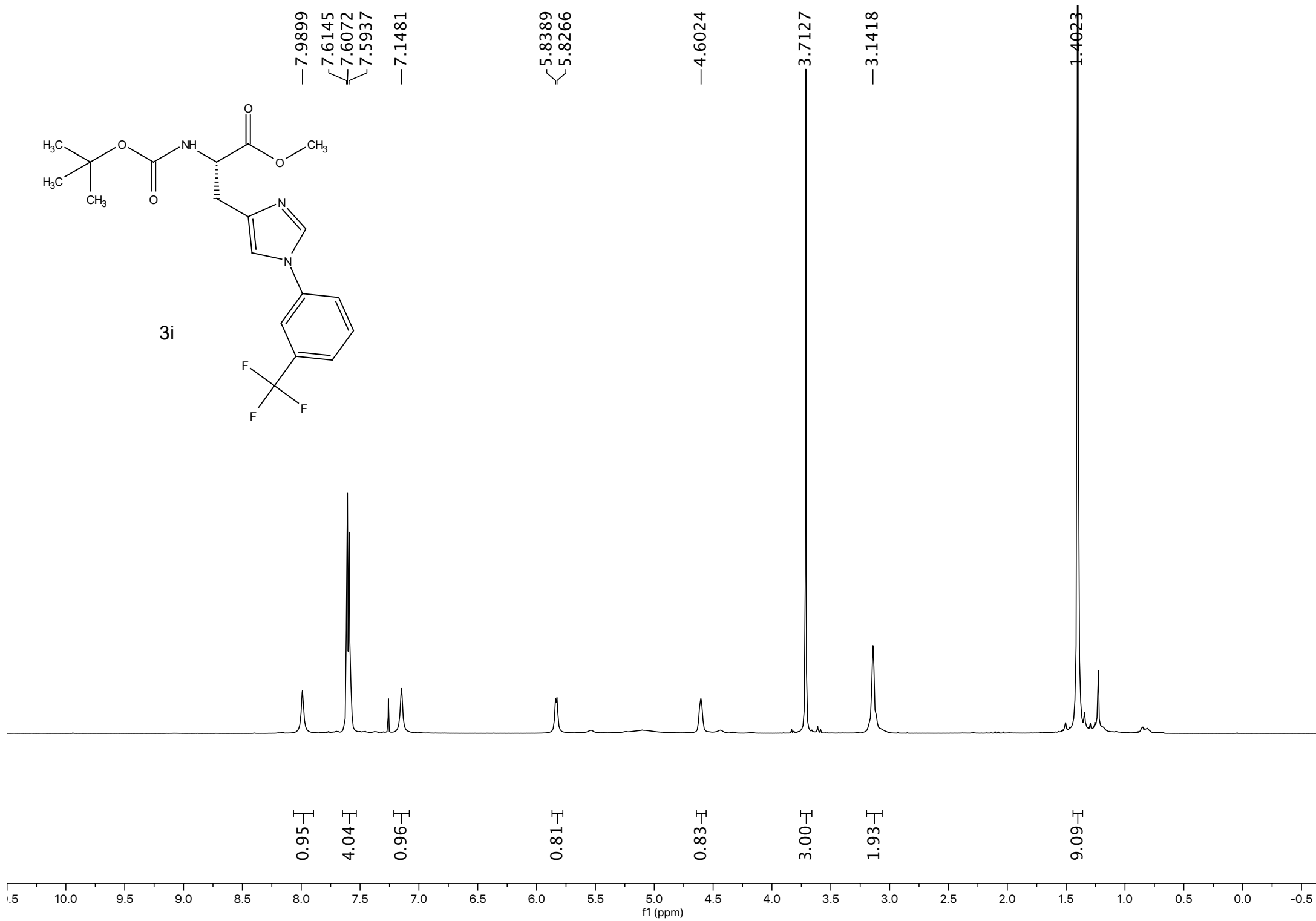
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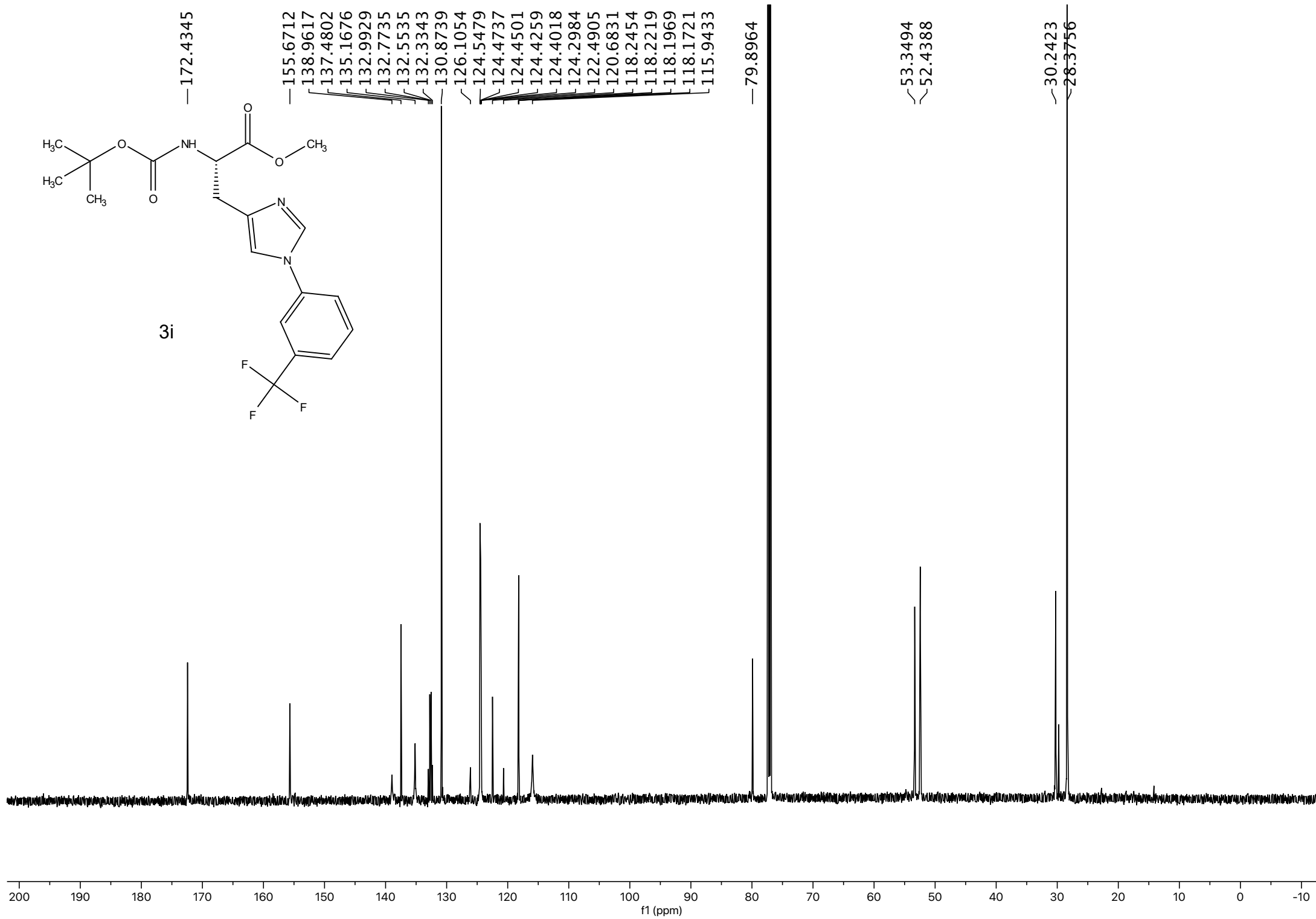


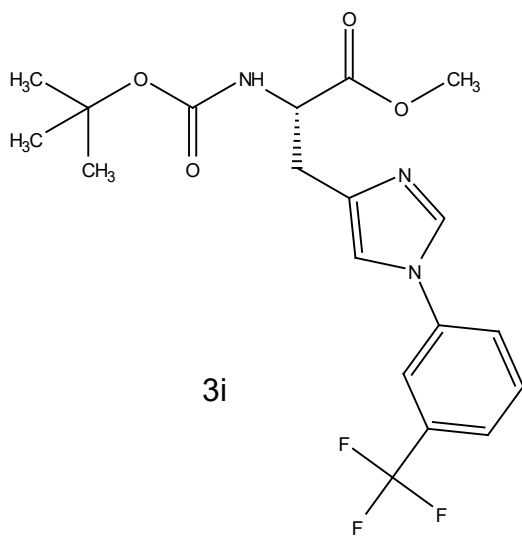






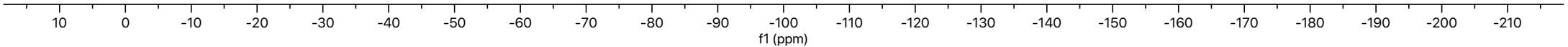




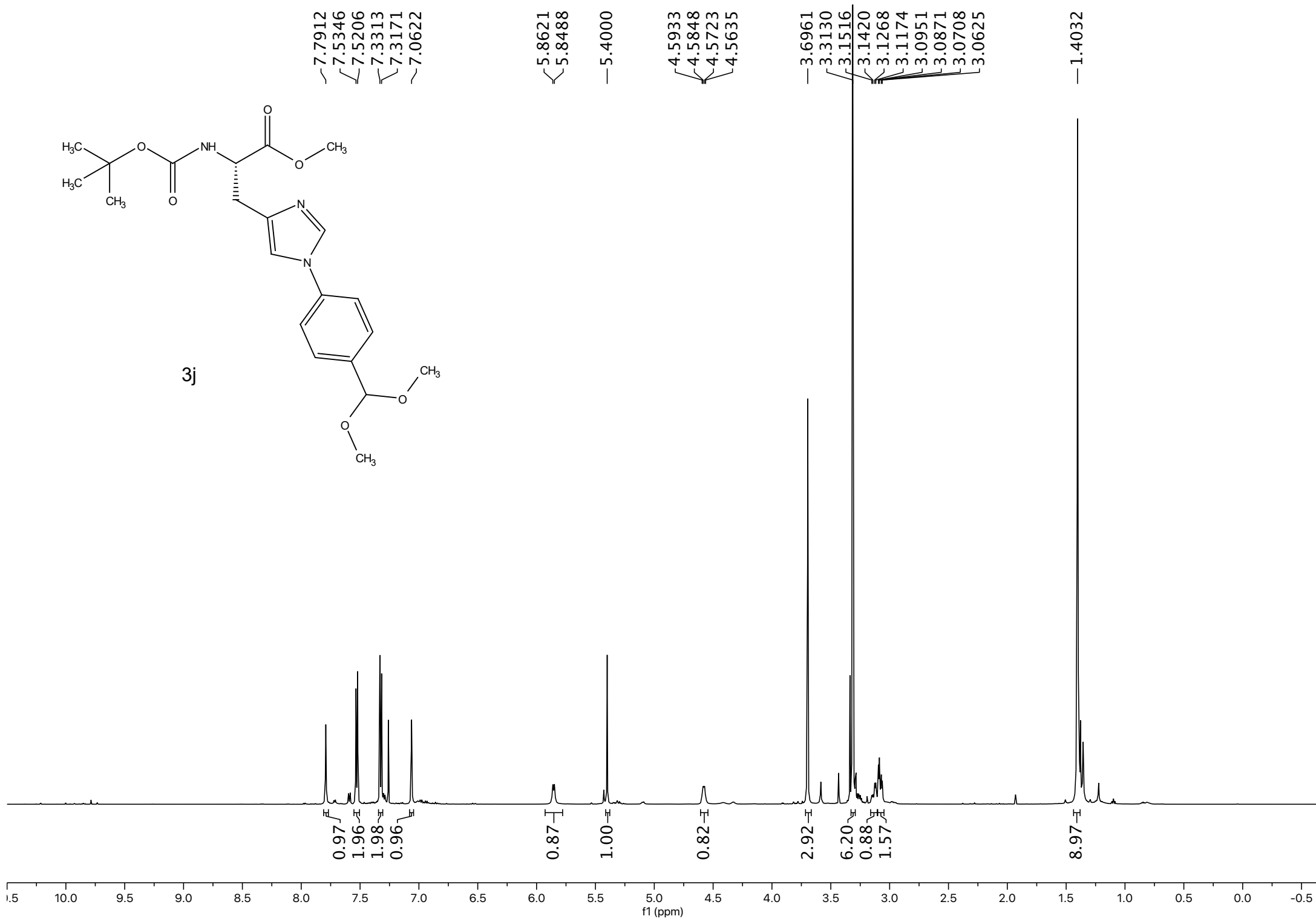
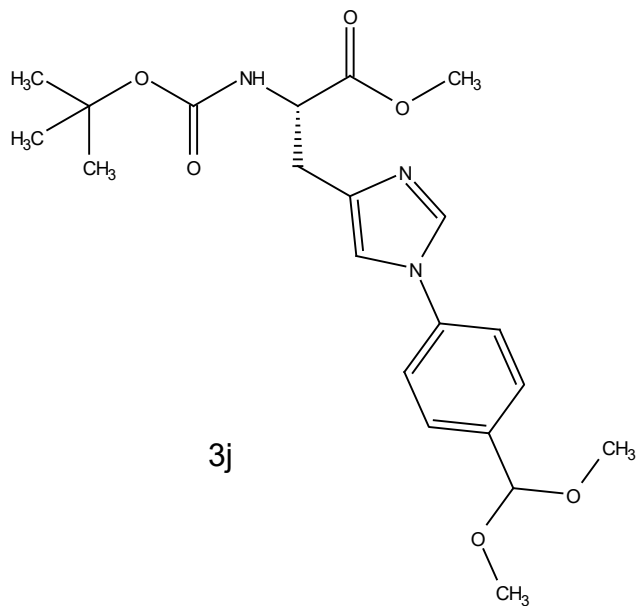


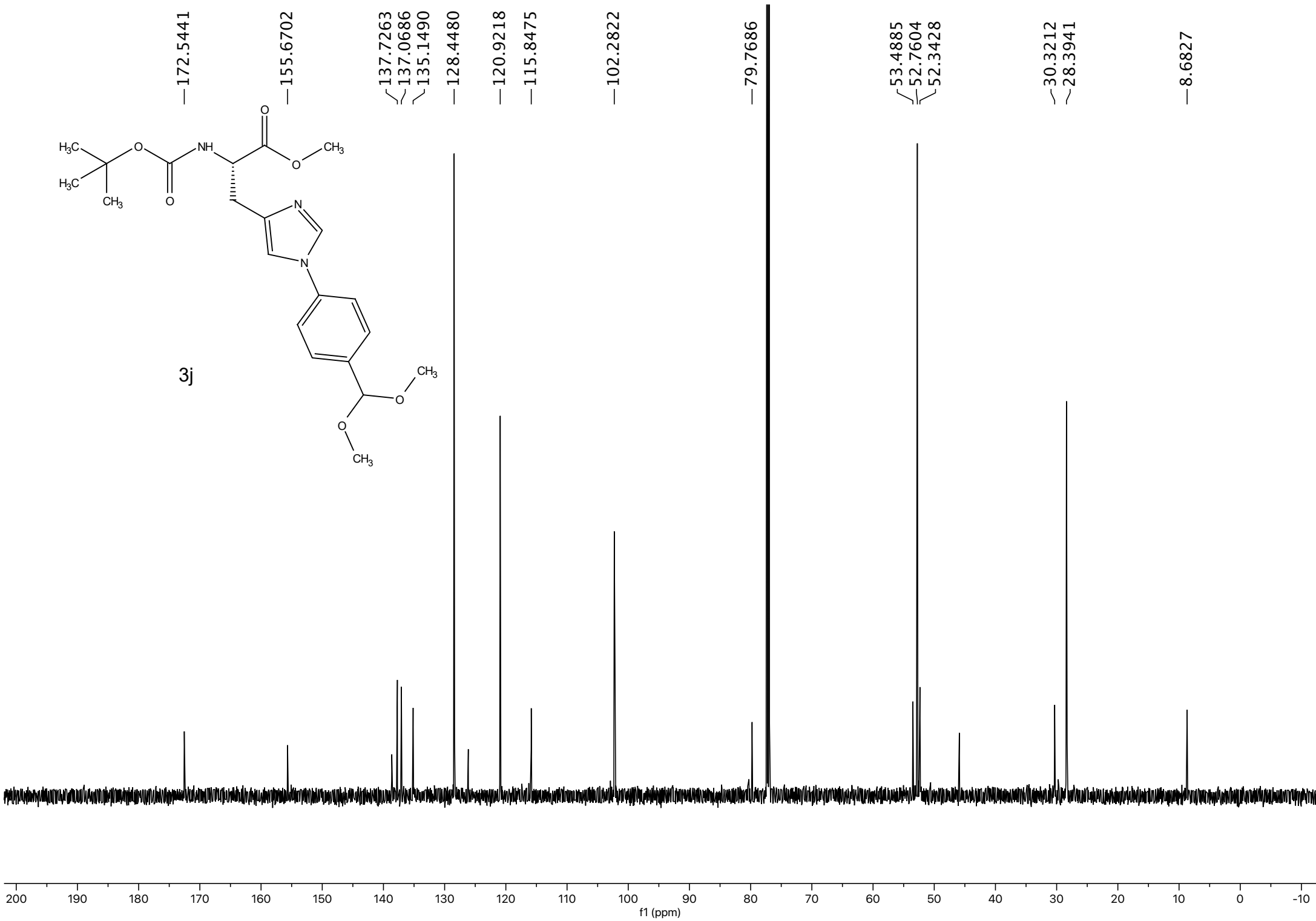
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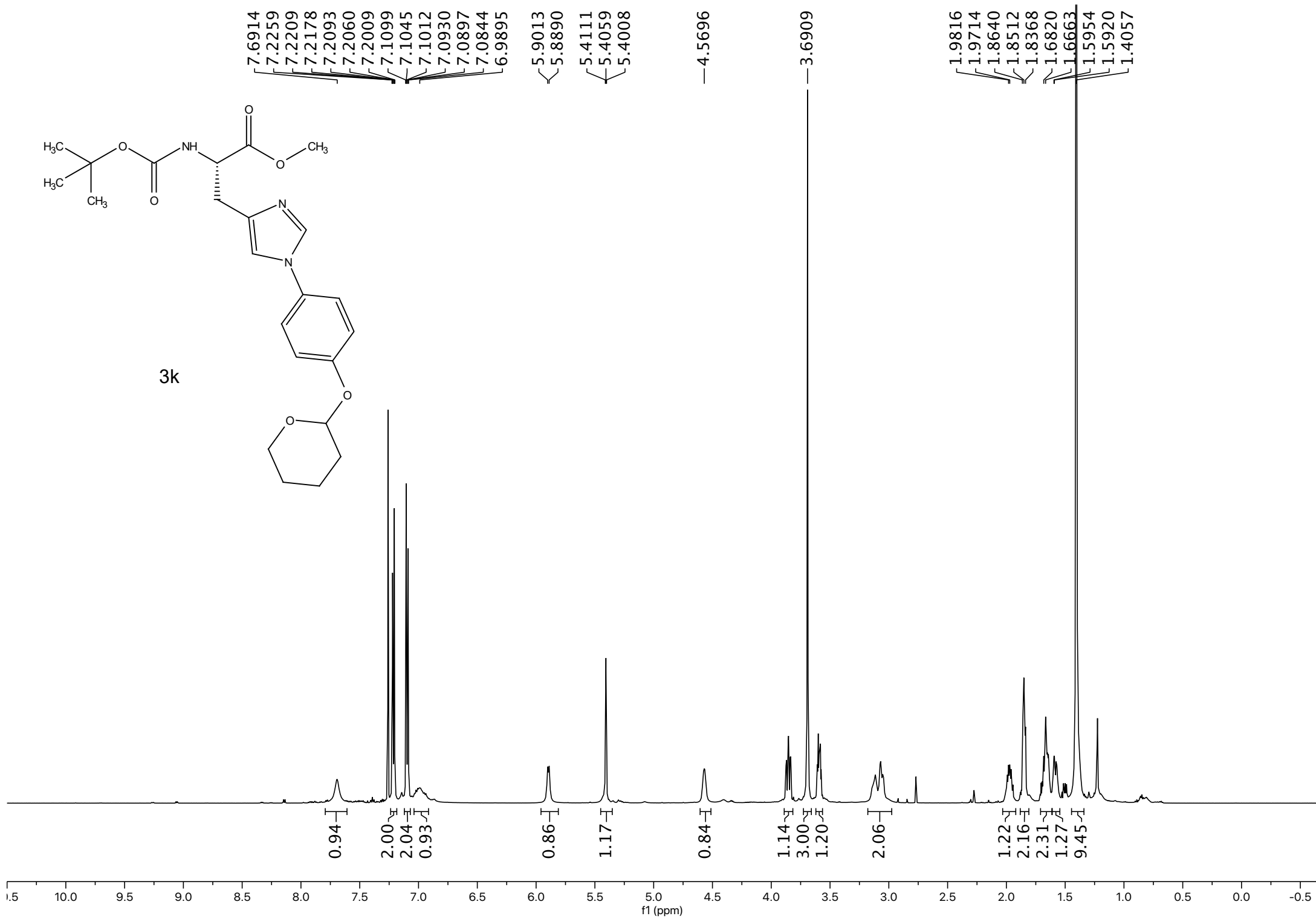
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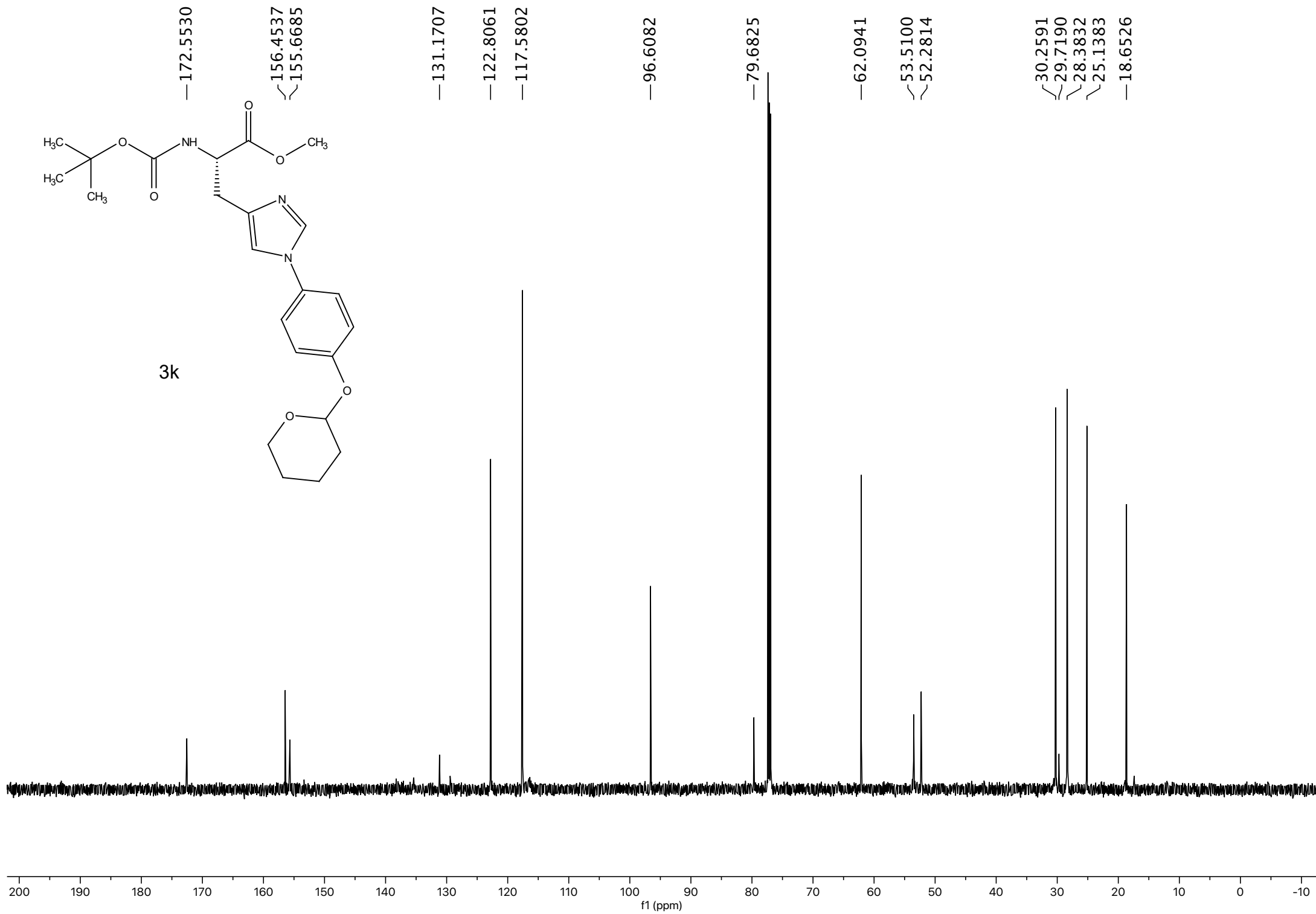


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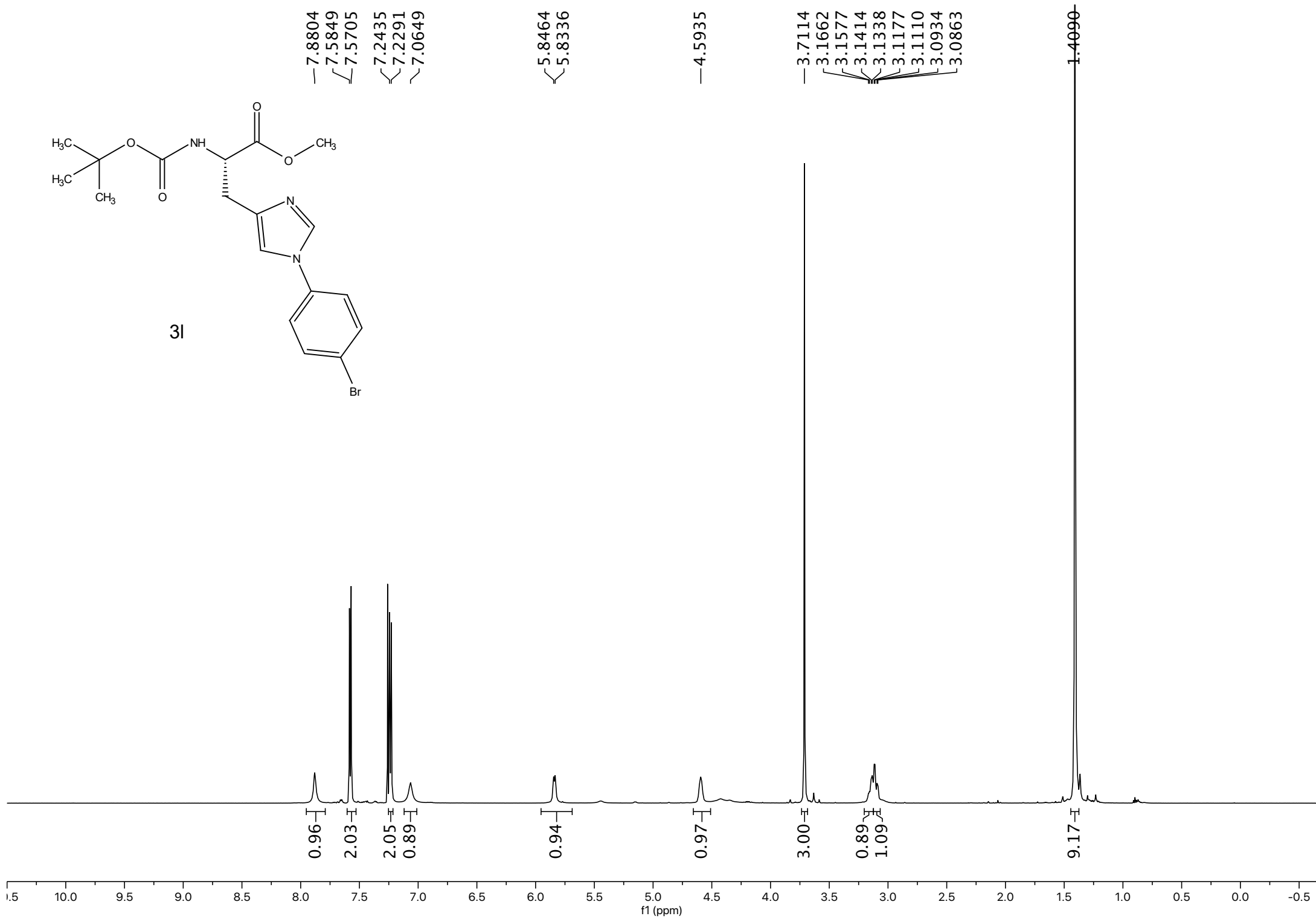
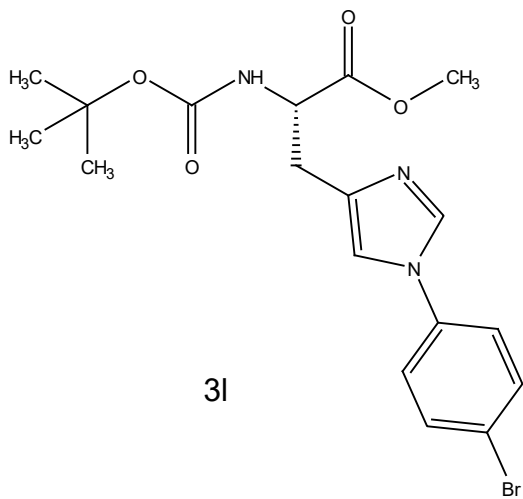


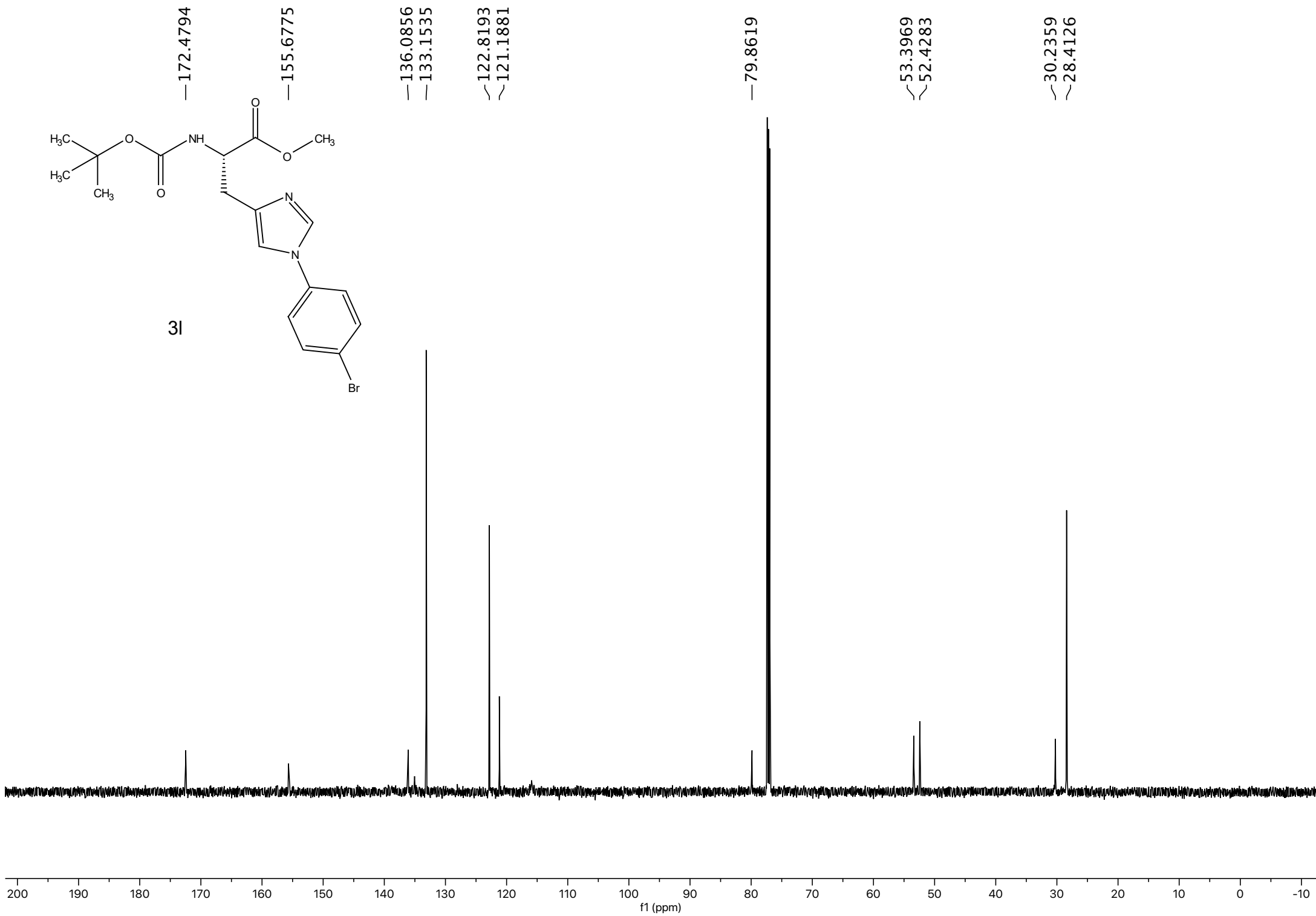


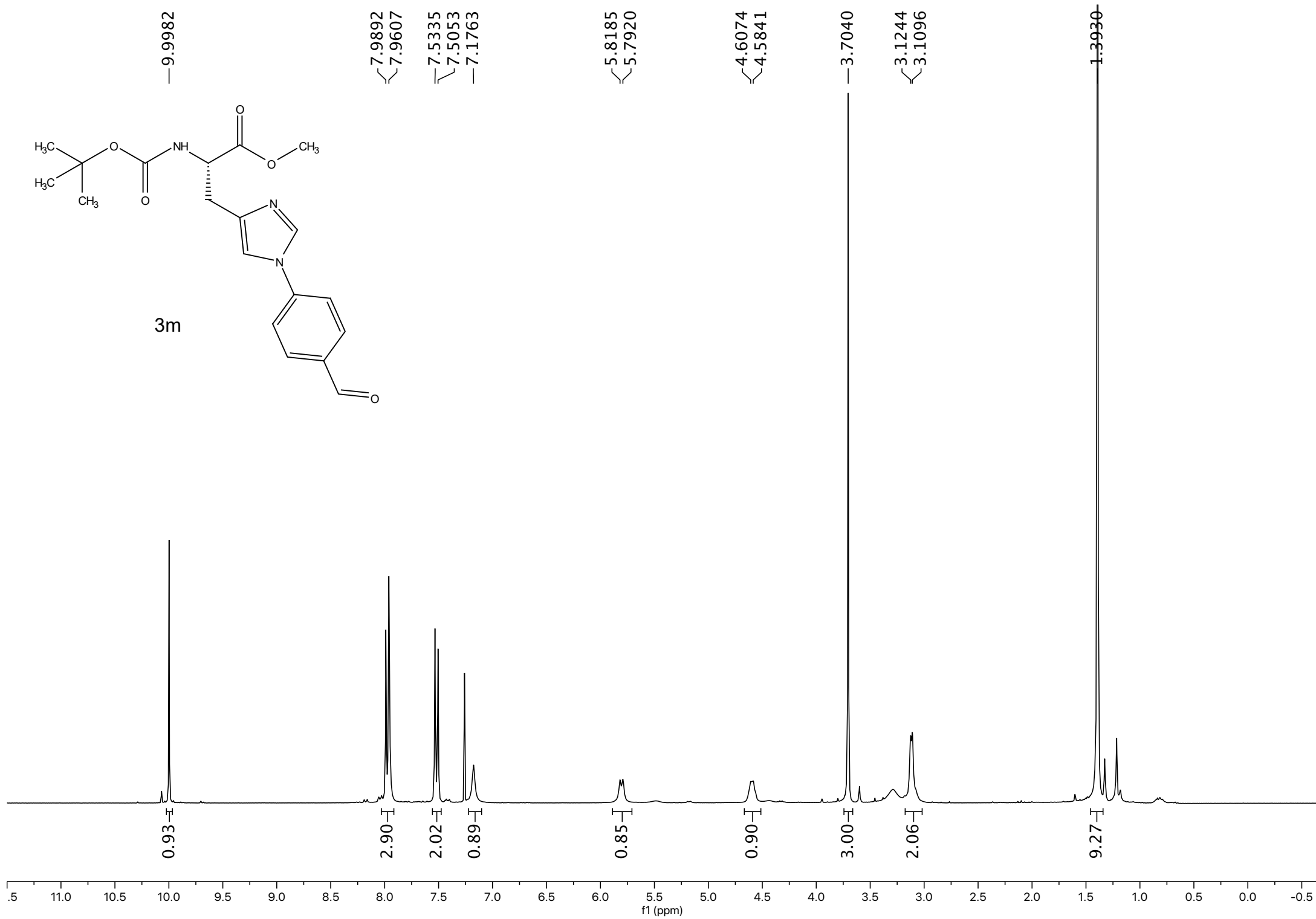


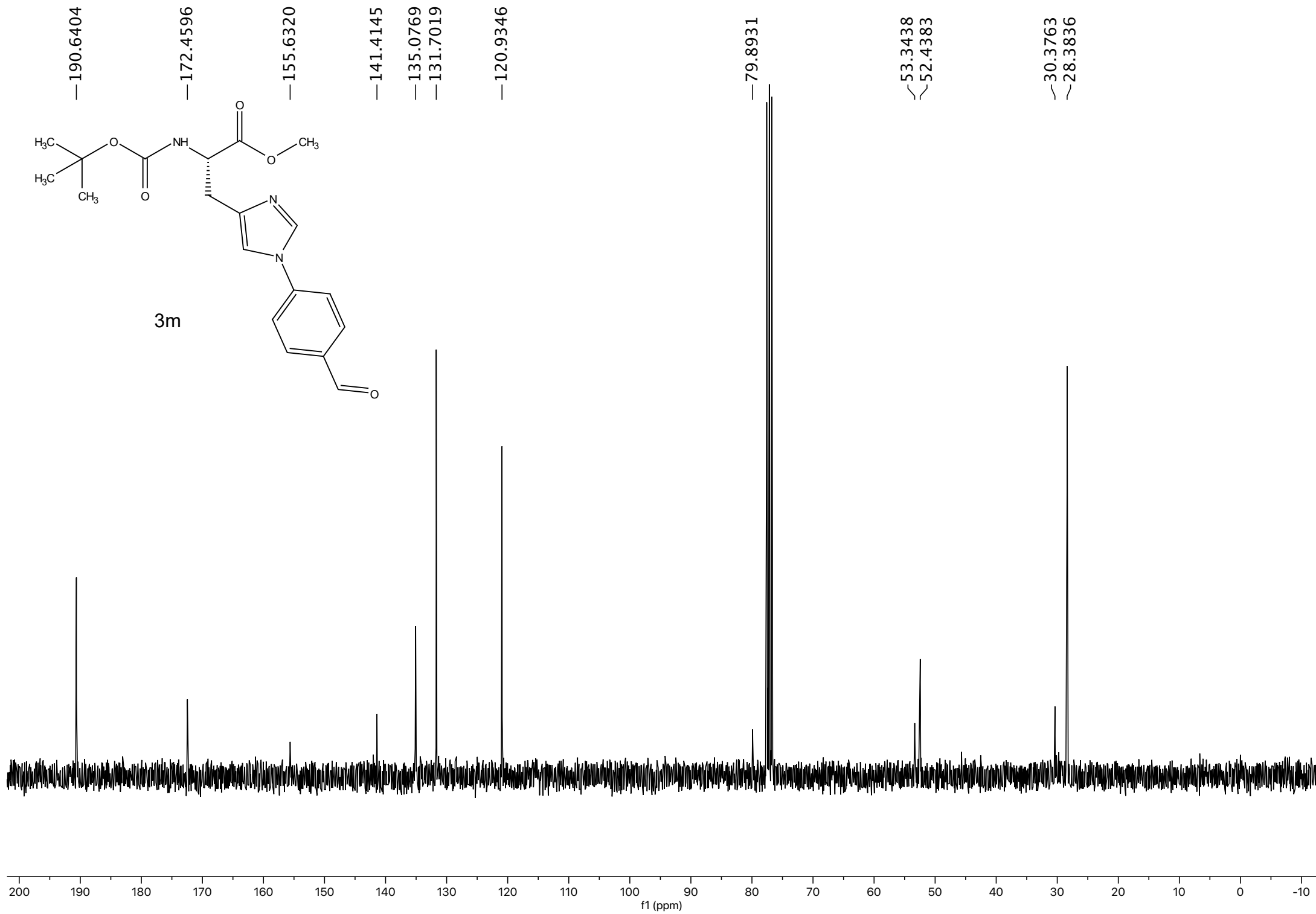


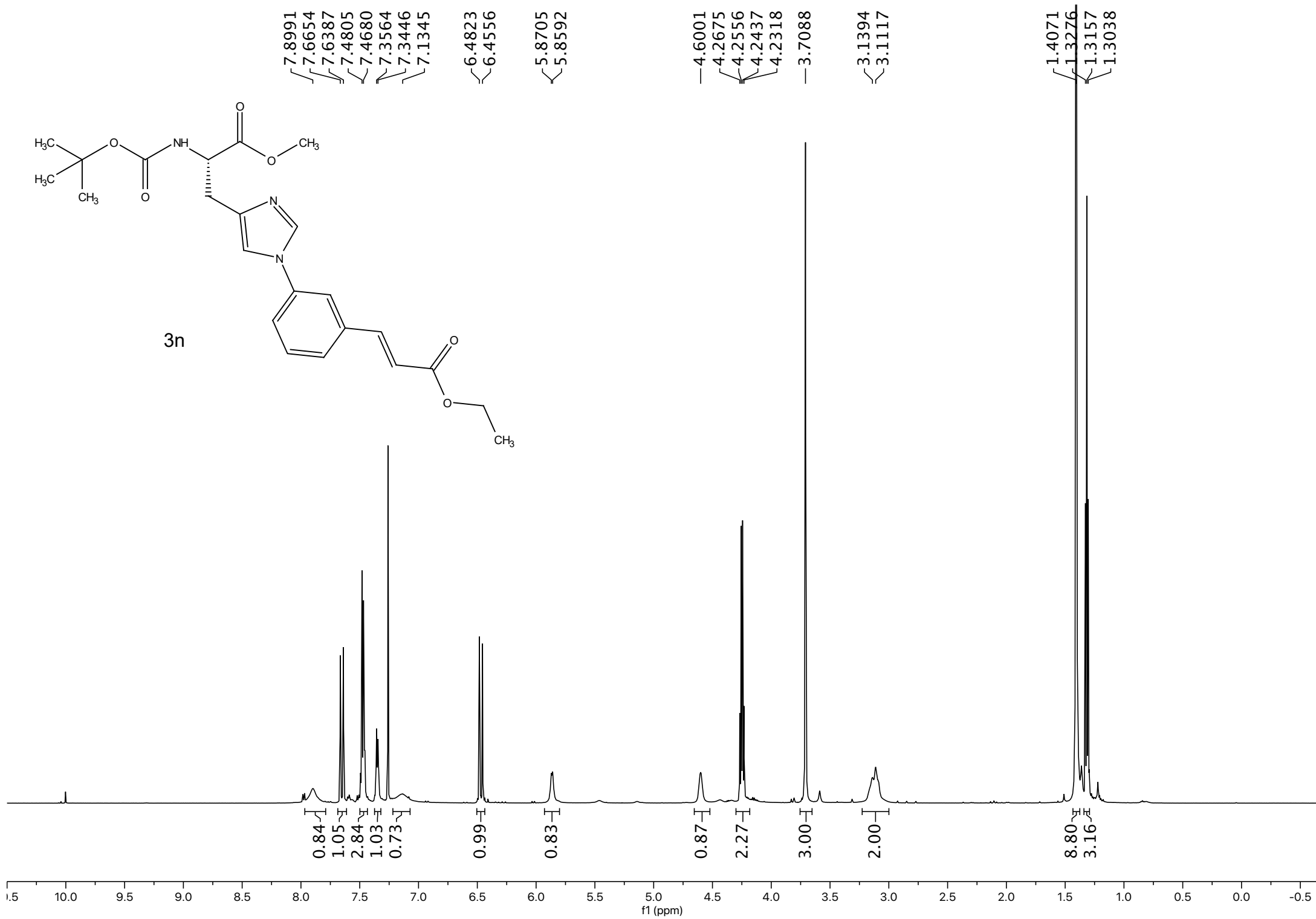
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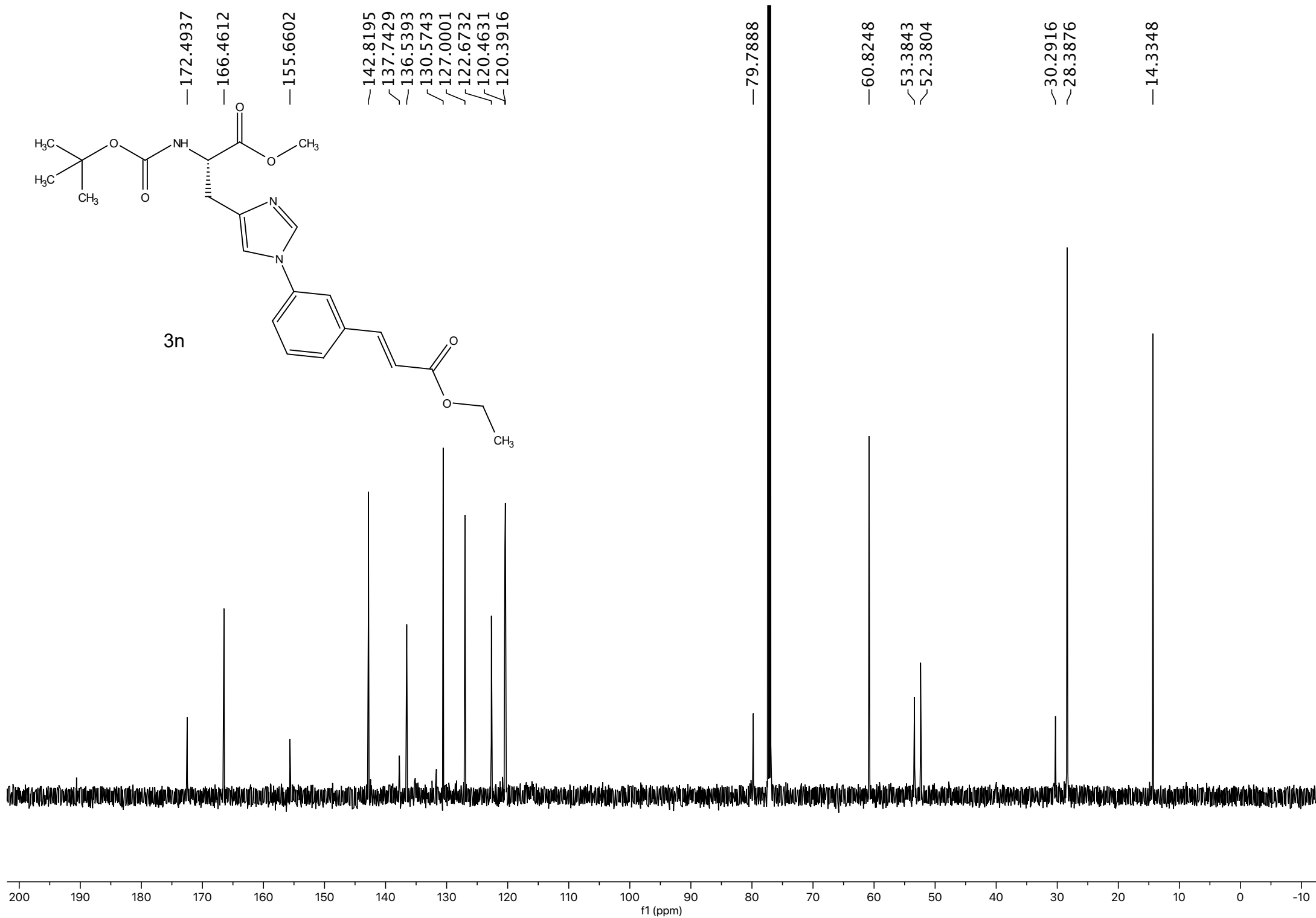


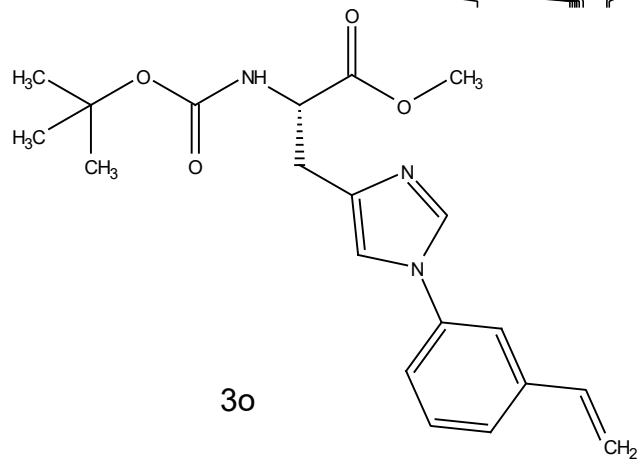




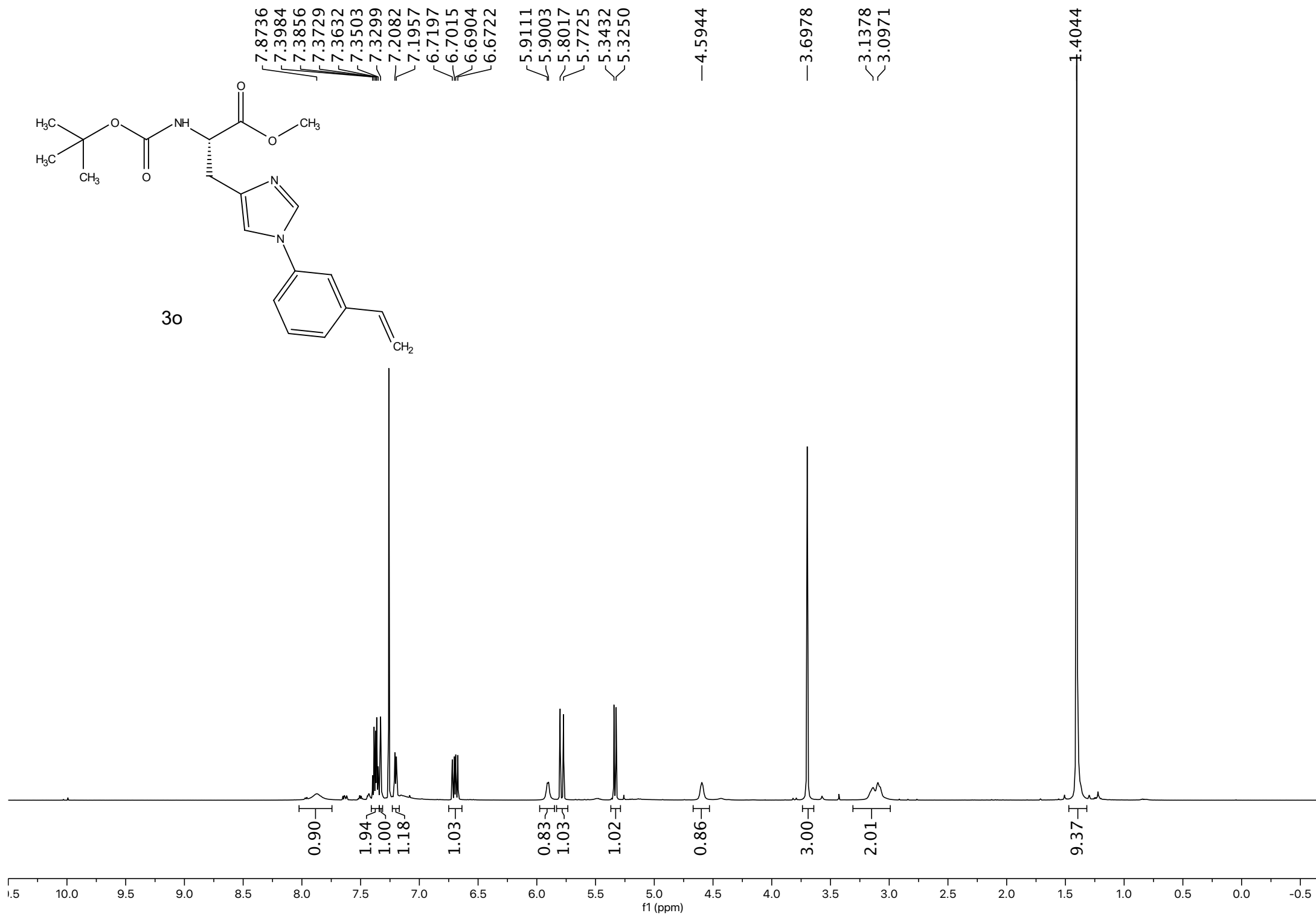


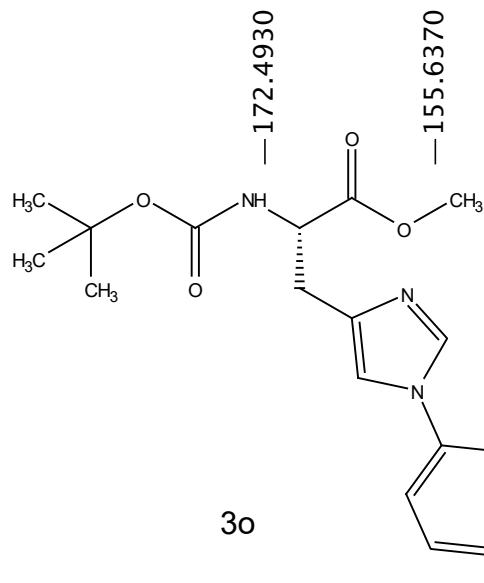




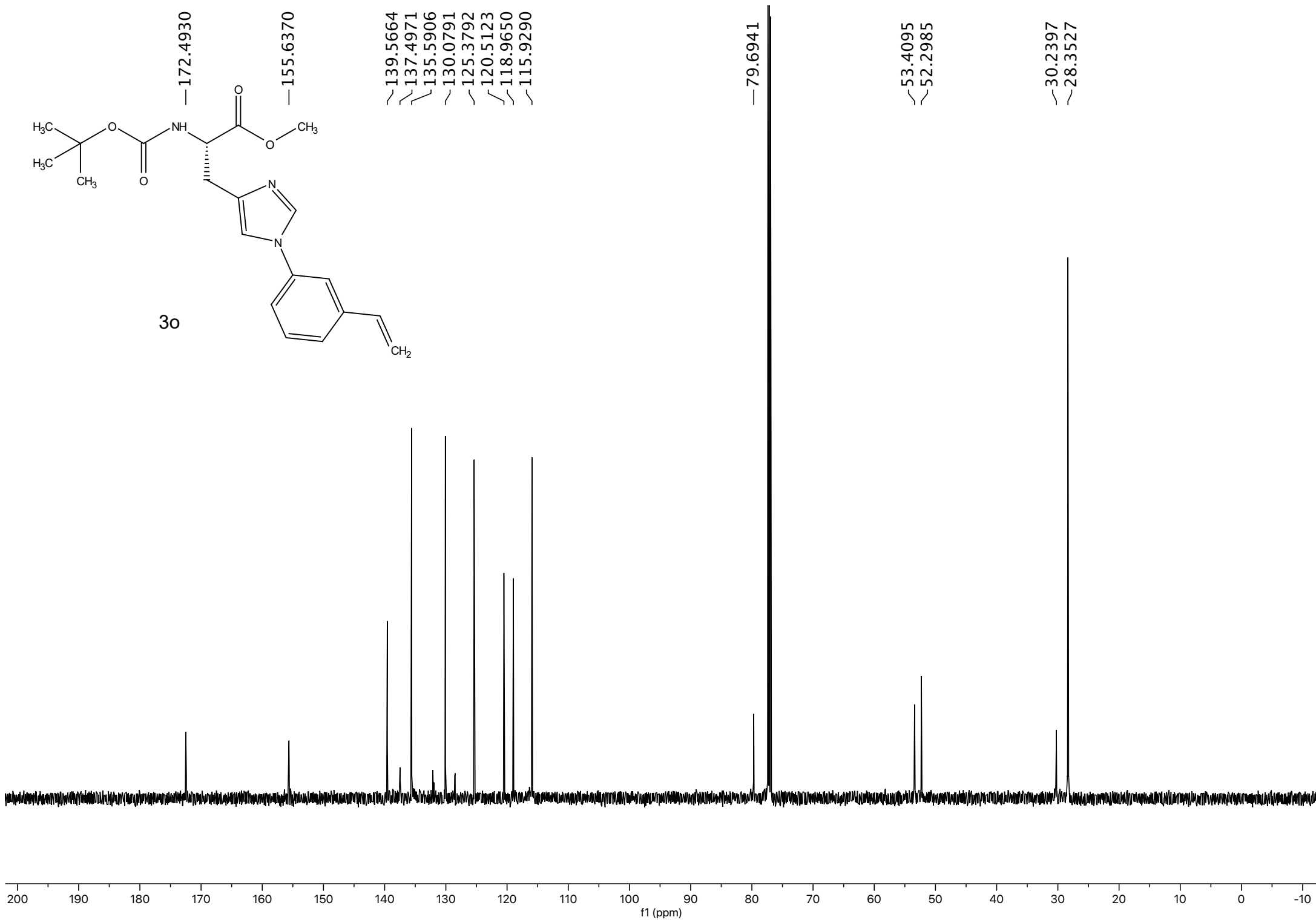


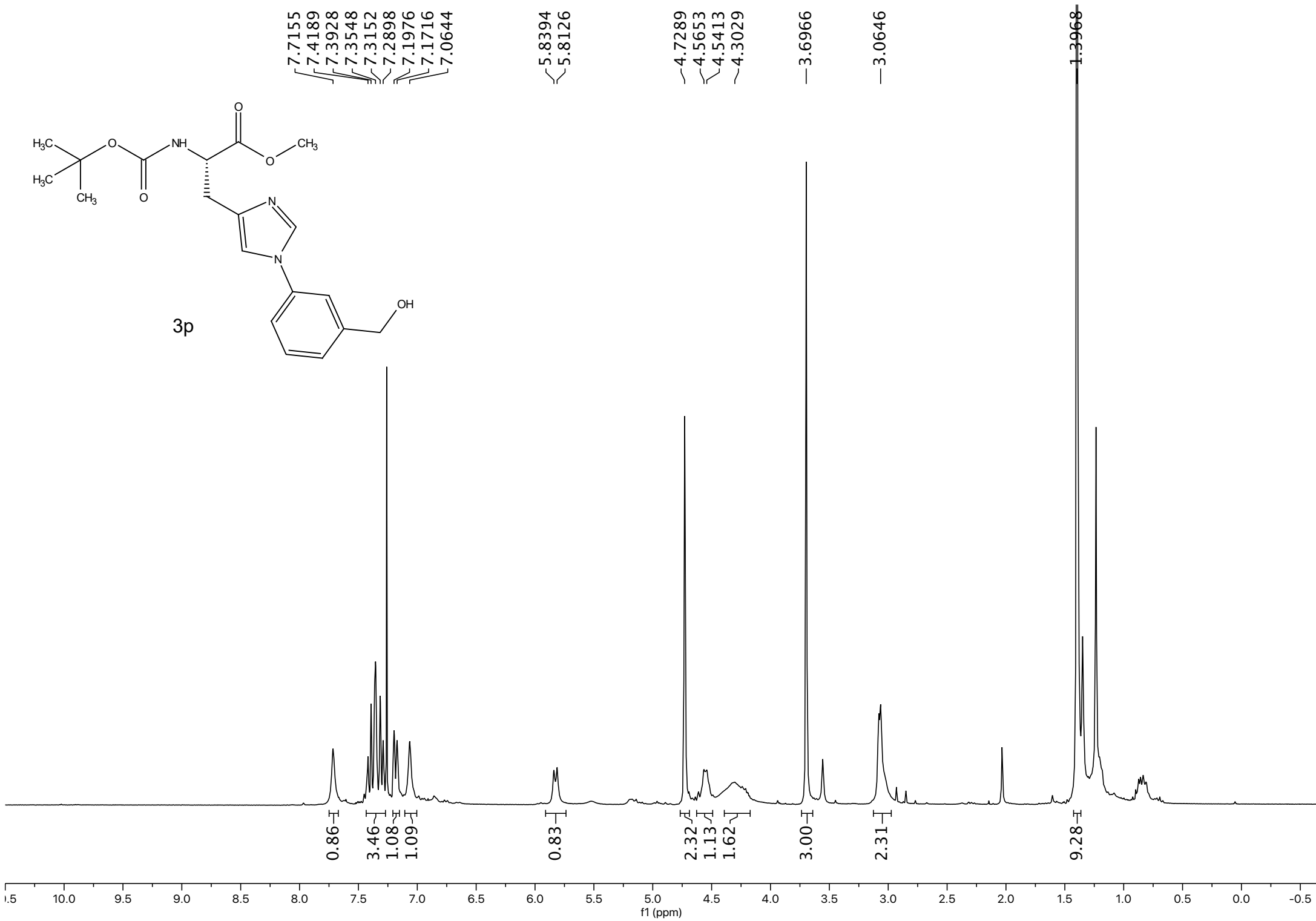
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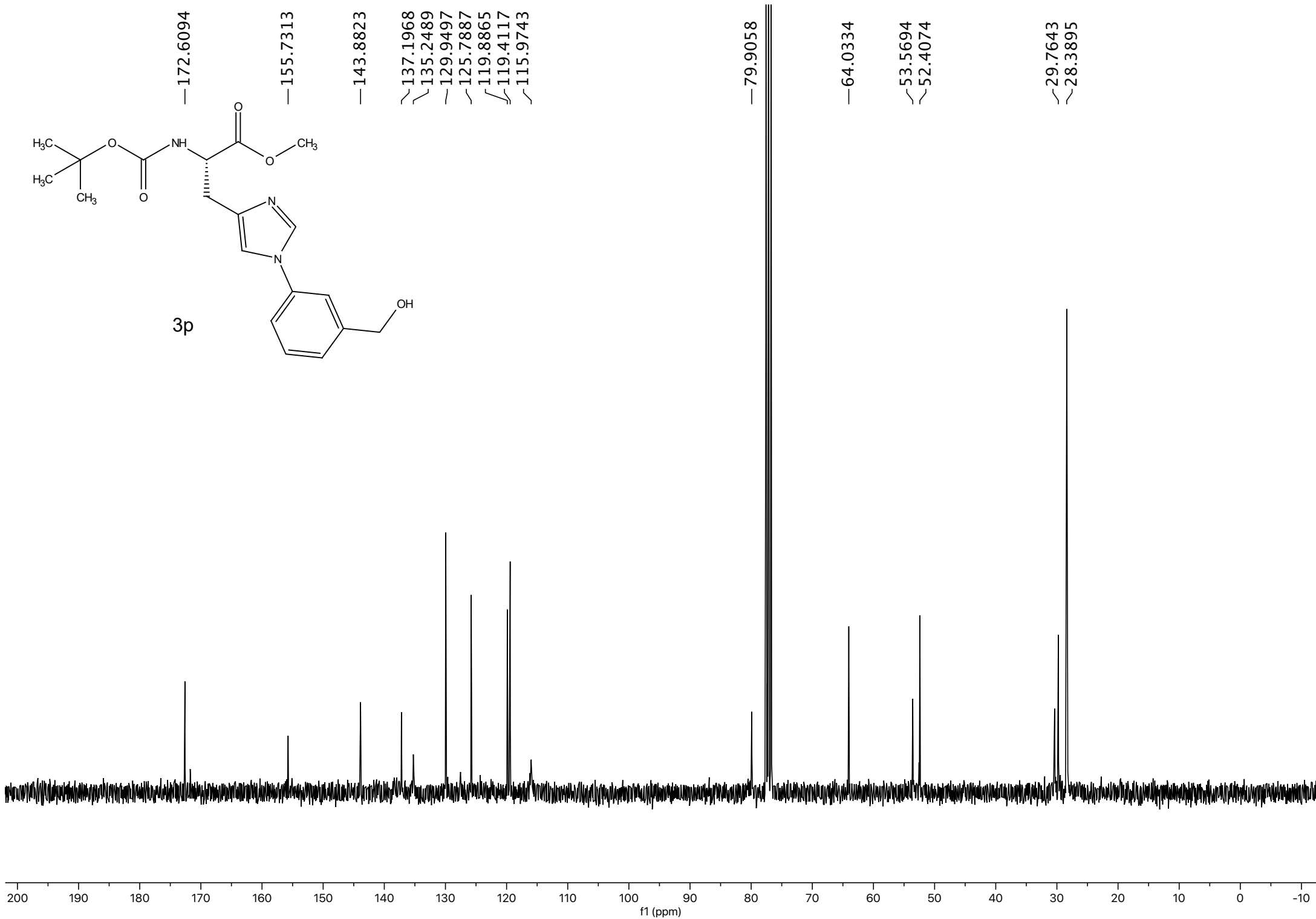
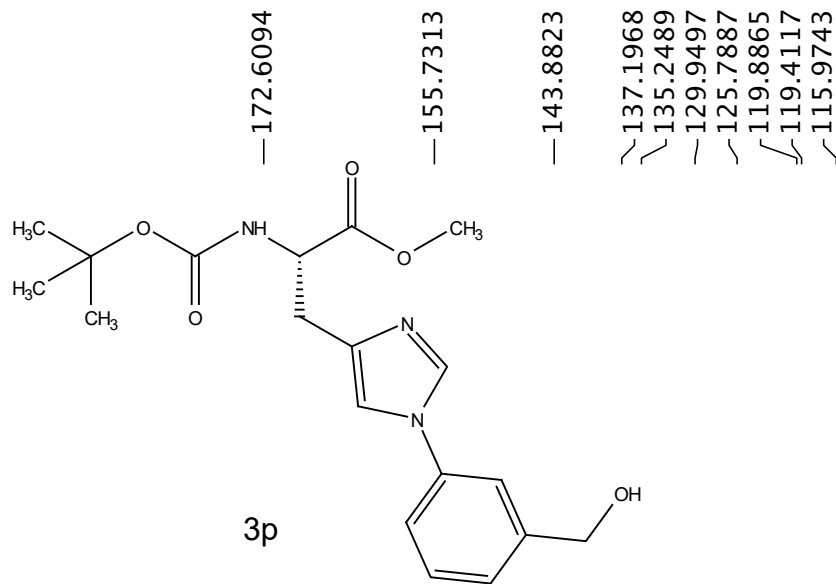


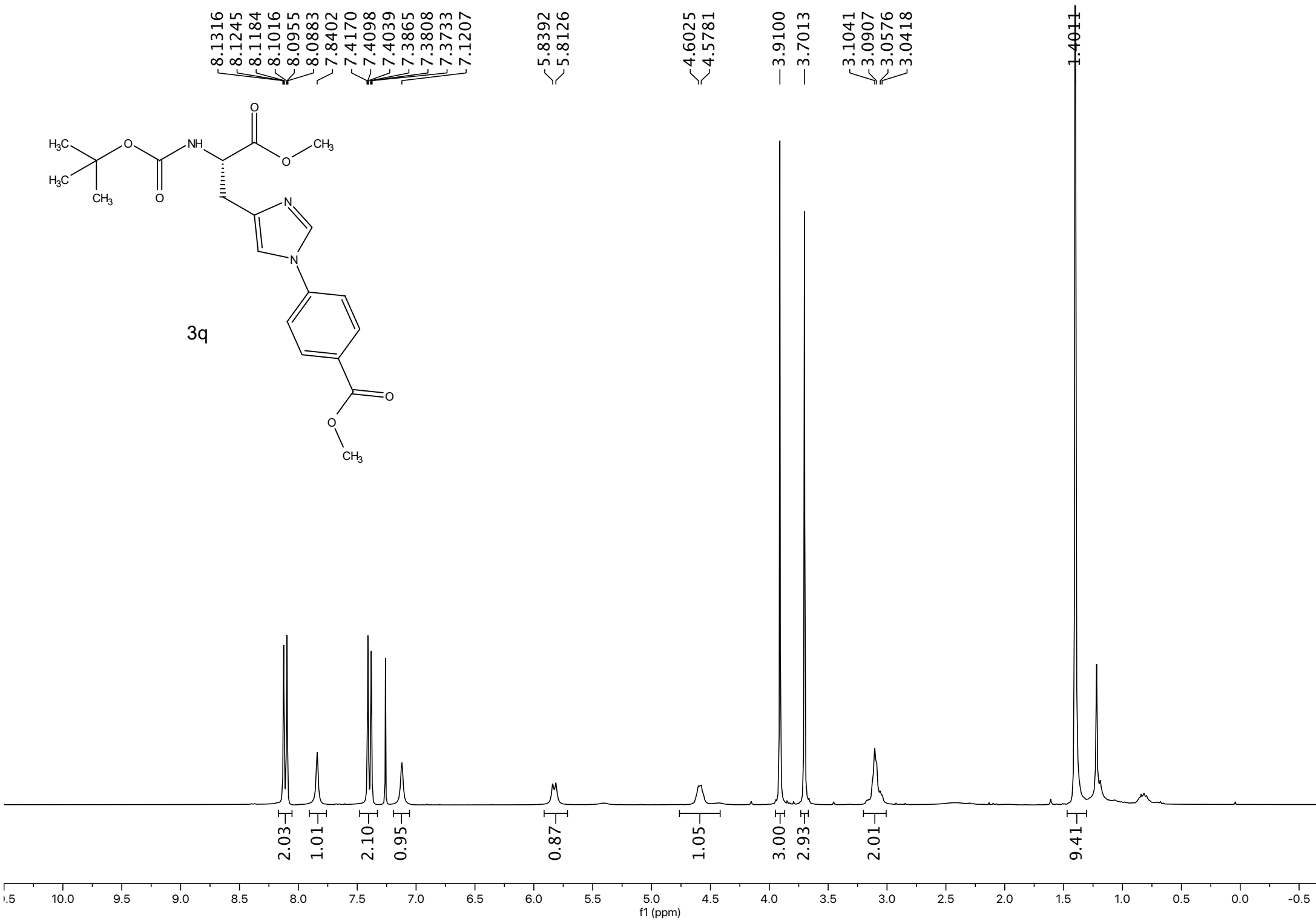


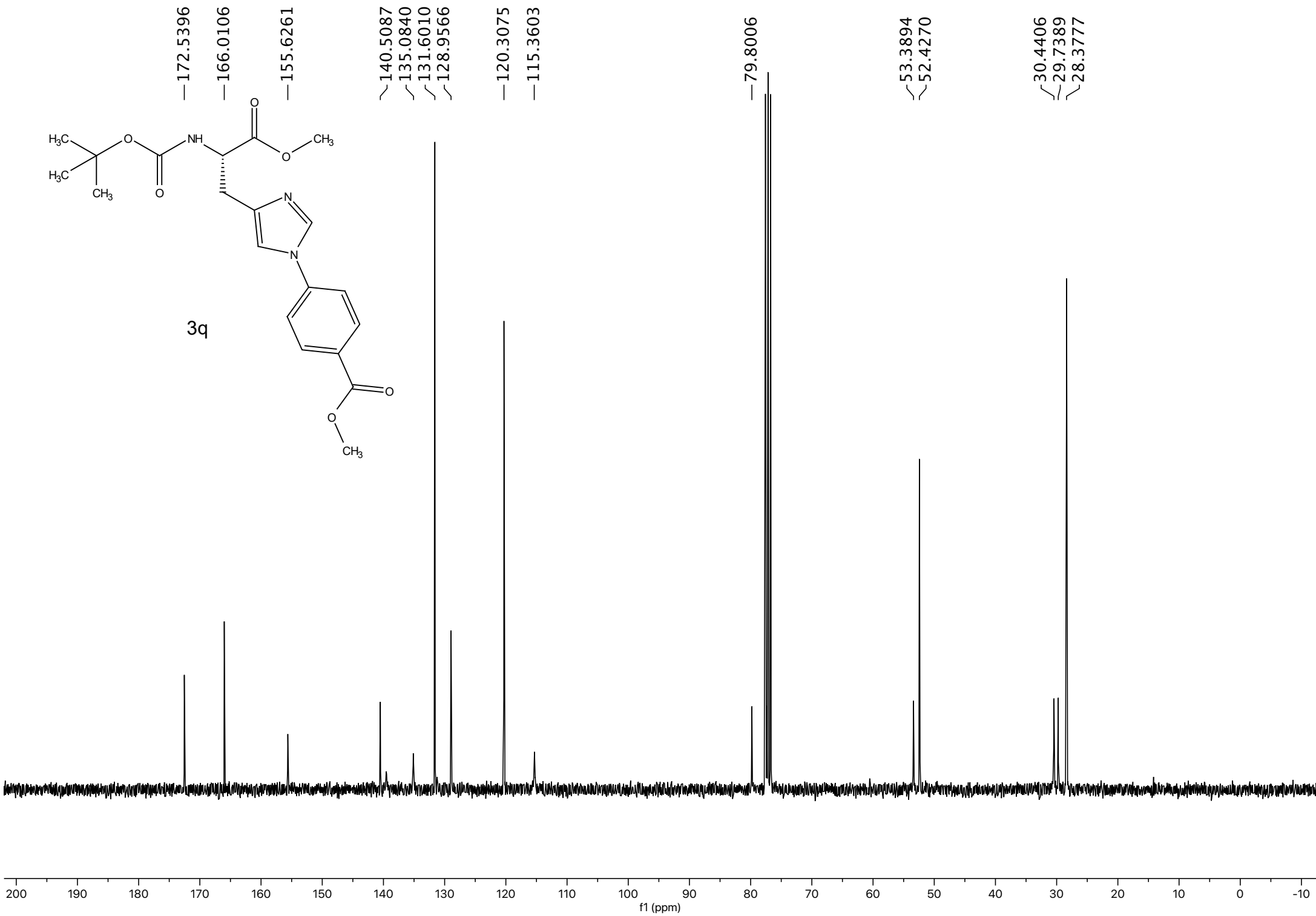
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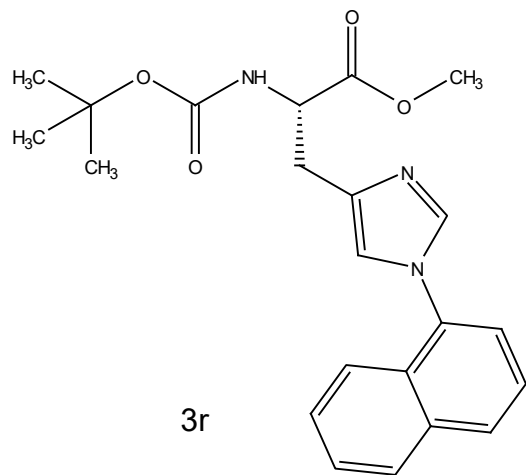








3r



7.9544
7.9297
7.9241
7.7089
7.5710
7.5632
7.5381
7.5228
7.5200
7.4334
7.4117
7.0438

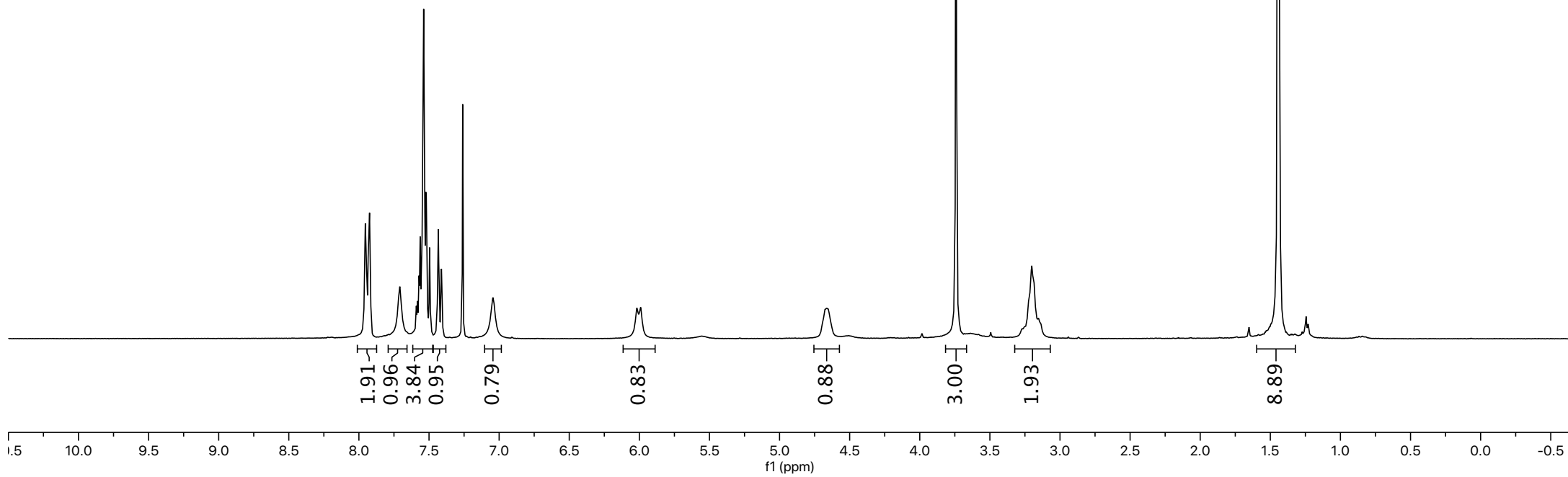
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5.9910

4.6628

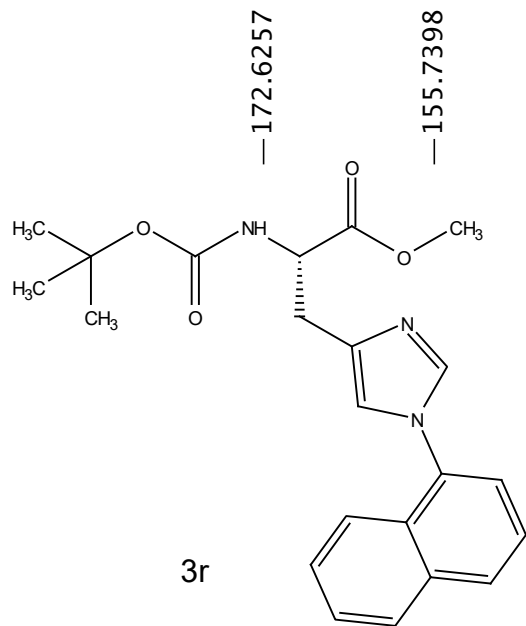
3.7414

3.2011

1.4430



3r



—172.6257

—155.7398

134.2621

133.8514

129.4955

129.4614

128.4322

127.7798

127.1247

125.2625

123.6768

122.2801

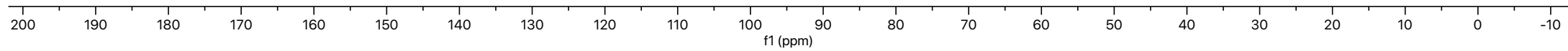
—79.8016

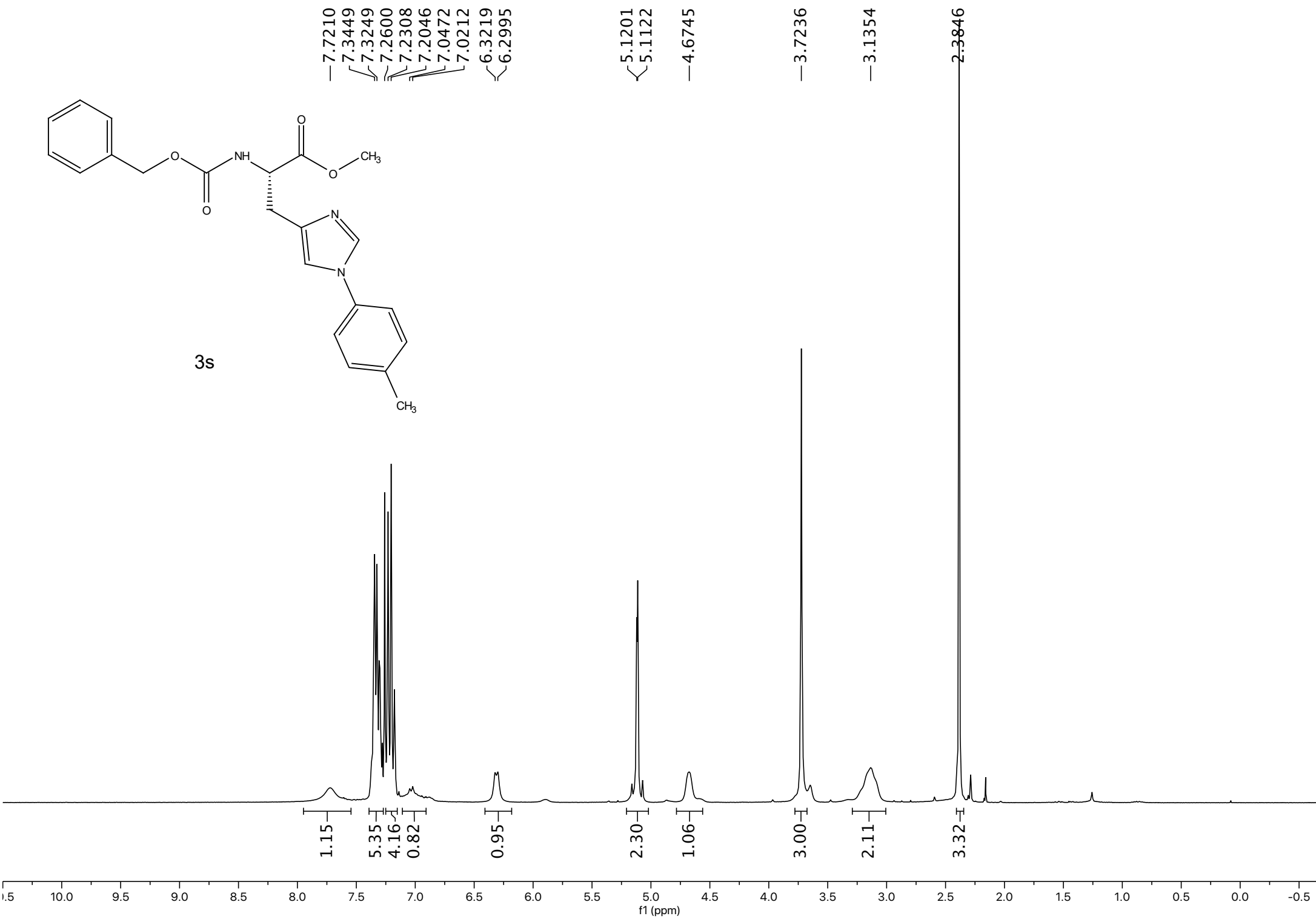
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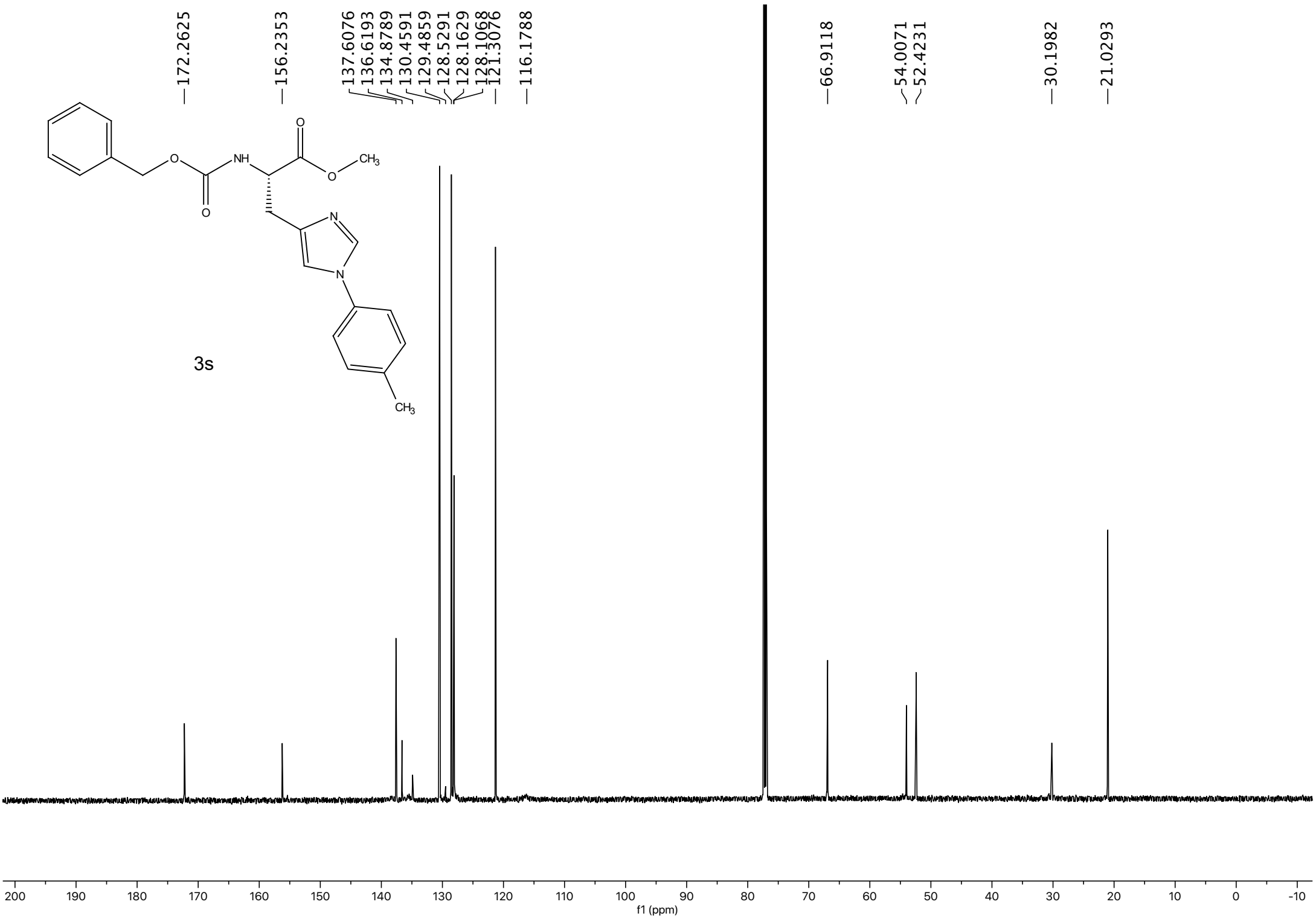
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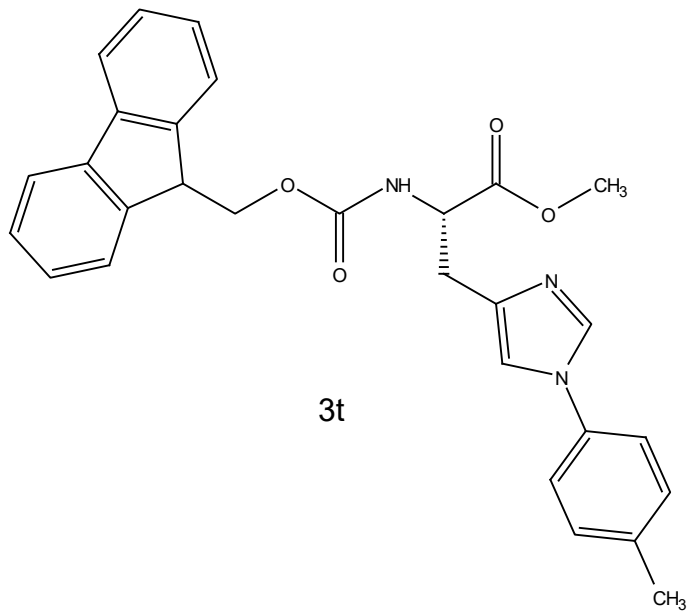
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~28.4646

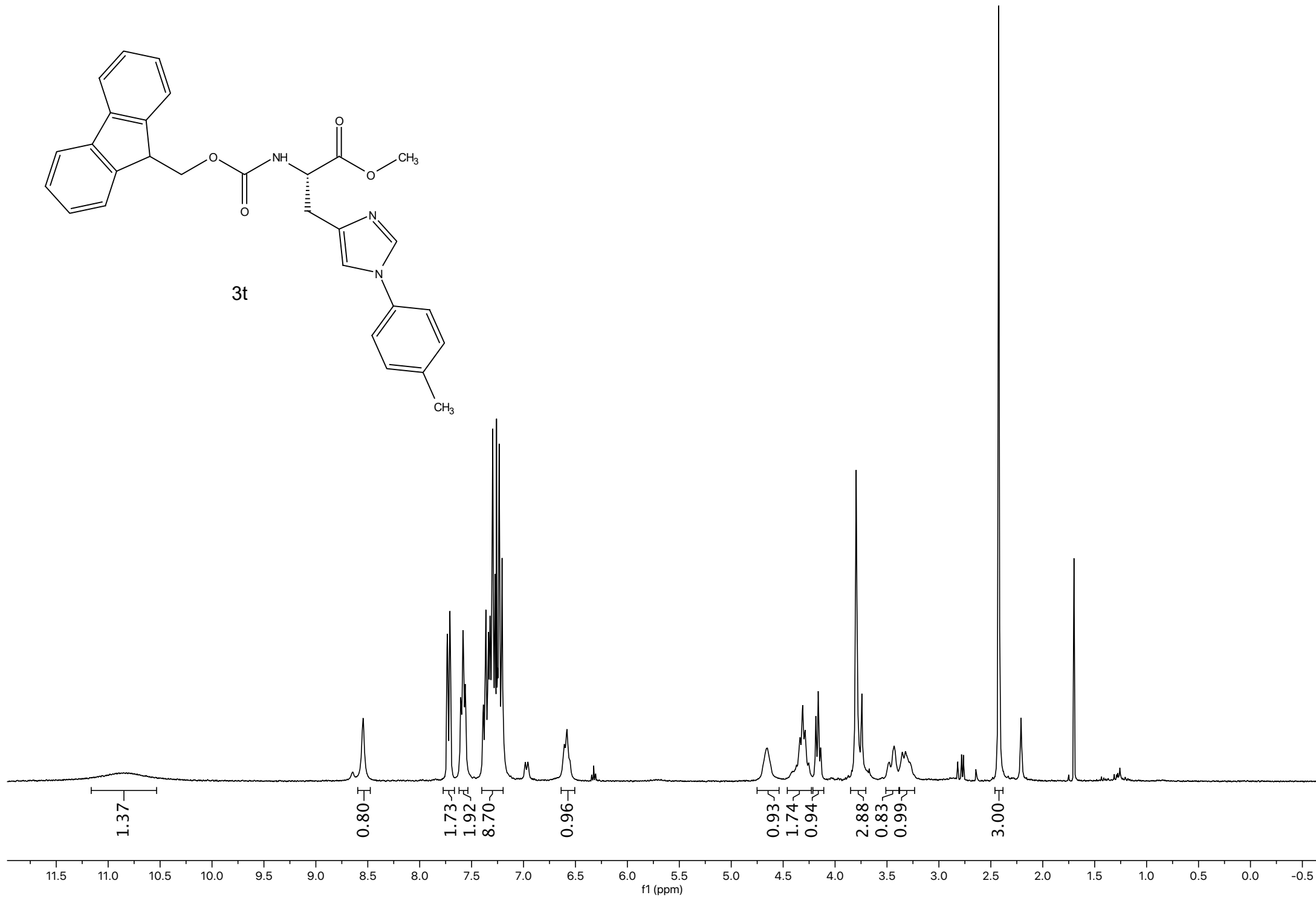


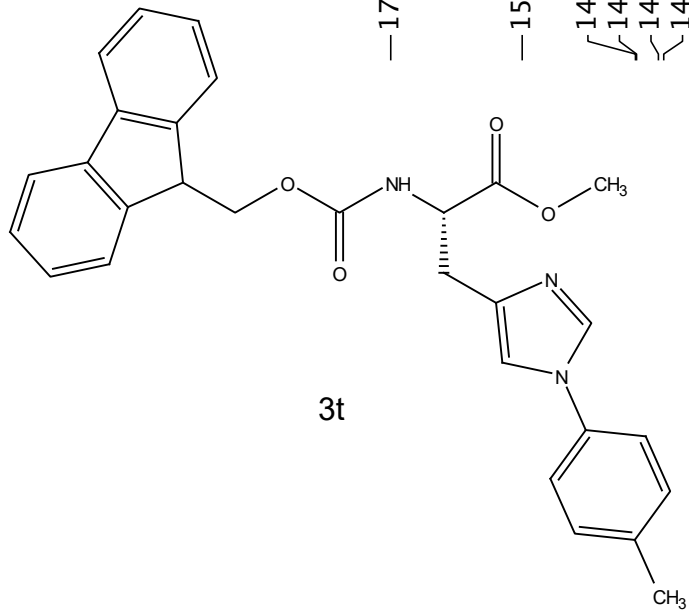




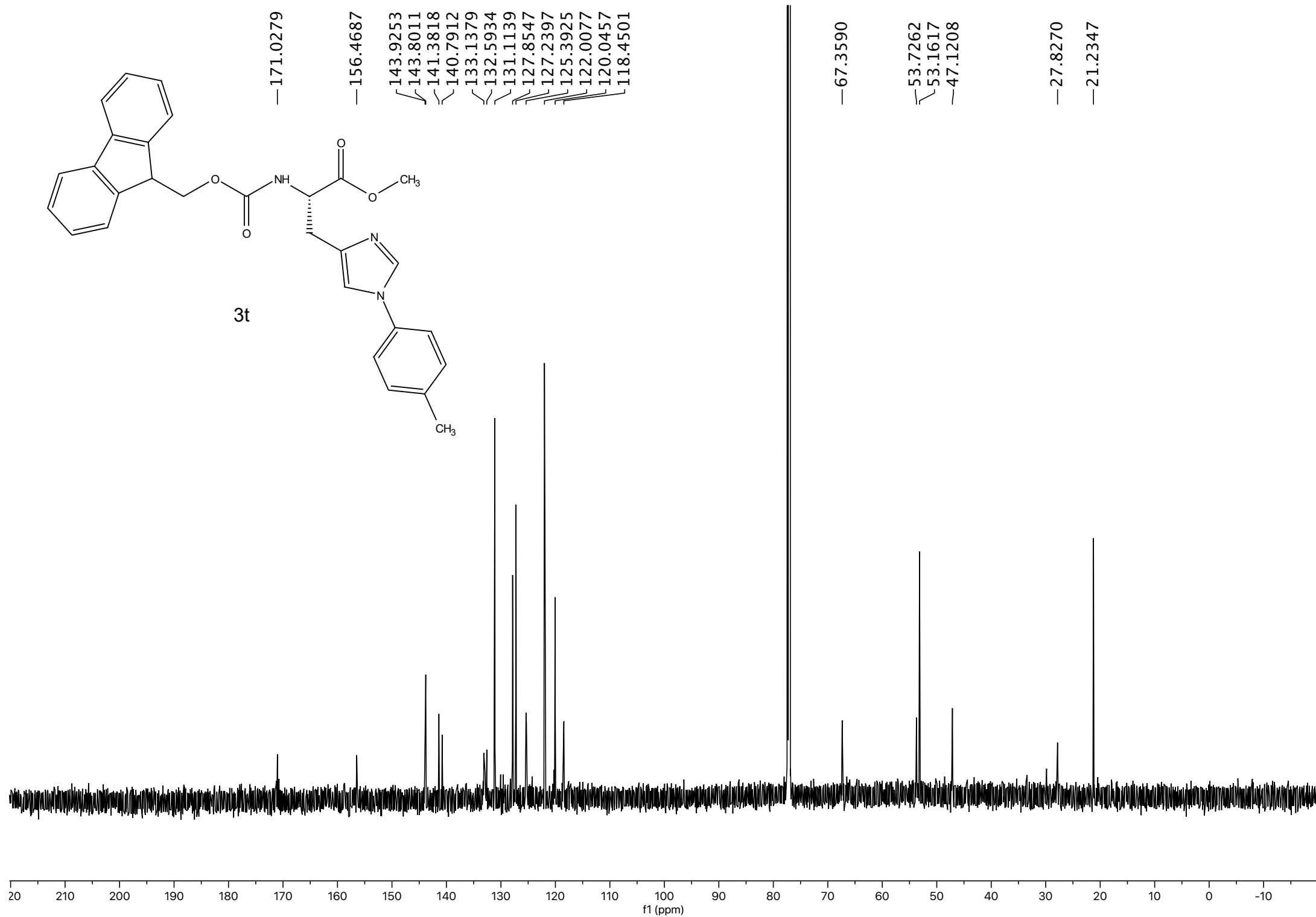


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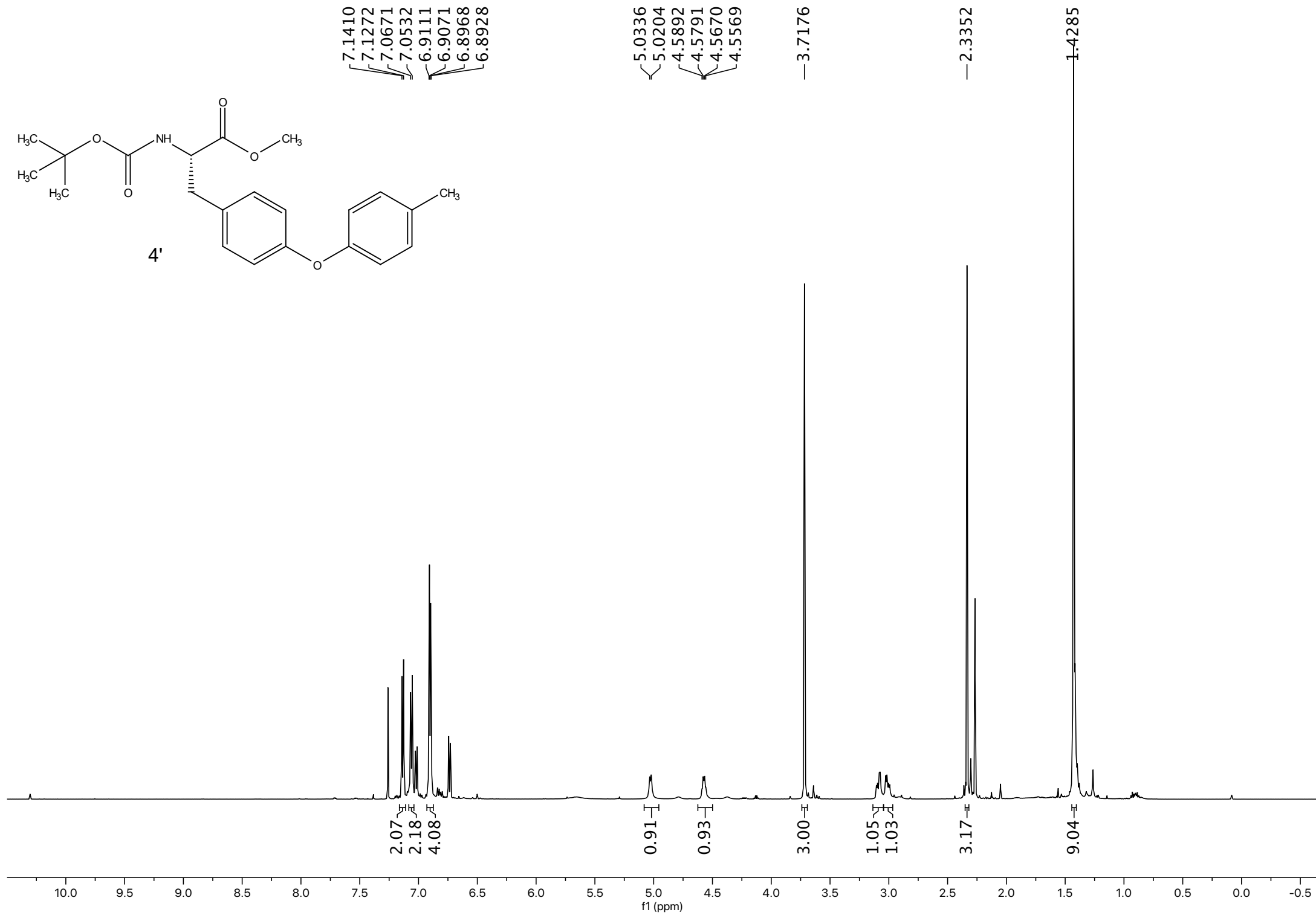
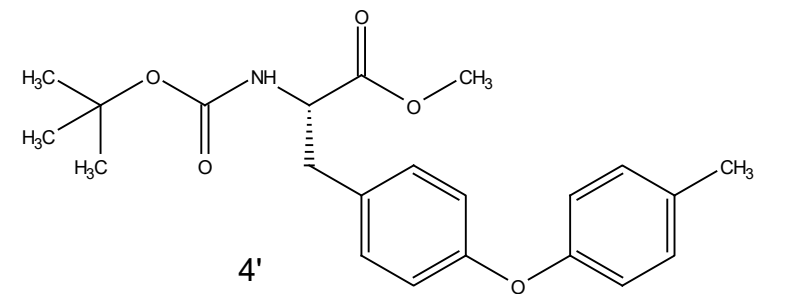


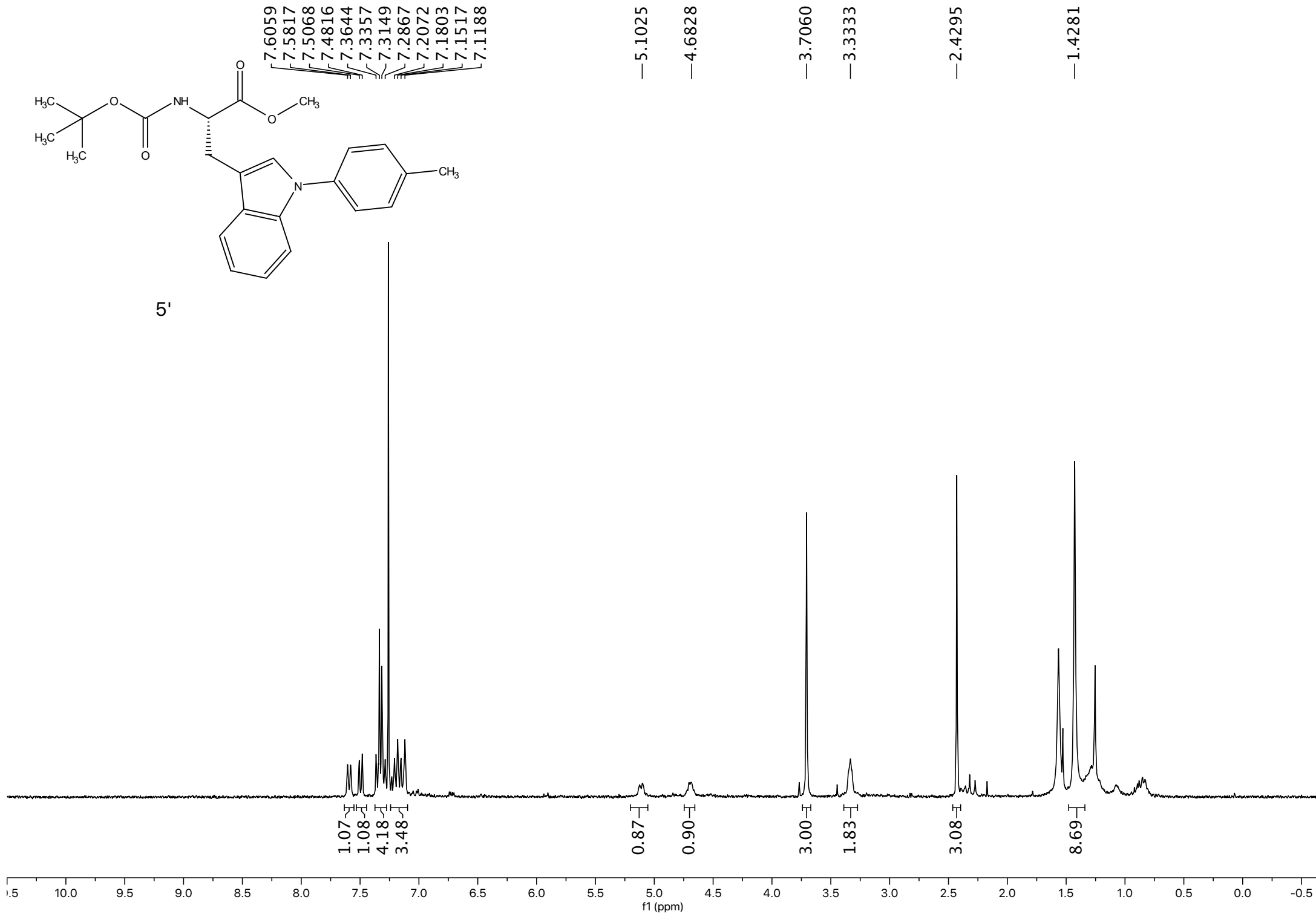
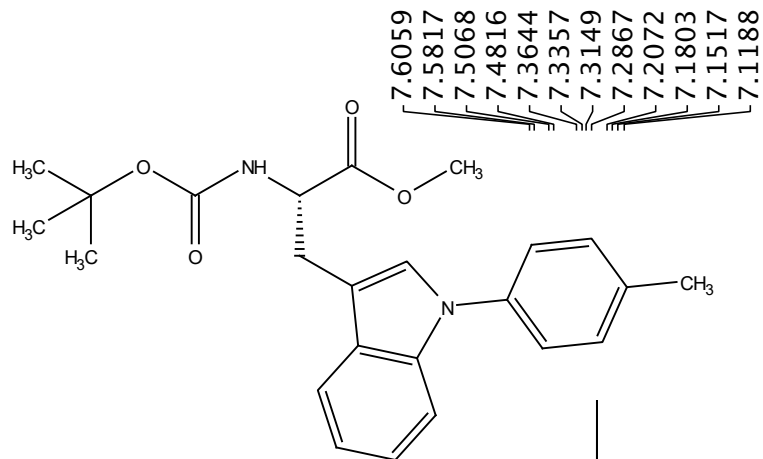


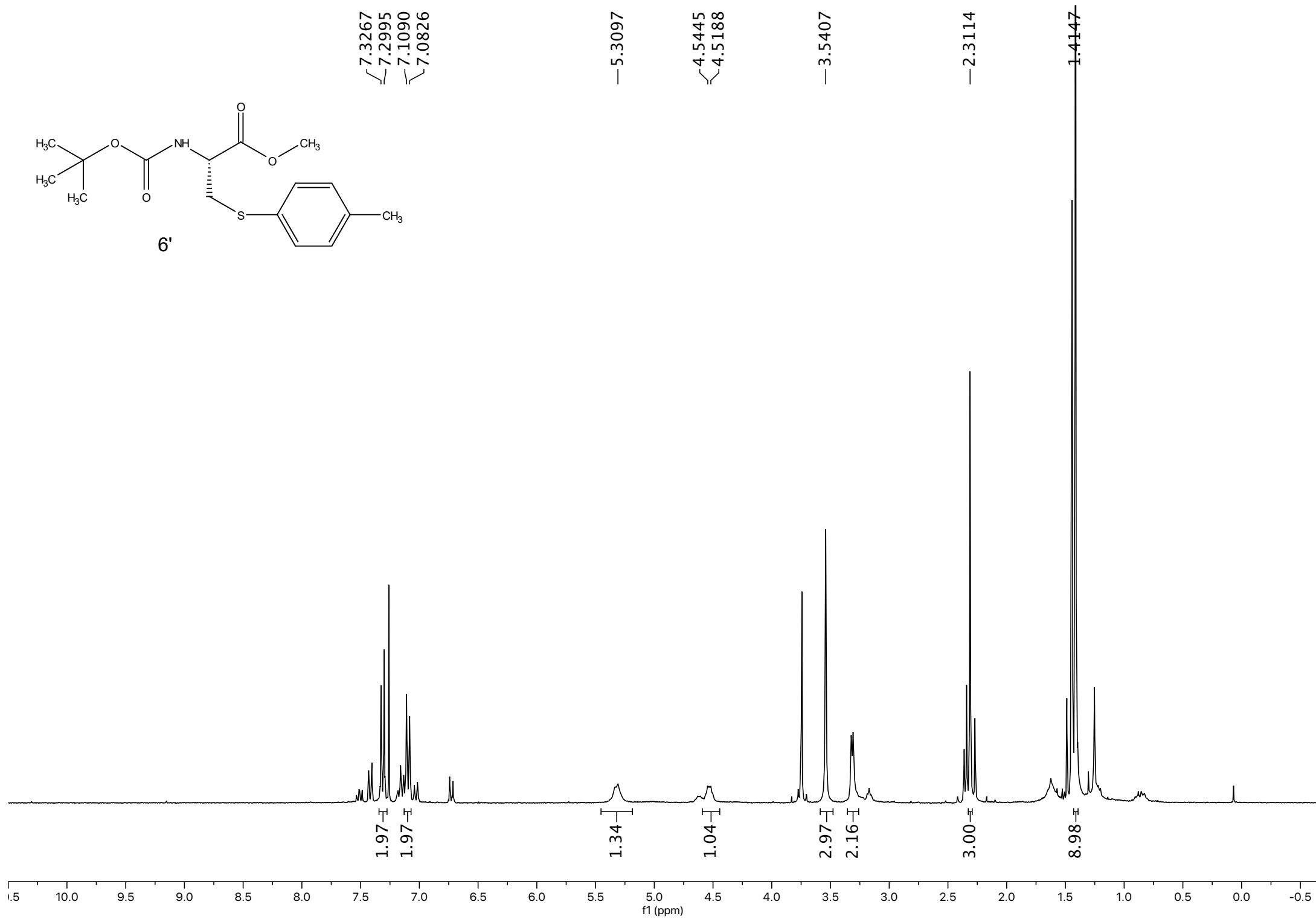
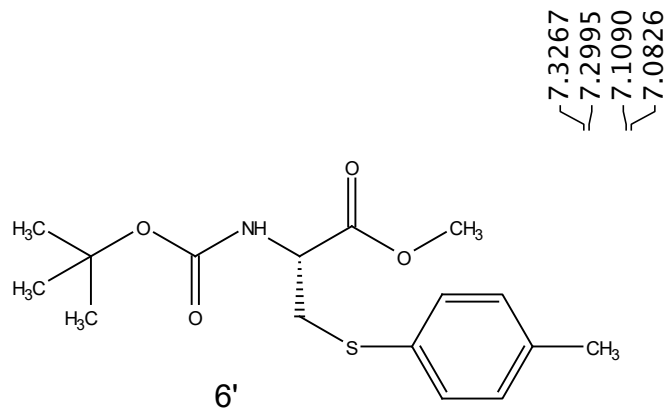
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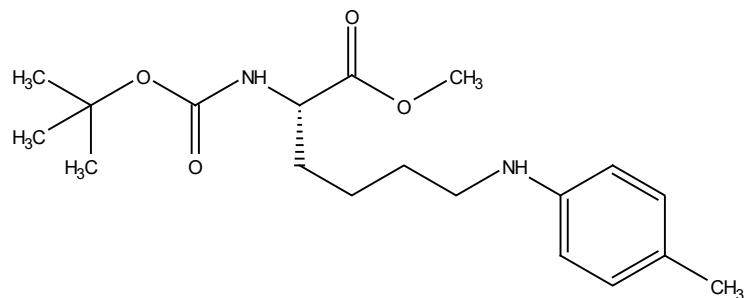


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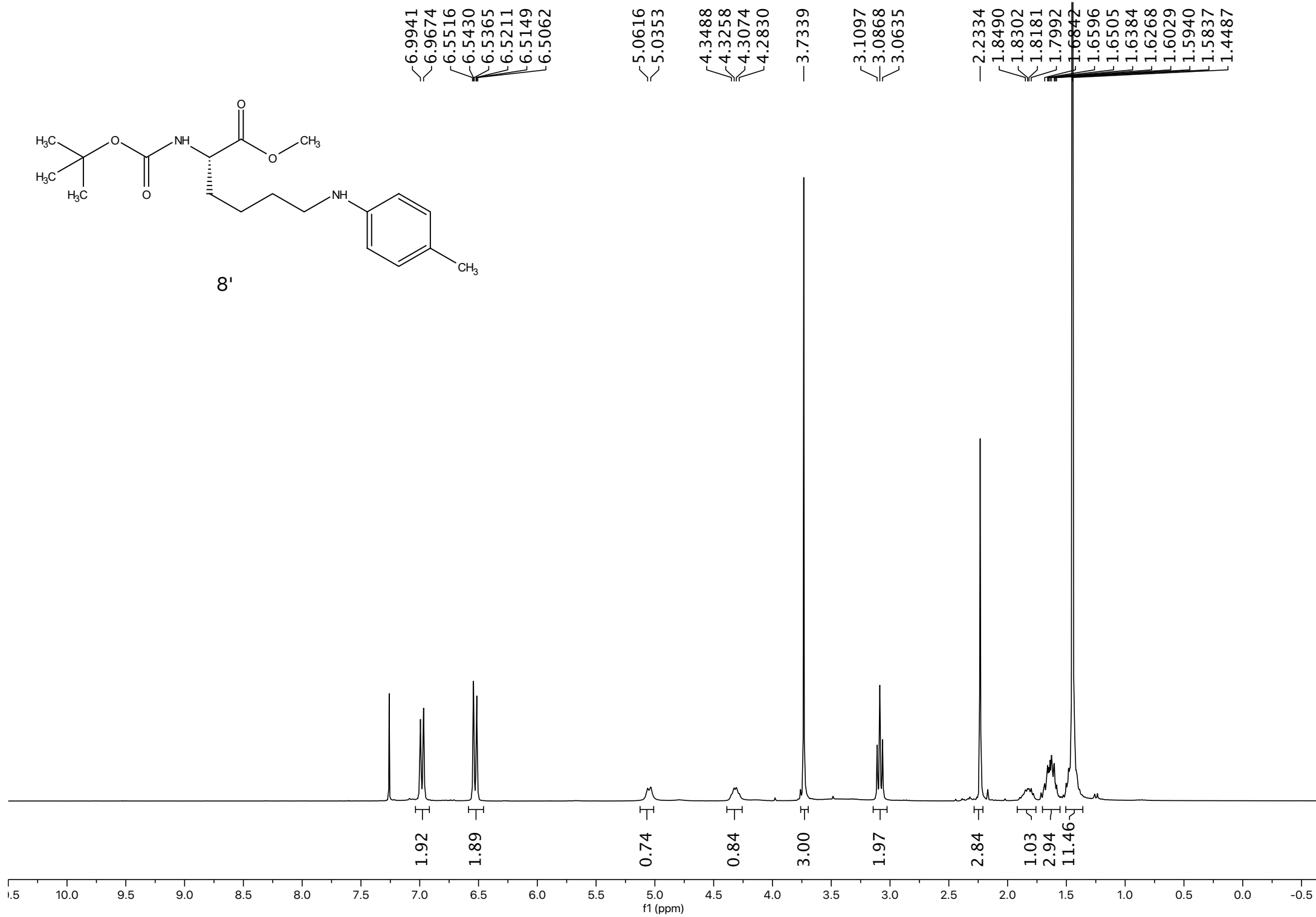




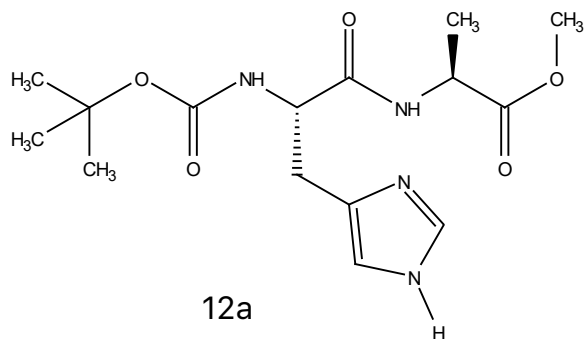




8'



12a



—7.7421

—7.5350

—6.7895

—5.9285

—5.9194

—4.4591

—4.4477

—4.4360

—4.4225

—4.4115

—4.4051

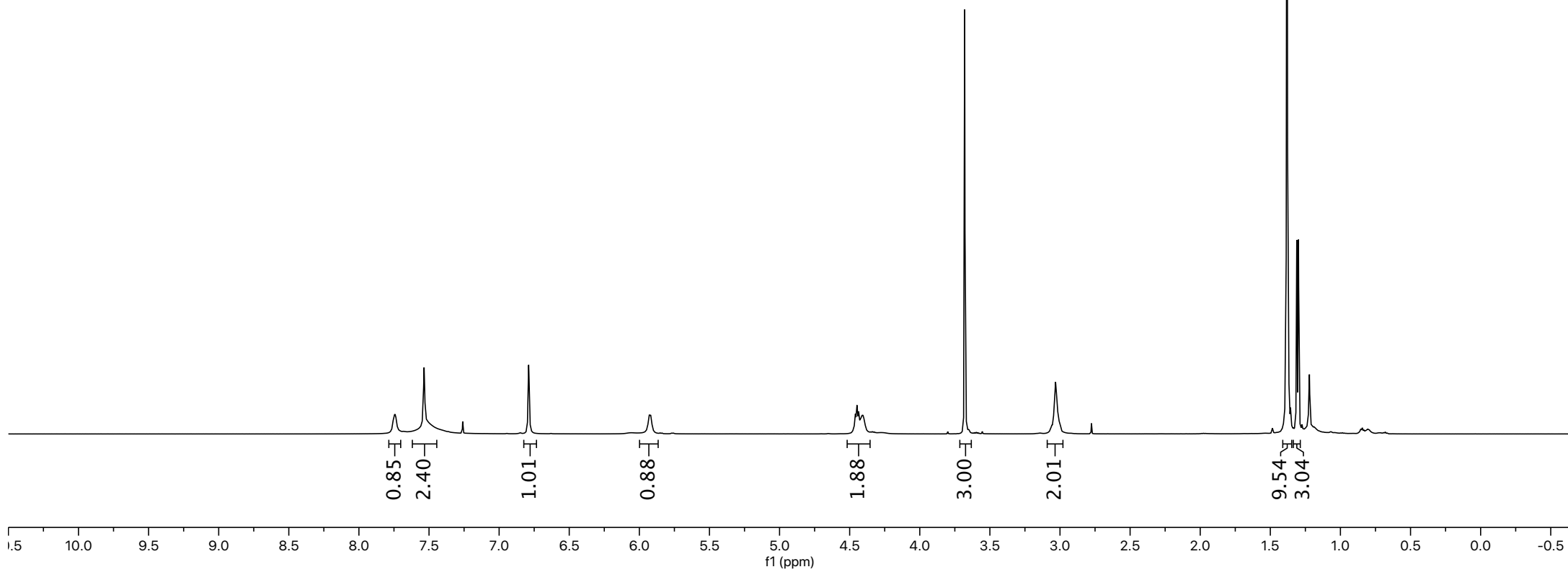
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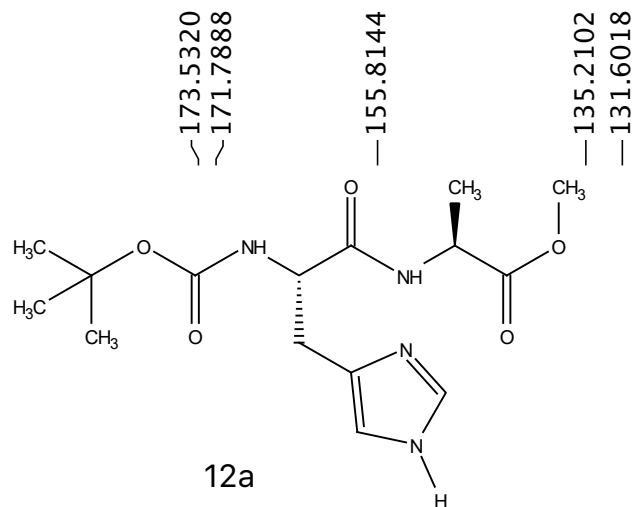
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—1.3810

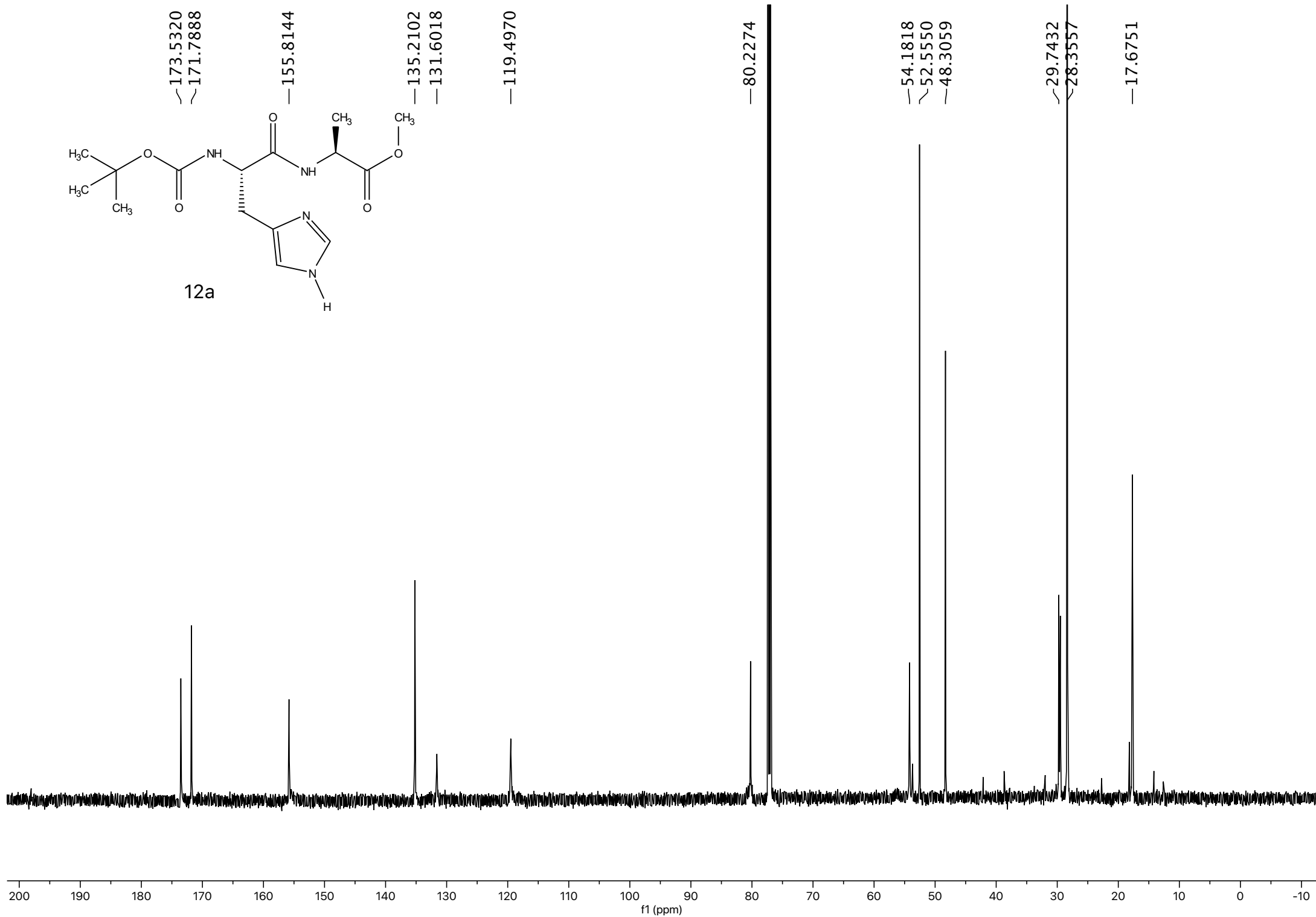
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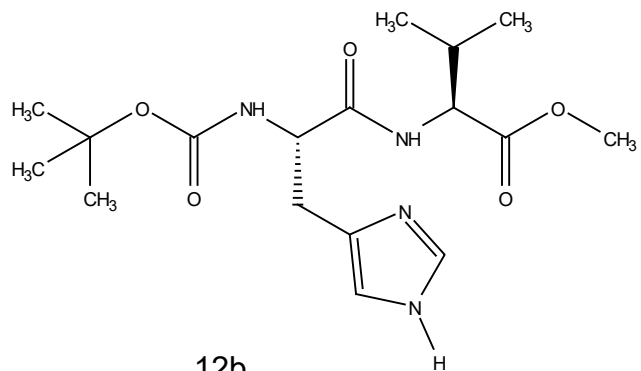
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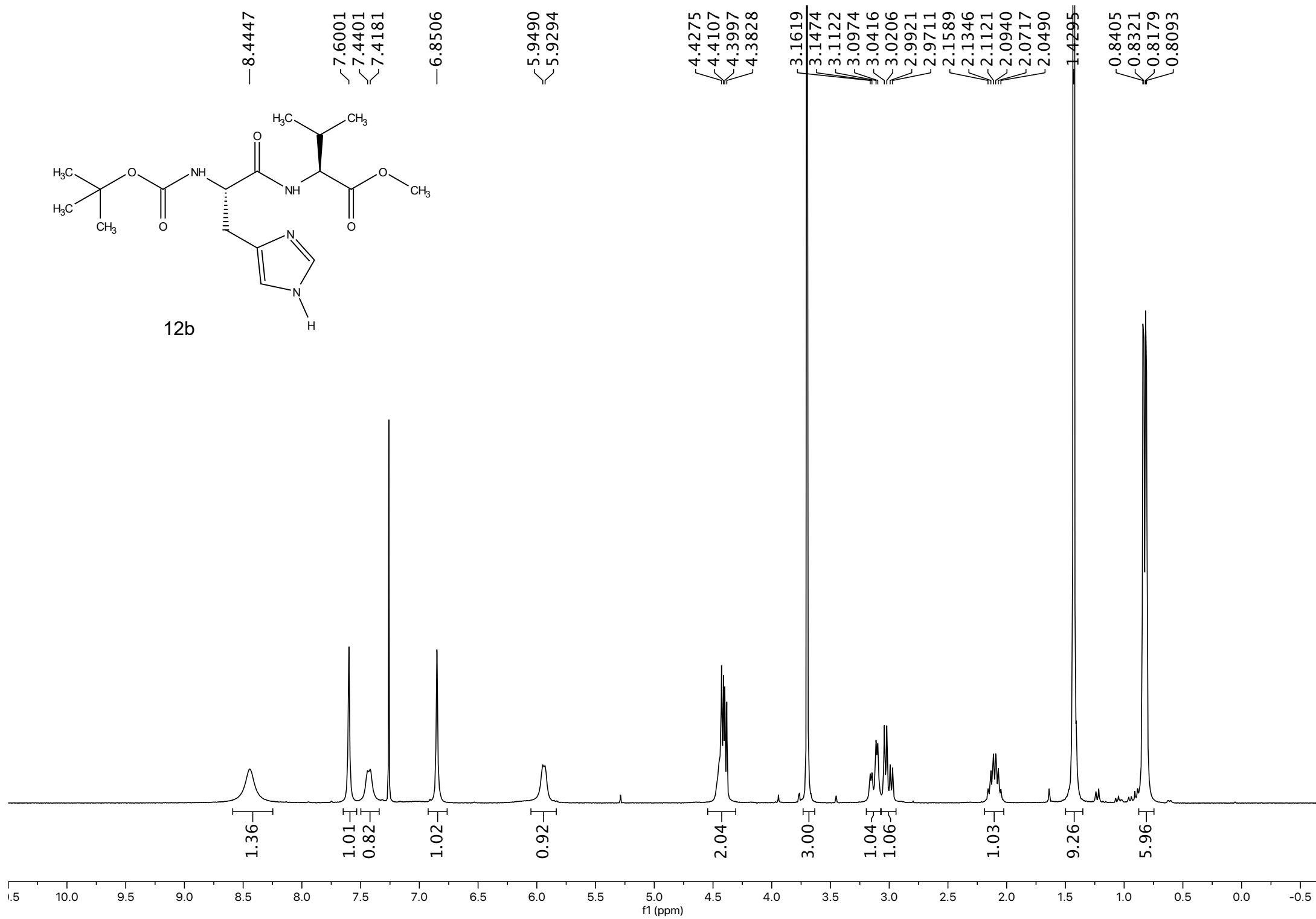


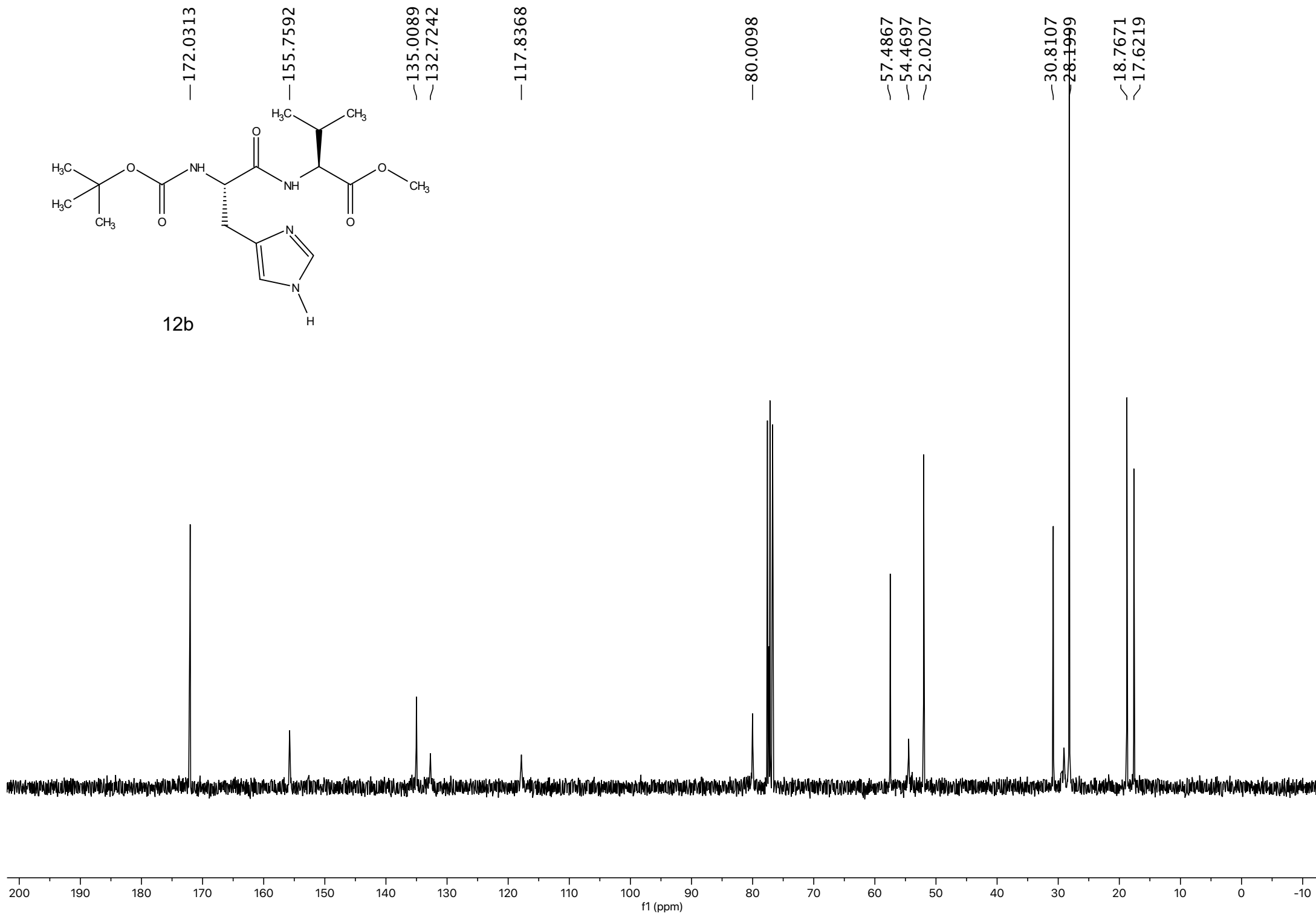
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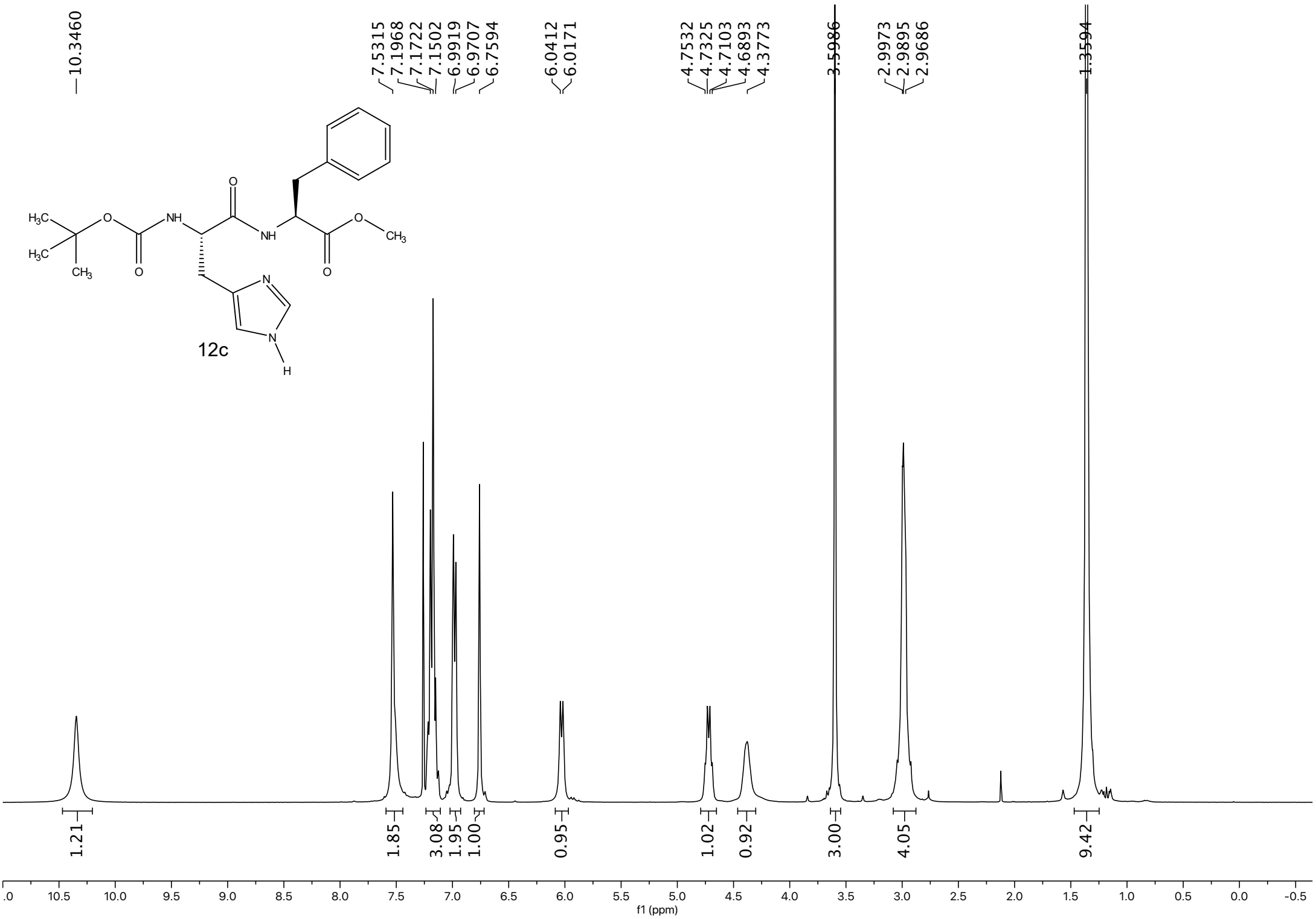


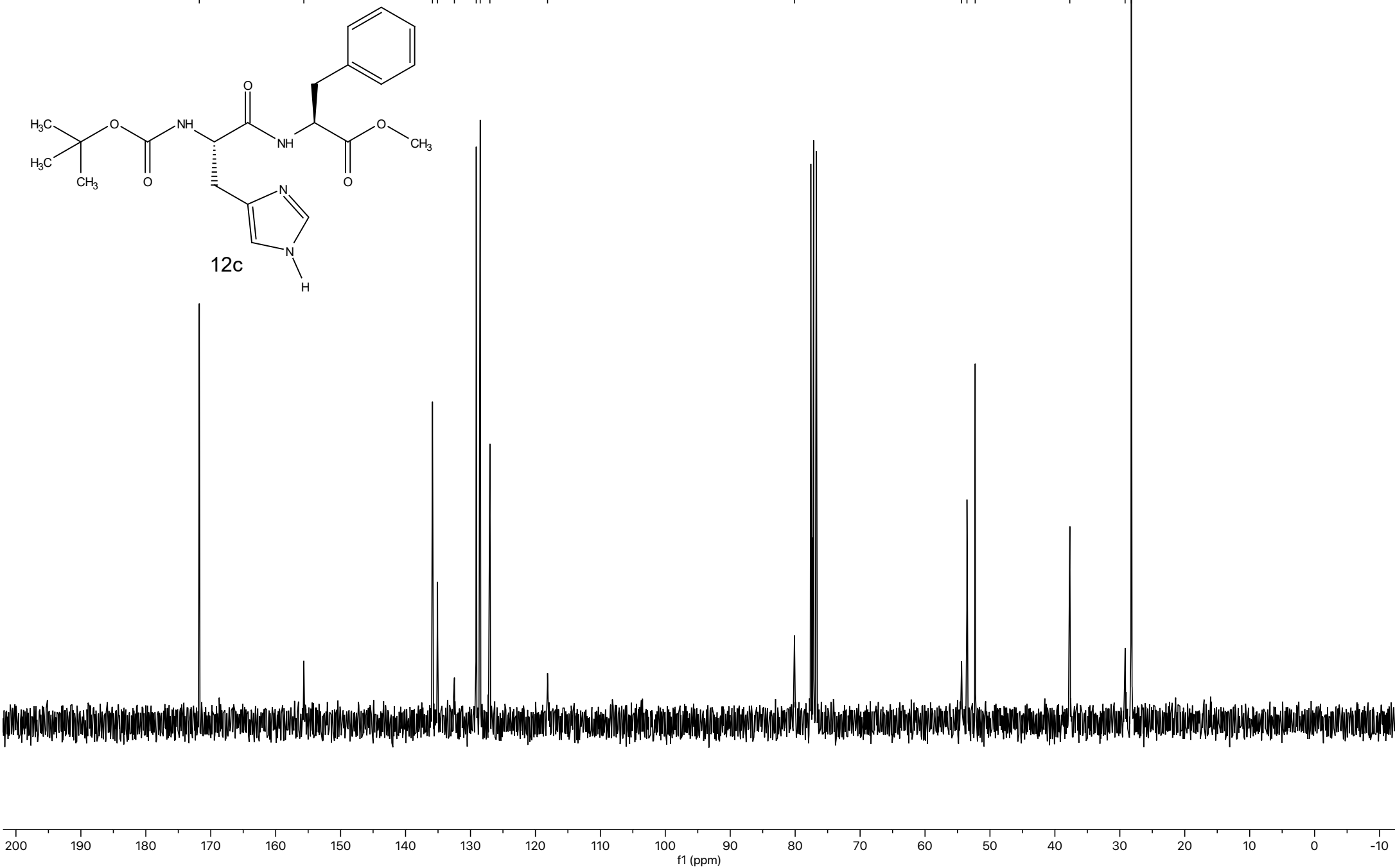
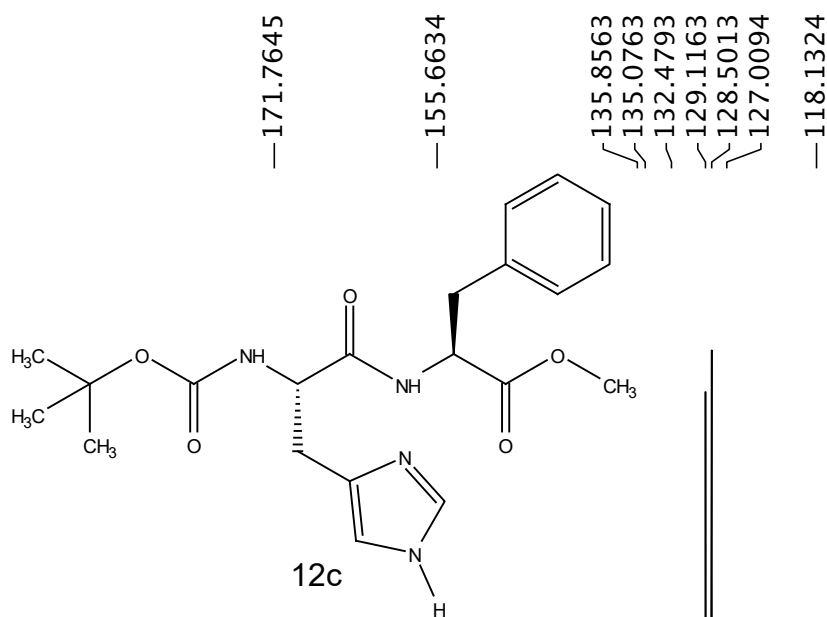


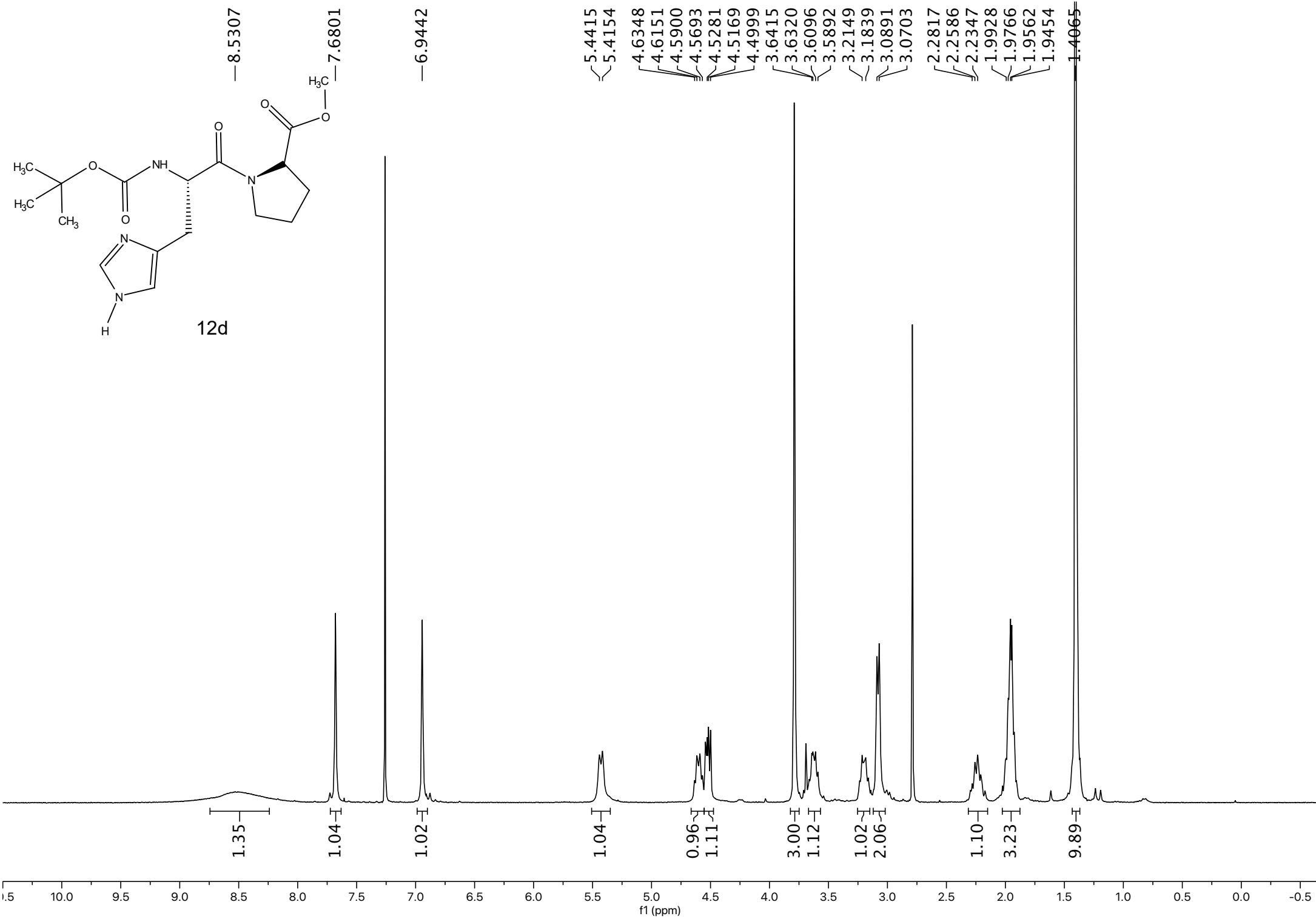
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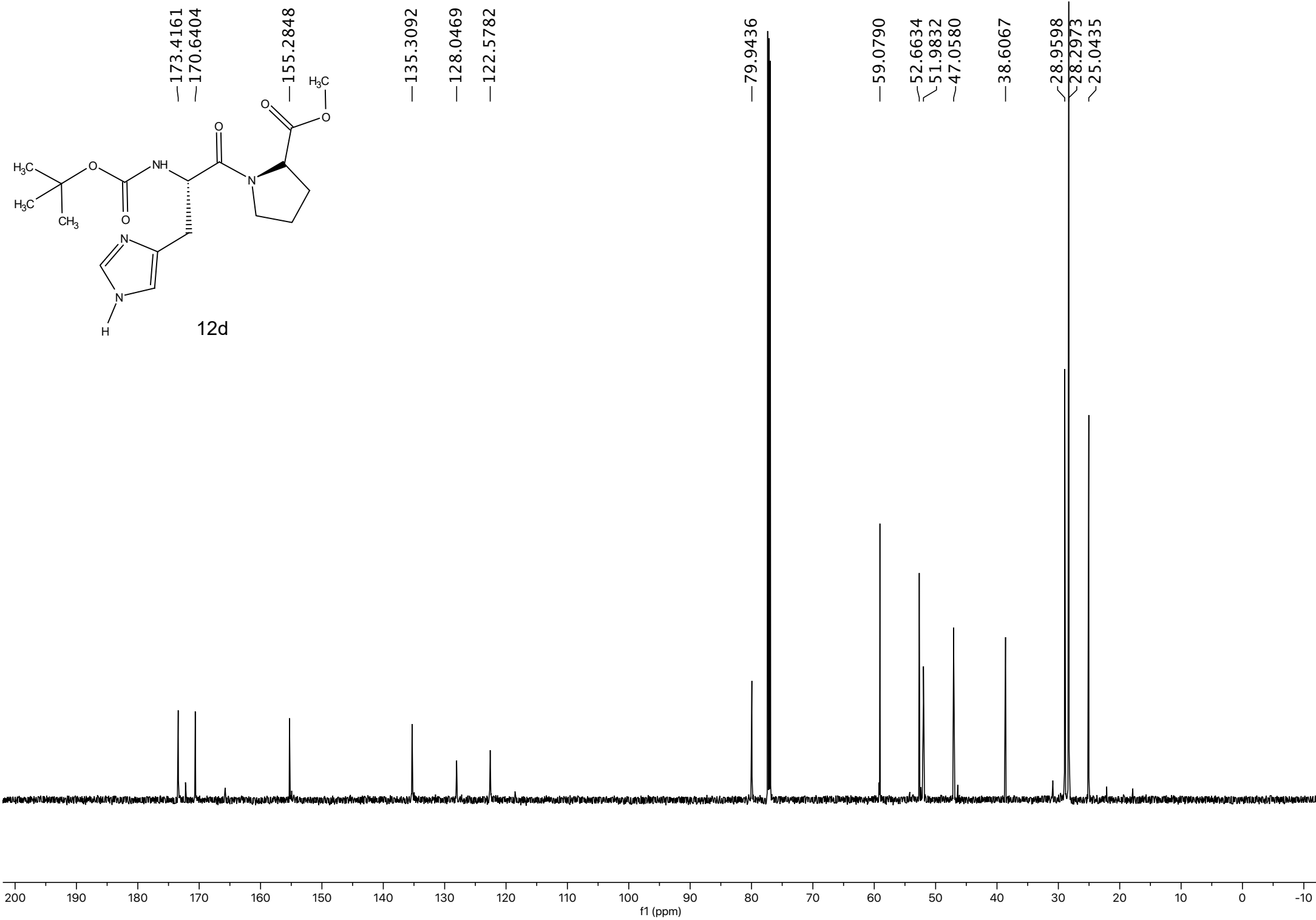


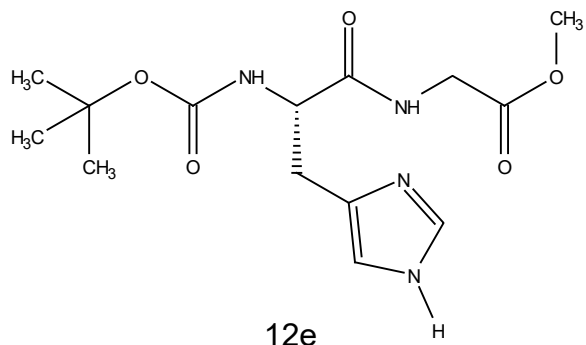












—9.3110

—7.8750

—7.5126

—6.7978

—5.9128

4.4347

4.0052

3.9966

3.9755

3.9667

3.9550

3.9246

3.6967

3.6875

3.0856

3.0452

3.0359

3.0191

3.0071

1.3787

1.01

0.85

1.01

1.00

0.95

0.98

2.05

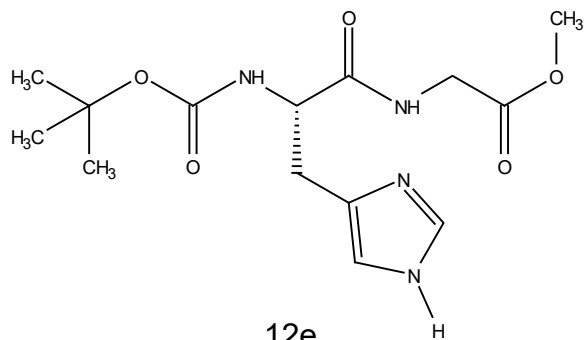
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0.95

1.17

9.57

f1 (ppm)



12e

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119.7881

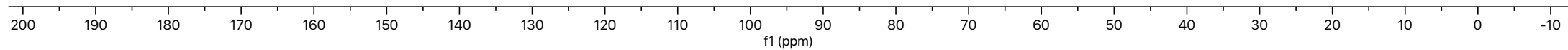
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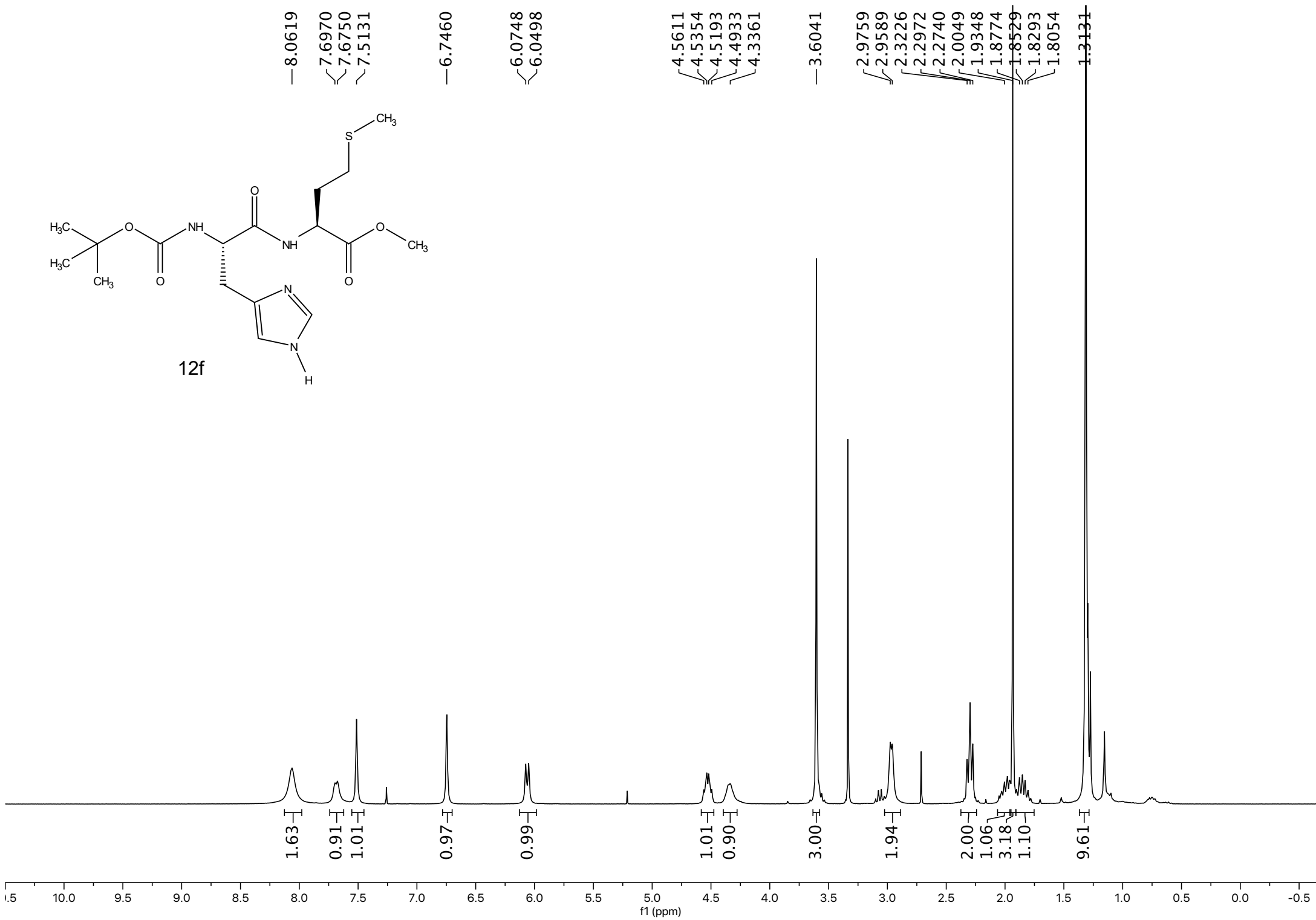
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52.4304

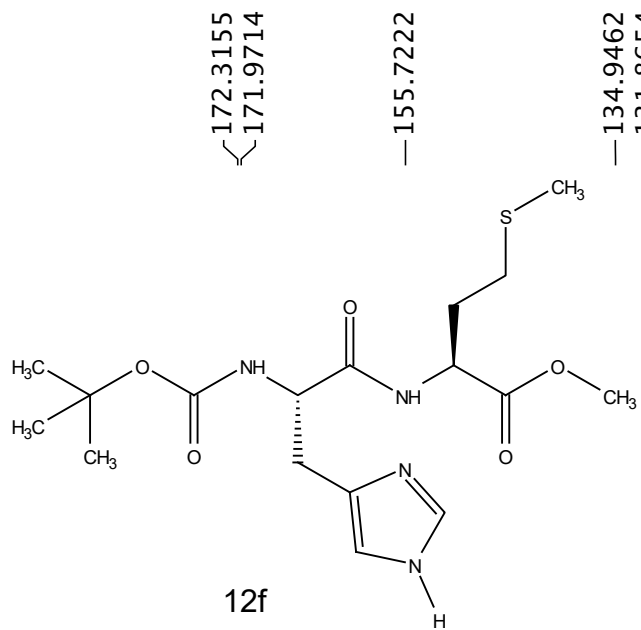
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18.3382







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155.7222

134.9462
131.8654

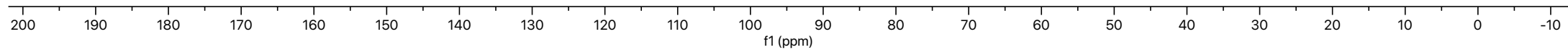
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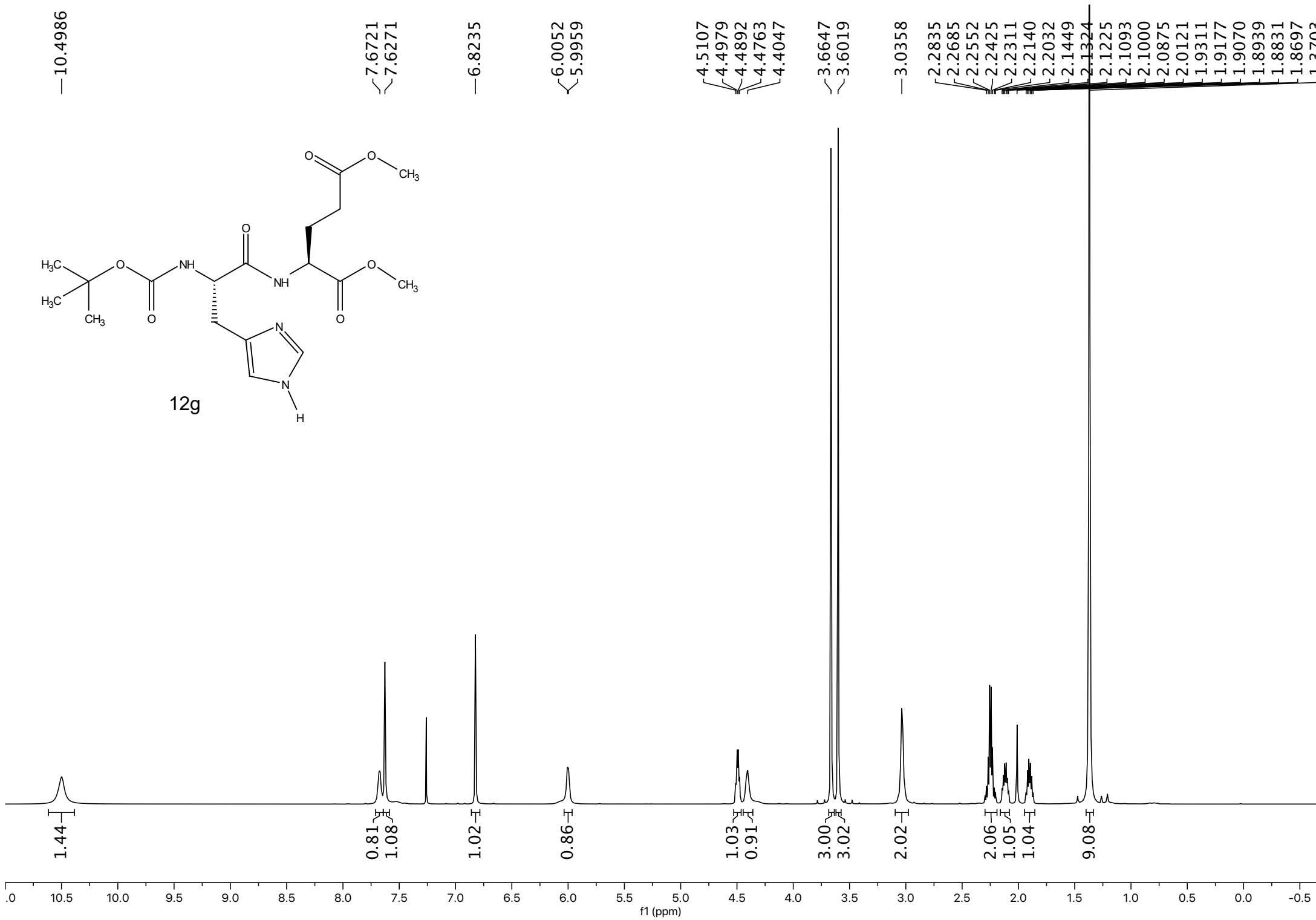
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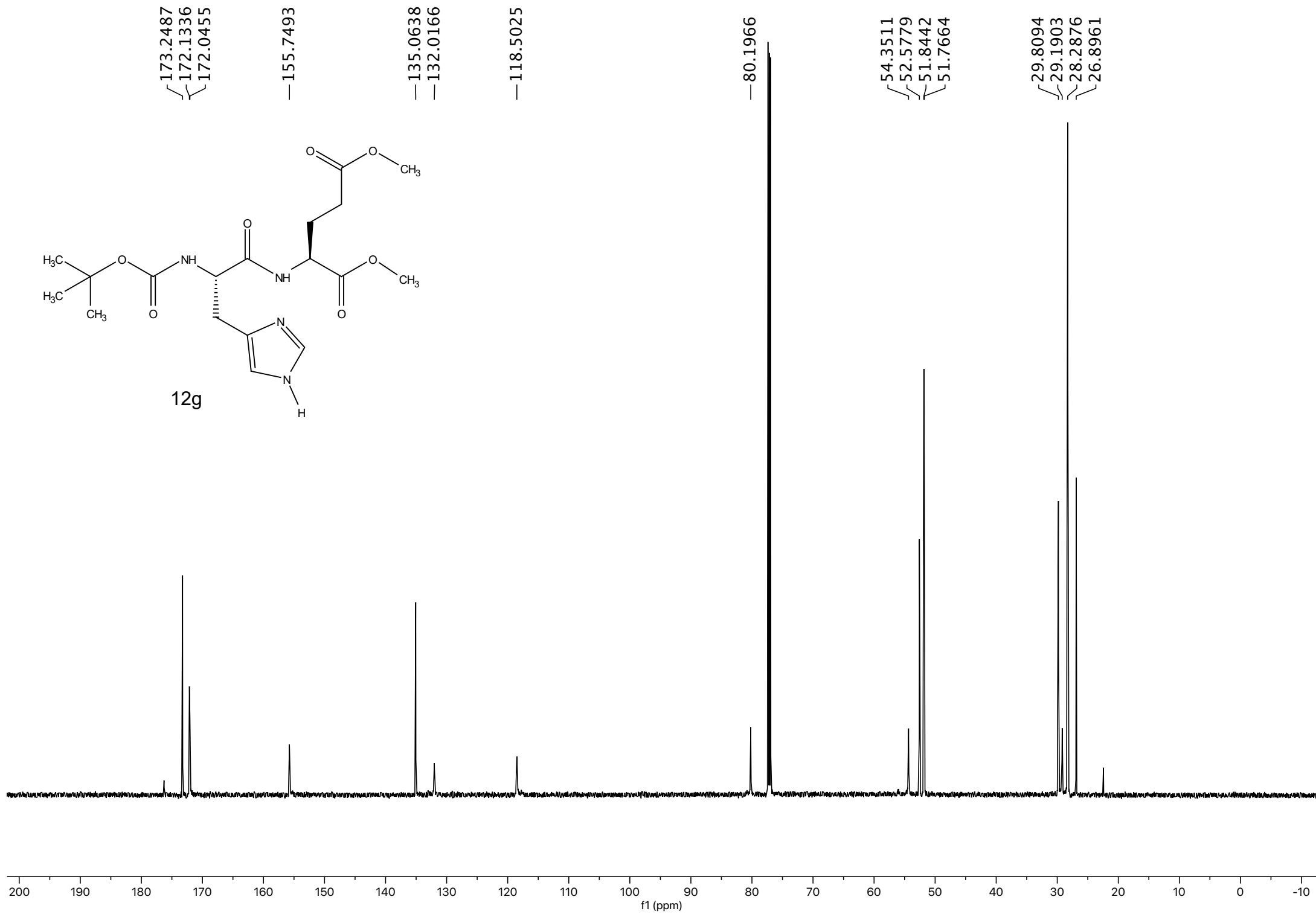
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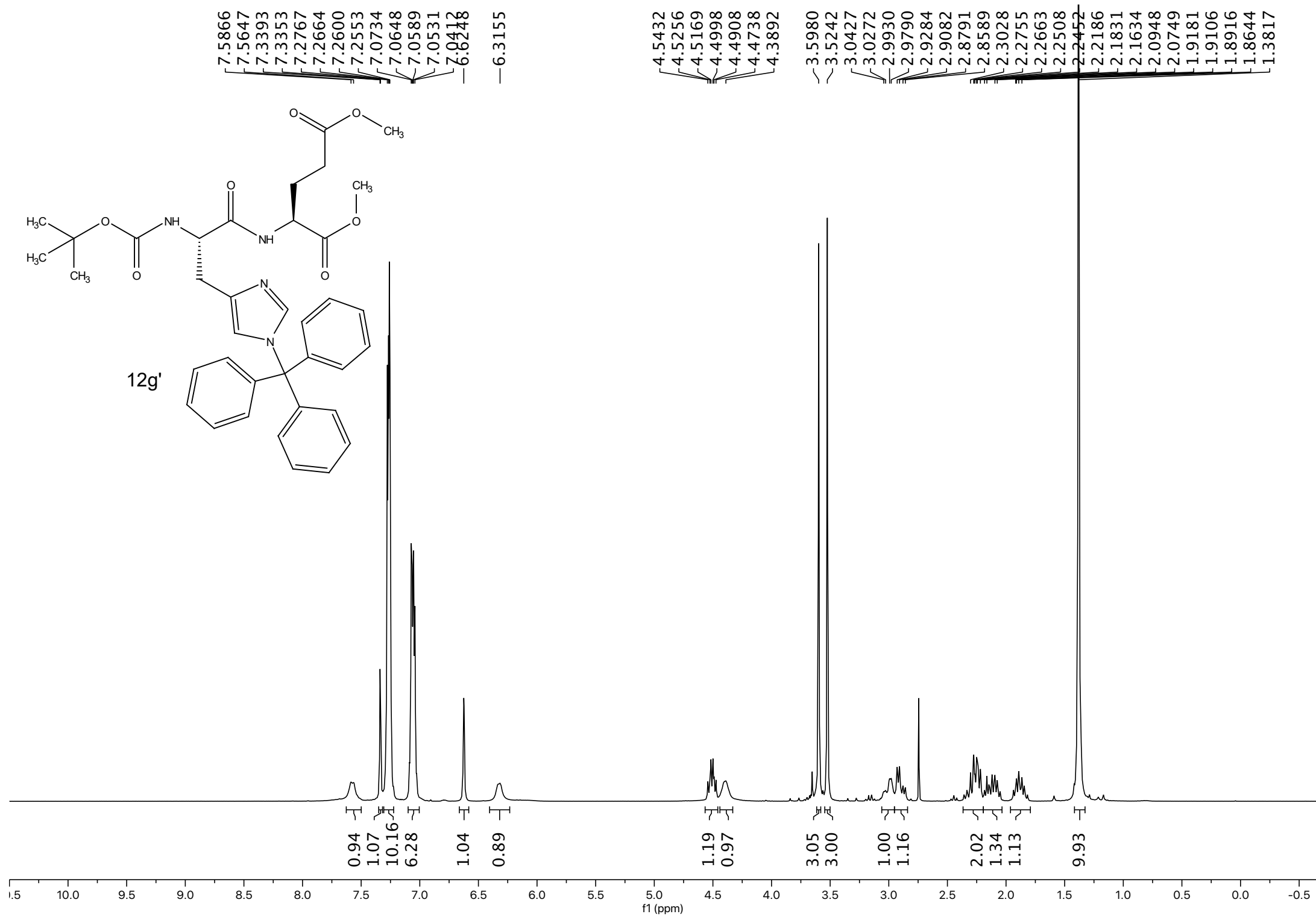
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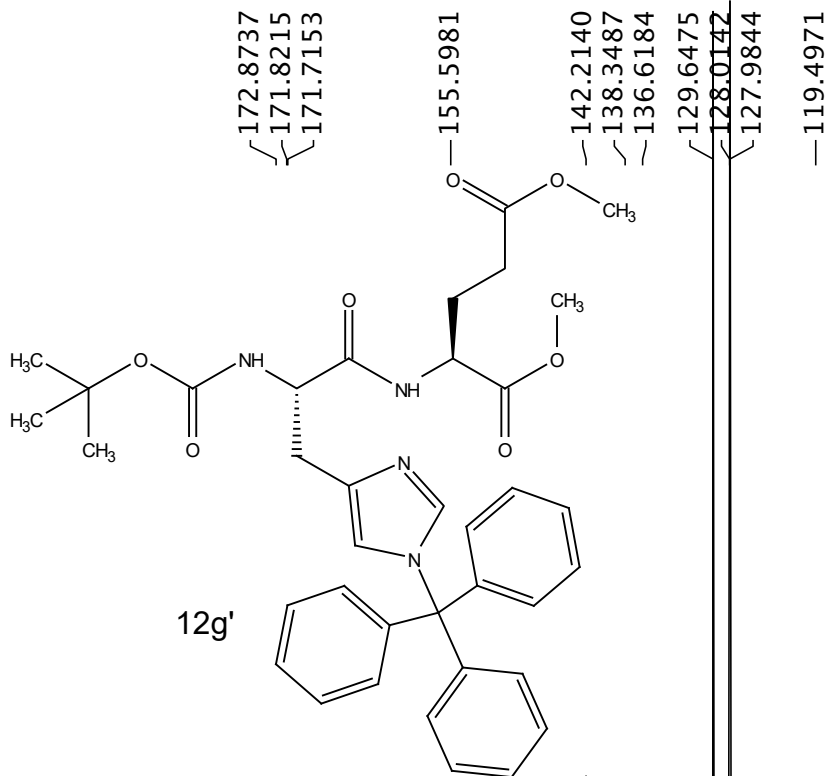








12g'



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171.8215
171.7153

155.5981

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136.6184

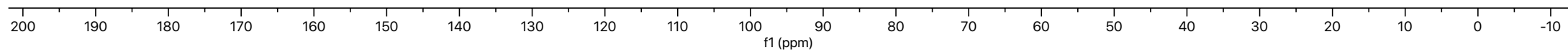
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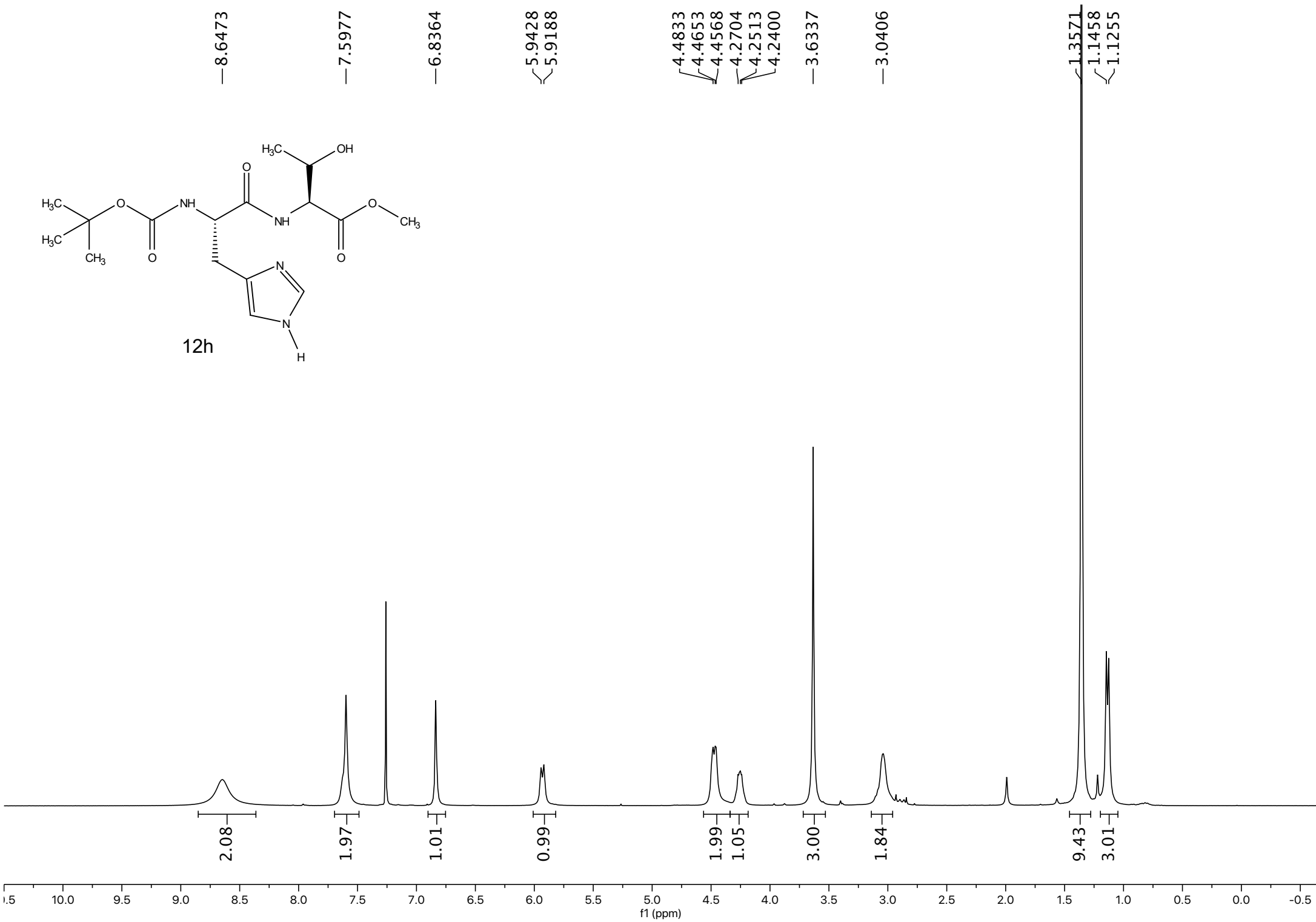
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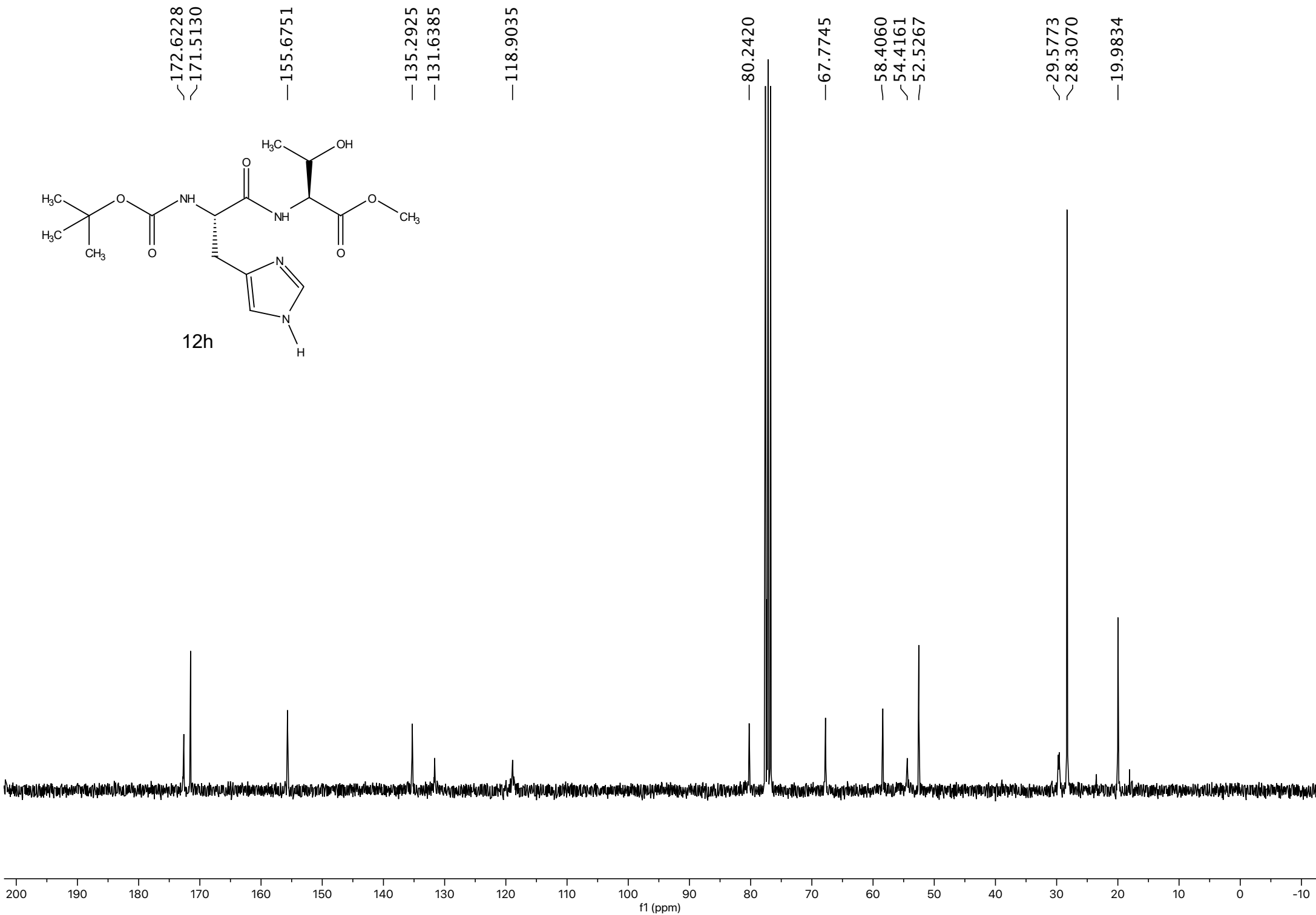
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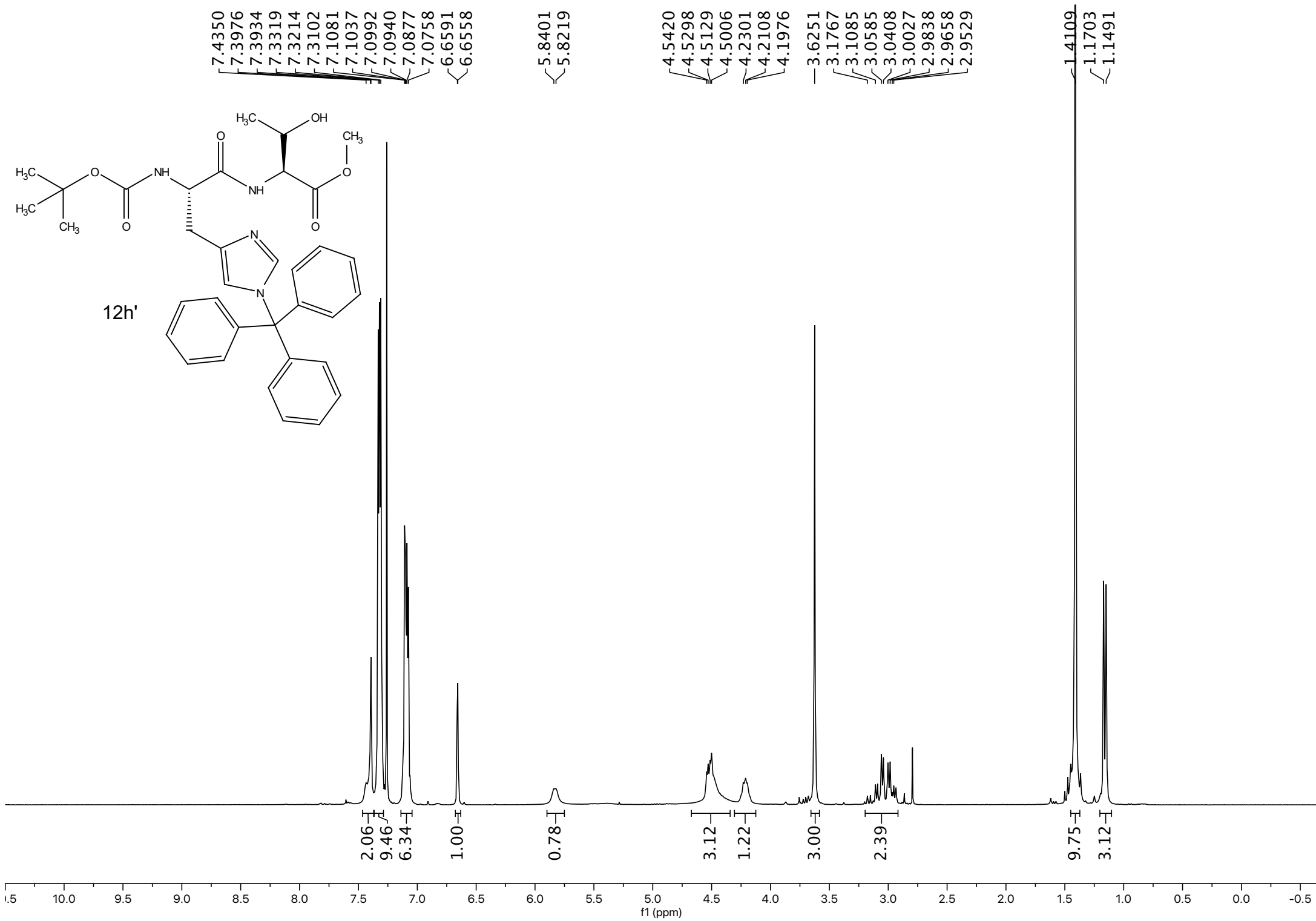
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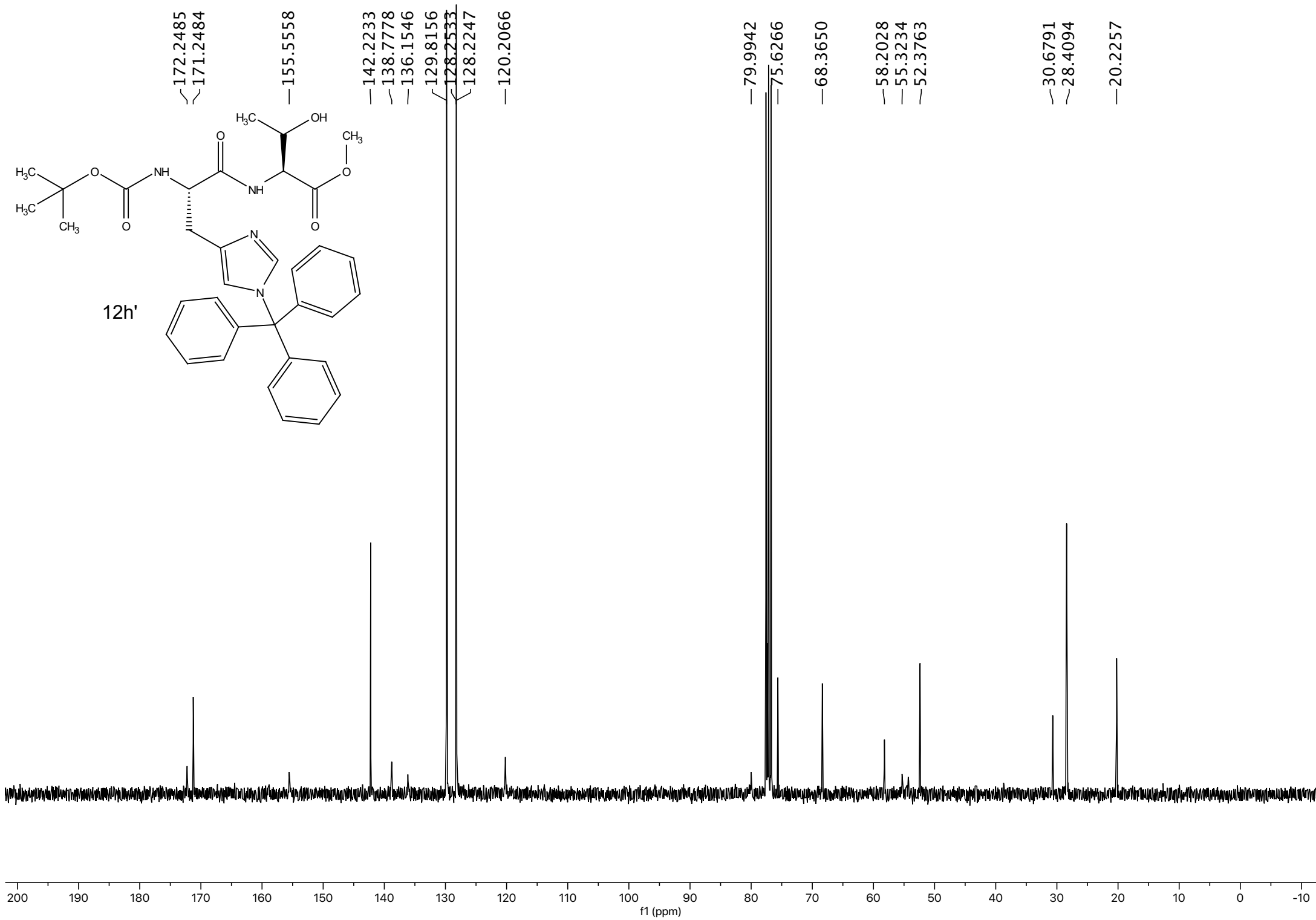
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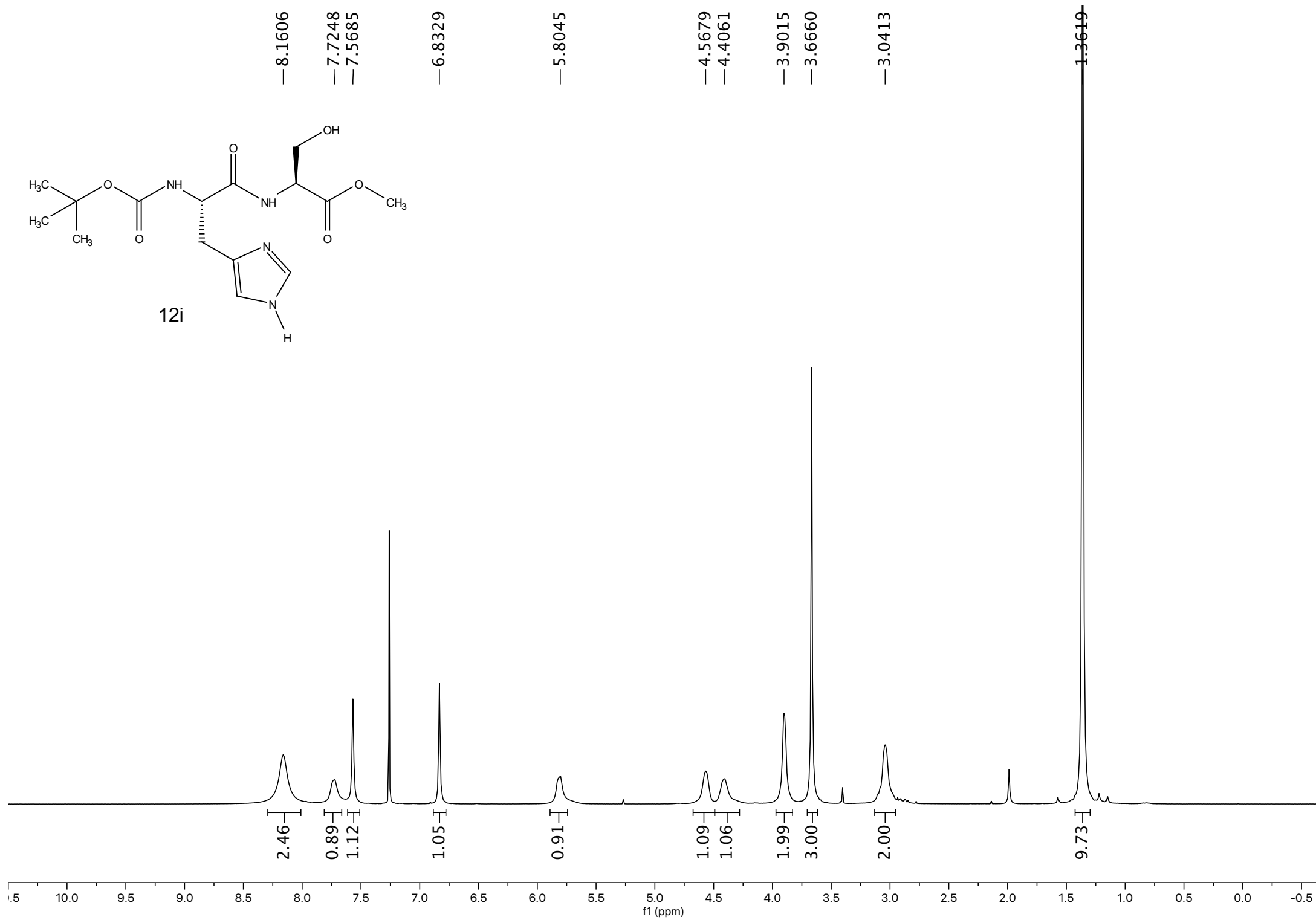
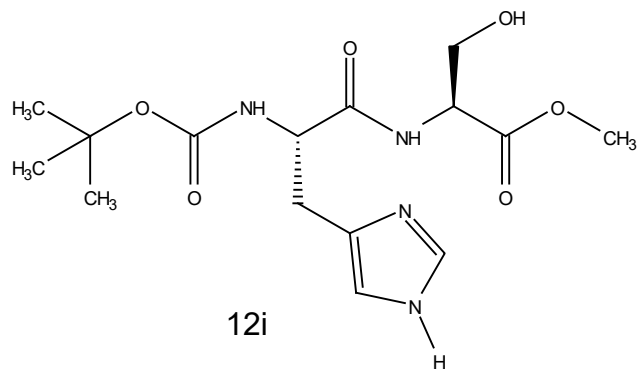


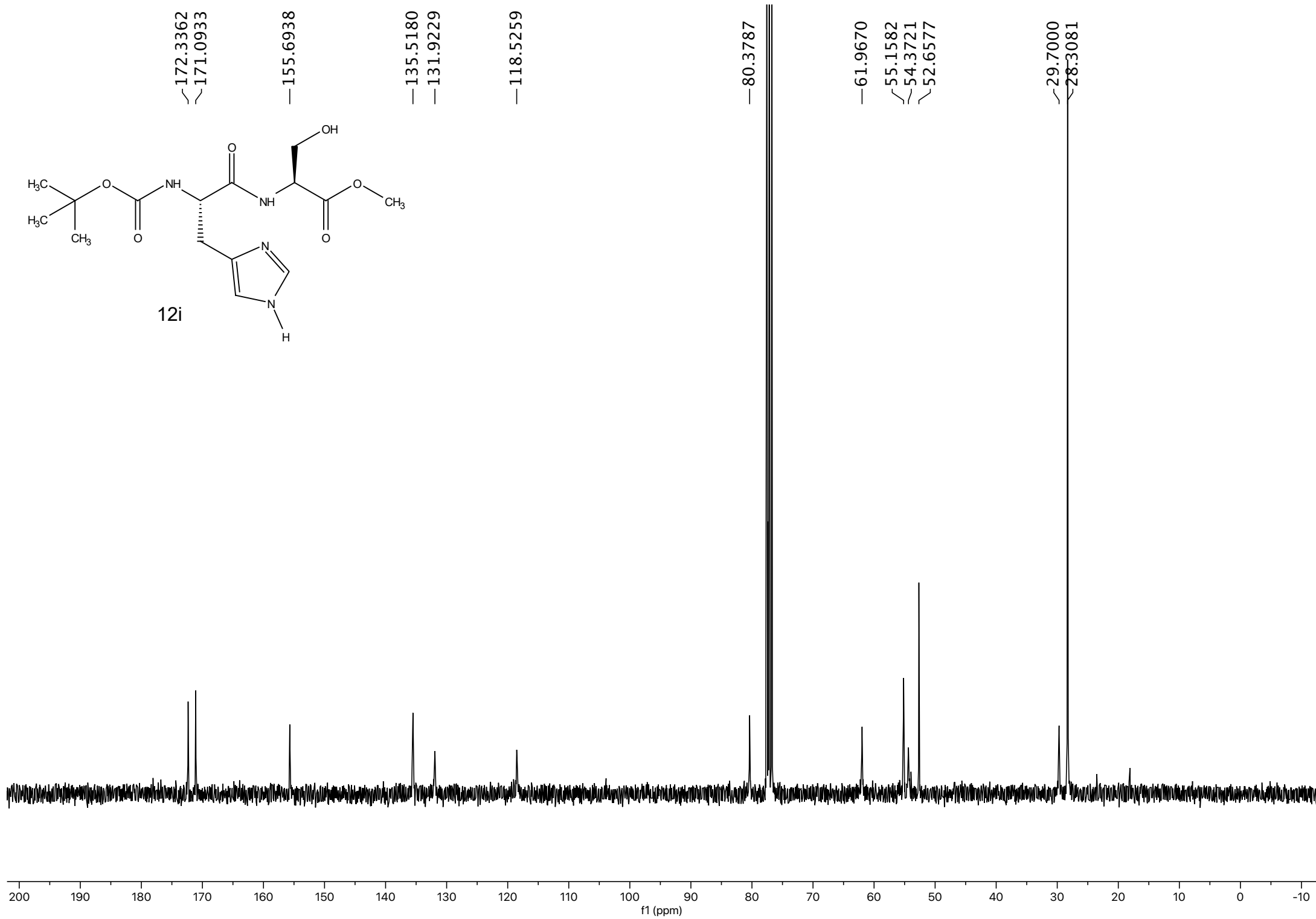


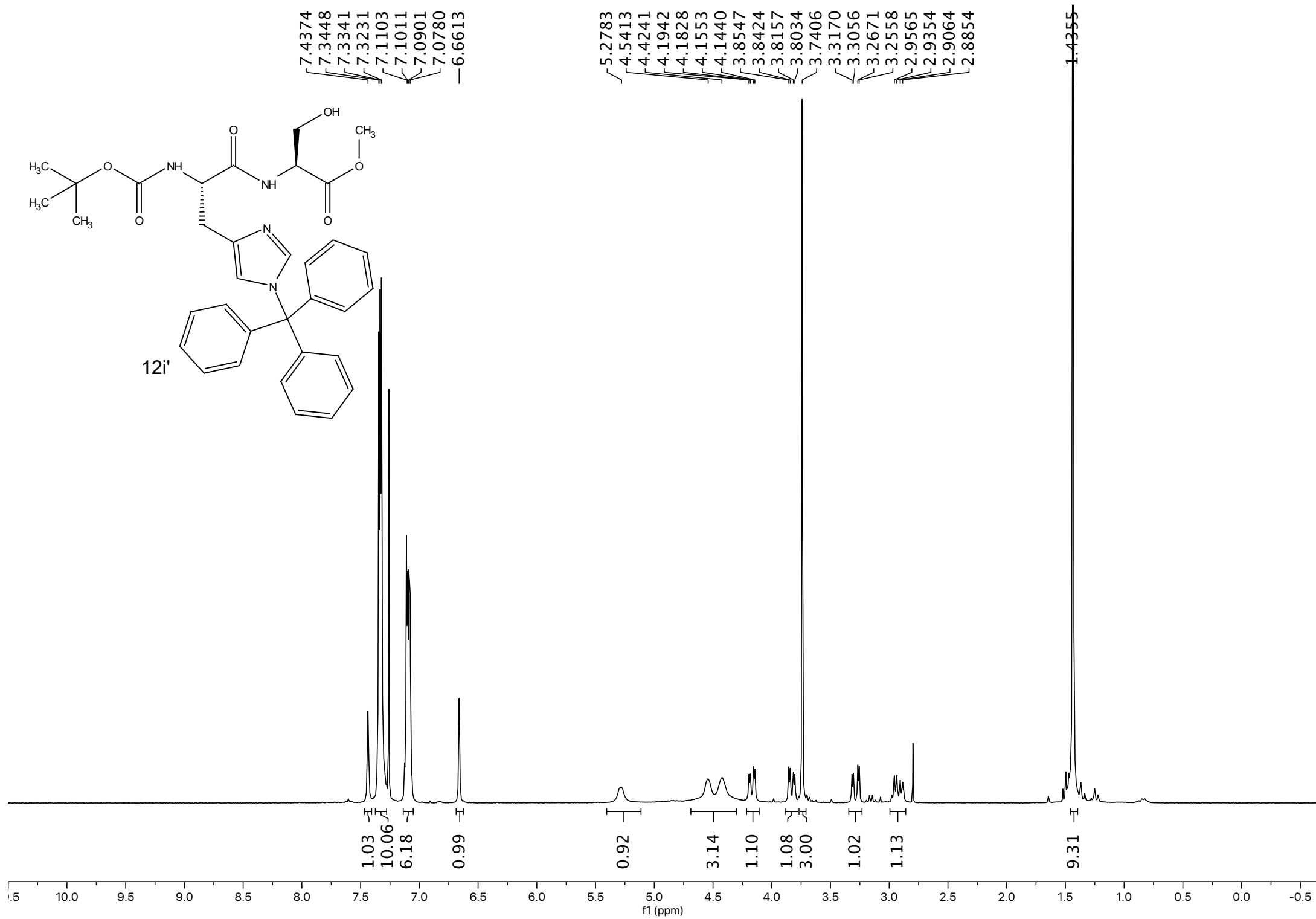


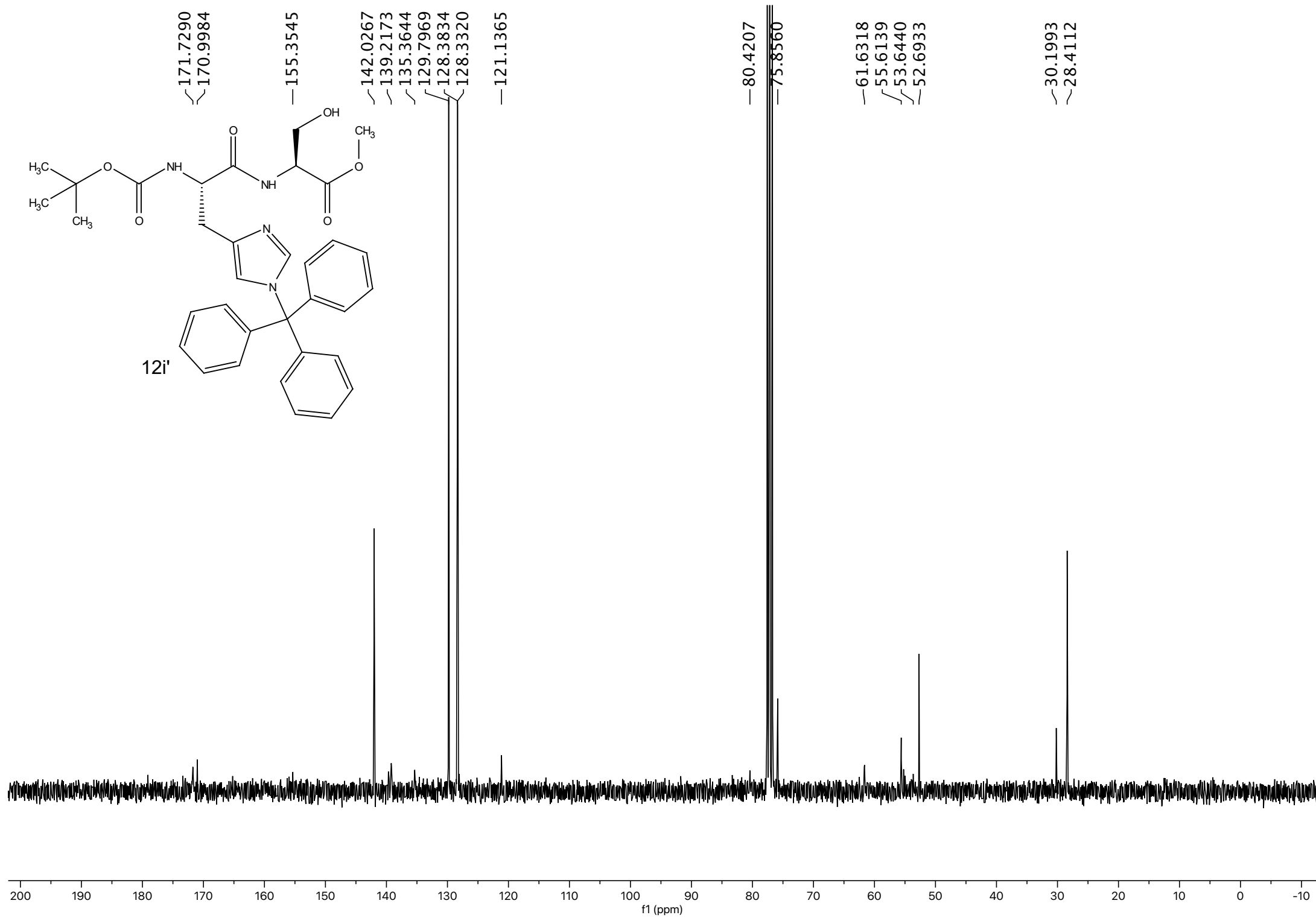


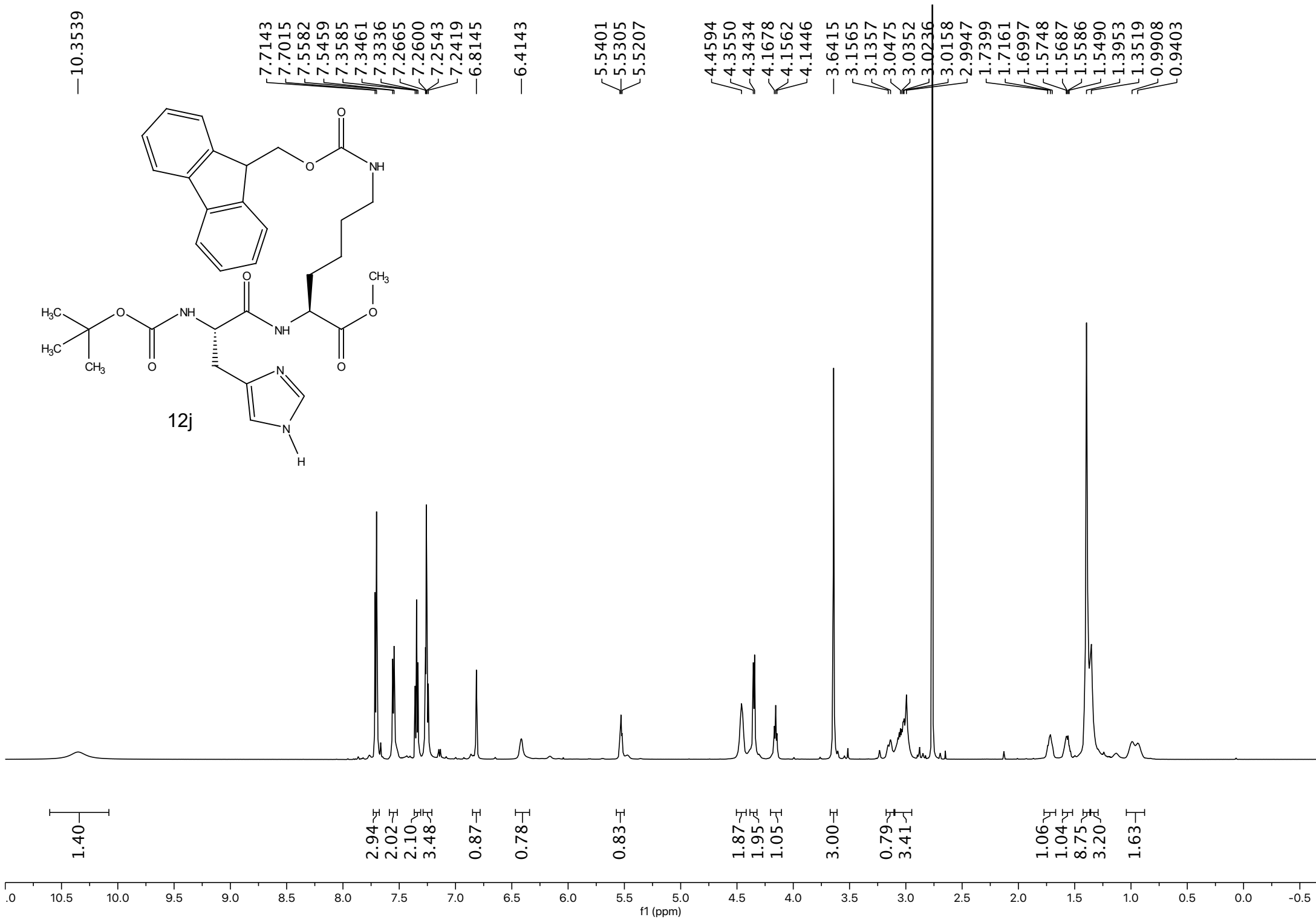


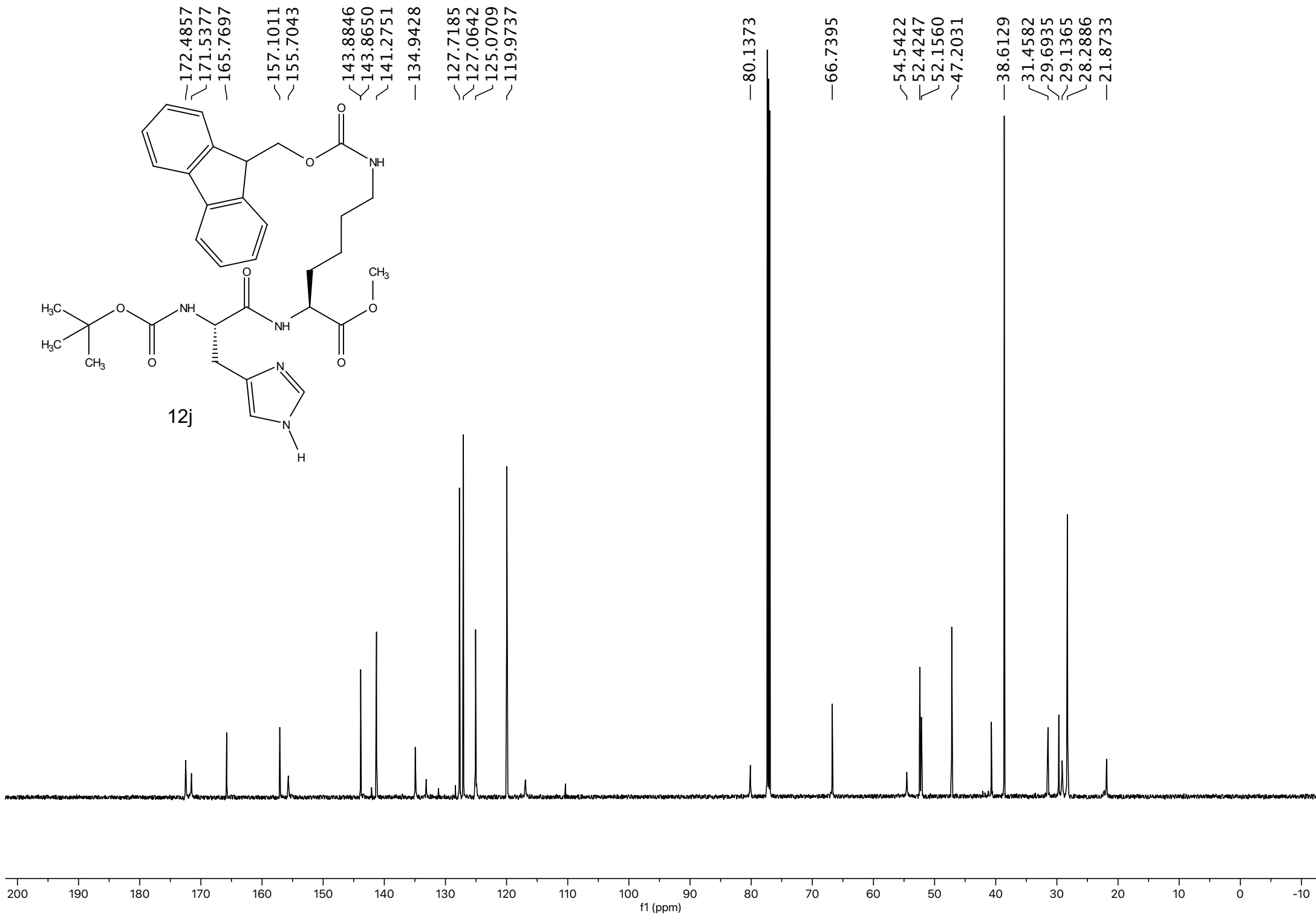


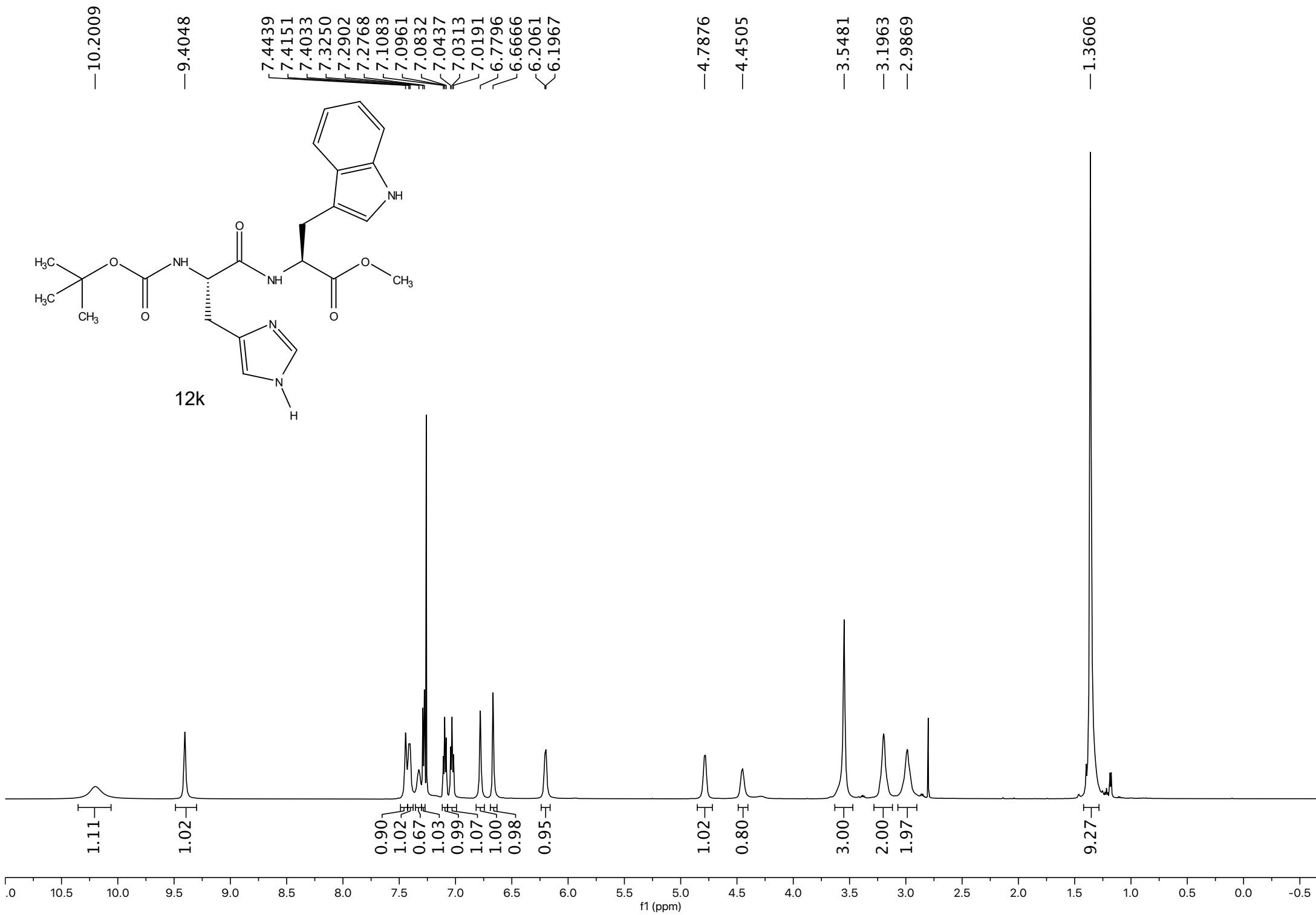


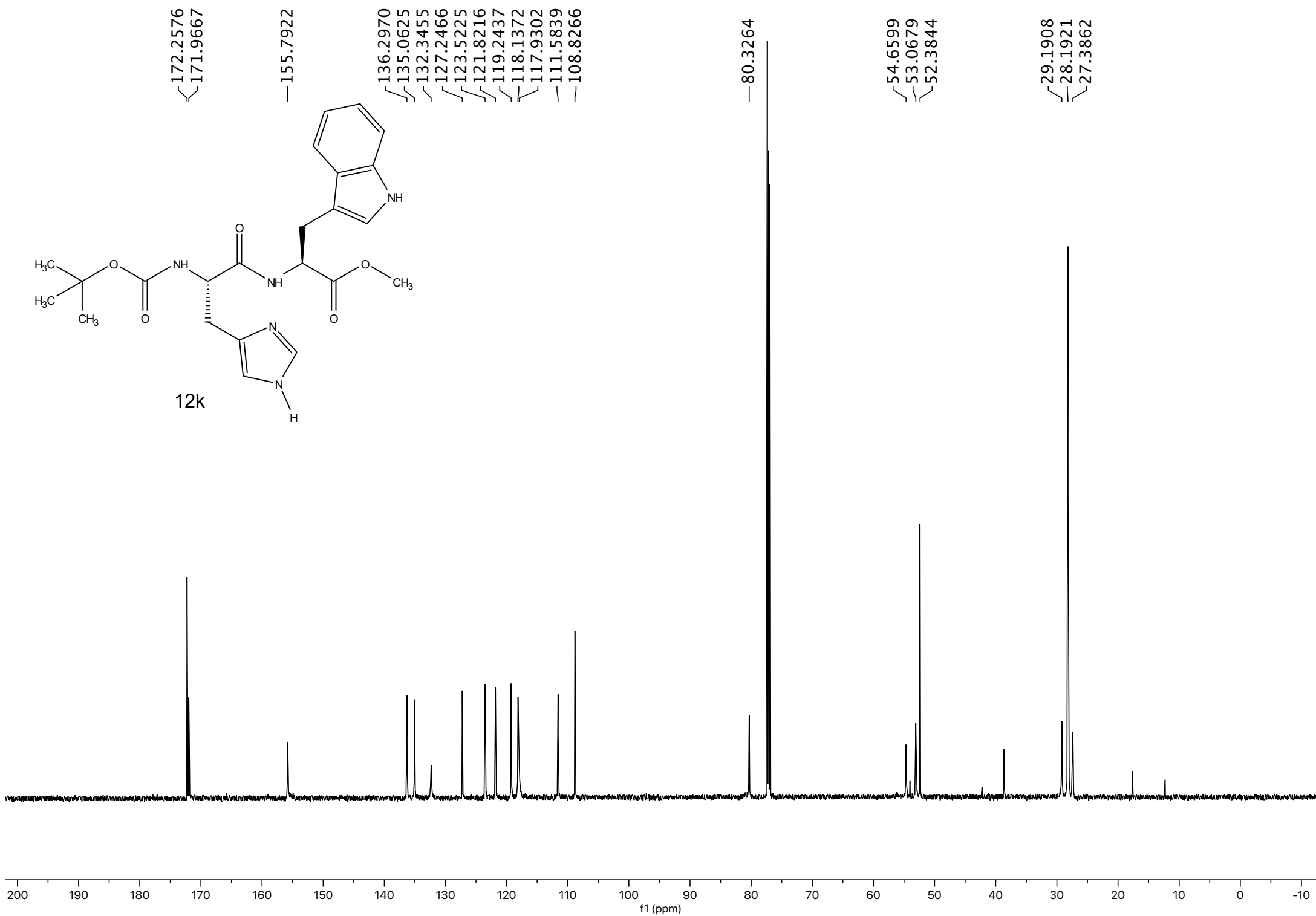
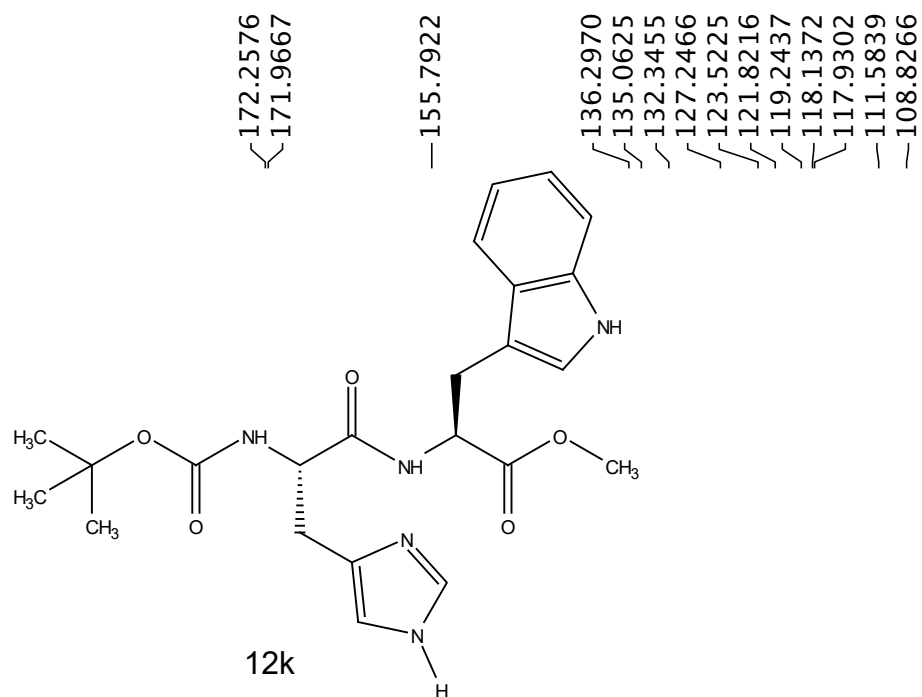


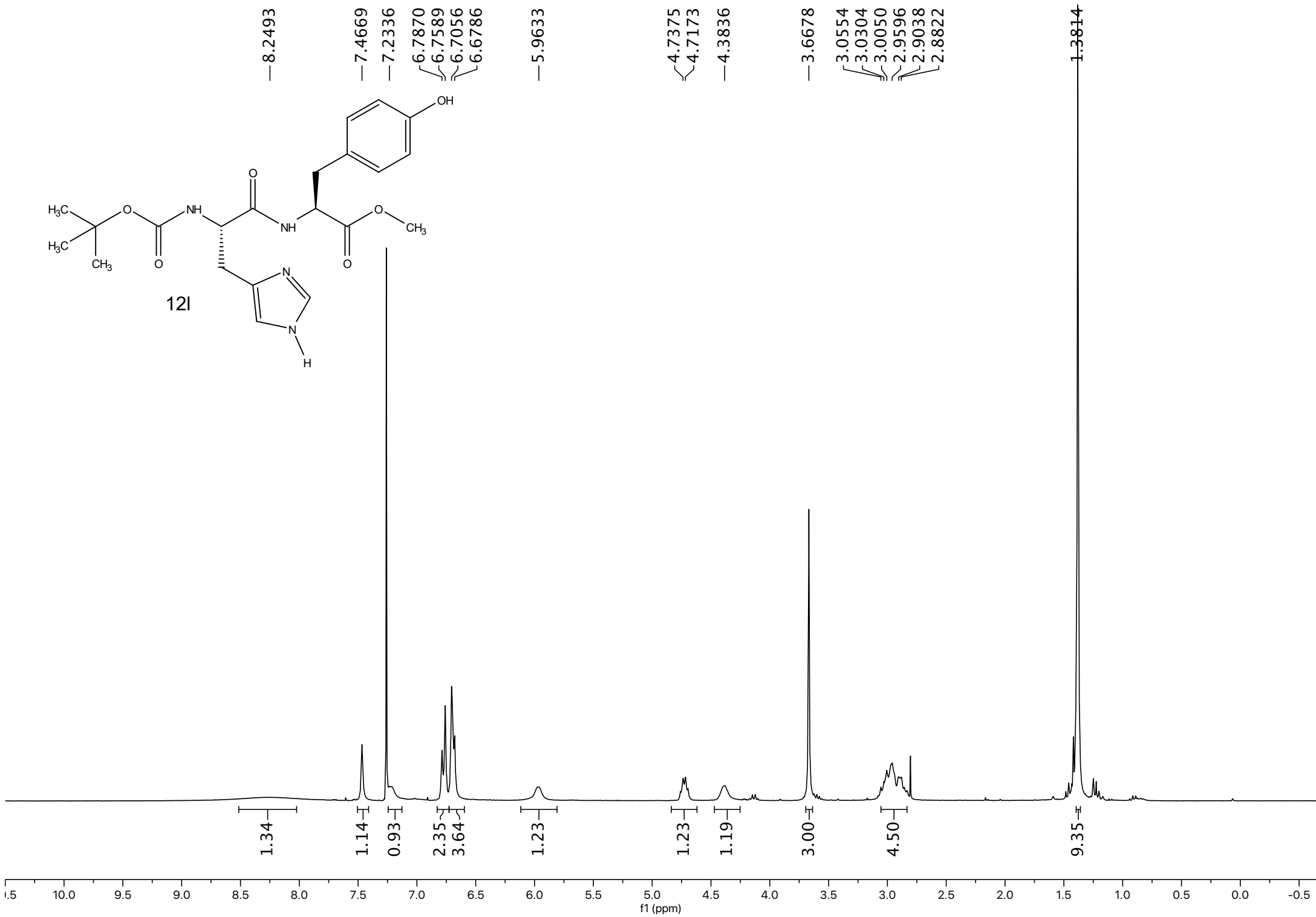


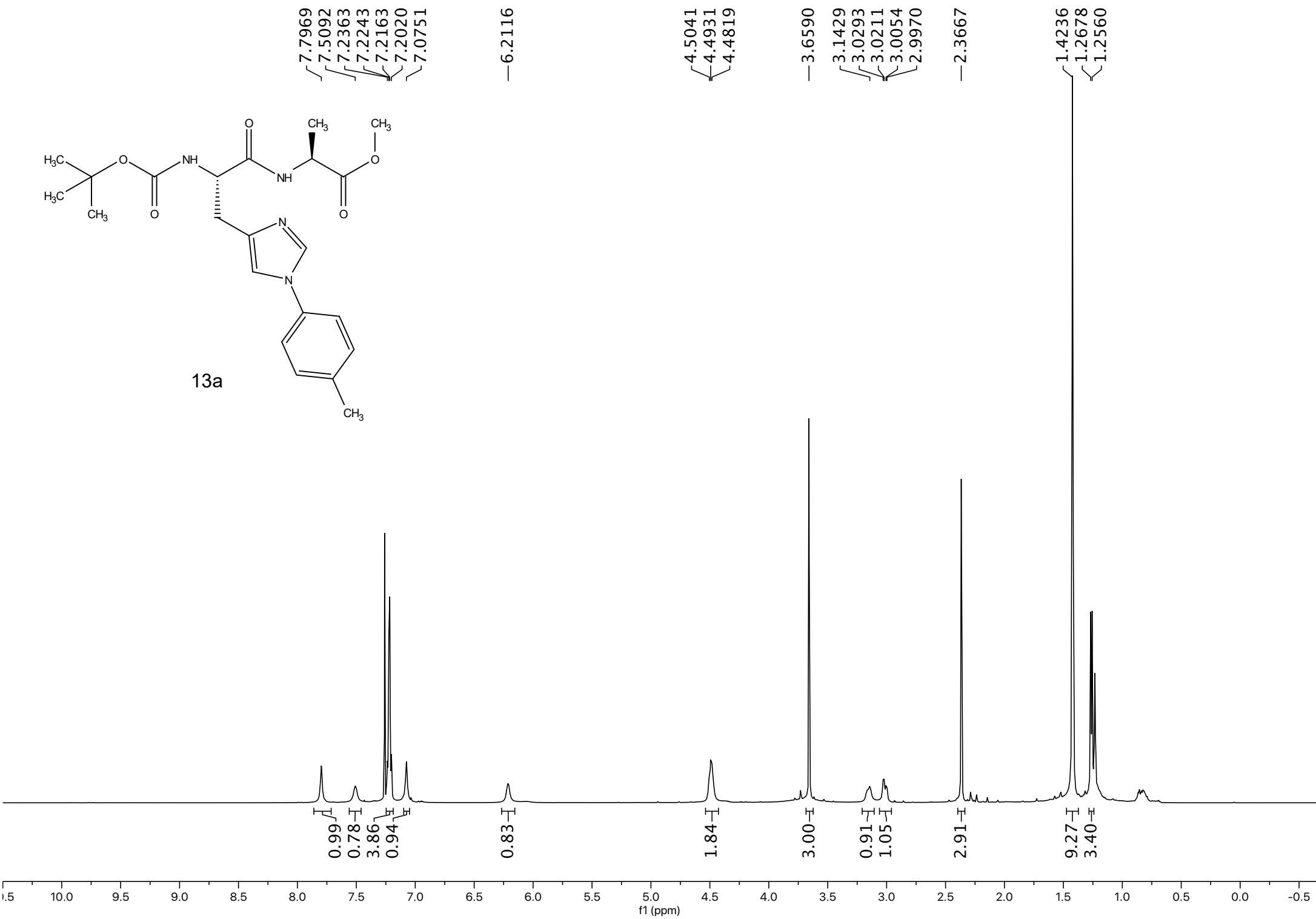


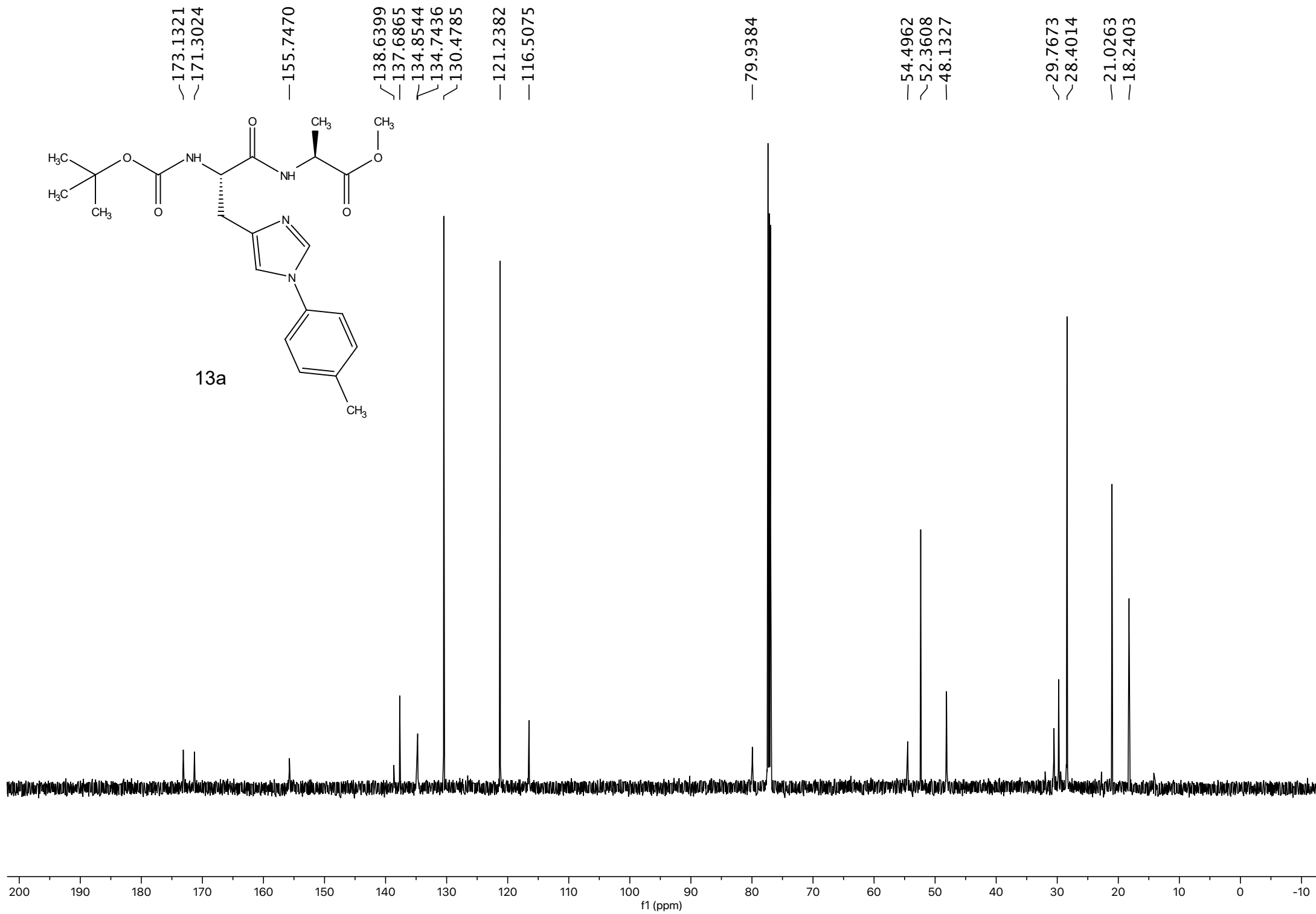




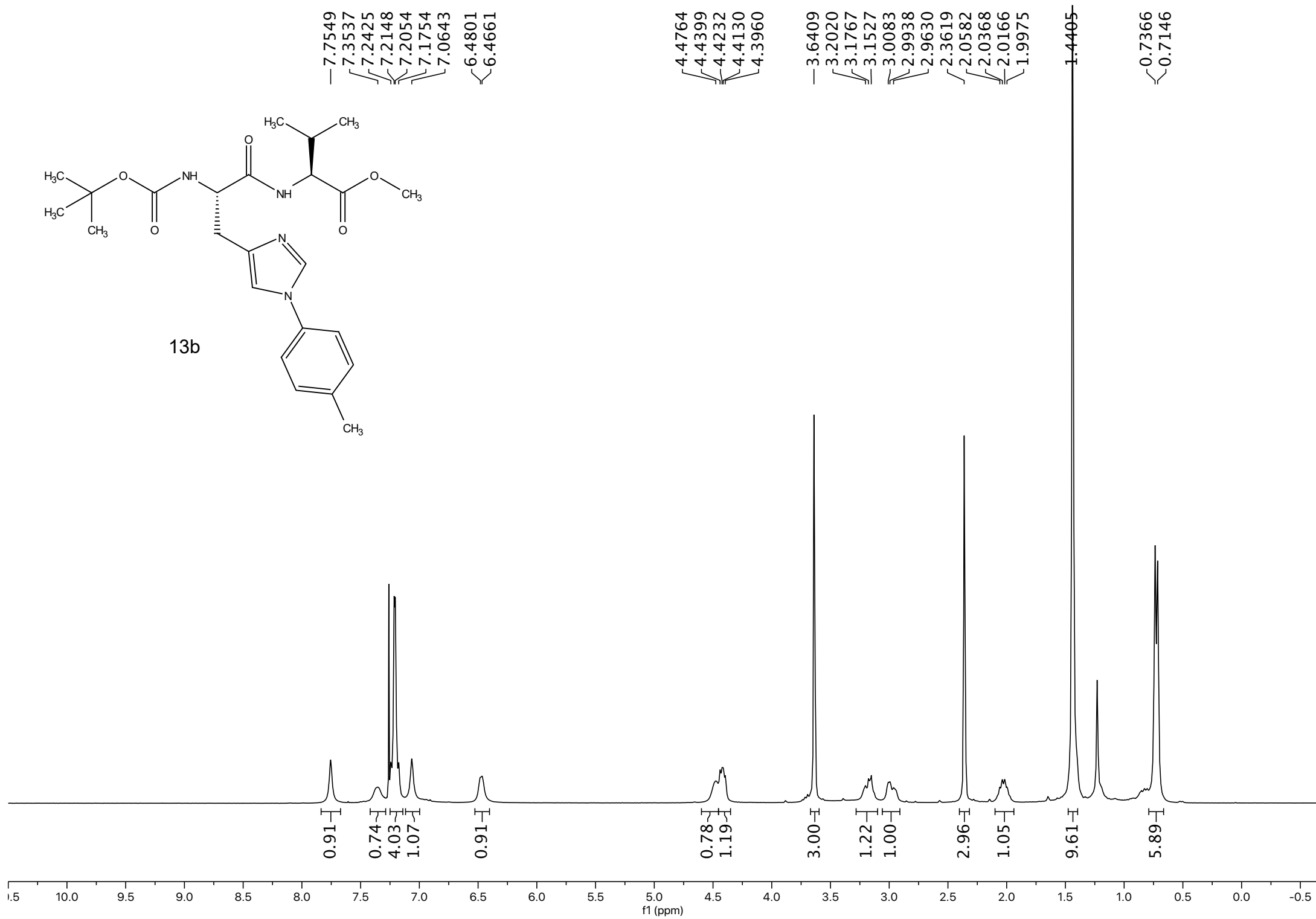
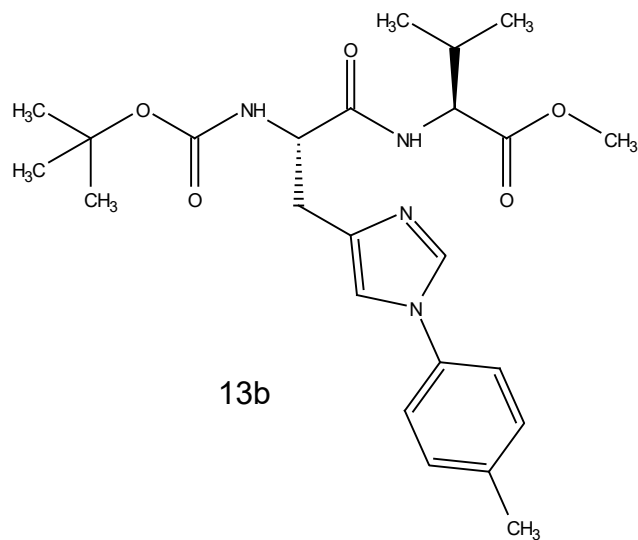


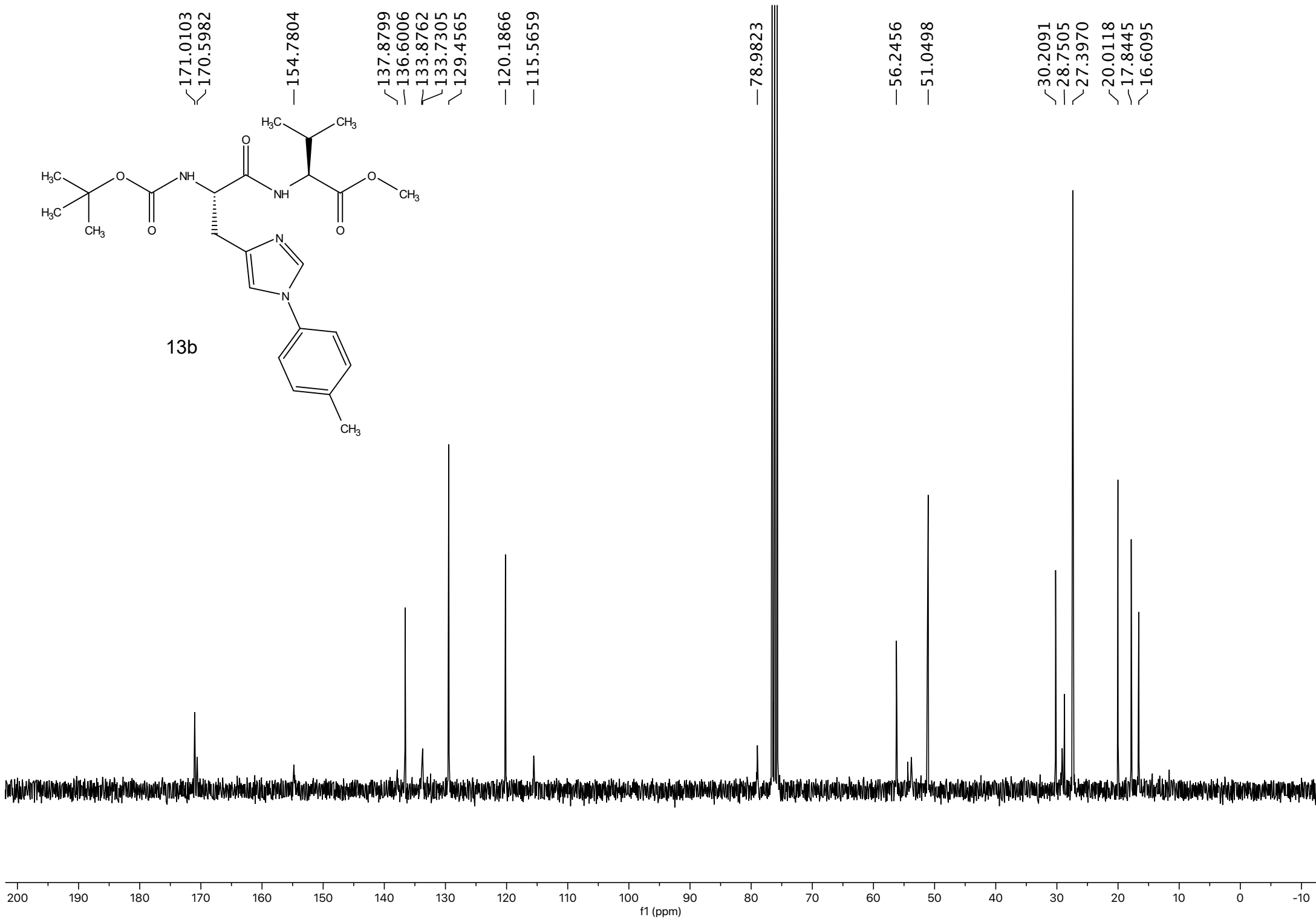


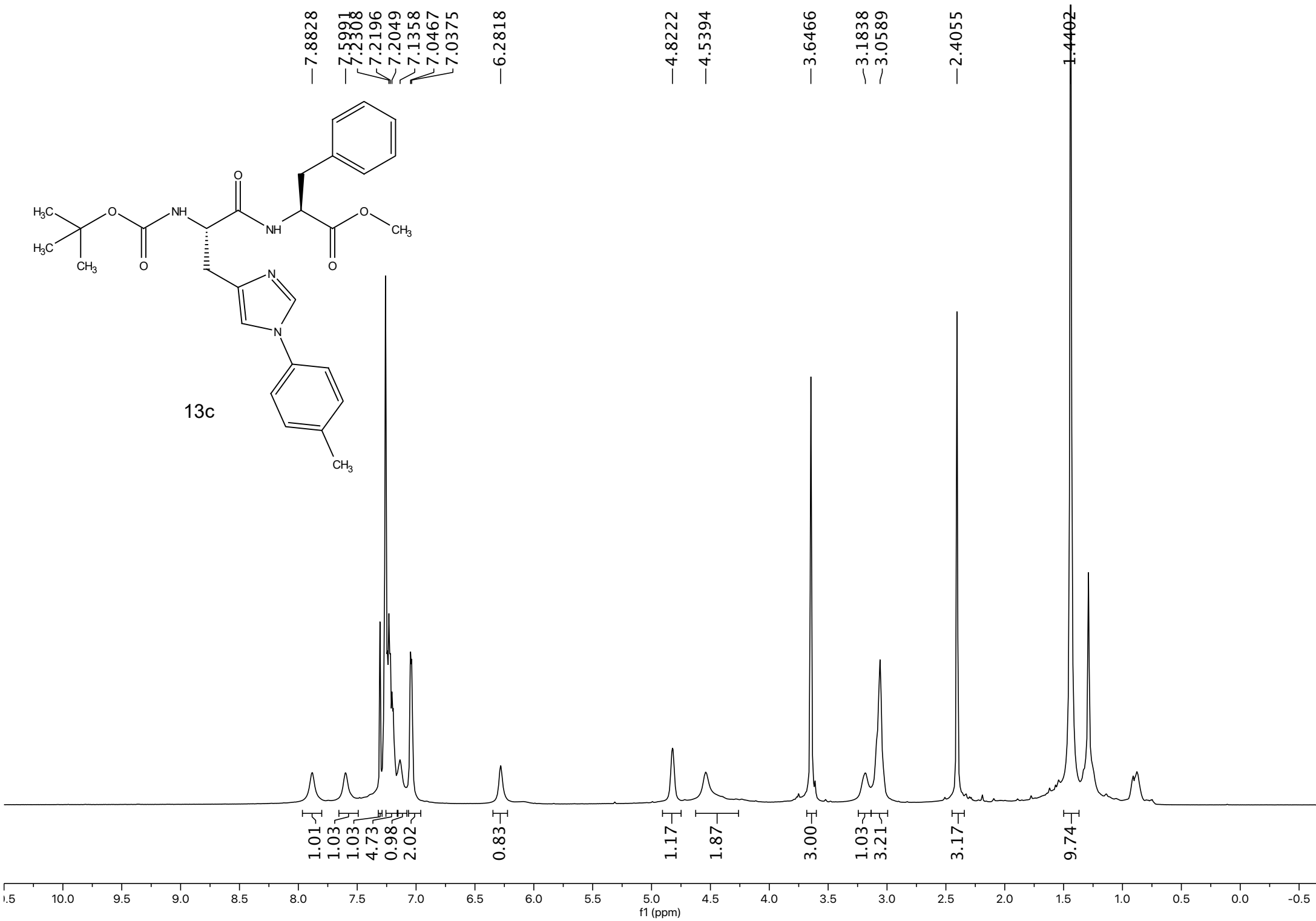


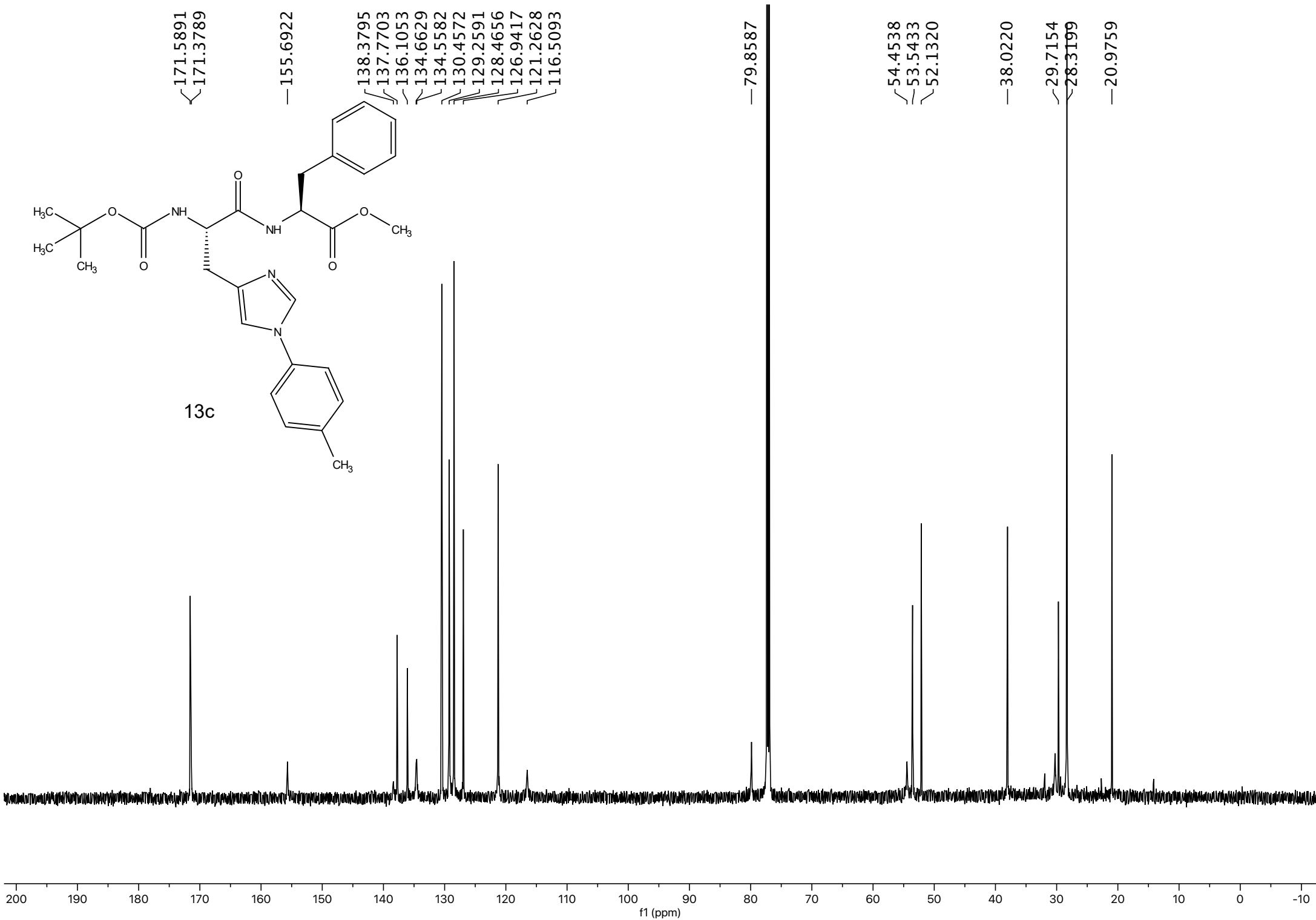


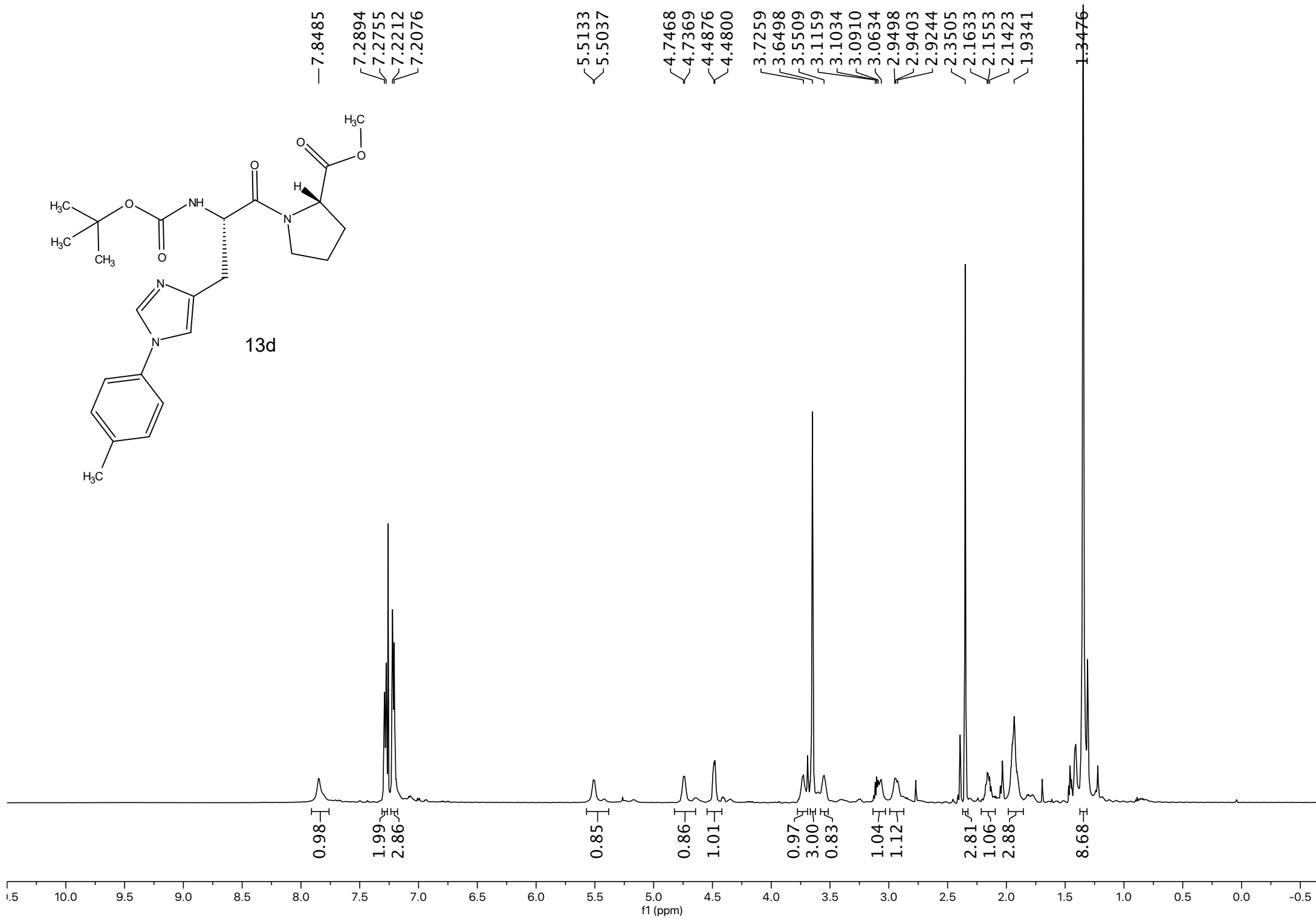
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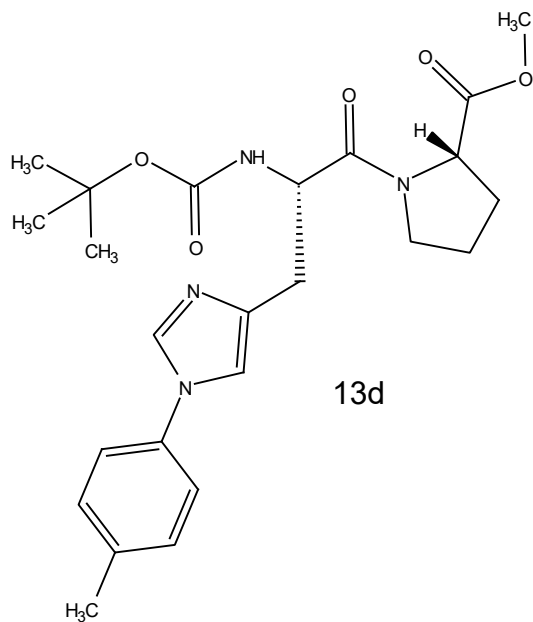












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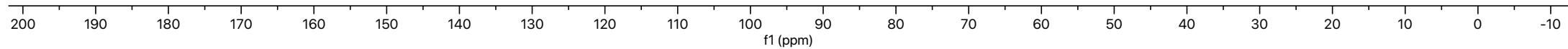
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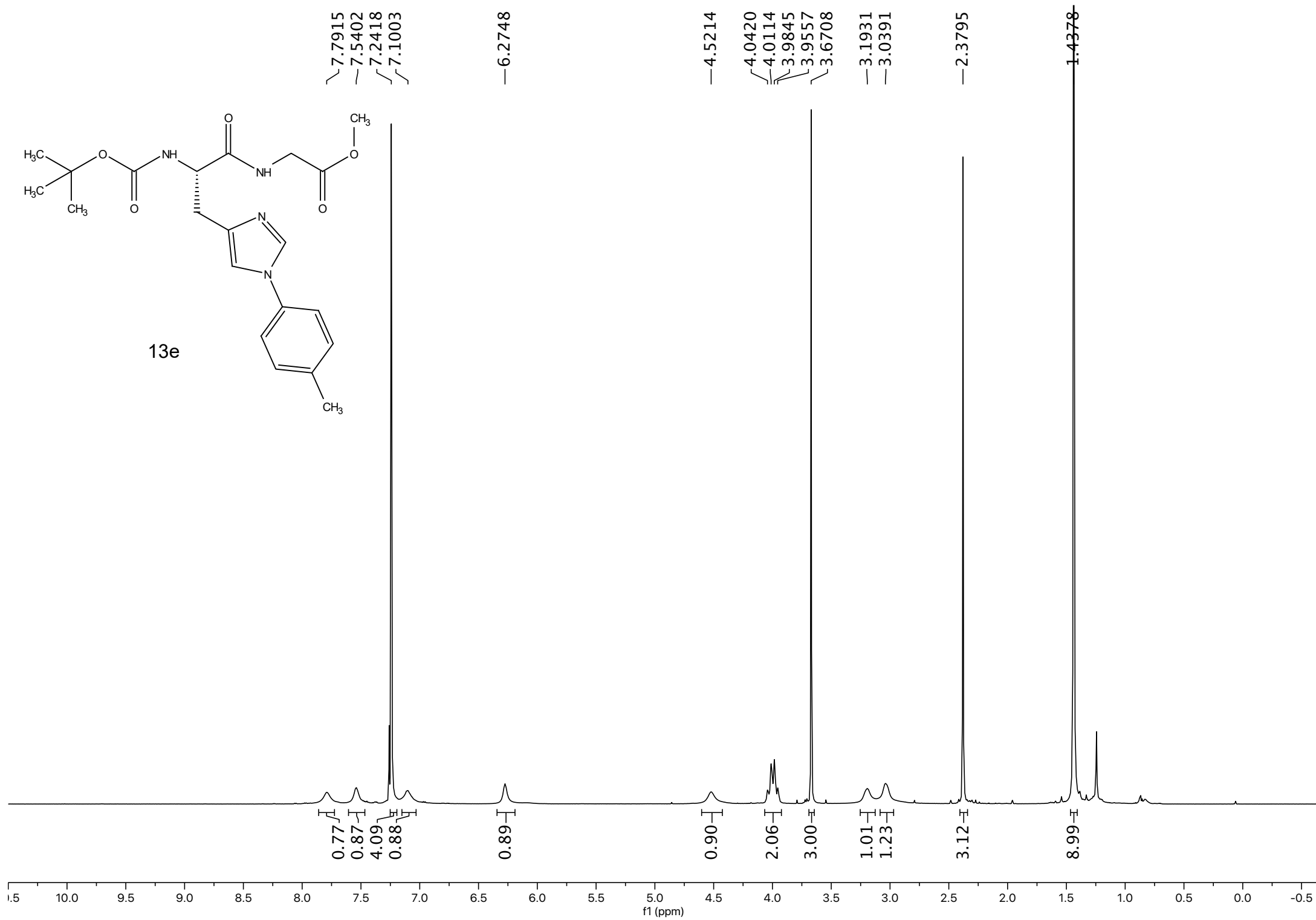
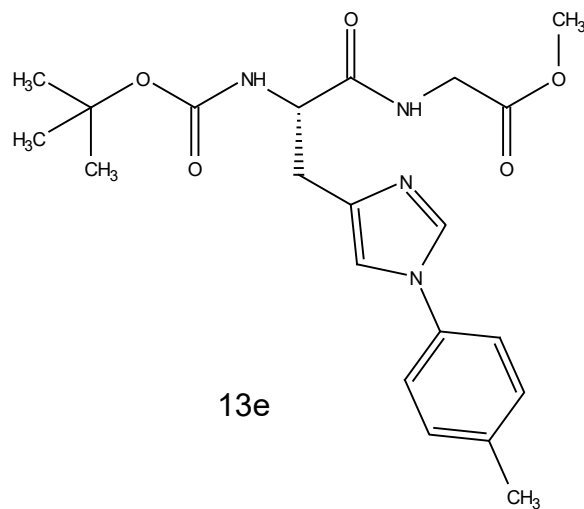
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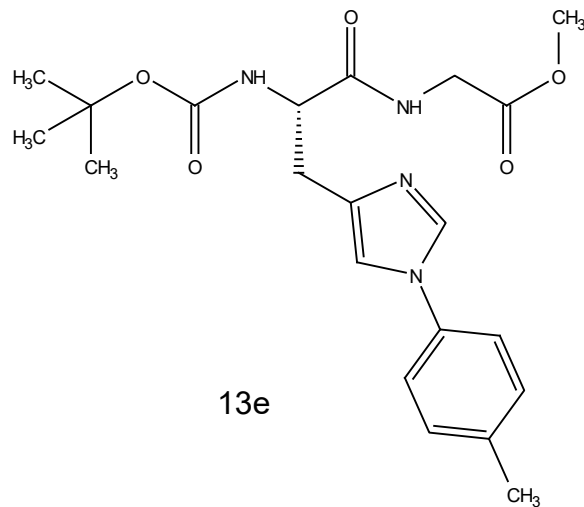
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13e



13e



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~130.4728

—121.3834

—116.4927

—80.0832

~54.5887

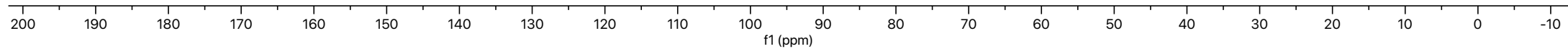
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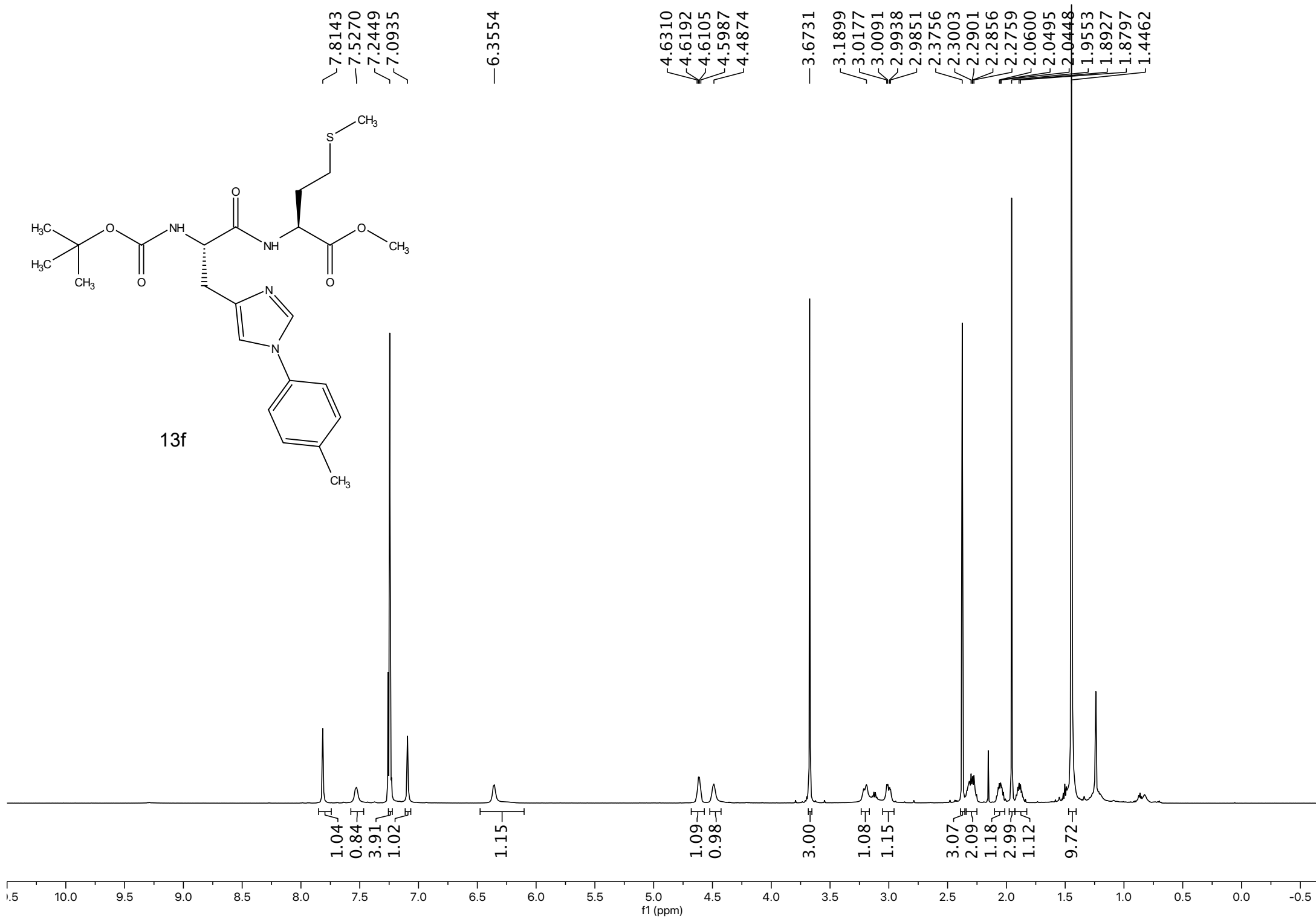
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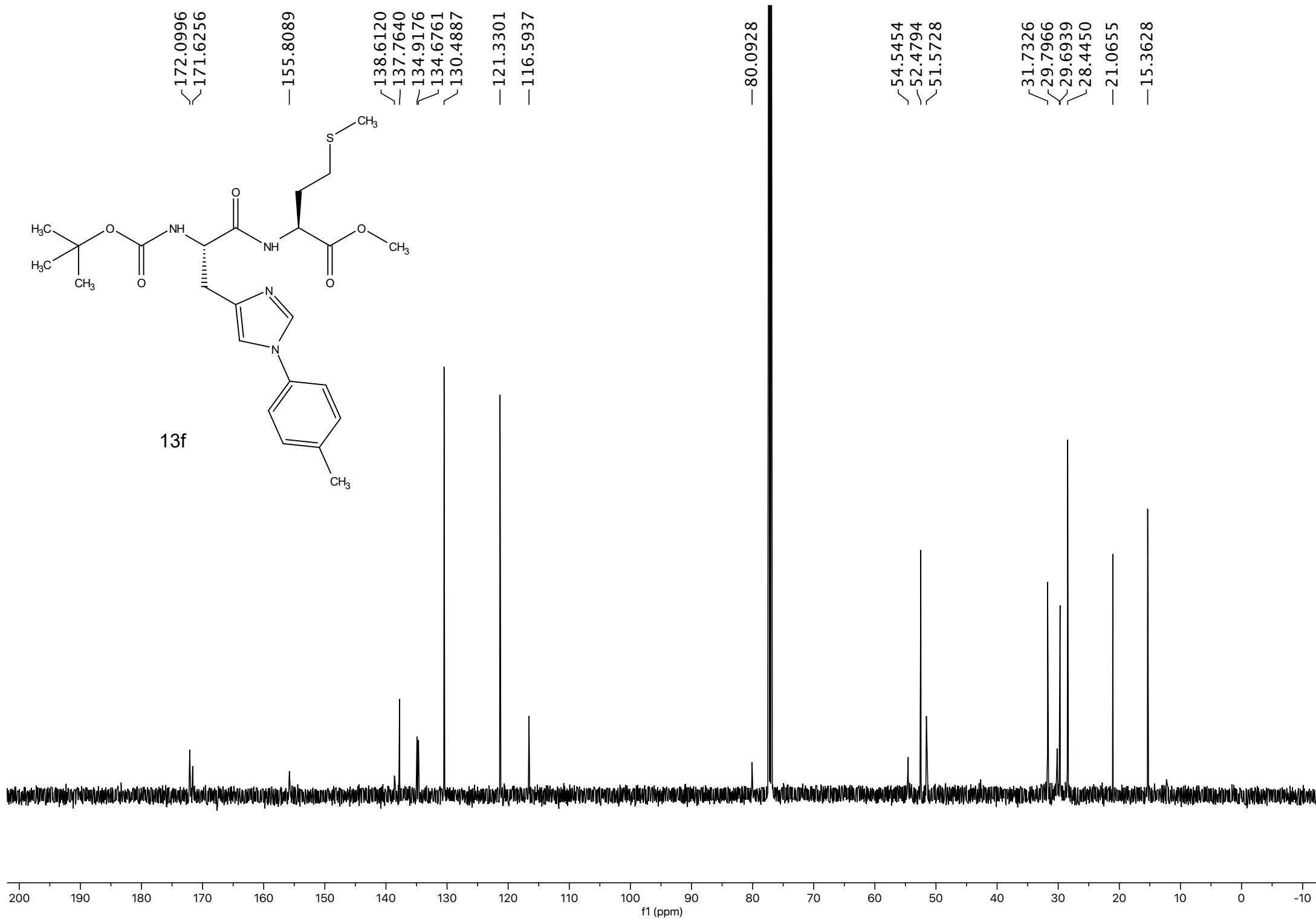
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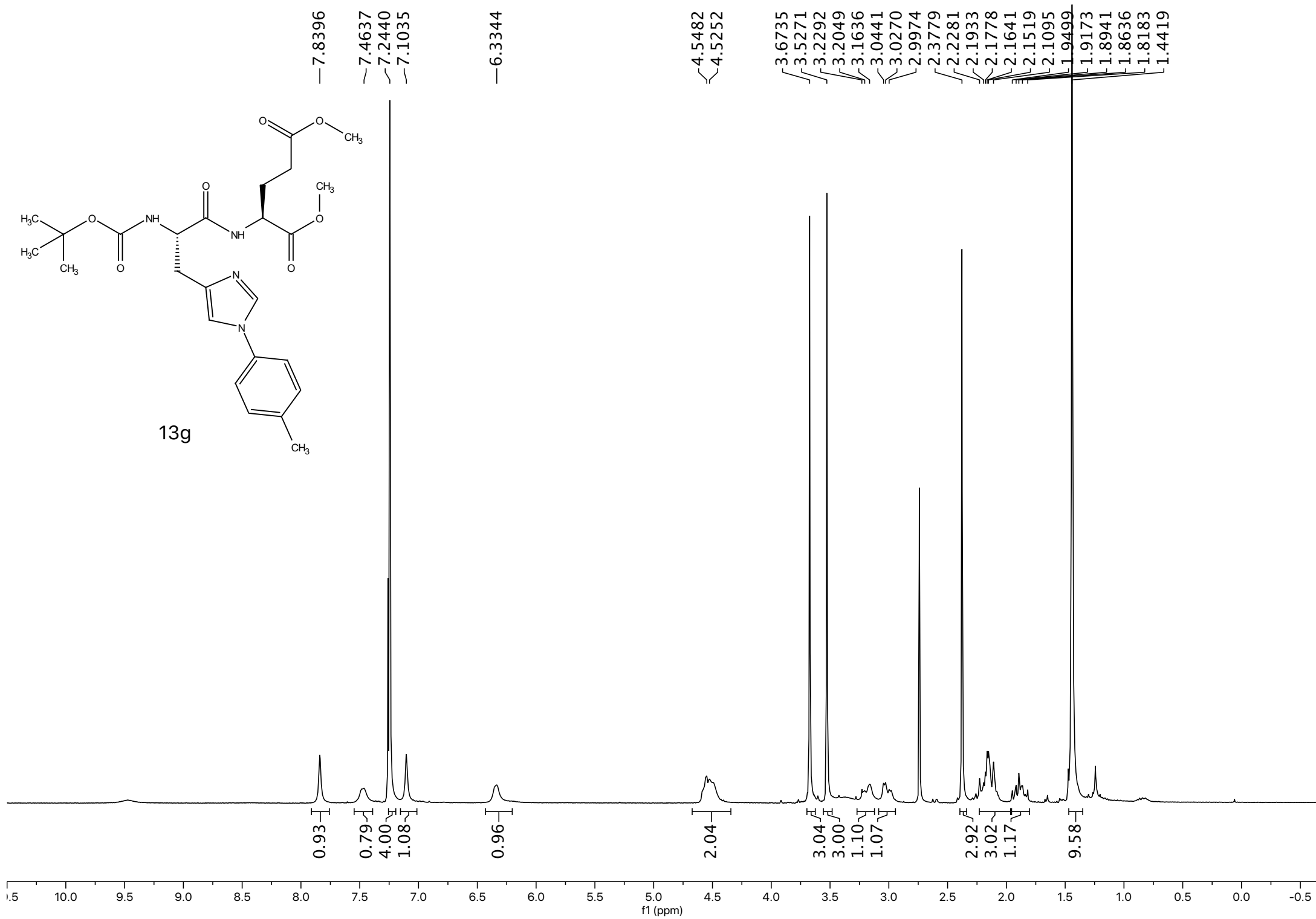
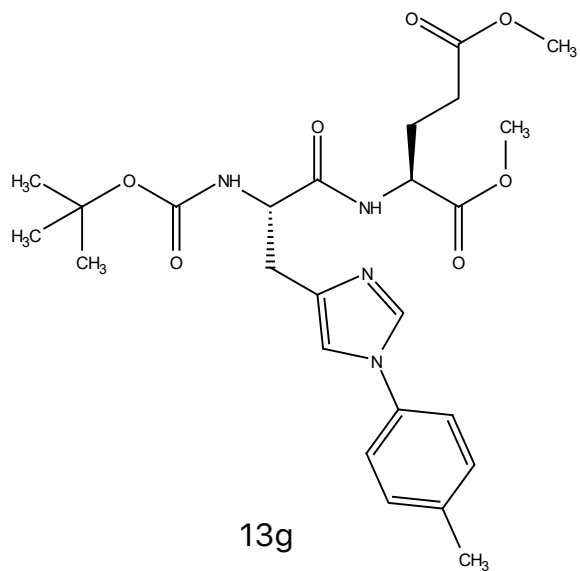
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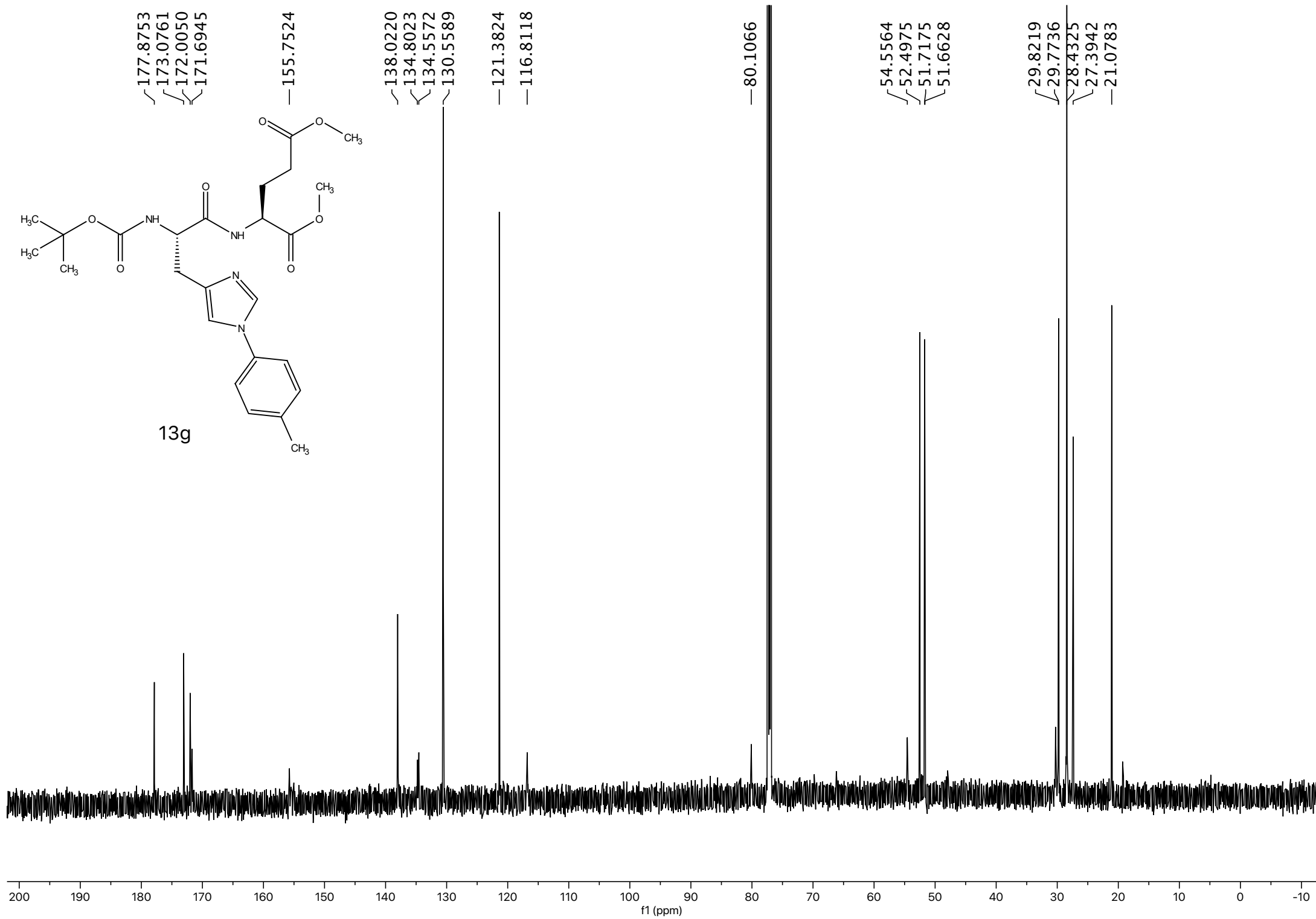
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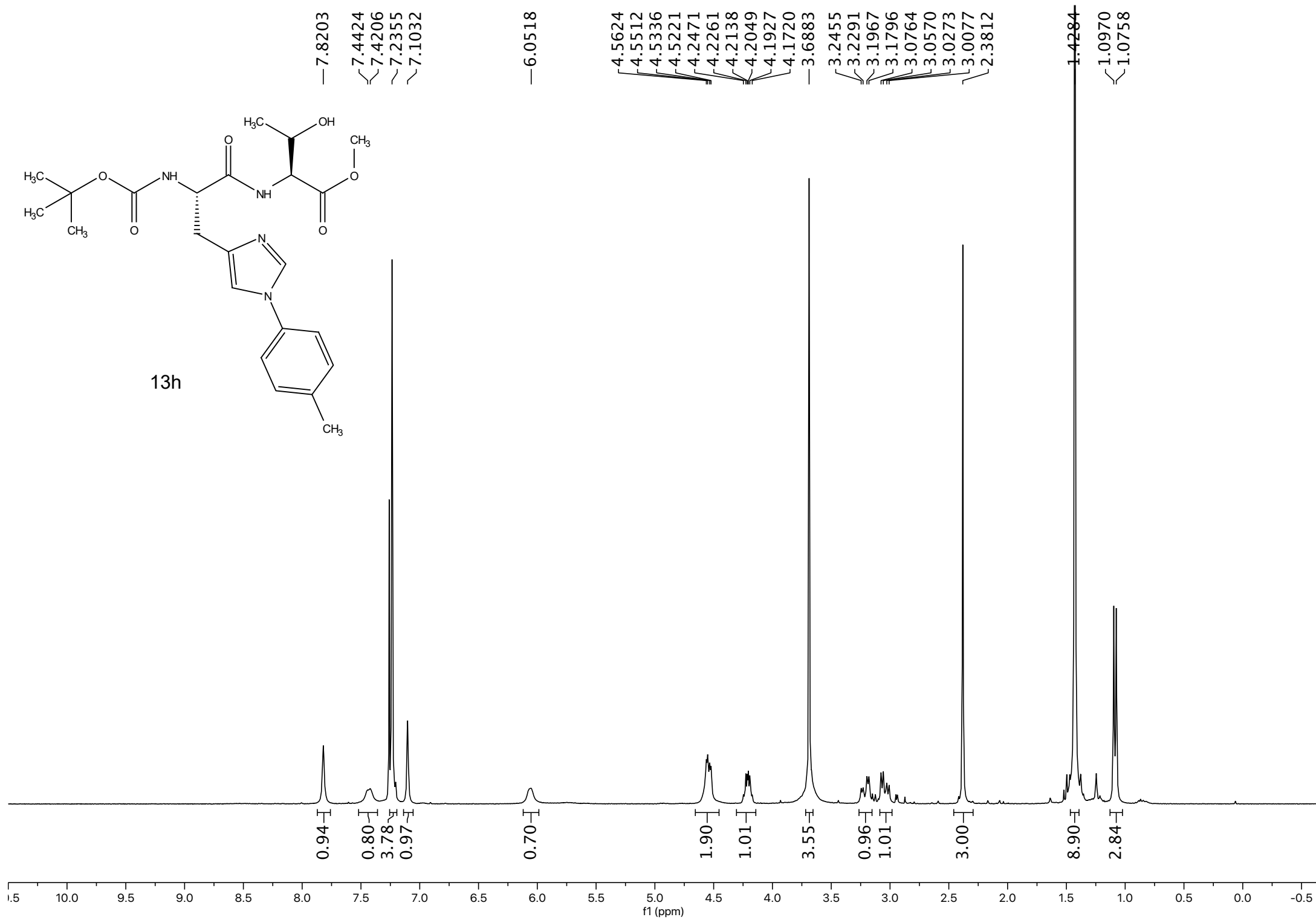
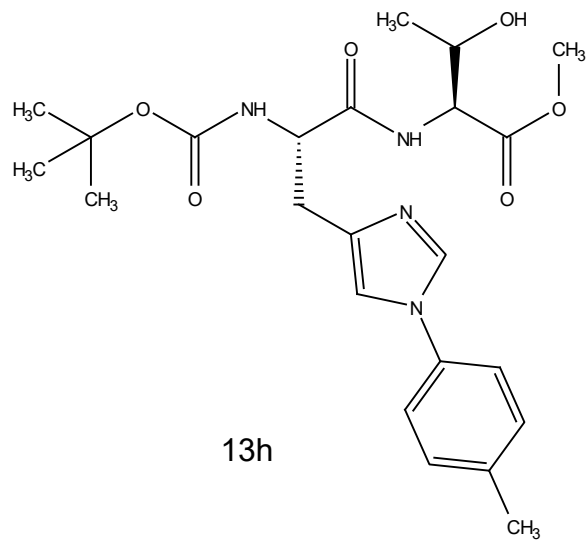




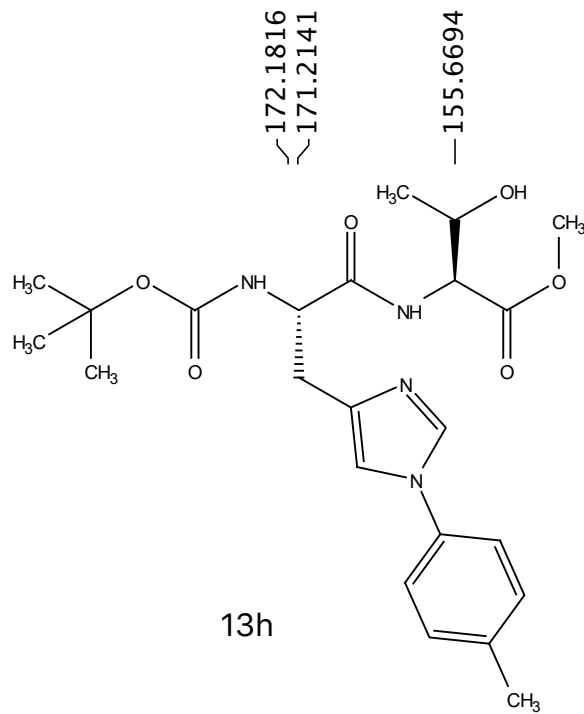




13h



13h



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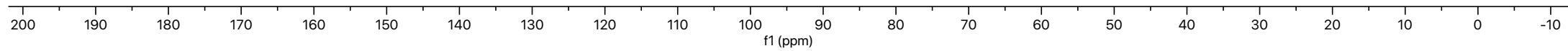
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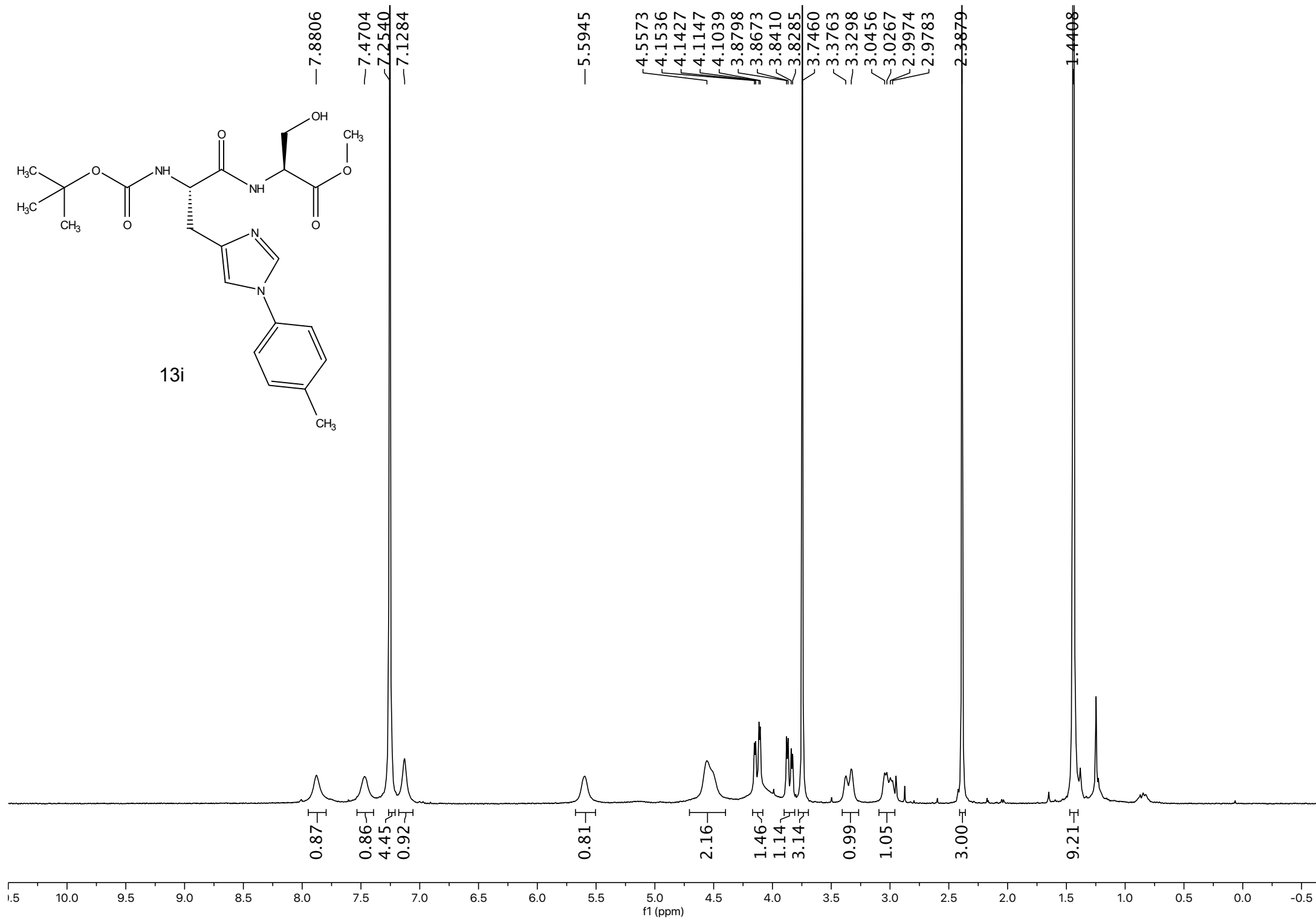
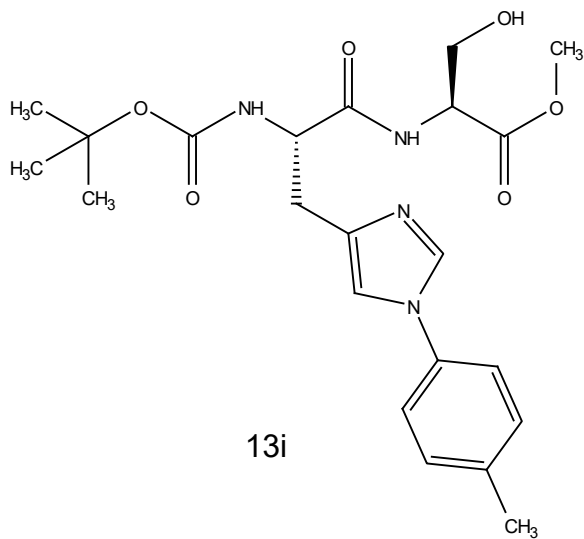
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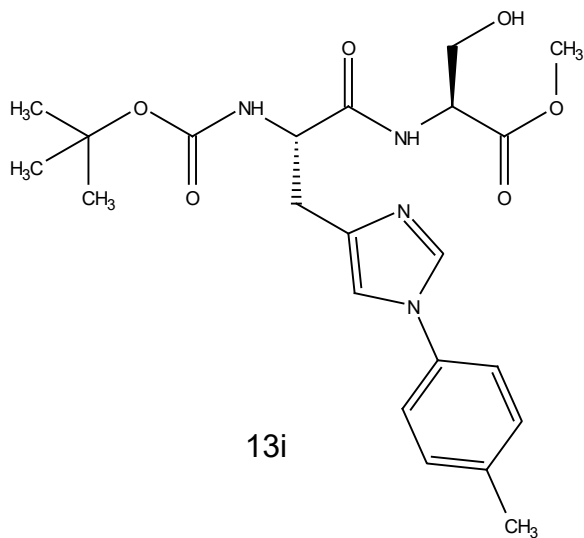
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13i



13i



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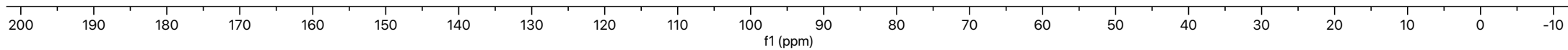
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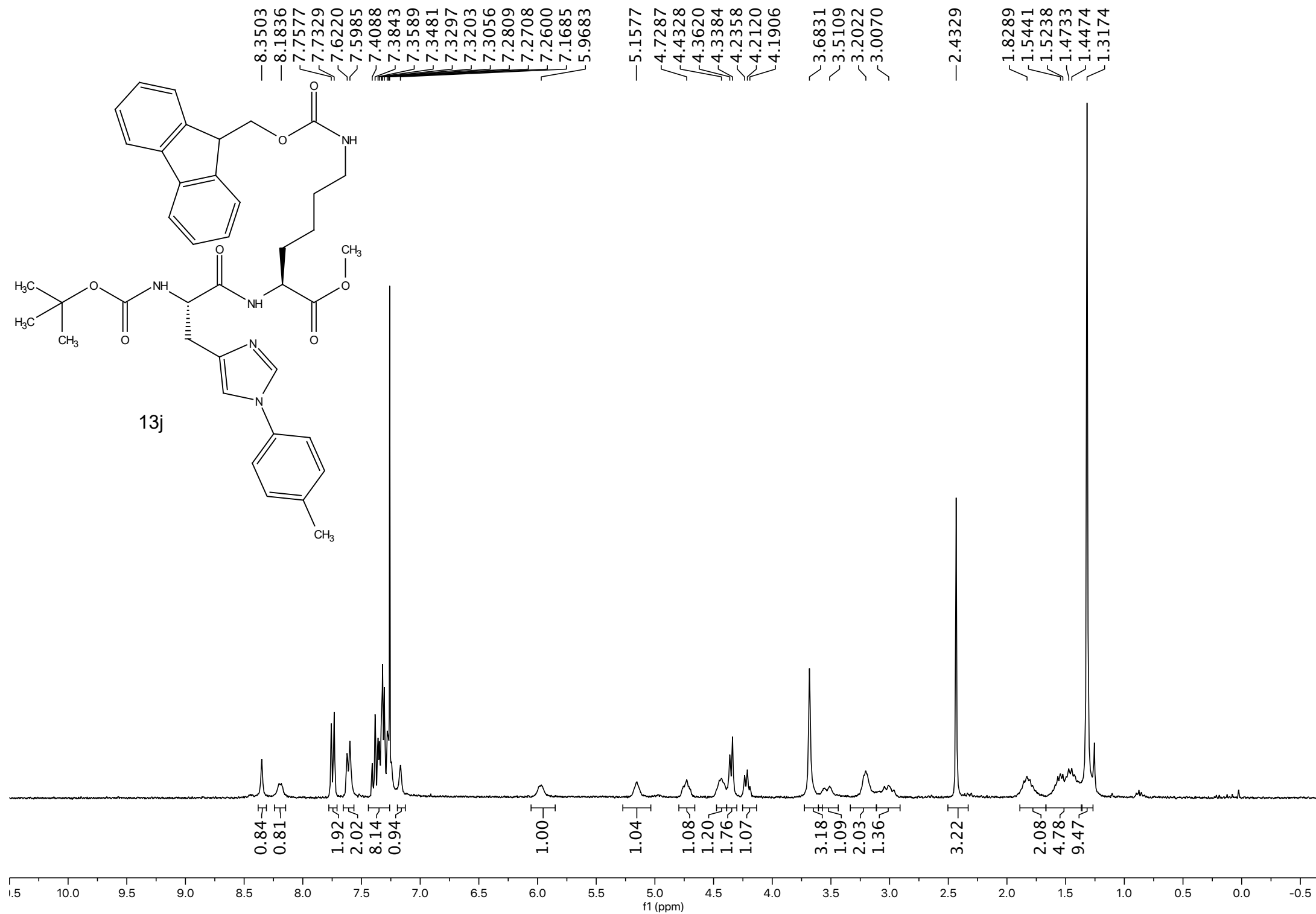
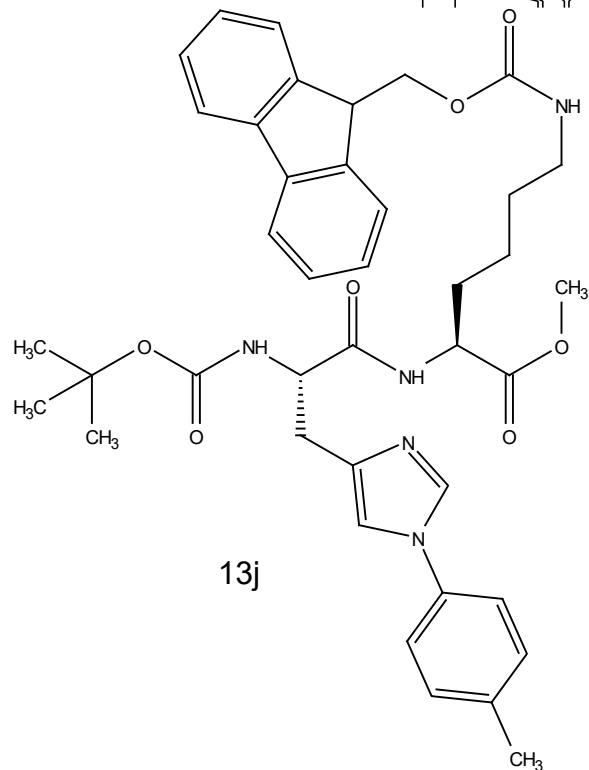
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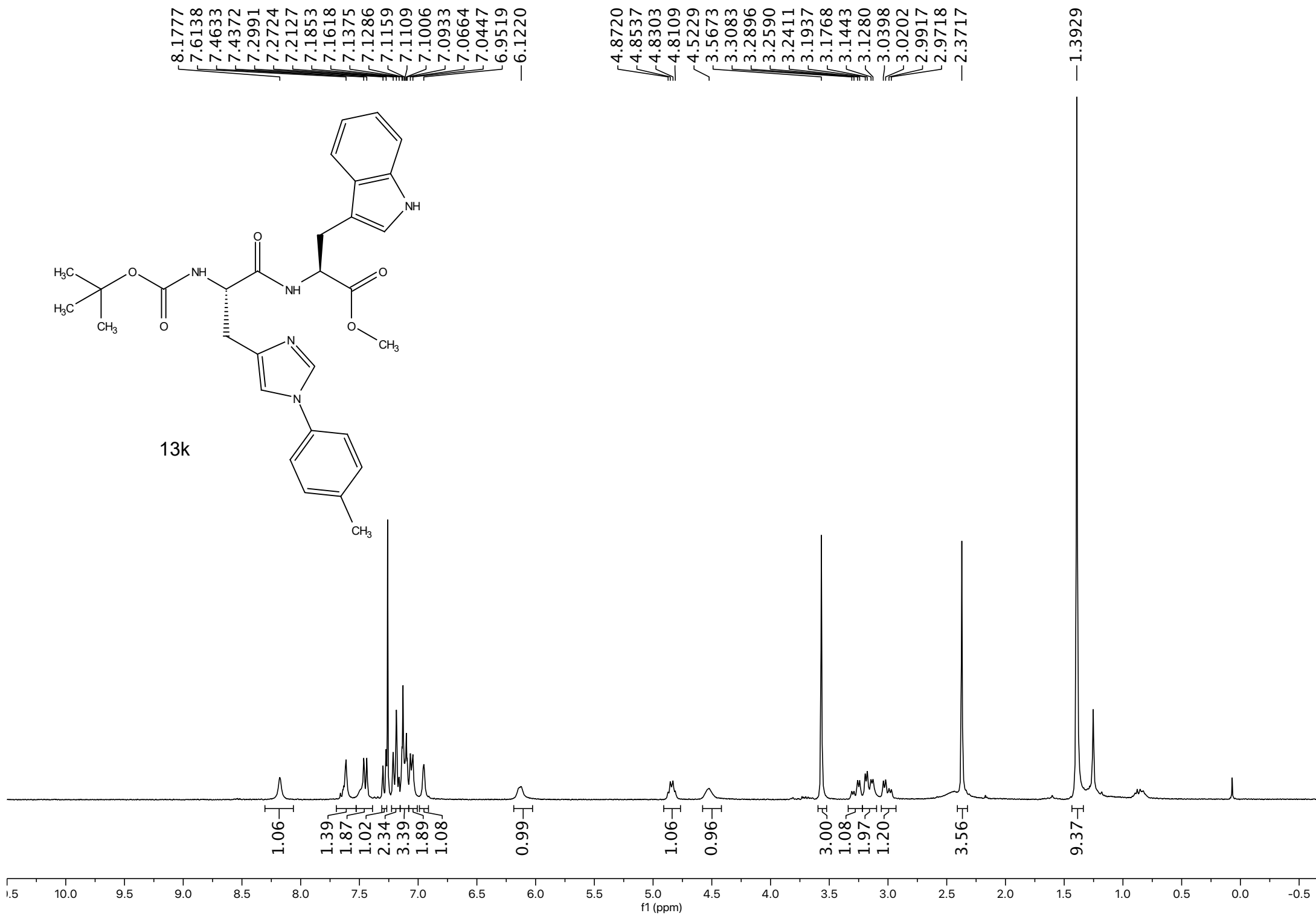
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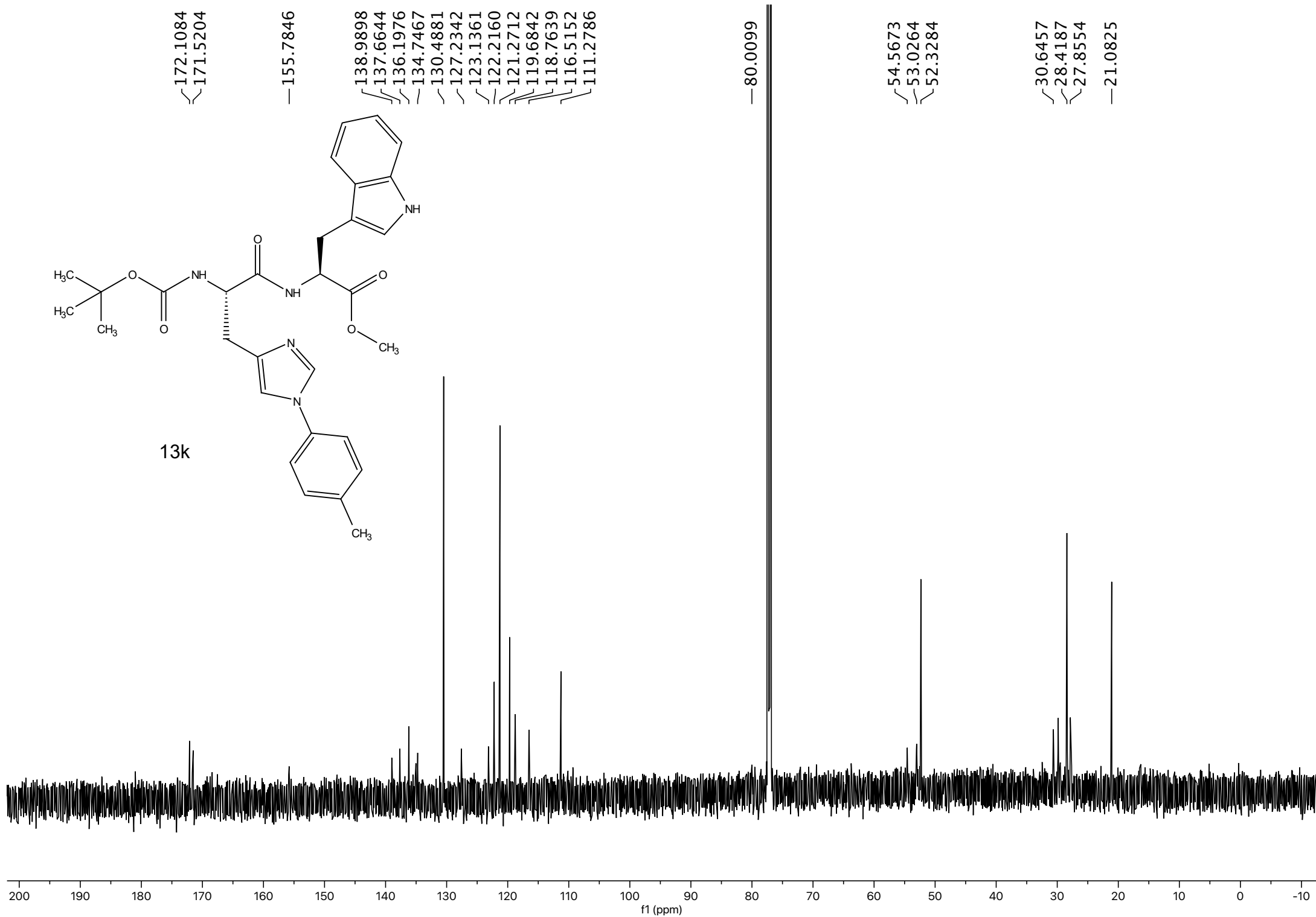
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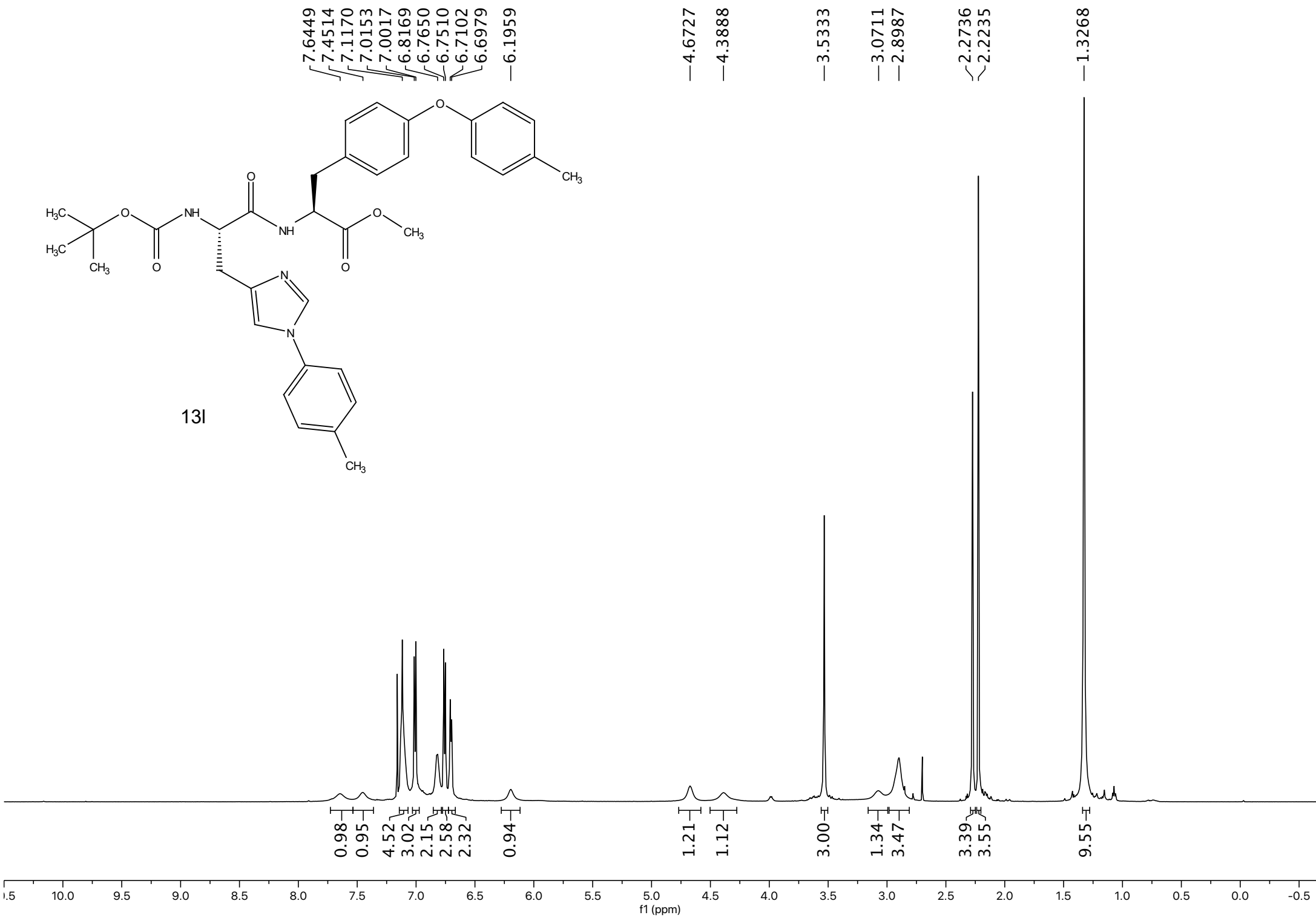


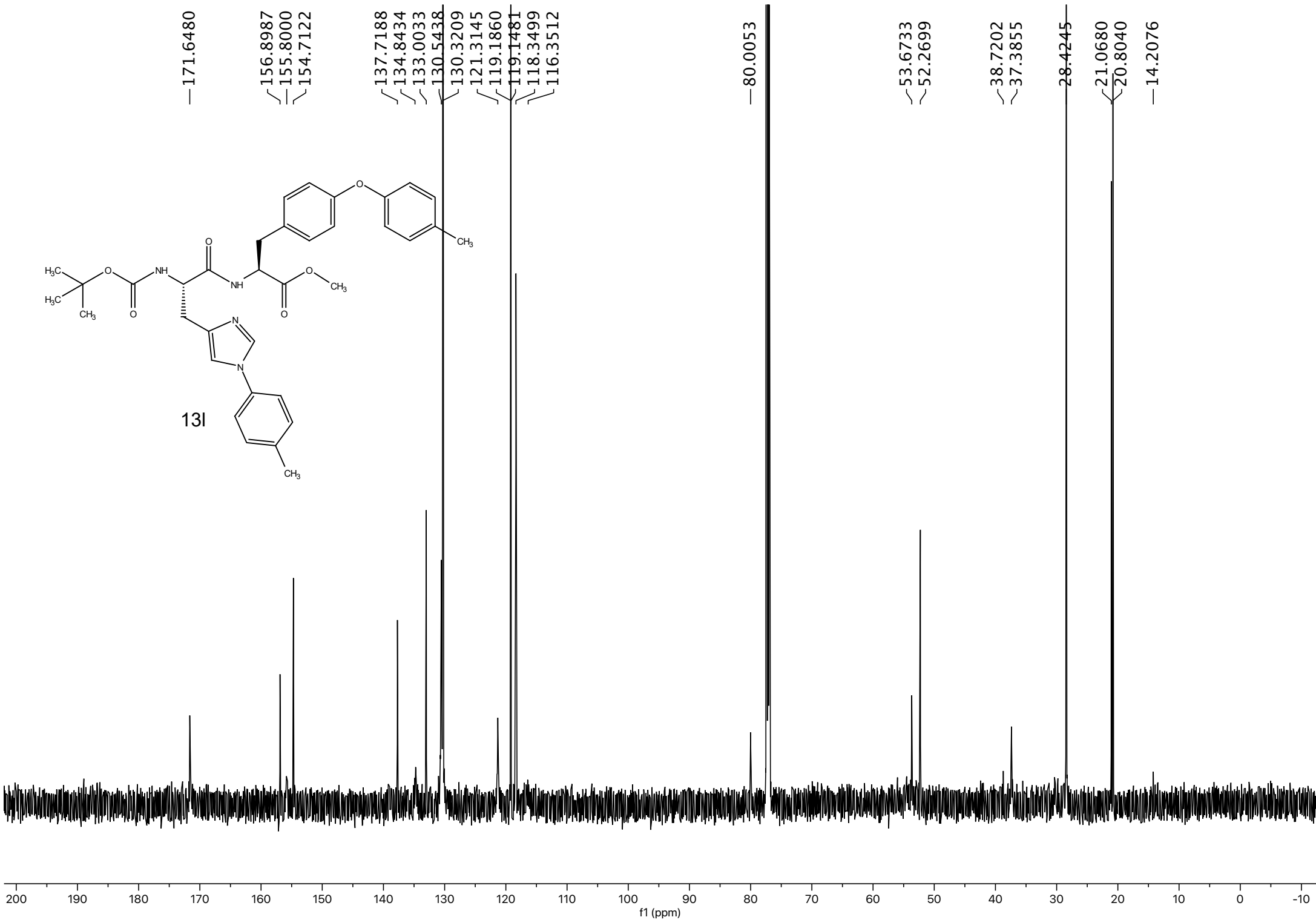
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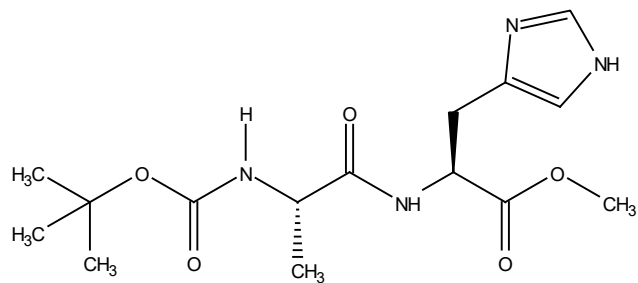




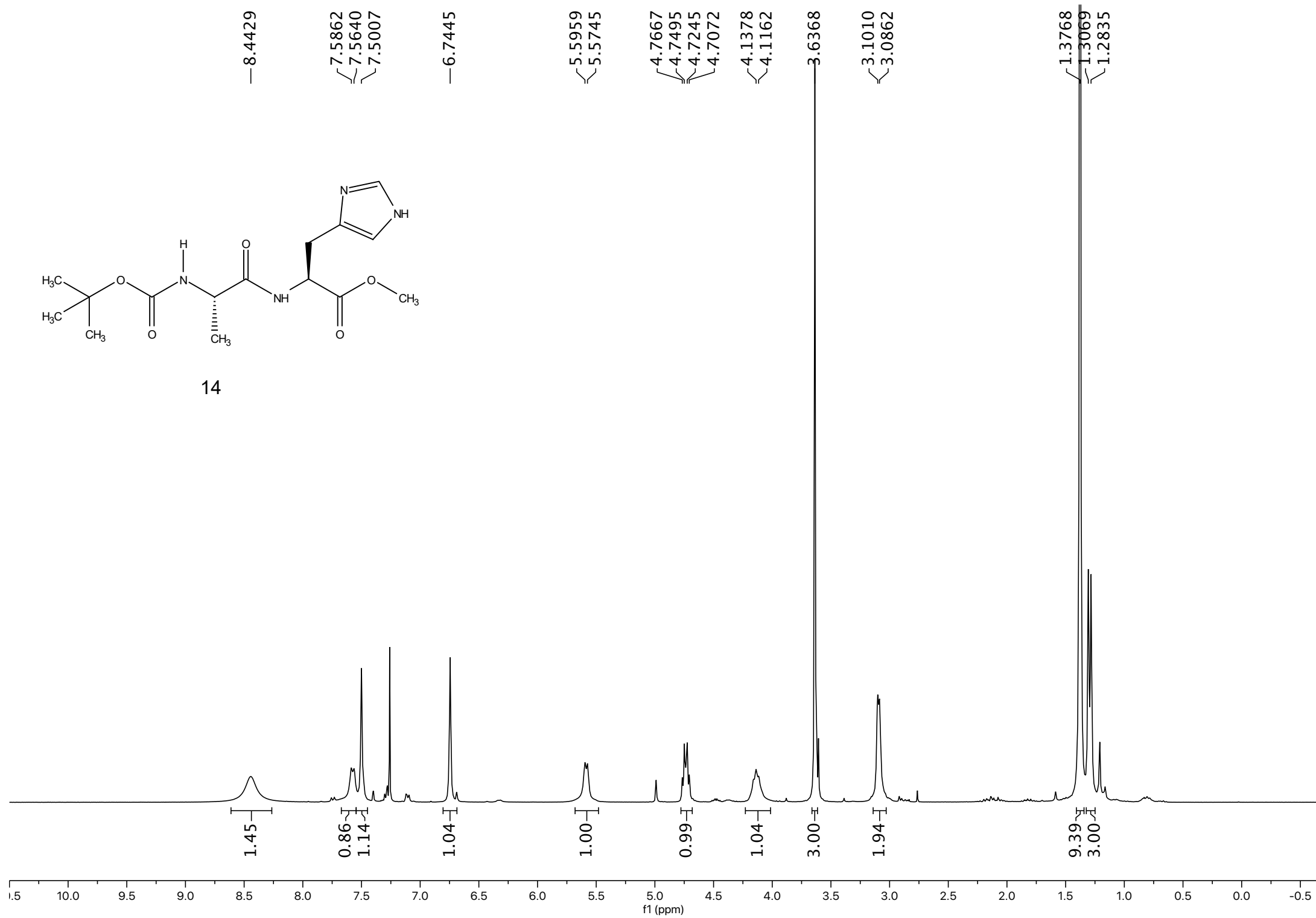


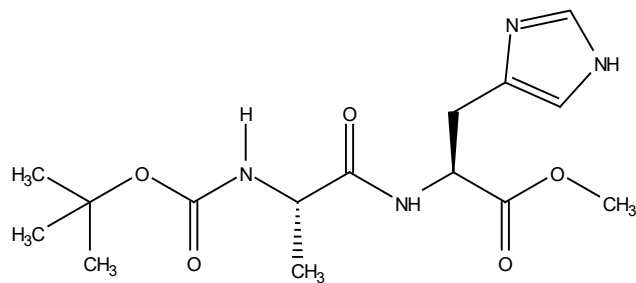




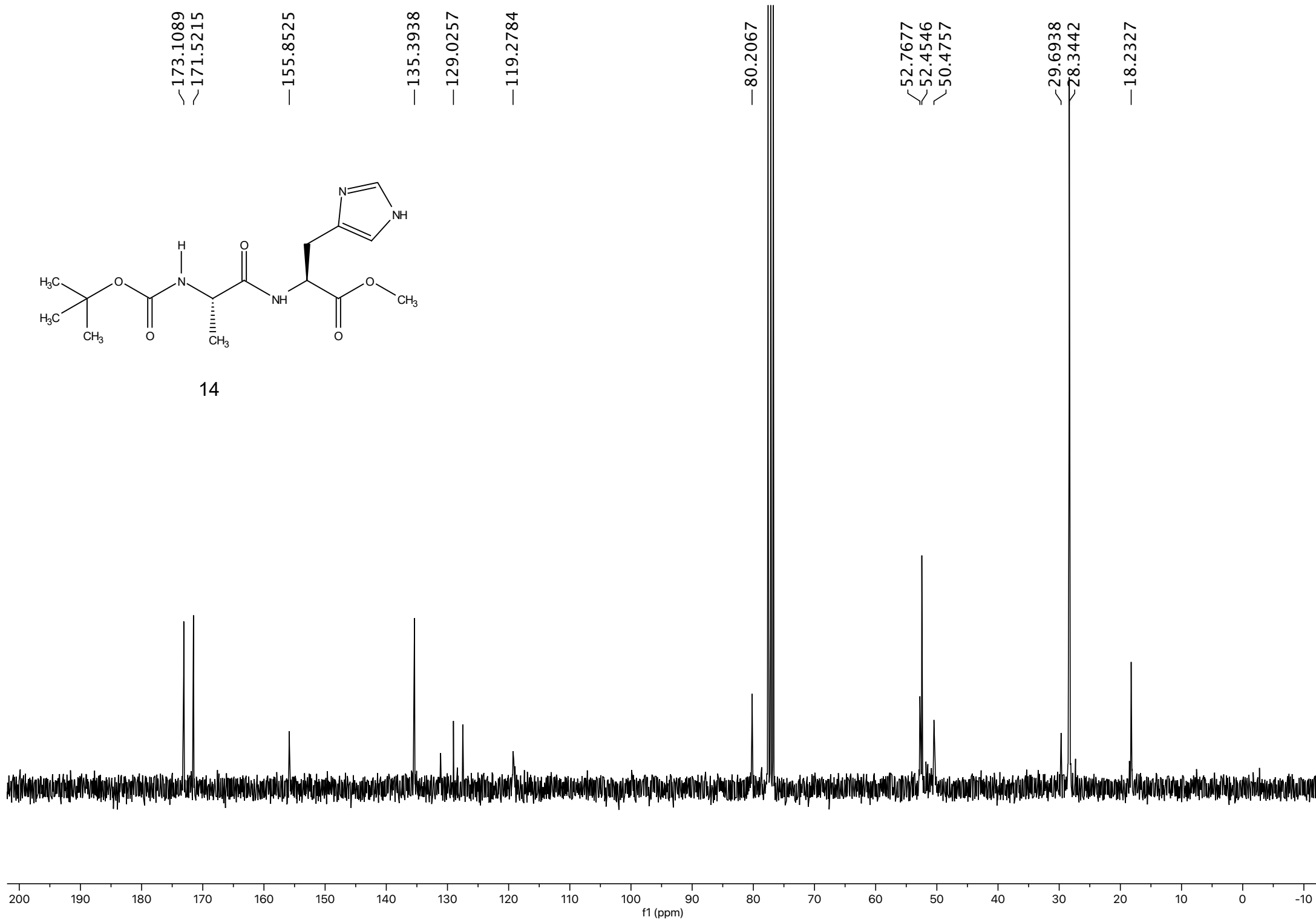


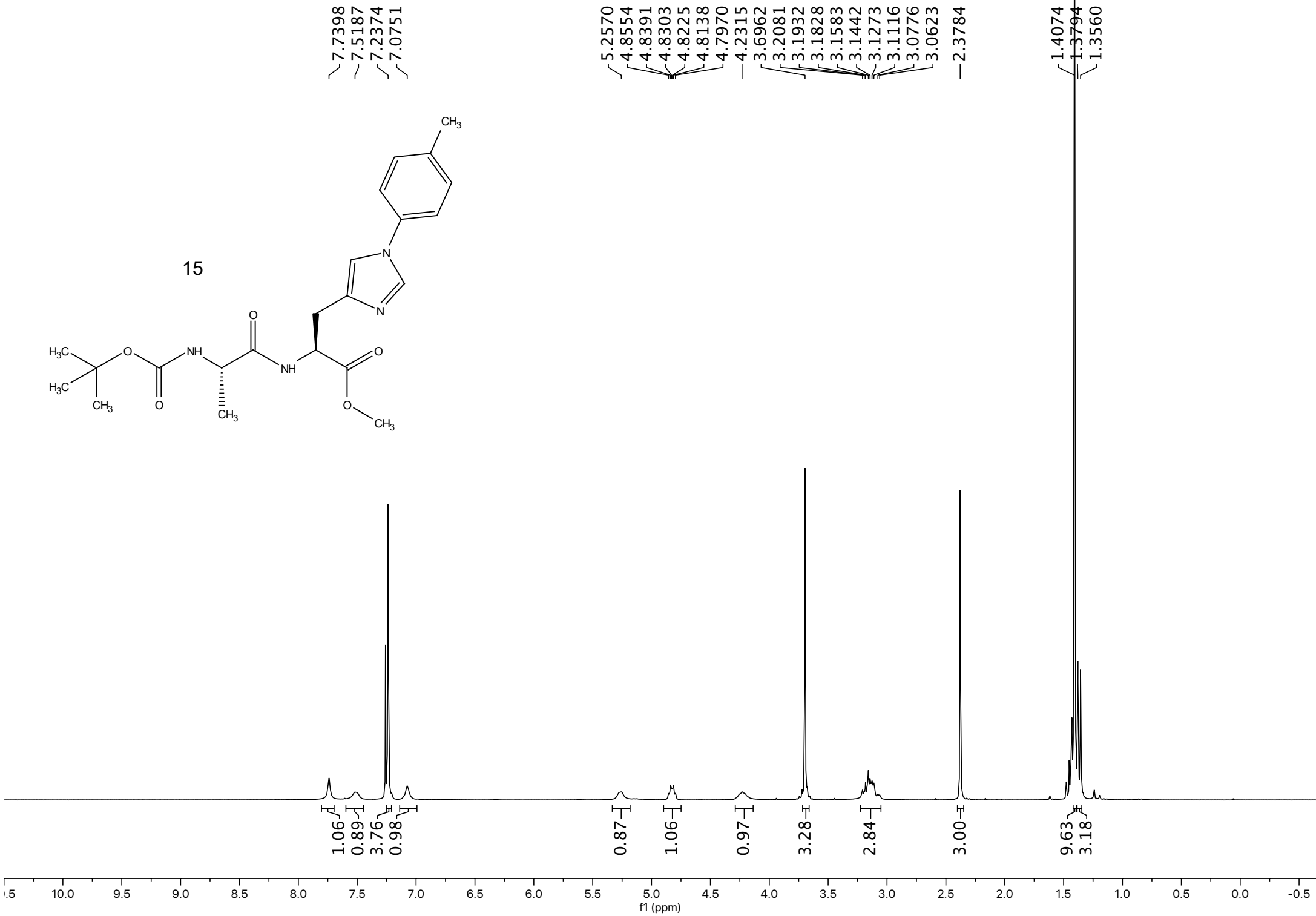
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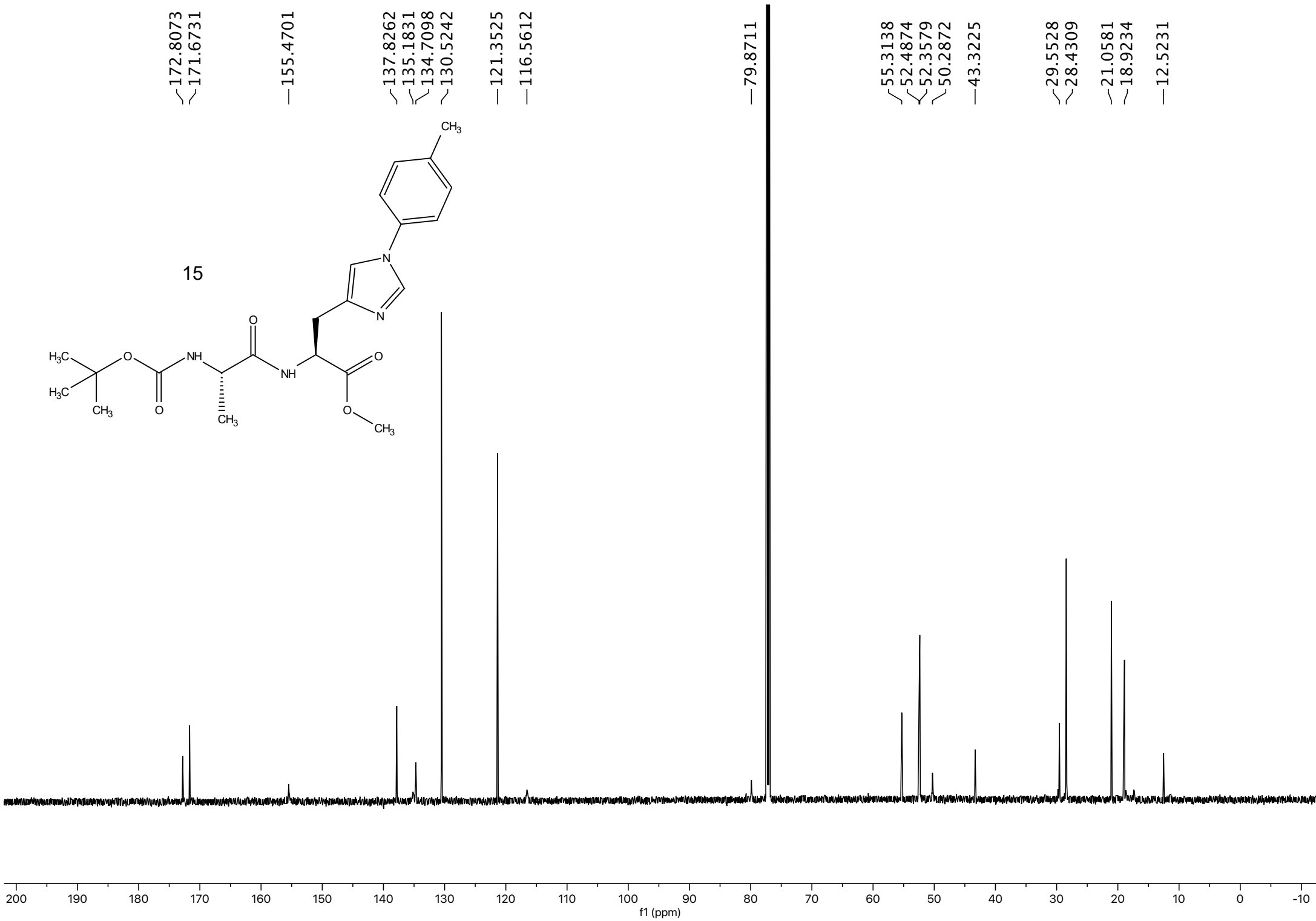


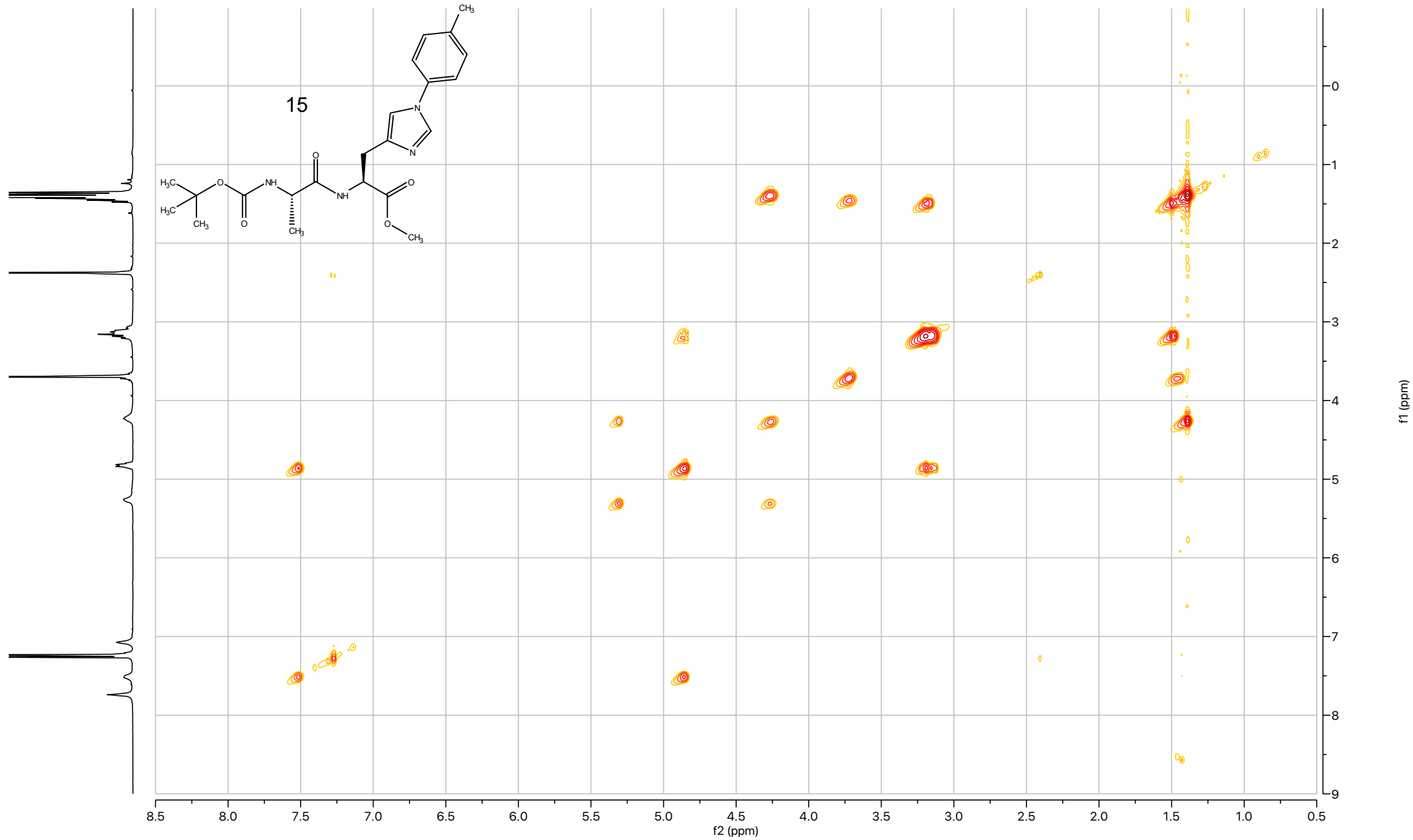


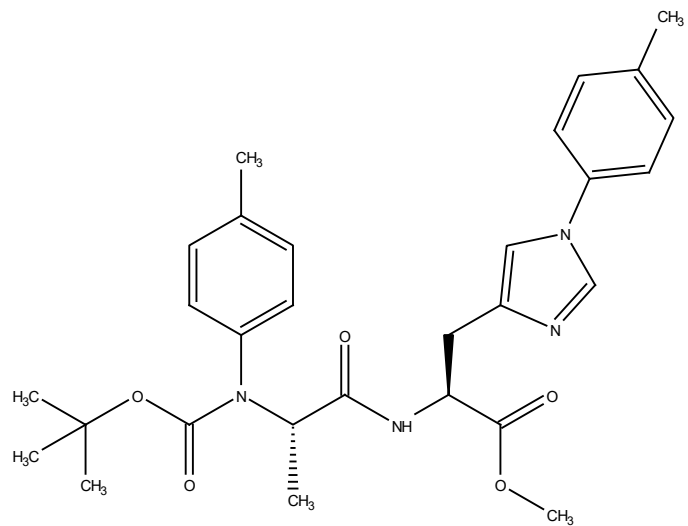
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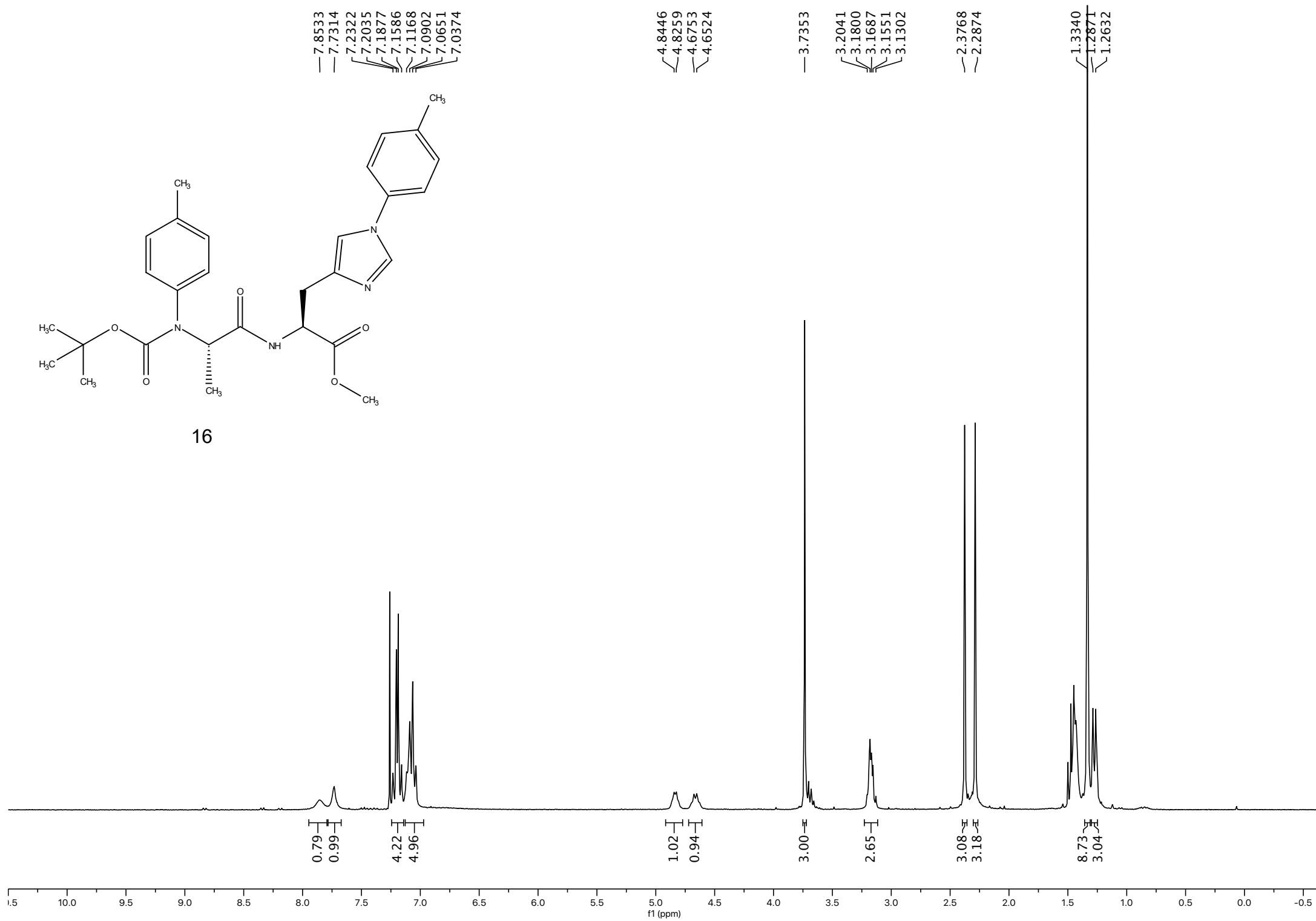


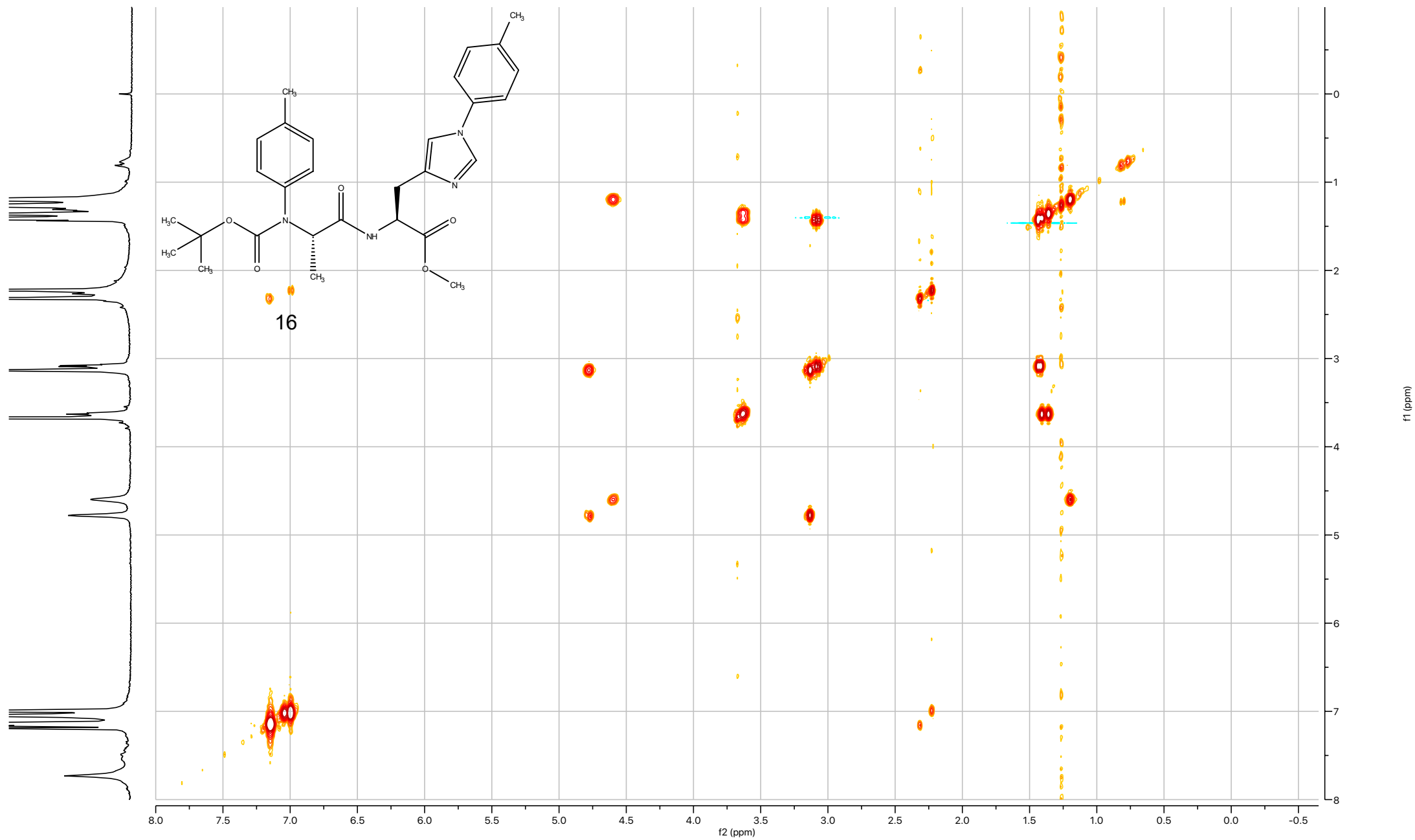
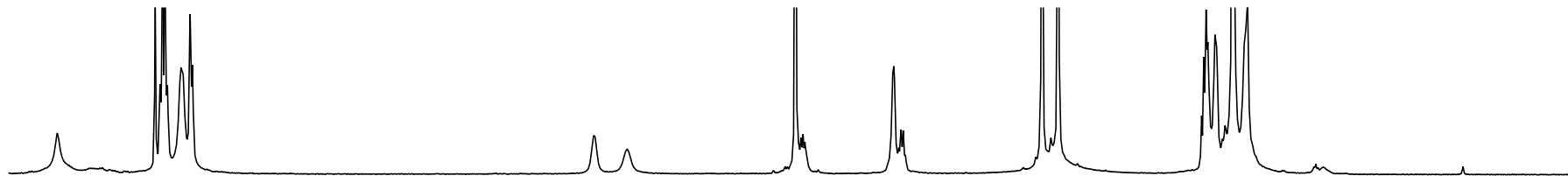




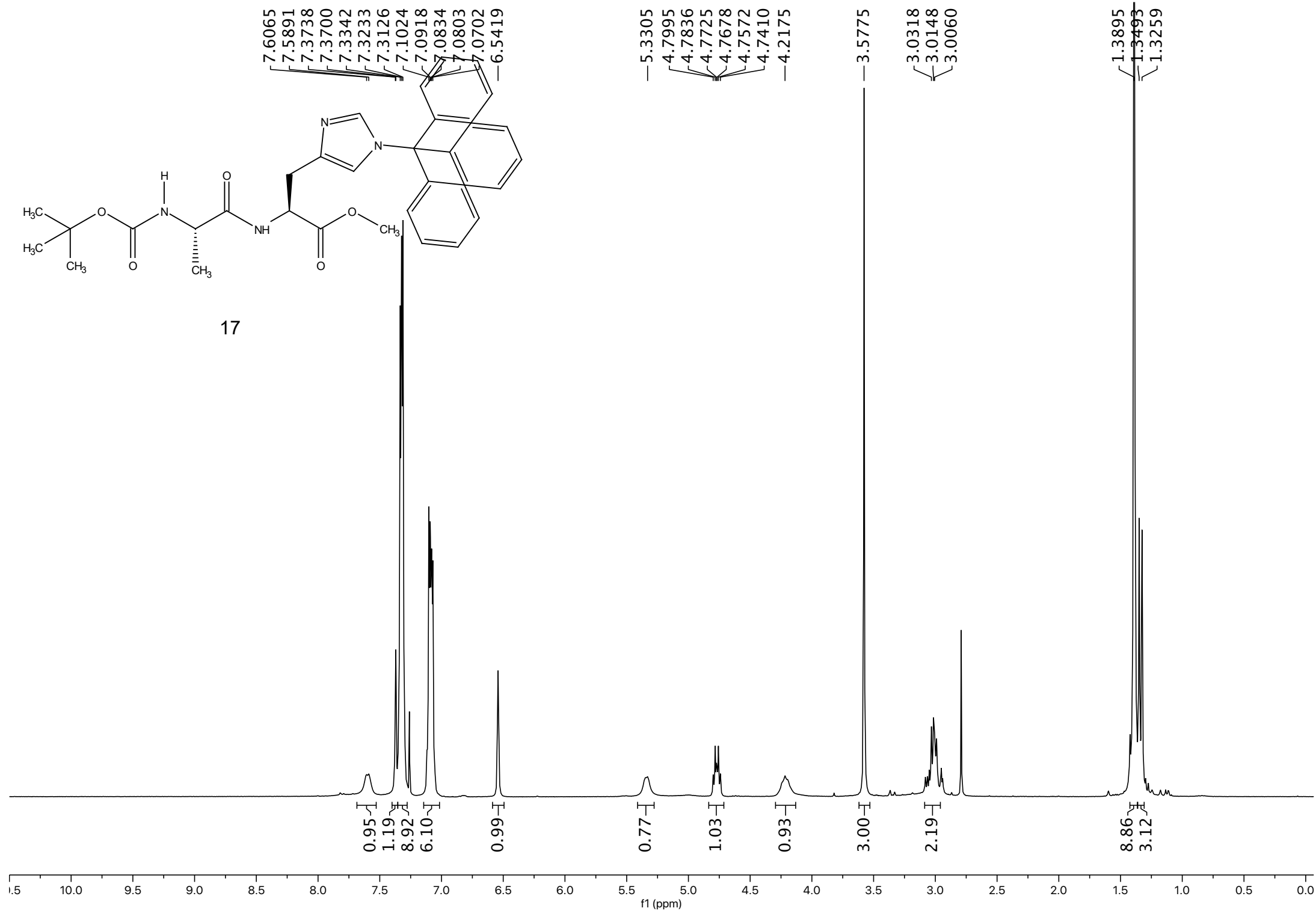
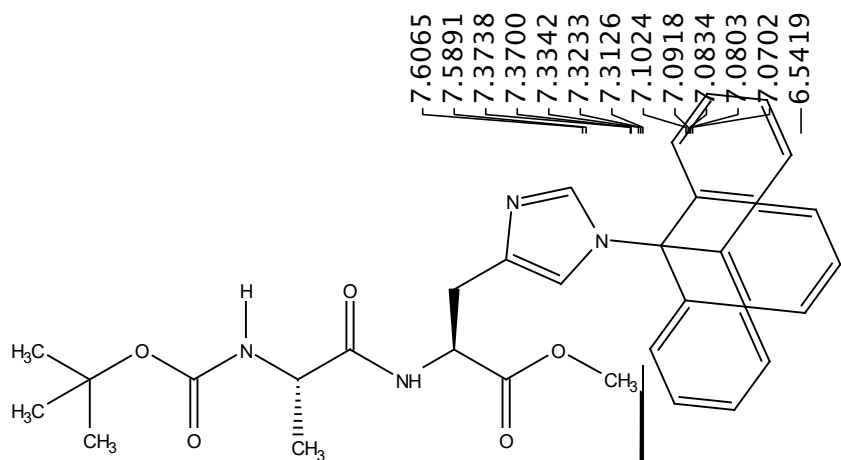


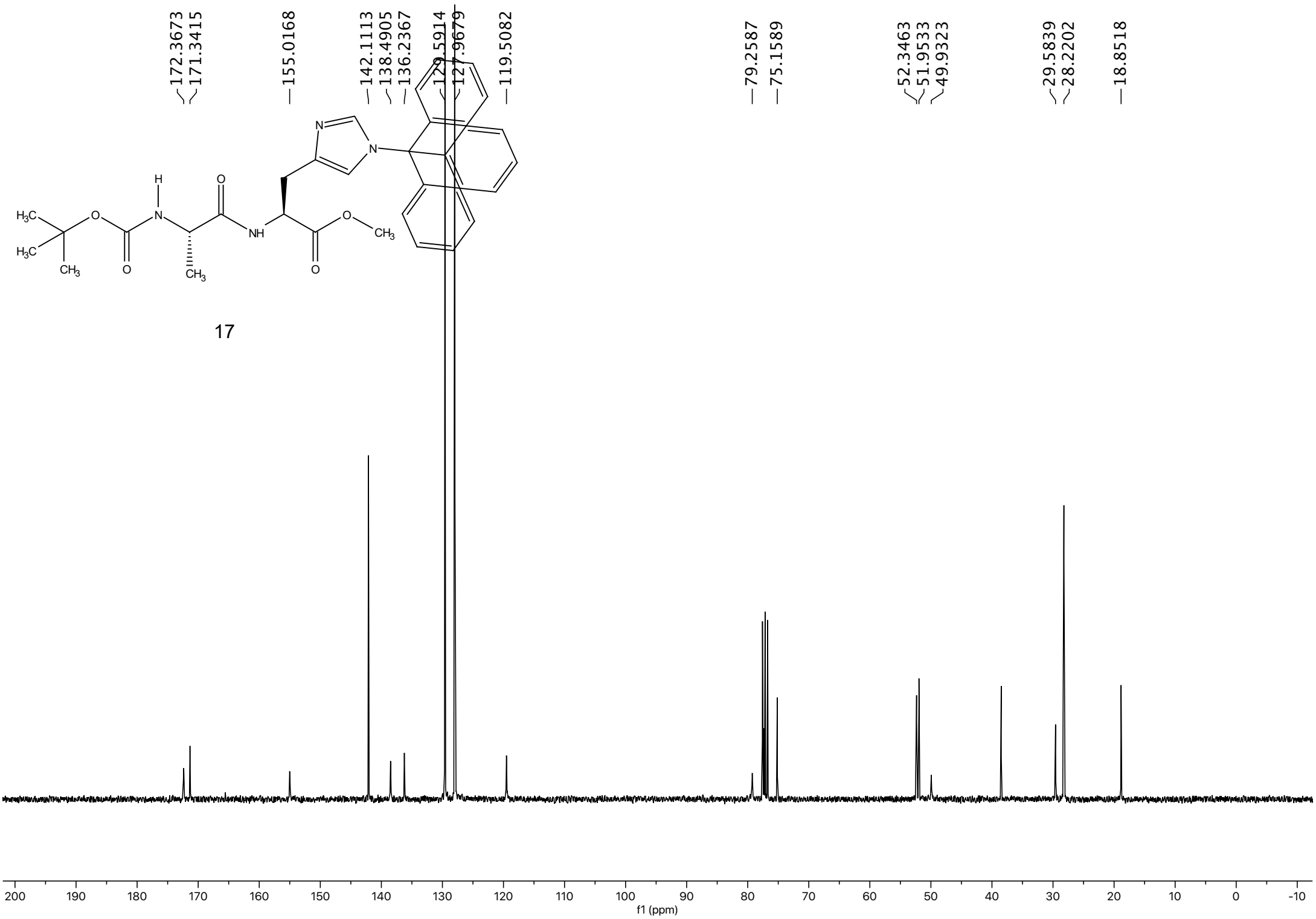
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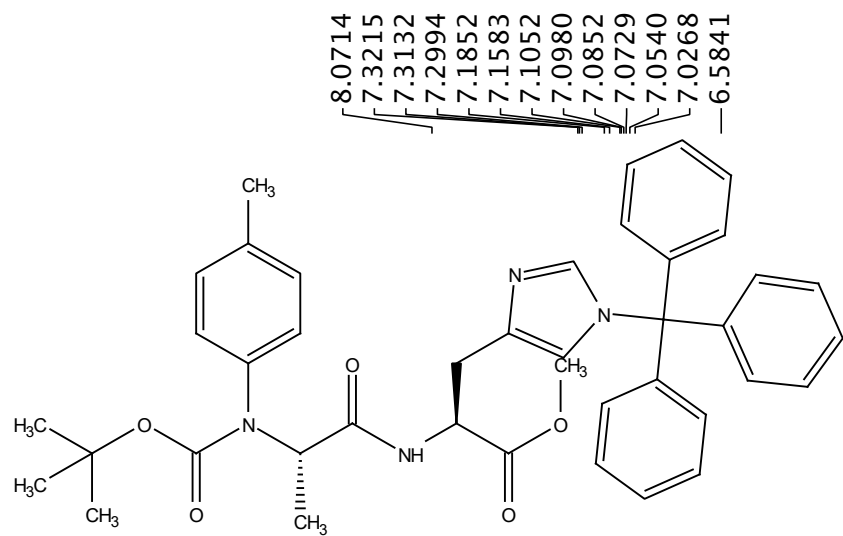




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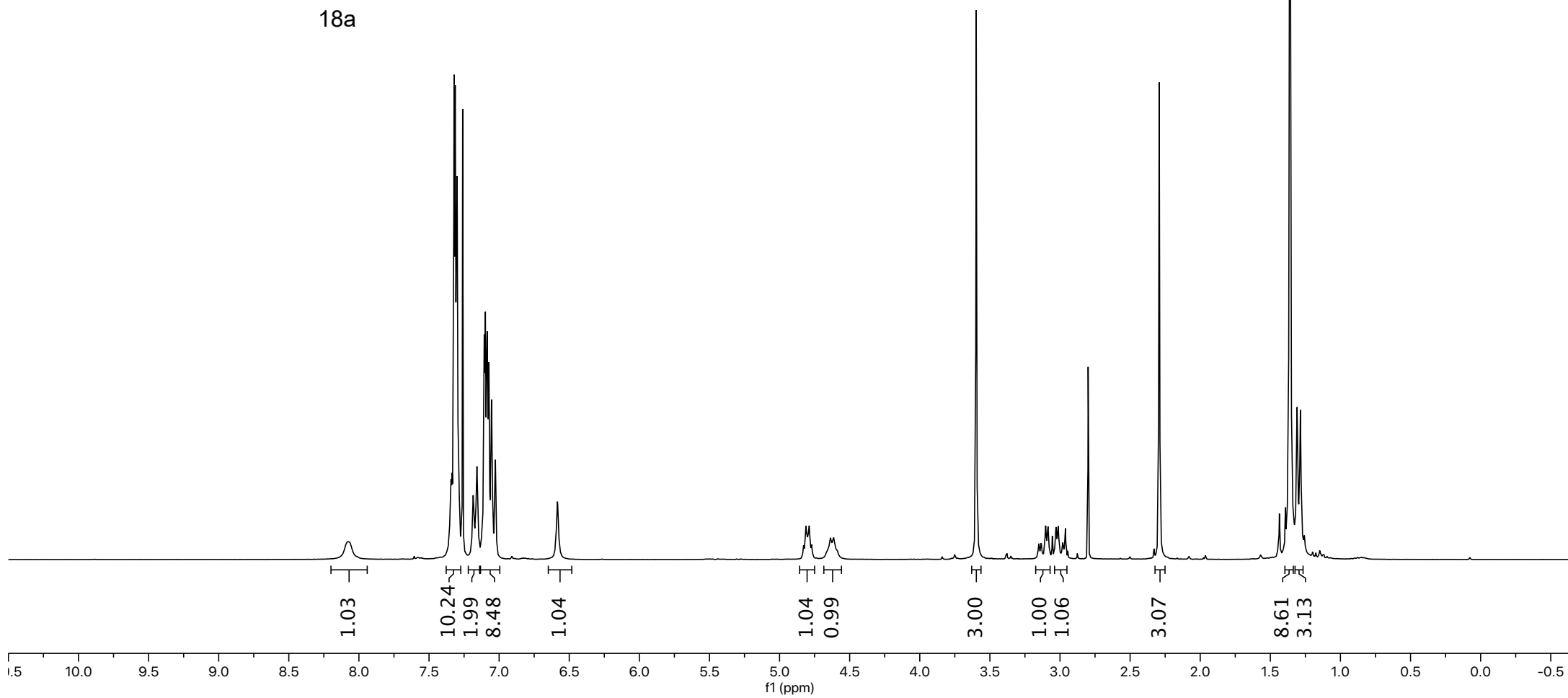
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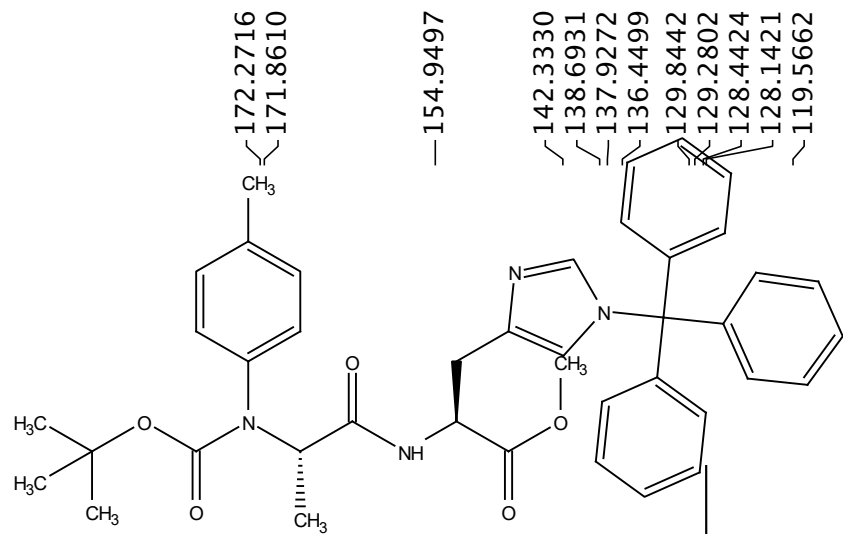
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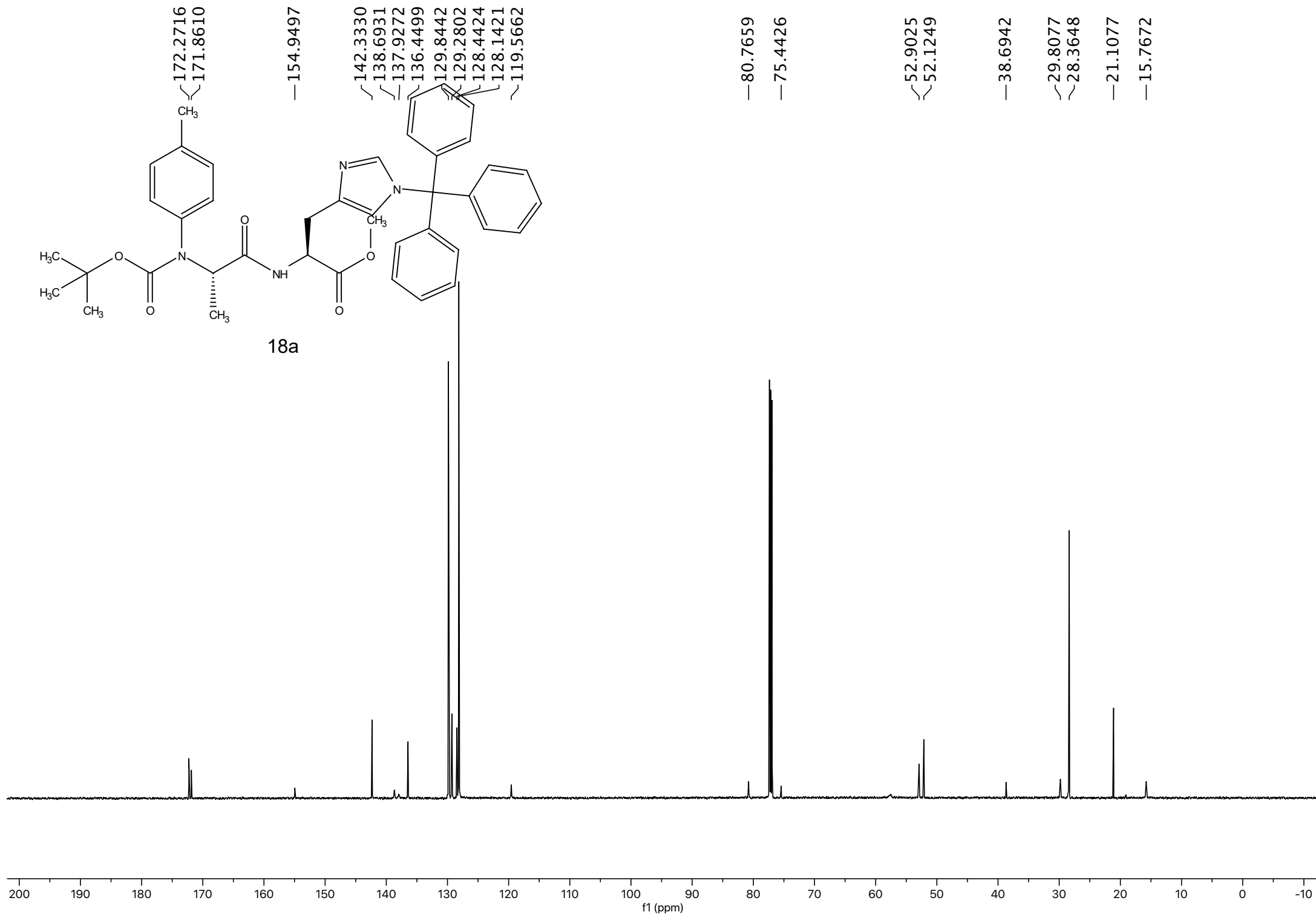
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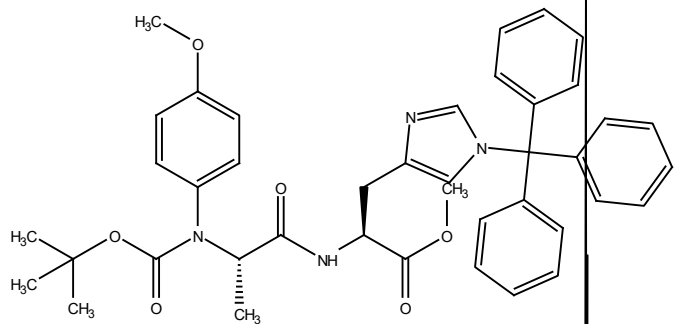
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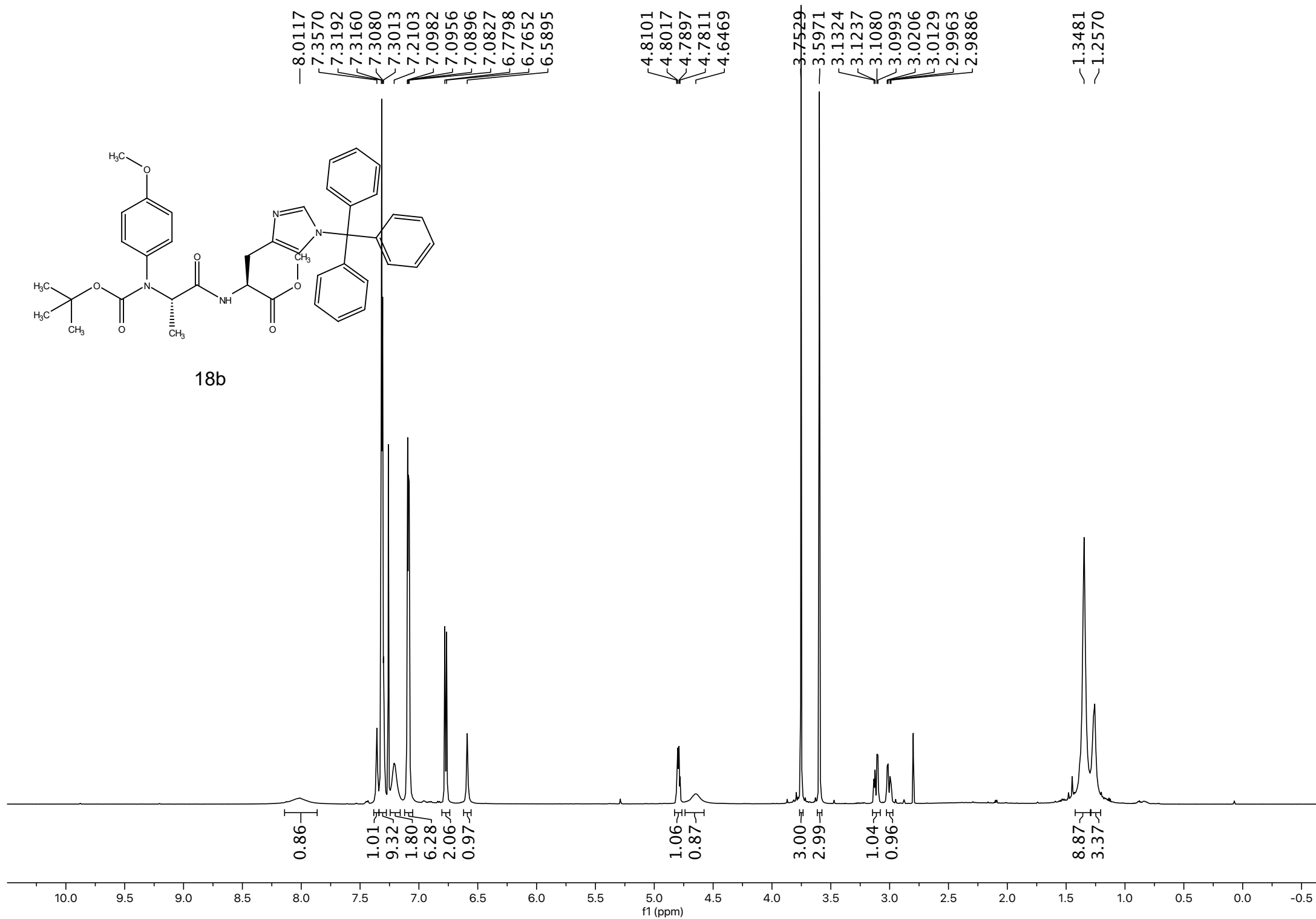


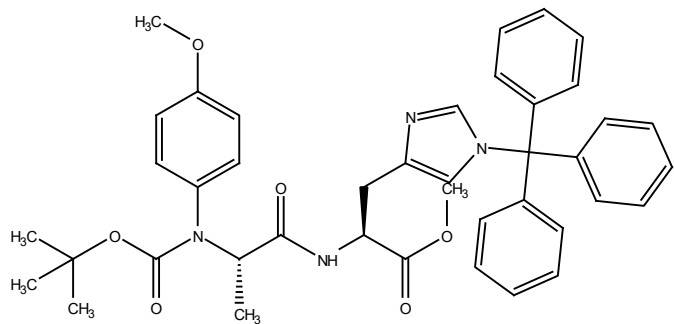
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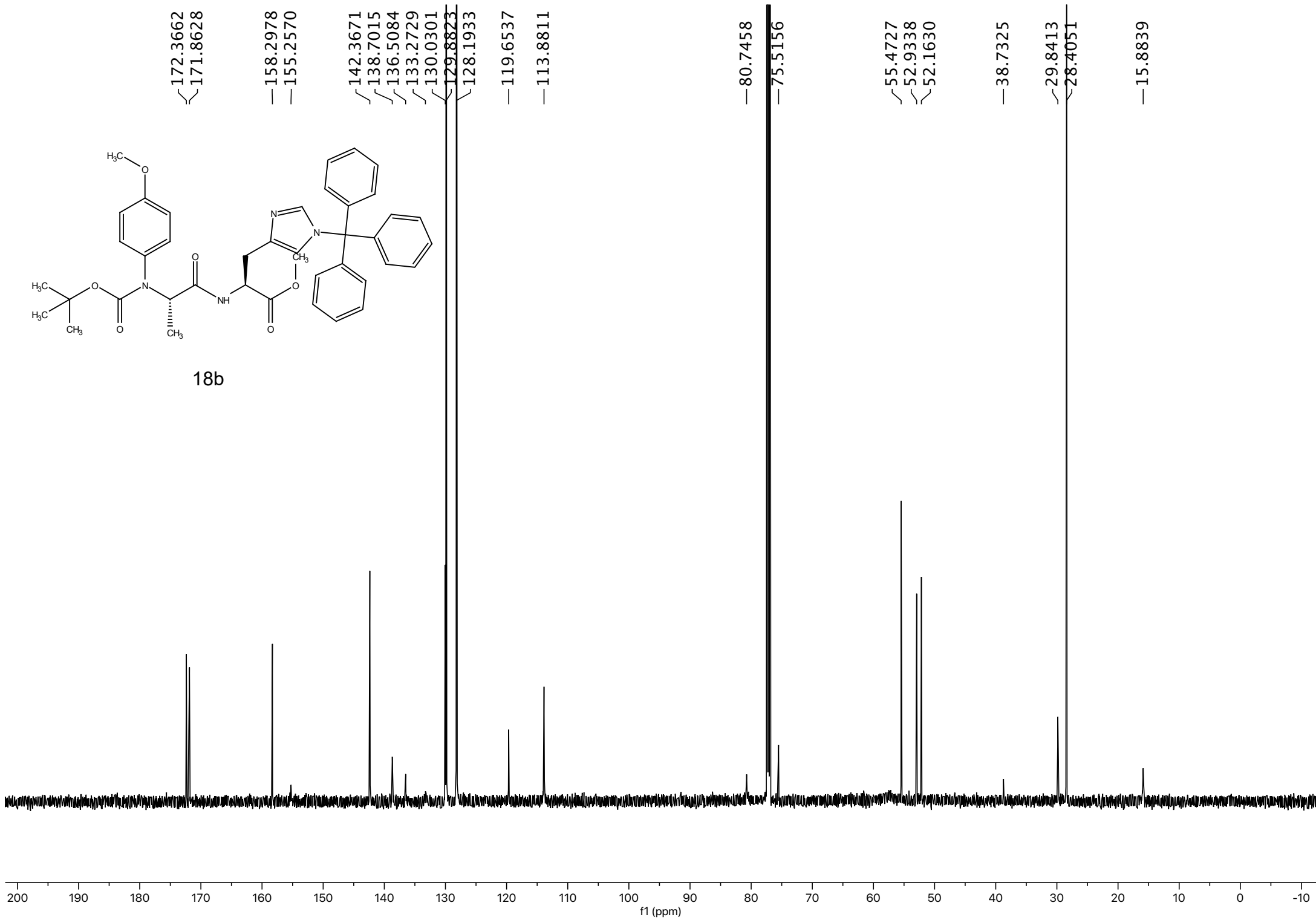


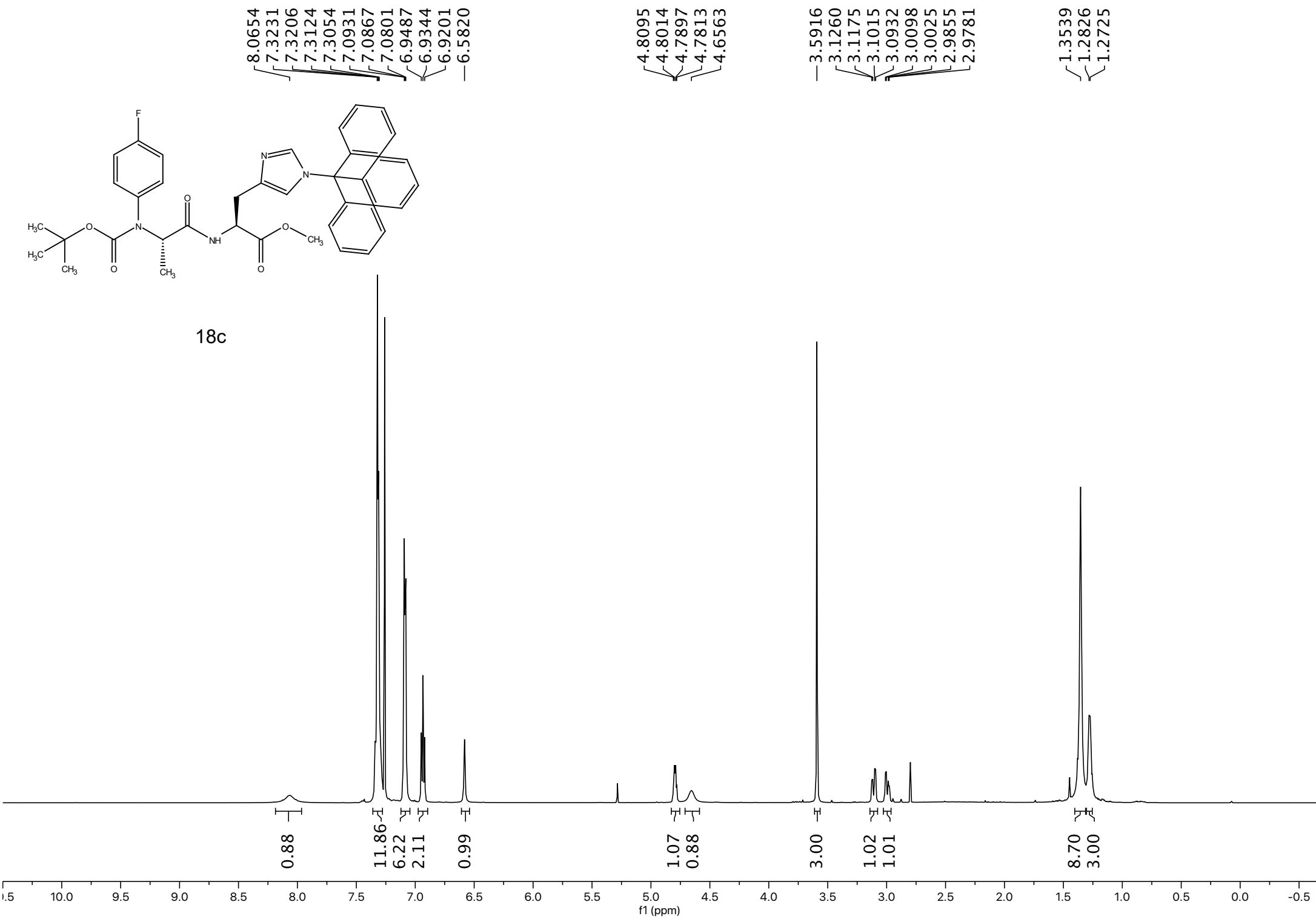
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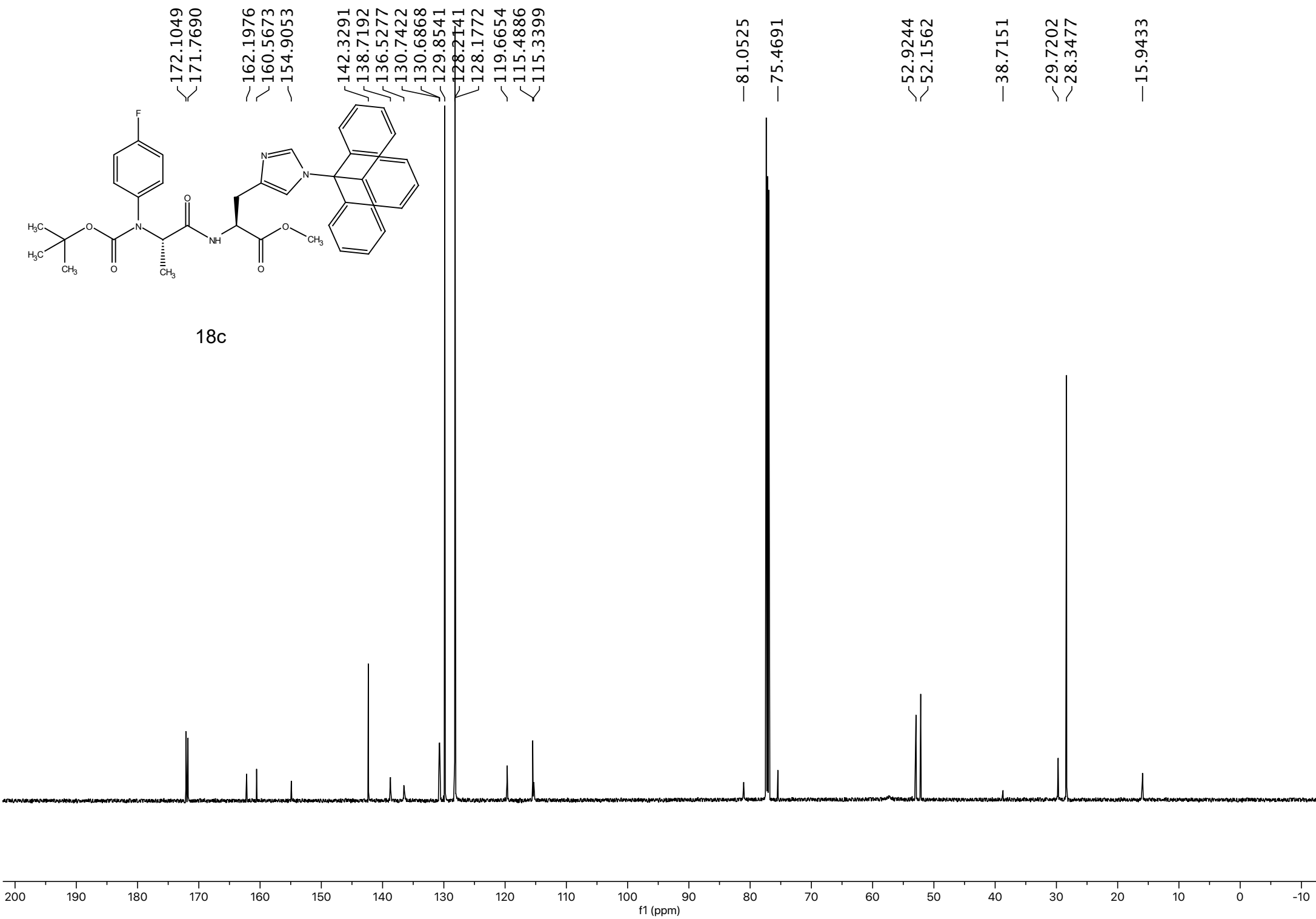


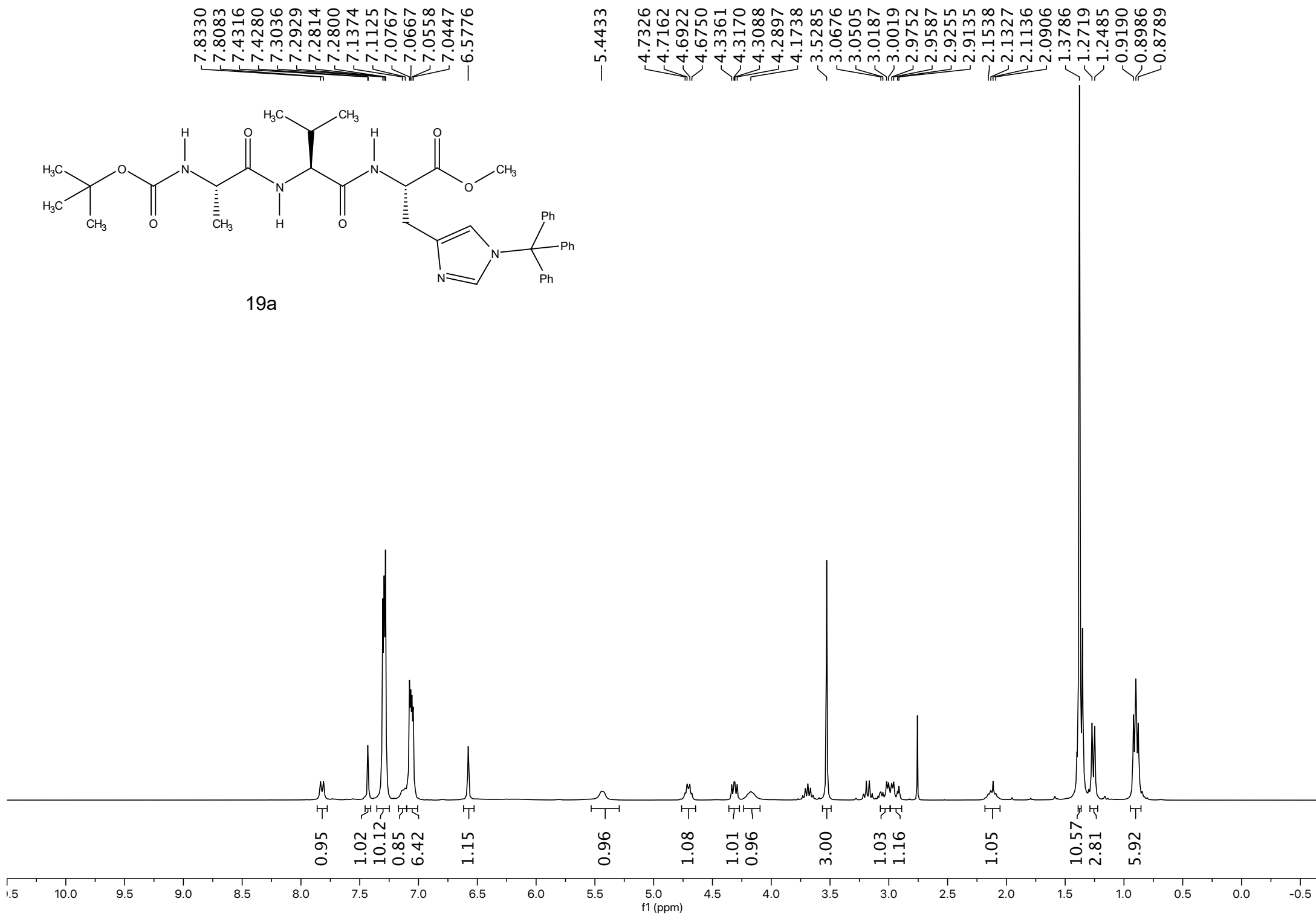


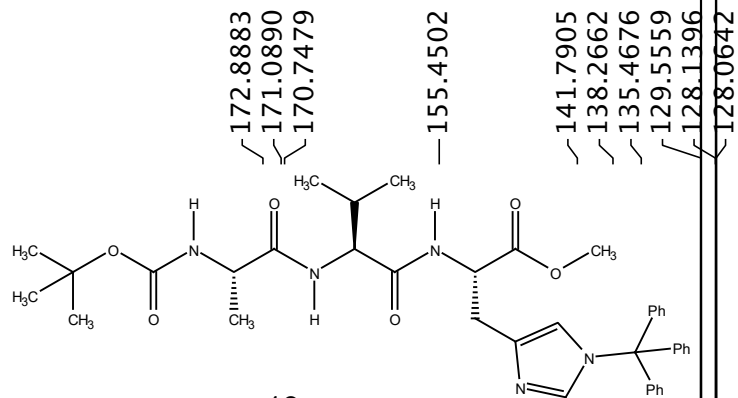
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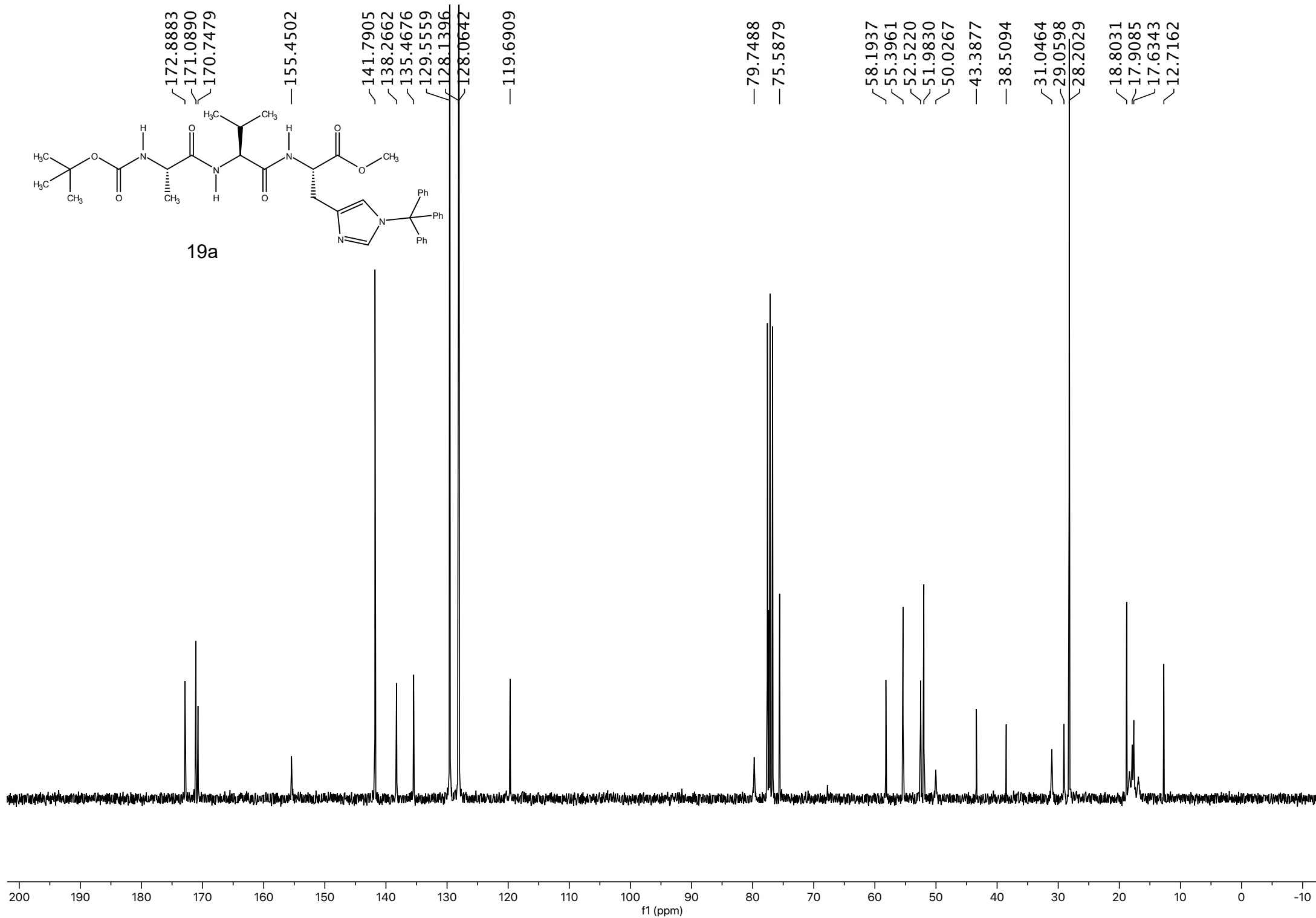


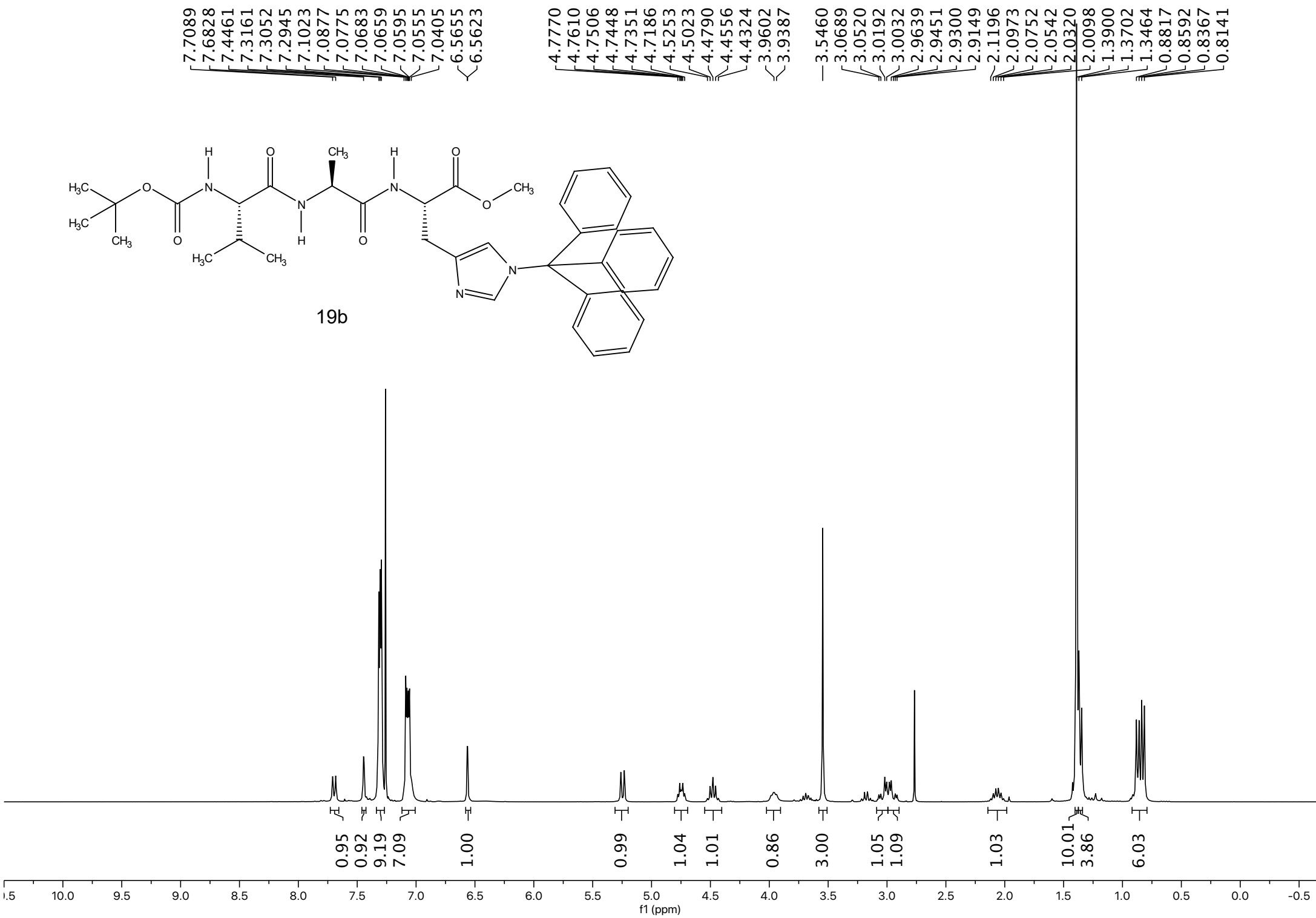


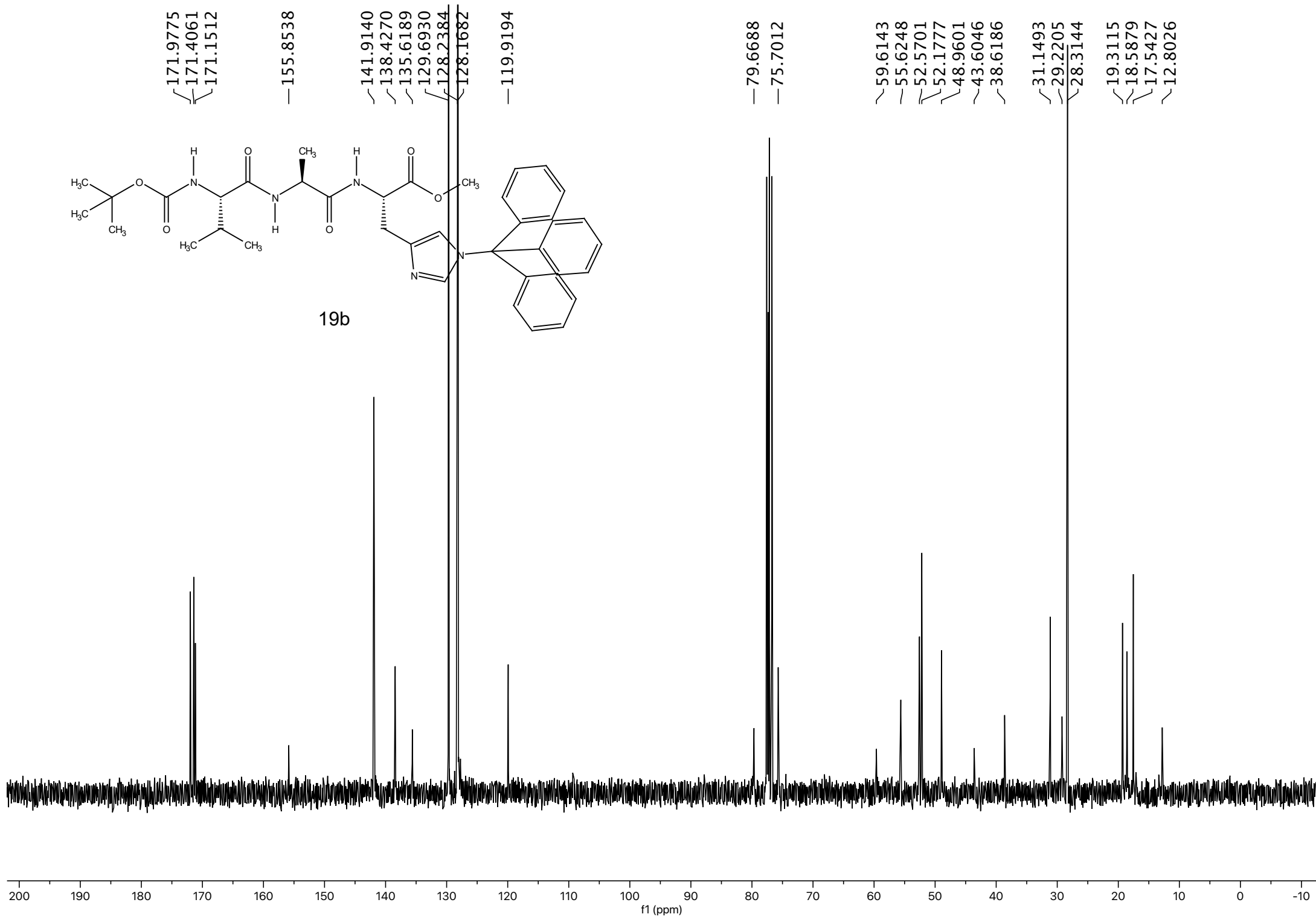


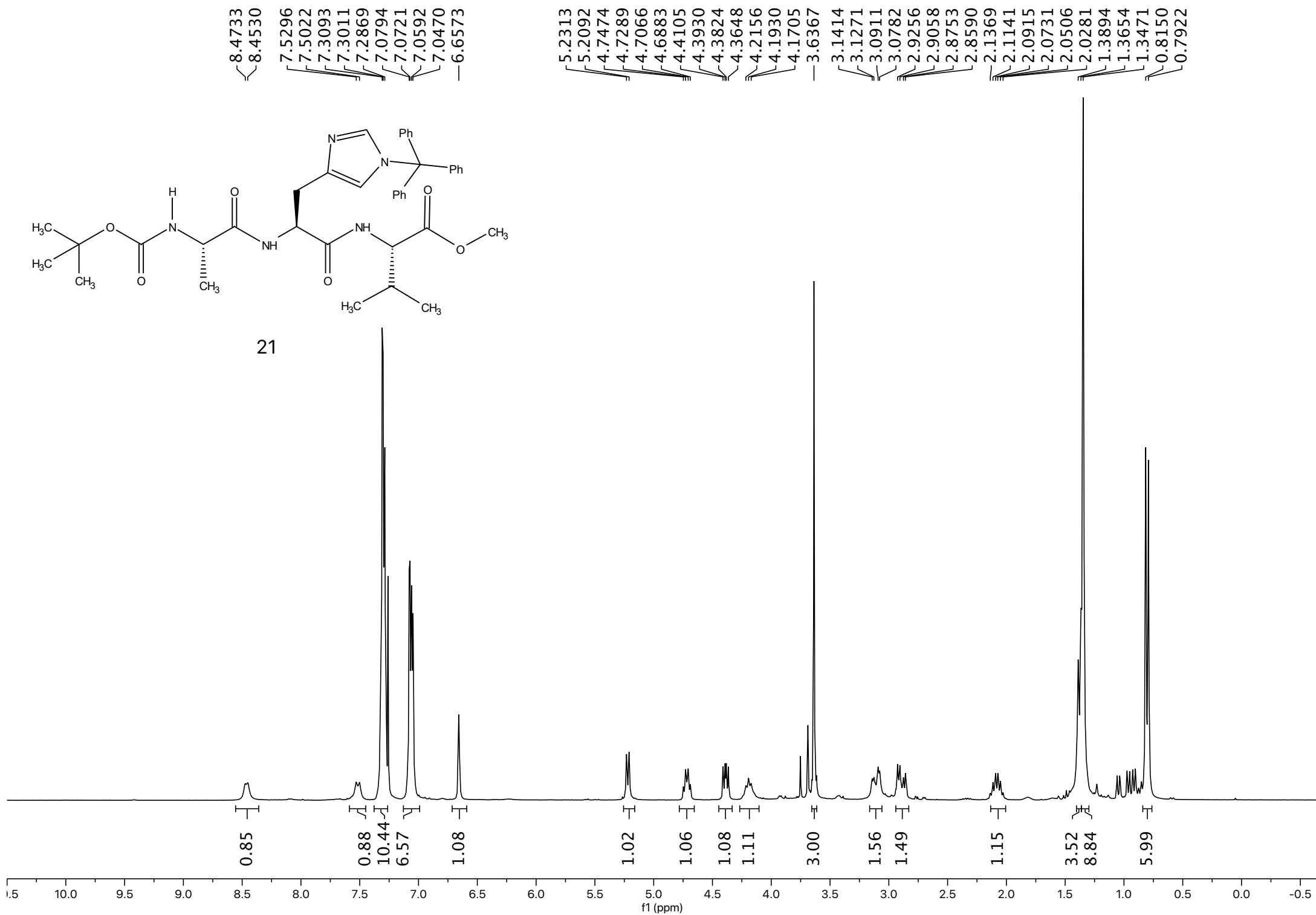


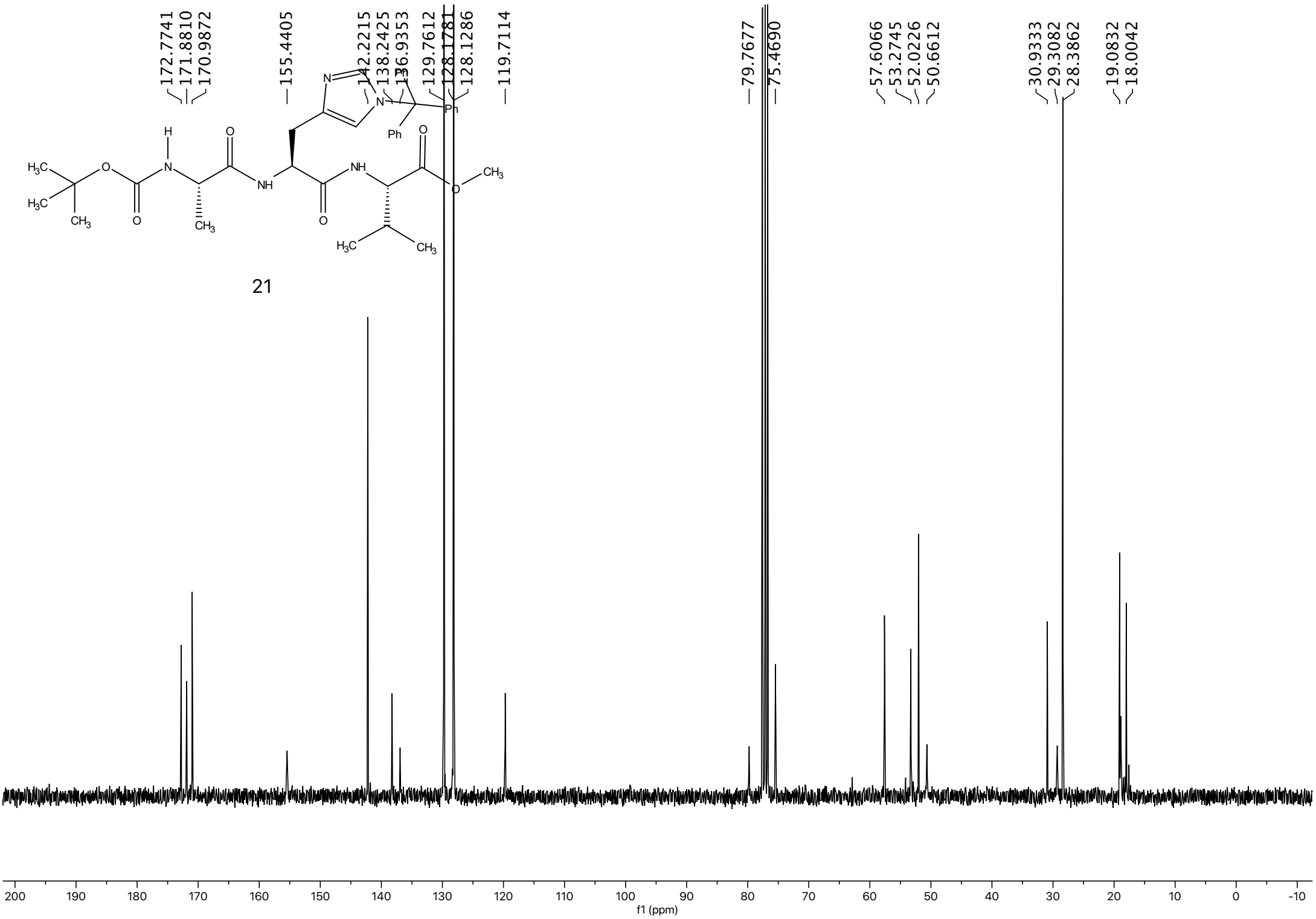
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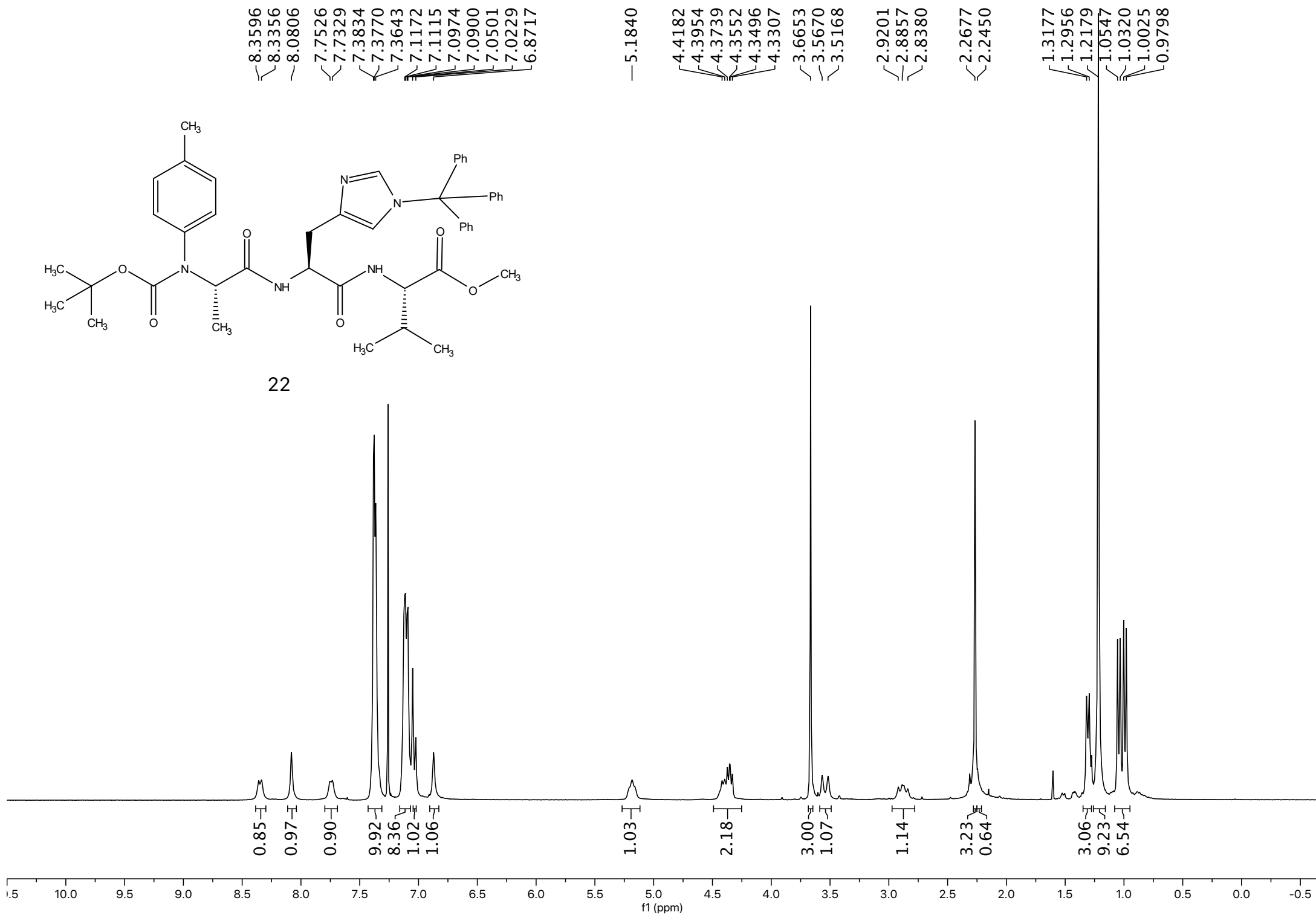


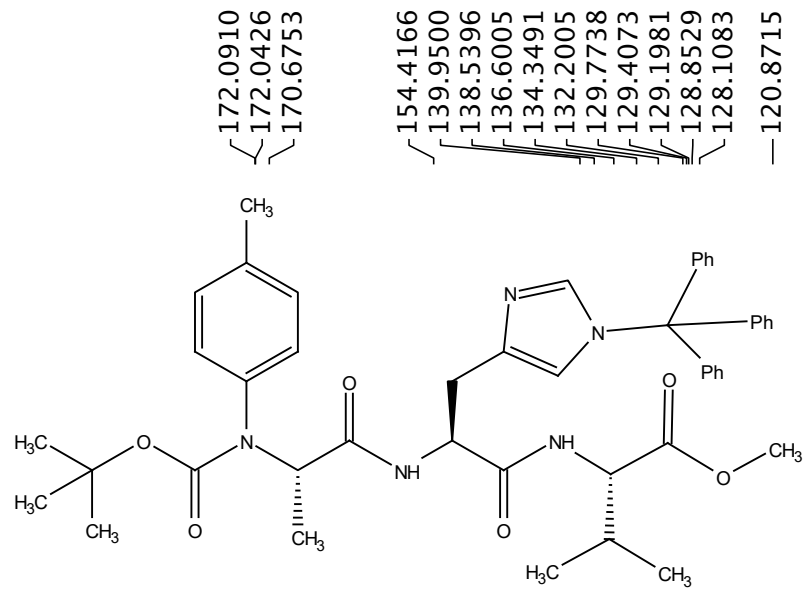




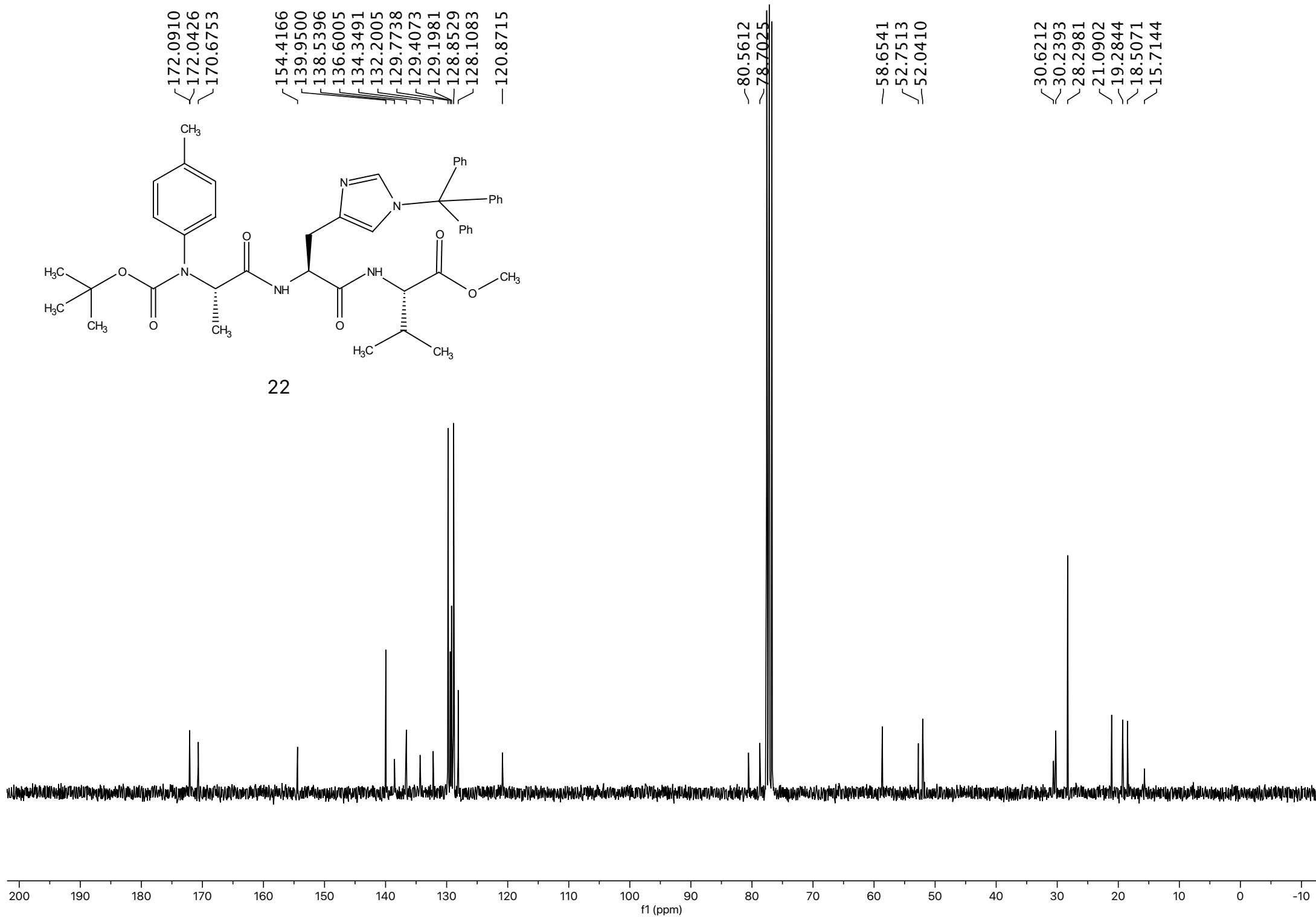


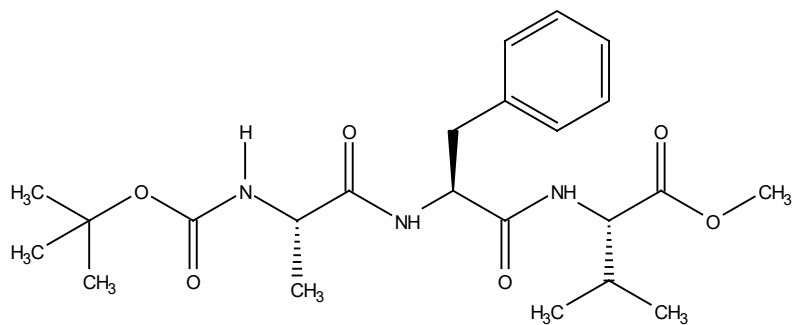




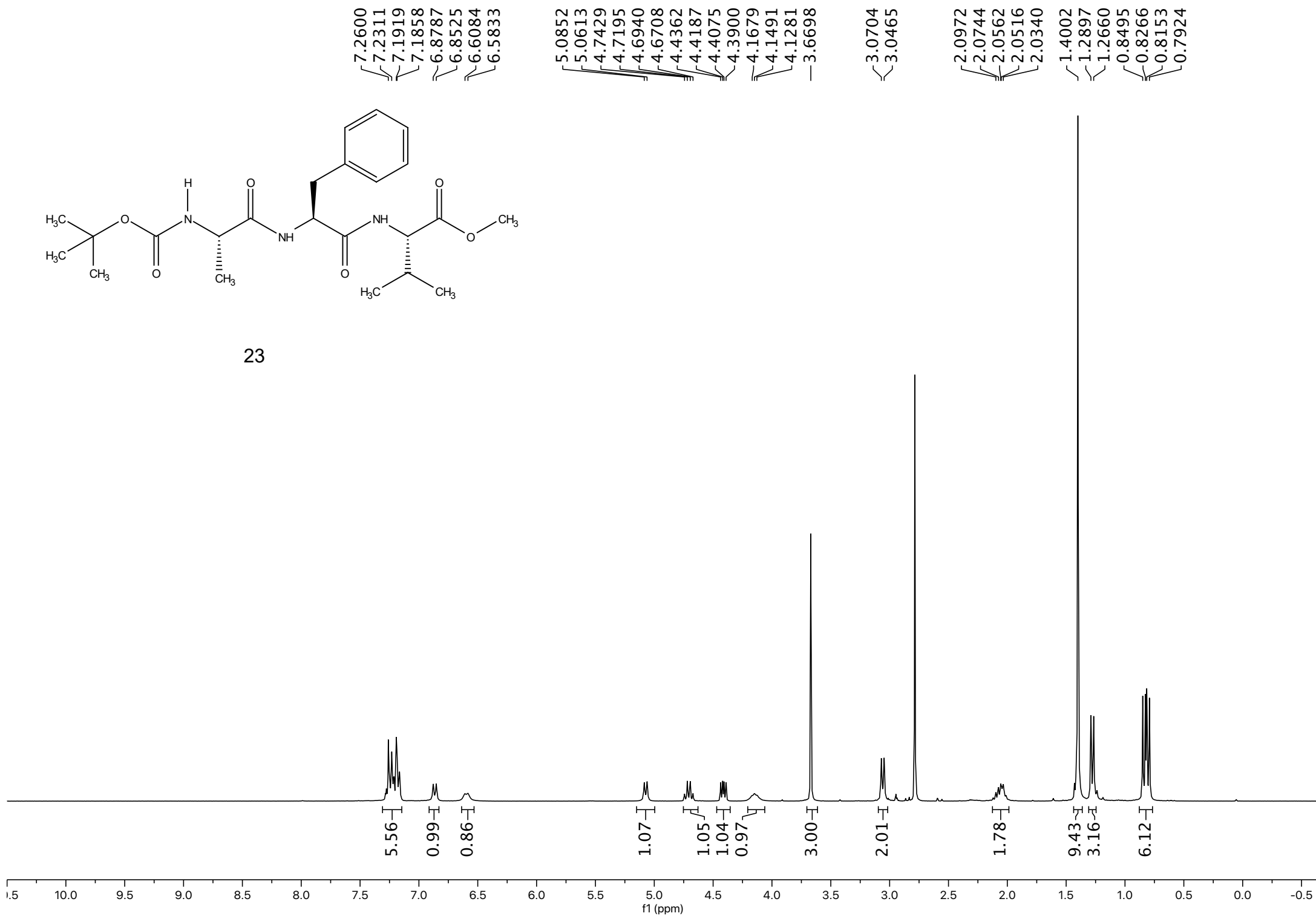


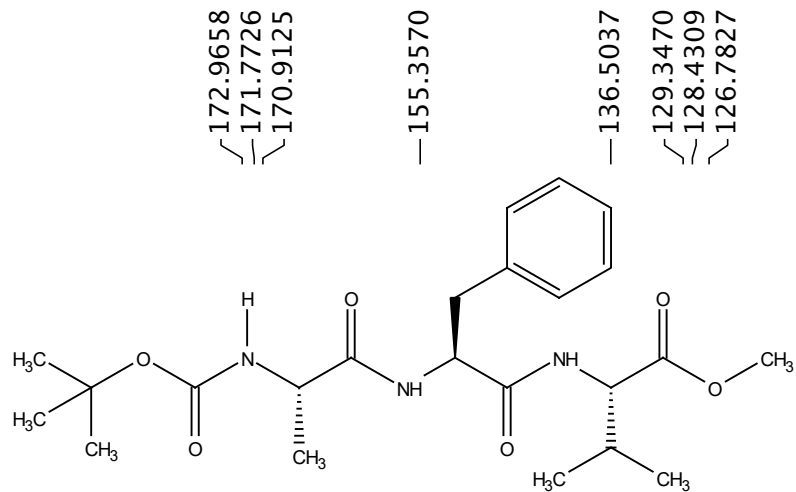
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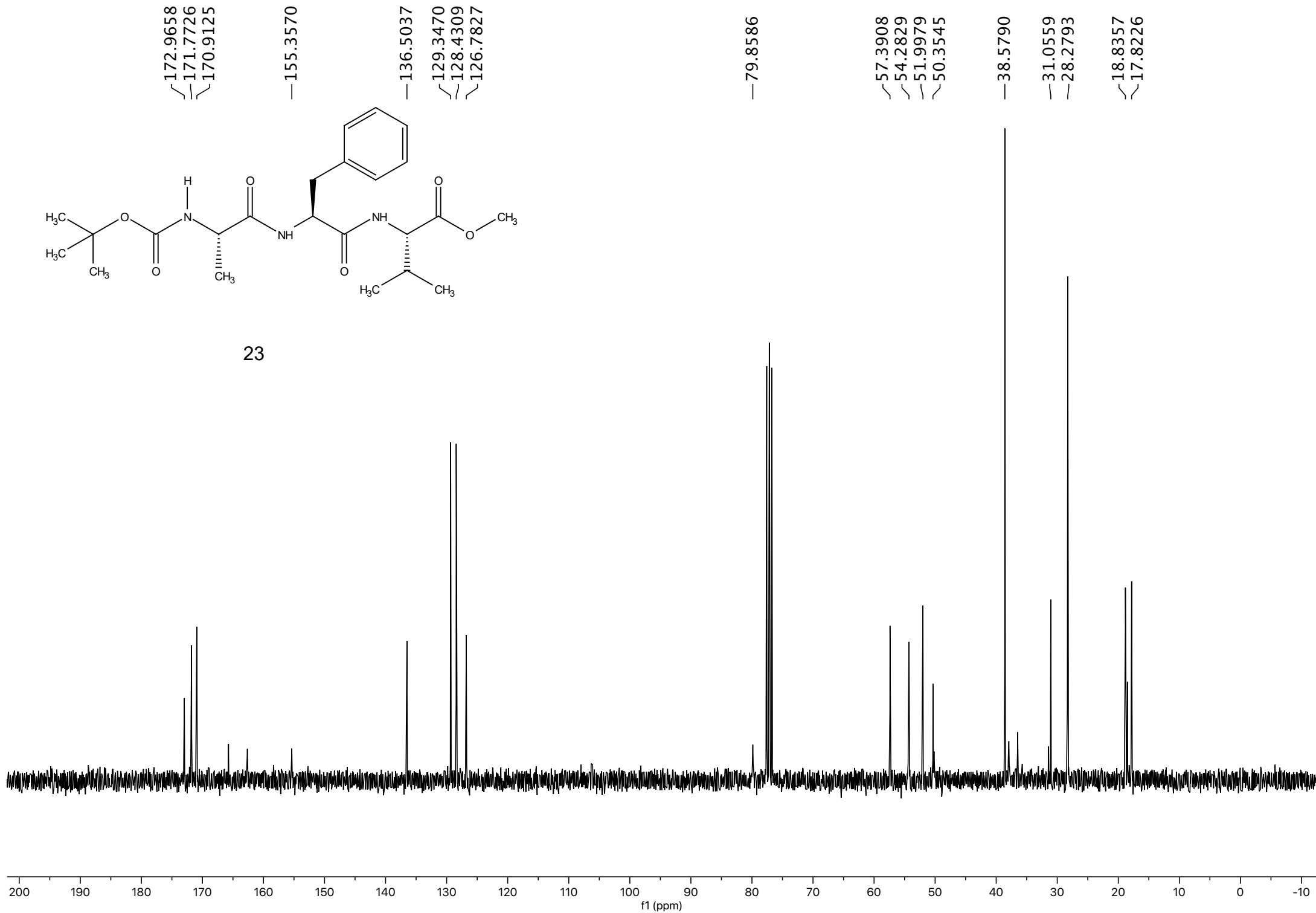


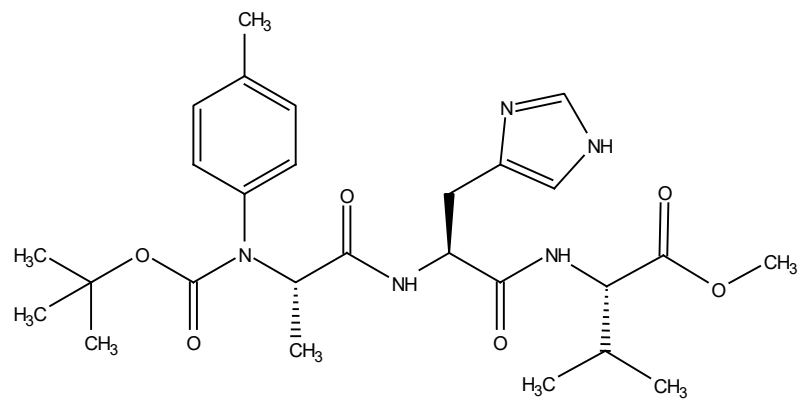
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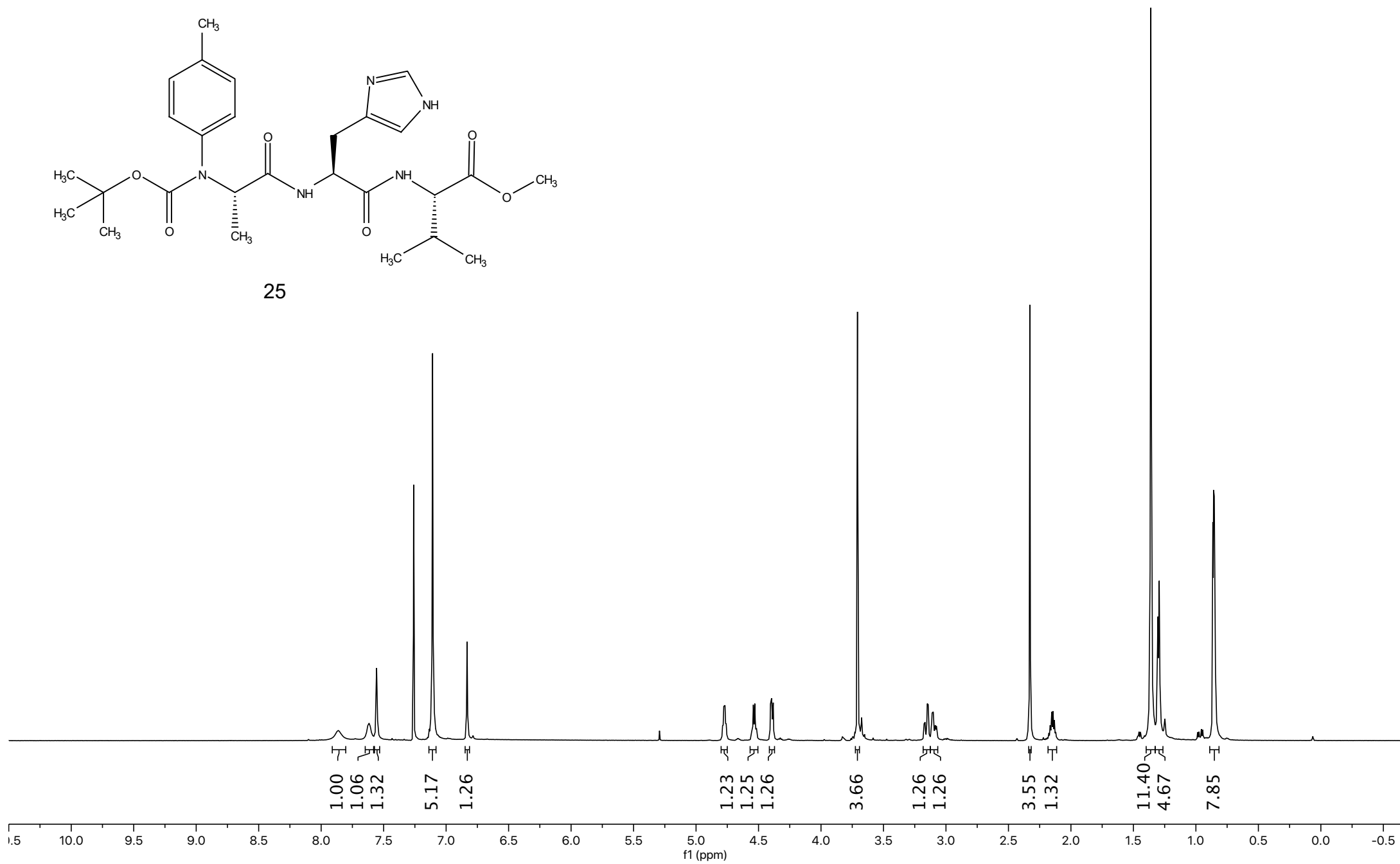


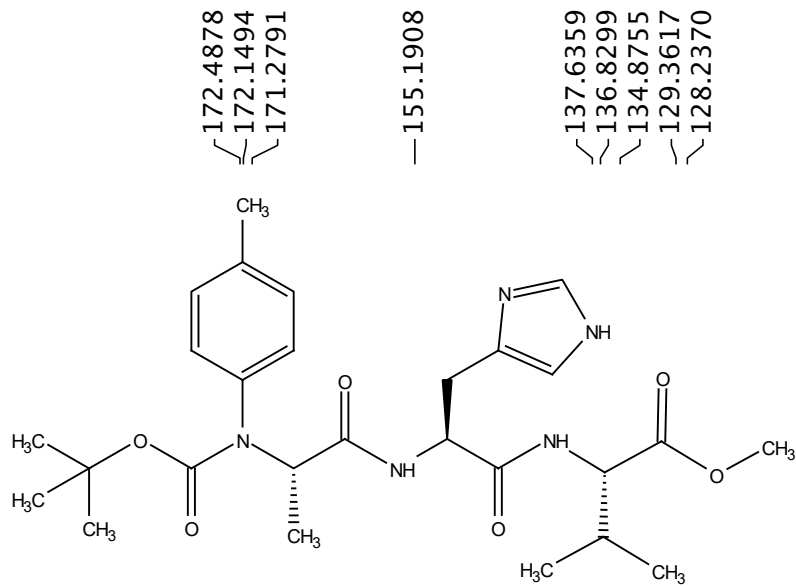
23





25





25

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171.2791

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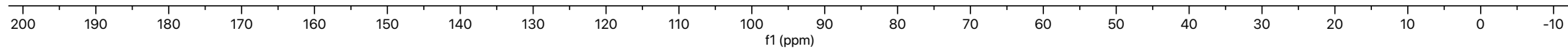
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17.8408

15.5390



Annexe B

Informations supplémentaires

*On the Copper-Promoted Backbone Arylation of the Histidine-Containing Peptides using
Triarylbi-muthines*

SUPPORTING INFORMATION

On the Copper-Promoted Arylation of the Backbone of Histidine-Containing

Peptides using Triarylbismuthines

Hwai-Chien Chan, Alexandre Gagnon*

Département de chimie
Université du Québec à Montréal
C.P. 8888, Succ. Centre-Ville, Montréal, Québec, Canada, H3C 3P8
gagnon.alexandre@uqam.ca

Table of contents

1. General information.....	B2
2. Triarylbismuthines (2) used in this work	B3
3. General procedures for peptides synthesis	B4
a. Synthesis of dipeptides Boc–A.A.–His(Trt)–OMe (1)	B4
b. Synthesis of tripeptides Boc–Ala–His(Trt)–A.A–OMe (4a–f)	B9
c. Synthesis of tripeptides Boc–A.A–His(Trt)–Val–OMe (4g–j)	B13
d. Synthesis of tripeptides Boc–A.A ₁ –A.A ₂ –His(Trt)–OMe (6a–e)	B16
e. Synthesis of tetrapeptide Boc–Ala–His(Trt)–Val–Gly–OMe (8)	B19
4. Procedure for the coupling between methyl ester N-protected histidine-containing peptides (1, 4, 6 and 8) and triarylbismuthines (2)	B20
a. Arylated compounds (3a–i) from dipeptides (1a–i) using tritolylbismuth (2a)	B21
b. Arylated compounds (5aa –an) from tripeptides (4a) using triarylbismuth (2).....	B26
c. Arylated compounds (5a –f) from tripeptides (4a–f) using tritolylbismuth (2a)	B35
d. Arylated compounds (5g–j) from tripeptides (4g–j) using tritolylbismuth (2a).....	B39
e. Arylated compounds (7) from tripeptides (6) using tritolylbismuth (2a)	B42
f. Arylated compounds (9) from tetrapeptide (8) using tritolylbismuth (2a)	B42
5. Procedure for selective deprotection of 5a	B43

1. General information

Unless otherwise indicated, all reactions were run under ambient atmosphere in non-flame dried glassware. For reactions performed under oxygen, 99.6% extra dry oxygen was used. Unless otherwise stated, commercial reagents were used without further purification. Anhydrous solvents were obtained using a MBRAUN (model MB-SPS 800) encapsulated solvent purification system. The evolution of reactions was monitored by analytical thin-layer chromatography using silica gel 60 F254 precoated plates. Flash chromatography was performed employing 220 mesh silica (Silicycle) using the indicated solvent system according to standard techniques. Nuclear magnetic resonance spectra ^1H were recorded on a BrukerAvance-III 300MHz spectrometer or a BrukerAvance-III 600MHz and ^{13}C were recorded on a BrukerAvance-III 75MHz spectrometer or a BrukerAvance-III 150MHz. Chemical shifts for ^1H -NMR spectra are recorded in parts per million from tetramethylsilane with the solvent resonance as the internal standard (chloroform, δ 7.26 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, p = quintuplet, m = multiplet, dd = doublet of doublet, dt = doublet of triplet, dq = doublet of quartet, tt = triplet of triplet, td = triplet of doublet, s(br) = broad singlet), coupling constant J in Hz and integration. Chemical shifts for ^{13}C -NMR spectra are recorded in parts per million from tetramethylsilane using the central peak of deuteriochloroform (77.16 ppm) as the internal standard. All ^{13}C -NMR spectra were obtained with complete proton decoupling. IR spectra were recorded on a Thermo Scientific Nicolet 6700 Smart ITR from thin films and are reported in reciprocal centimeters (cm^{-1}). Melting points were measured using Büchi M560 melting point apparatus in open capillaries.

2. Triarylbismuthines (2) used in this work

The organobismuthines used in this publication are illustrated in **Figure S1**. The procedures for synthesis of these organobismuthines can be found in the following references: ^a Hébert, M.; Petiot, P.; Benoit, E.; Dansereau, J.; Ahmad, T.; Le Roch, A.; Ottenwaelder, X.; A. Gagnon, *Eur. J. Org. Chem.*, **2016**, 81, 5401; ^b Petiot, P.; A. Gagnon, *Eur. J. Org. Chem.*, **2013**, 5282; ^c X. Marset, J. Torregrosa-Crespo, R. M. Martínez-Espinosa, G. Guillena, D. J. Ramón, *Green Chem.*, **2019**, 21, 4127; ^d R. Ding, C.-S. Ge, Y.-J. Chen, D. Wanga, C.-J. Li, *Tetrahedron Lett.*, **2002**, 43, 7789.

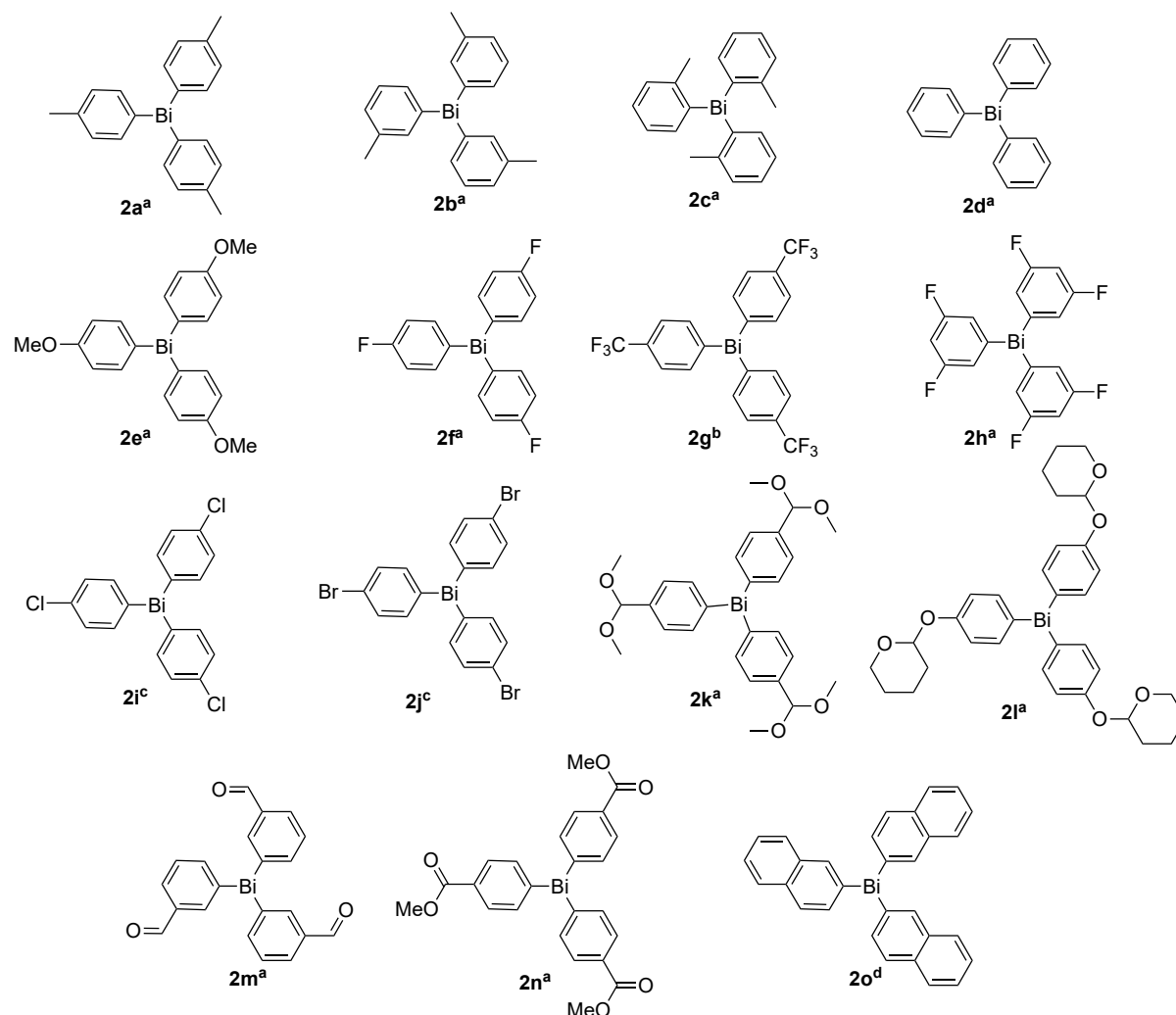
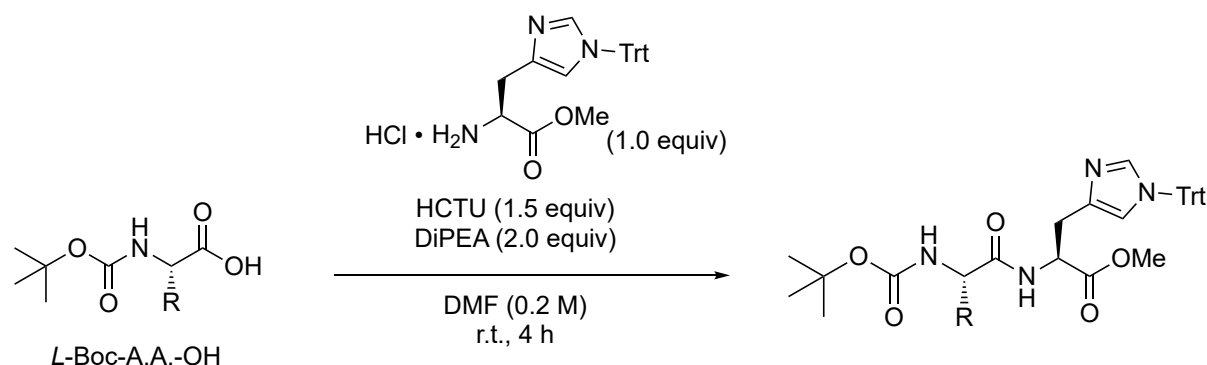


Figure S1. Functionalized organobismuthines used in this publication

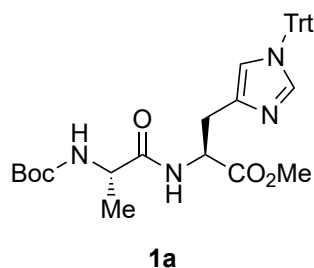
3. General procedures for peptides synthesis

a. Synthesis of dipeptides Boc-A.A.-His(Trt)-OMe (1)



Manuscript, Scheme 2.: In a 50 mL flask, DIPEA (155 μL , 893 mmol, 2.0 equiv) was added to $\text{L-H-His(Trt)-OMe} \cdot \text{HCl}$ (200 mg, 446 μmol , 1.0 equiv), L-Boc-A.A.-OH (446 μmol , 1.0 equiv) and HCTU (277 mg, 670 mmol, 1.5 equiv) in DMF (2 mL, 0.2 M). The mixture was stirred for 4h at room temperature before being extracted twice with dichloromethane. The combined organic phases were successively washed with sat. aq. NaHCO_3 (2x20 mL), brine (2x20 mL), dried over Na_2SO_4 , filtered and concentrated under reduce pressure. The resulting crude product was purified by column chromatography using the indicated eluent system to afford the desired pure product.

Methyl N^α -((*tert*-butoxycarbonyl)-*L*-alanyl)- N^ϵ -trityl-*L*-histidinate (1a)



The general procedure was followed starting from L-Boc-Ala-OH (85 mg, 449 μmol). The resulting crude product was purified on silica gel (Hexane-EtOAc, gradient from 80:20 to 30:70) to afford **1a** as a white solid (250 mg, 96%). **Mp** 74 – 76 $^\circ\text{C}$; **R_f** = 0.42 (DCM-MeOH, 95:5).

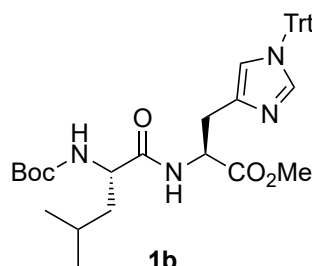
IR (ATR)/ $\bar{\nu}$: 3284, 2977, 1744, 1711, 1673, 1492, 1445, 1365, 1243, 1163, 1133, 1033, 1001, 870, 848, 748, 700, 660, 638 cm^{-1} .

$^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 7.60 (d, J = 5.2, 1H), 7.37 (d, J = 1.1 Hz, 1H), 7.35 – 7.27 (m, 9H), 7.15 – 6.98 (m, 6H), 6.54 (s, 1H), 5.33 (s, 1H), 4.77 (dt, J = 8.0, 4.7 Hz, 1H), 4.29 – 4.13 (m, 1H), 3.58 (s, 3H), 3.09 – 2.97 (m, 2H), 1.39 (s, 9H), 1.34 (d, J = 7.0 Hz, 3H).

$^{13}\text{C-NMR}$ (75 MHz, CDCl_3) δ 172.4, 171.3, 155.0, 142.1, 138.5, 136.2, 129.6, 128.0, 119.5, 79.3, 75.2, 52.3, 52.0, 49.9, 29.6, 28.2, 18.9.

HRMS–ESI: m/z $[M + H]^+$ calcd. for $C_{34}H_{38}N_4O_5$: 583.2915; found 583.2943.

Methyl N^α -((*tert*-butoxycarbonyl)-*L*-leucyl)- N^ϵ -trityl-*L*-histidinate (1b**)**



The general procedure was followed starting from *L*-Boc-Leu-OH (103 mg, 446 μ mol). The resulting crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 98:2) to afford **1b** as a white solid (192 mg, 69%). **Mp** 67 – 69 °C; **R_f** = 0.33 (DCM–MeOH, 95:5).

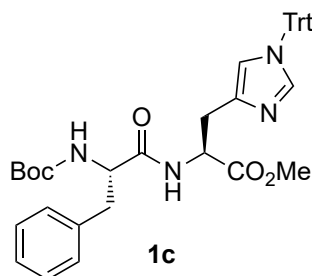
IR (ATR)/ $\bar{\nu}$: 3292, 2954, 1745, 1711, 1669, 1493, 1445, 1365, 1245, 1164, 1132, 1036, 1001, 871, 823, 748, 700, 660, 638 cm^{-1} .

1H -NMR (300 MHz, $CDCl_3$) δ 7.69 (d, J = 6.8 Hz, 1H), 7.37 – 7.27 (m, 10H), 7.13 – 7.03 (m, 6H), 6.54 (s, 1H), 5.09 (d, J = 7.9 Hz, 1H), 4.74 (dt, J = 7.7, 4.8 Hz, 1H), 4.26 – 4.09 (m, 1H), 3.56 (s, 3H), 3.04 (dd, J = 14.6, 4.9 Hz, 1H), 2.95 (dd, J = 14.6, 4.7 Hz, 1H), 1.79 – 1.58 (m, 2H), 1.43 – 1.39 (m, 1H), 1.37 (s, 9H), 0.89 (dd, J = 6.2, 3.5 Hz, 6H).

^{13}C -NMR (75 MHz, $CDCl_3$) δ 172.3, 171.4, 165.7, 155.3, 142.2, 138.6, 136.3, 129.7, 128.1, 119.6, 79.4, 75.2, 53.0, 52.5, 52.0, 42.1, 29.6, 28.3, 24.6, 22.9.

HRMS–ESI: m/z $[M + H]^+$ calcd. for $C_{37}H_{44}N_4O_5$: 625.3385; found 625.3396.

Methyl N^α -((*tert*-butoxycarbonyl)-*L*-phenylalanyl)- N^ϵ -trityl-*L*-histidinate (1c**)**



The general procedure was followed starting from *L*-Boc-Phe-OH (118 mg, 446 μ mol). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 80:20 to 30:70) to afford **1c** as a white solid (147 mg, 50%). **Mp** 76 – 78 °C; **R_f** = 0.68 (Hexane–EtOAc, 20:80).

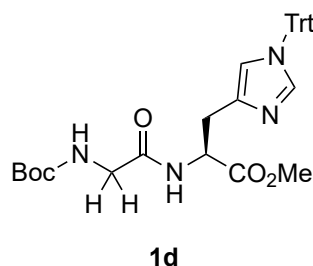
IR (ATR)/ $\bar{\nu}$: 3289, 2976, 1743, 1711, 1673, 1492, 1444, 1390, 1365, 1244, 1163, 1132, 1022, 1001, 870, 748, 699, 659, 638 cm^{-1} .

1H -NMR (300 MHz, $CDCl_3$) δ 7.62 – 7.50 (m, 1H), 7.37 – 7.28 (m, 10H), 7.21 (d, J = 4.4 Hz, 4H), 7.14 – 7.02 (m, 7H), 6.49 (s, 1H), 5.27 (d, J = 7.3 Hz, 1H), 4.77 (dt, J = 7.8, 4.7 Hz, 1H), 4.55 – 4.36 (m, 1H), 3.58 (s, 3H), 3.14 (dd, J = 13.9, 5.9 Hz, 1H), 3.08 – 2.87 (m, 3H), 1.34 (s, 9H).

^{13}C -NMR (75 MHz, $CDCl_3$) δ 171.3, 171.1, 155.2, 142.2, 138.6, 136.9, 136.2, 129.7, 129.5, 128.4, 128.1, 128.1, 126.6, 119.6, 79.6, 75.3, 55.5, 52.6, 52.1, 38.6, 29.9, 28.3.

HRMS–ESI: m/z $[M + H]^+$ calcd. for $C_{40}H_{42}N_4O_5$: 659.3228; found 659.3234.

Methyl N^α -((*tert*-butoxycarbonyl)glycyl)- N^ϵ -trityl-L-histidinate (1d**)**



The general procedure was followed starting from Boc–Gly–OH (78 mg, 446 μ mol). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 80:20 to 30:70) to afford **1d** as a white solid (165 mg, 61%). **Mp** 80 – 82 $^\circ$ C; **R_f** = 0.43 (DCM–MeOH, 95:5).

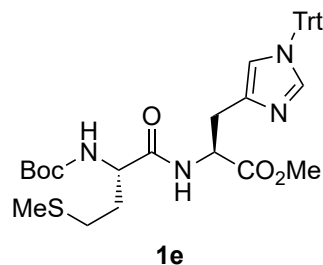
IR (ATR)/ $\bar{\nu}$: 3288, 2978, 2923, 1668, 1494, 1445, 1367, 1279, 1244, 1158, 1035, 1001, 942, 838, 748, 701, 660 cm^{-1} .

$^1\text{H-NMR}$ (600 MHz, CDCl_3) δ 7.75 – 7.70 (m, 1H), 7.36 (s, 1H), 7.35 – 7.29 (m, 9H), 7.12 – 7.06 (m, 6H), 6.53 (s, 1H), 5.30 (s, 1H), 4.82 – 4.76 (m, 1H), 3.84 – 3.80 (m, 2H), 3.58 (s, 3H), 3.04 (dd, J = 14.7, 5.0 Hz, 1H), 3.00 – 2.93 (m, 1H), 1.39 (s, 9H).

$^{13}\text{C-NMR}$ (150 MHz, CDCl_3) δ 171.7, 169.4, 156.0, 142.3, 138.8, 136.4, 129.8, 128.2, 128.2, 119.8, 79.9, 75.5, 52.6, 52.3, 44.2, 29.7, 28.4.

HRMS–ESI: m/z [$\text{M} + \text{H}$] $^+$ calcd. for $\text{C}_{33}\text{H}_{36}\text{N}_4\text{O}_5$: 569.2759; found 569.2764.

Methyl N^α -((*tert*-butoxycarbonyl)-L-methionyl)- N^ϵ -trityl-L-histidinate (1e**)**



The general procedure was followed starting from L–Boc–Met–OH (111 mg, 446 μ mol). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 100:0 to 50:50) to afford **1e** as a white solid (169 mg, 59%). **Mp** 55 – 57 $^\circ$ C; **R_f** = 0.49 (DCM–MeOH, 95:5).

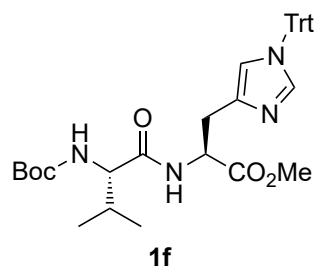
IR (ATR)/ $\bar{\nu}$: 3288, 2920, 1745, 1711, 1673, 1492, 1444, 1365, 1239, 1164, 1132, 1024, 1001, 908, 869, 748, 700, 660, 637 cm^{-1} .

$^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 7.79 (d, J = 7.7 Hz, 1H), 7.34 (s, 1H), 7.32 – 7.22 (m, 9H), 7.12 – 6.99 (m, 6H), 6.51 (s, 1H), 5.48 (d, J = 8.1 Hz, 1H), 4.72 (dt, J = 7.7, 4.8 Hz, 1H), 4.36 – 4.27 (m, 1H), 3.53 (s, 3H), 3.09 – 2.86 (m, 2H), 2.52 (t, J = 7.5 Hz, 2H), 2.15 – 2.01 (m, 1H), 1.99 (s, 3H), 1.96 – 1.78 (m, 1H), 1.36 (s, 9H).

$^{13}\text{C-NMR}$ (75 MHz, CDCl_3) δ 171.3, 171.2, 165.7, 155.3, 142.2, 138.7, 136.2, 129.7, 128.1, 119.6, 79.5, 75.4, 53.4, 52.6, 52.1, 32.6, 29.7, 29.5, 28.4, 15.2.

HRMS–ESI: m/z [$\text{M} + \text{H}$] $^+$ calcd. for $\text{C}_{36}\text{H}_{42}\text{N}_4\text{O}_5\text{S}$: 643.2949; found 643.2963.

Methyl N^α -((*tert*-butoxycarbonyl)-L-valyl)- N^ϵ -trityl-L-histidinate (1f**)**



The general procedure was followed starting from *L*-Boc-Val-OH (97 mg, 446 μ mol). The resulting crude product was purified on silica gel (Hexane-EtOAc, gradient from 80:20 to 30:70) to afford **1f** as a white solid (160 mg, 59%). **Mp** 61 – 63 °C; **R_f** = 0.25 (Hexane-EtOAc, 50:50).

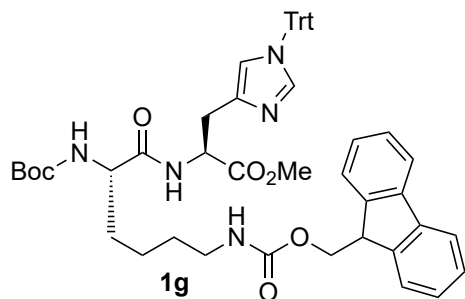
IR (ATR)/ $\bar{\nu}$: 3303, 2964, 1745, 1712, 1673, 1492, 1445, 1365, 1238, 1160, 1133, 1088, 1037, 1001, 909, 871, 841, 748, 700, 660, 637 cm^{-1} .

$^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 7.66 (d, J = 7.6 Hz, 1H), 7.36 (d, J = 1.4 Hz, 1H), 7.34 – 7.27 (m, 9H), 7.13 – 7.01 (m, 6H), 6.54 (s, 1H), 5.30 (d, J = 8.8 Hz, 1H), 4.75 (dt, J = 7.6, 4.9 Hz, 1H), 4.12 – 4.00 (m, 1H), 3.55 (s, 3H), 3.04 (dd, J = 14.7, 5.2 Hz, 1H), 2.93 (dd, J = 14.4, 5.0 Hz, 1H), 2.12 (h, J = 6.4 Hz, 1H), 1.38 (s, 9H), 0.90 (dd, J = 22.7, 6.8 Hz, 6H).

$^{13}\text{C-NMR}$ (75 MHz, CDCl_3) δ 171.5, 171.4, 155.8, 142.2, 138.7, 136.3, 129.8, 128.2, 128.1, 119.7, 79.5, 75.4, 59.4, 52.6, 52.1, 31.5, 29.6, 28.4, 19.0.

HRMS-ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{36}\text{H}_{42}\text{N}_4\text{O}_5$: 611.3228; found 611.3244.

Methyl *N* $^{\alpha}$ -(*N* 6 -(((9*H*-fluoren-9-yl)methoxy)carbonyl)-*N* 2 -(*tert*-butoxycarbonyl)-*L*-lysyl)-*N* $^{\epsilon}$ -trityl-*L*-histidinate (1g**)**



The general procedure was followed starting from *L*-Boc-Lys(Fmoc)-OH (209 mg, 446 μ mol). The resulting crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to 98:2) to afford **1g** as a white solid (224 mg, 58%). **Mp** 86 – 88 °C; **R_f** = 0.36 (DCM-MeOH, 95:5).

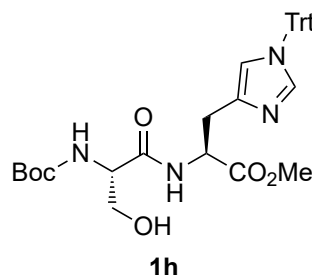
IR (ATR)/ $\bar{\nu}$: 3296, 2931, 1709, 1494, 1446, 1365, 1243, 1163, 1134, 1088, 1023, 1001, 842, 742, 701, 660, 637 cm^{-1} .

$^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 7.81 (d, J = 7.7 Hz, 1H), 7.76 (d, J = 7.5 Hz, 2H), 7.58 (d, J = 7.3 Hz, 2H), 7.38 (t, J = 7.1 Hz, 3H), 7.35 – 7.28 (m, 11H), 7.16 – 7.02 (m, 6H), 6.55 (s, 1H), 5.36 (d, J = 7.8 Hz, 1H), 5.32 – 5.22 (m, 1H), 4.79 (dt, J = 7.7, 4.7 Hz, 1H), 4.44 – 4.30 (m, 2H), 4.28 – 4.13 (m, 2H), 3.56 (s, 3H), 3.27 – 2.92 (m, 4H), 2.02 – 1.77 (m, 1H), 1.75 – 1.47 (m, 5H), 1.41 (s, 9H).

$^{13}\text{C-NMR}$ (75 MHz, CDCl_3) δ 171.8, 171.6, 156.6, 155.5, 144.1, 142.3, 141.4, 138.8, 136.4, 129.8, 128.2, 128.2, 127.7, 127.1, 125.2, 120.0, 119.7, 79.7, 75.4, 66.6, 54.1, 52.6, 52.3, 47.4, 40.6, 32.8, 29.5, 29.3, 28.4, 22.0.

HRMS–ESI: m/z $[M + H]^+$ calcd. for $C_{52}H_{55}N_5O_7$: 862.4174; found 862.4177.

Methyl N^α -((*tert*-butoxycarbonyl)-*L*-seryl)- N^ϵ -trityl-*L*-histidinate (1h**)**



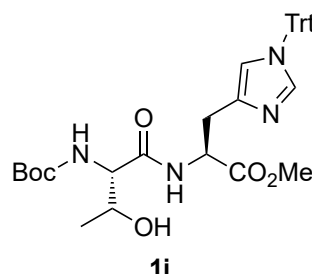
The general procedure was followed starting from *L*-Boc-Ser-OH (92 mg, 446 μ mol). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 80:20 to 20:80) to afford **1h** as a white solid (76 mg, 28%). **Mp** 86 – 88 °C; **R_f** = 0.21 (Hexane–EtOAc, 30:70).

IR (ATR)/ $\bar{\nu}$: 3300, 2933, 1713, 1667, 1492, 1445, 1390, 1365, 1240, 1159, 1133, 1086, 1056, 1034, 1002, 840, 748, 701, 660, 638 cm^{-1} .

1H -NMR (600 MHz, $CDCl_3$) δ 7.36 (s, 1H), 7.35 – 7.29 (m, 9H), 7.26 – 7.20 (m, 1H), 7.10 – 7.03 (m, 6H), 6.50 (s, 1H), 5.76 (s, 1H), 4.77 (s, 1H), 4.12 – 4.06 (m, 2H), 3.66 (dd, J = 12.0, 4.7 Hz, 1H), 3.56 (s, 3H), 3.29 – 3.23 (m, 1H), 2.84 (dd, J = 14.6, 5.9 Hz, 1H), 1.42 (s, 9H).

^{13}C -NMR (150 MHz, $CDCl_3$) δ 171.4, 171.2, 155.5, 142.1, 138.8, 135.9, 129.8, 128.3, 128.2, 120.0, 80.1, 75.6, 63.0, 57.3, 53.3, 52.4, 29.2, 28.4.

Methyl N^α -((*tert*-butoxycarbonyl)-*L*-threonyl)- N^ϵ -trityl-*L*-histidinate (1i**)**



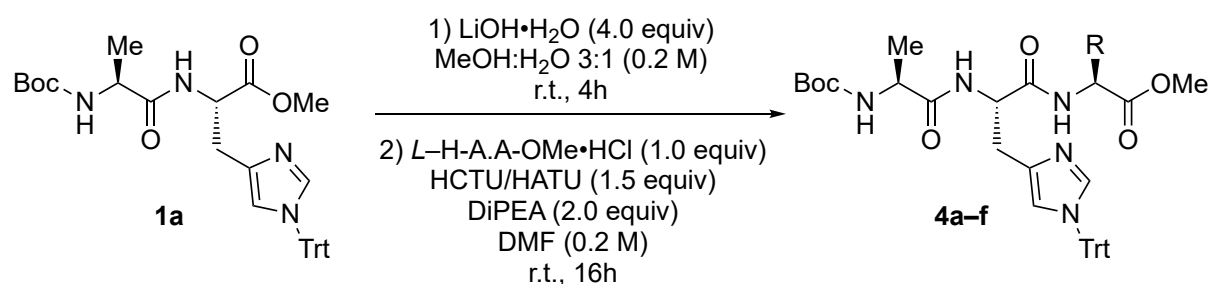
The general procedure was followed starting from *L*-Boc-Thr-OH (98 mg, 446 μ mol). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 80:20 to 20:80) to afford **1i** as a white solid (152 mg, 56%). **Mp** 68 – 70 °C; **R_f** = 0.14 (Hexane–EtOAc, 30:70).

IR (ATR)/ $\bar{\nu}$: 3312, 2975, 2931, 1742, 1712, 1672, 1491, 1445, 1366, 1240, 1160, 1133, 1088, 1063, 1036, 1000, 871, 841, 748, 701, 661, 638 cm^{-1} .

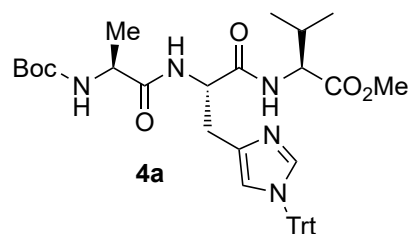
1H -NMR (600 MHz, $CDCl_3$) δ 7.26 (s, 1H), 7.23 – 7.19 (m, 9H), 7.02 (d, J = 7.1 Hz, 1H), 6.99 – 6.93 (m, 6H), 6.36 (s, 1H), 5.47 (d, J = 7.3 Hz, 1H), 4.65 (s, 1H), 4.29 (d, J = 6.6 Hz, 1H), 3.87 (d, J = 7.3 Hz, 1H), 3.44 (s, 3H), 3.27 (d, J = 13.9 Hz, 1H), 2.72 (dd, J = 14.5, 4.9 Hz, 1H), 1.34 (s, 9H), 1.12 (d, J = 6.4 Hz, 3H)./1

^{13}C -NMR (150 MHz, $CDCl_3$) δ 171.4, 171.3, 156.0, 142.1, 138.8, 135.9, 130.0, 128.2, 120.0, 80.0, 75.5, 67.3, 61.0, 53.4, 52.3, 28.9, 28.4, 20.0.

HRMS–ESI: m/z $[M + H]^+$ calcd. for $C_{35}H_{40}N_4O_6$: 613.3021; found 613.3025.

b. Synthesis of tripeptides Boc-Ala-His(Trt)-A.A-OMe (4a-f)

Manuscript, Scheme 4.: In a 50 mL flask, lithium hydroxide monohydrate (288 mg, 6.86 mmol, 4.0 equiv) was added to a solution of methyl *N*^α-((*tert*-butoxycarbonyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidinate (**1a**) (1g, 1.72 mmol, 1.0 equiv) in methanol:water 3:1 (9 mL, 0.2 M). The mixture was stirred for 4h at room temperature before concentrated to water under reduce pressure. The residue was diluted with a small amount of water before being acidify to pH 3 with a solution of HCl 1 M, a white precipitate appears. The suspension was extract twice with EtOAc (2x20 mL). The combined organic phases were washed with brine (20 mL), dried over Na₂SO₄, filtered and concentrated under reduce pressure. The white solid residue was used with no further purification step. In a 50 mL flask, DIPEA (2.0 equiv) was added to Boc-Ala-His(Trt)-OH (1.0 equiv), *L*-H-A.A-OMe·HCl (1.0 equiv) and HCTU or HATU (1.5 equiv) in DMF (0.2 M). The mixture was stirred for 16h at room temperature before being extracted twice with dichloromethane. The combined organic phases were successively washed with sat. aq. NaHCO₃ (2x20 mL), brine (2x20 mL), dried over Na₂SO₄, filtered and concentrated under reduce pressure. The resulting crude product was purified by column chromatography using the indicated eluent system to afford the desired pure product.

Methyl *N*^α-((*tert*-butoxycarbonyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (4a**)**

The general procedure was followed starting from Boc-Ala-His(Trt)-OH (640 mg, 1.13 mmol), *L*-H-Val-OMe·HCl (189 mg, 1.13 mmol), HCTU (698 mg, 1.69 mmol) and DIPEA (385 μL, 2.25 mmol) in DMF (6 mL). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 90:10 up to 20:80) to afford **4a** as a white solid (501 mg, 65%). **Mp** 79 – 81 °C; **R_f** = 0.42 (DCM–MeOH, 95:5).

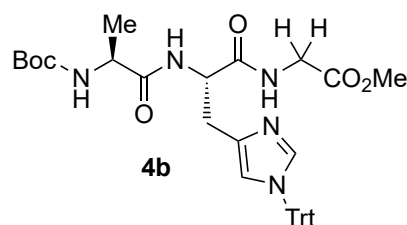
IR (ATR)/ $\bar{\nu}$: 3275, 2970, 1740, 1659, 1493, 1445, 1366, 1245, 1205, 1159, 1001, 841, 748, 701, 659, 638 cm⁻¹.

¹H-NMR (300 MHz, CDCl₃) δ 8.46 (d, *J* = 6.1 Hz, 1H), 7.52 (d, *J* = 8.2 Hz, 1H), 7.38 – 7.27 (m, 10H), 7.13 – 7.00 (m, 6H), 6.66 (s, 1H), 5.22 (d, *J* = 6.6 Hz, 1H), 4.78 – 4.65 (m, 1H), 4.39 (dd, *J* = 8.4, 5.3 Hz, 1H), 4.24 – 4.12 (m, 1H), 3.64 (s, 3H), 3.11 (A of ABX, *J* = 15.1, 4.3 Hz, 1H), 2.89 (B of ABX, *J* = 15.1, 5.9 Hz, 1H), 2.14 – 2.01 (m, 1H), 1.38 (d, *J* = 7.2 Hz, 3H), 1.35 (s, 9H), 0.80 (d, *J* = 6.8 Hz, 6H).

¹³C-NMR (75 MHz, CDCl₃) δ 172.8, 171.9, 171.0, 155.4, 142.2, 138.2, 136.9, 129.8, 128.2, 128.1, 119.7, 79.8, 75.5, 57.6, 53.3, 52.0, 50.7, 30.9, 29.3, 28.4, 19.1, 18.0.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₃₉H₄₇N₅O₆: 682.3599, found 682.3594.

Methyl *N*^α-((*tert*-butoxycarbonyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidylglycinate (4b**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-OH (170 mg, 0.3 mmol), H-Gly-OMe•HCl (38 mg, 0.3 mmol), HCTU (186 mg, 0.45 mmol) and DiPEA (105 μL, 0.6 mmol) in DMF (2 mL). The resulting crude product was

purified on silica gel (DCM–MeOH, gradient from 100:0 to 97:3) to afford **4b** as a white solid (119 mg, 62%). **Mp** 93 – 95 °C; **R_f** = 0.13 (DCM–MeOH, 95:5).

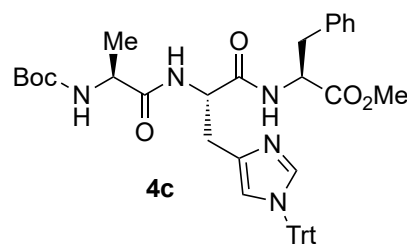
IR (ATR)/ $\bar{\nu}$: 3290, 2977, 1753, 1660, 1520, 1492, 1445, 1365, 1245, 1203, 1162, 1133, 1036, 1001, 844, 748, 700, 660, 638 cm⁻¹.

¹H-NMR (300 MHz, CDCl₃) δ 8.61 (d, *J* = 7.7 Hz, 1H), 7.56 (t, *J* = 5.6 Hz, 1H), 7.33 (d, *J* = 1.4 Hz, 1H), 7.32 – 7.26 (m, 9H), 7.15 – 7.01 (m, 6H), 6.64 (s, 1H), 5.35 (d, *J* = 5.3 Hz, 1H), 4.70 (dt, *J* = 7.7, 5.0 Hz, 1H), 4.15 – 4.00 (m, 1H), 3.97 – 3.79 (m, 2H), 3.62 (s, 3H), 3.13 (dd, *J* = 14.1, 4.2 Hz, 1H), 2.87 (dd, *J* = 14.7, 5.4 Hz, 1H), 1.36 (d, *J* = 7.0 Hz, 3H), 1.29 (s, 9H).

¹³C-NMR (75 MHz, CDCl₃) δ 172.8, 171.5, 169.6, 156.1, 142.3, 138.3, 136.8, 129.8, 128.1, 128.1, 119.9, 80.1, 75.4, 53.2, 52.0, 51.4, 41.1, 29.1, 28.3, 18.1.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₃₆H₄₁N₅O₆: 640.3130, found 640.3129.

Methyl *N*^α-((*tert*-butoxycarbonyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-phenylalaninate (4c**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-OH (250 mg, 0.44 mmol), *L*-H-Phe-OMe•HCl (95 mg, 0.44 mmol), HCTU (273 mg, 0.66 mmol) and DiPEA (150 μL, 0.76 mmol) in DMF (2 mL). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from

90:10 to 30:70) to afford **4c** as a white solid (212 mg, 66%). **Mp** 87 – 89 °C; **R_f** = 0.41 (DCM–MeOH, 95:5).

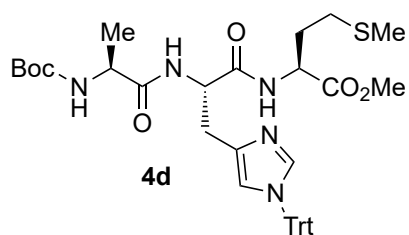
IR (ATR)/ $\bar{\nu}$: 3290, 2976, 1744, 1657, 1493, 1444, 1365, 1243, 1212, 1163, 1131, 1087, 1031, 1001, 856, 747, 699, 659, 638 cm⁻¹.

¹H-NMR (300 MHz, CDCl₃) δ 8.09 (d, *J* = 7.3 Hz, 1H), 7.63 (d, *J* = 7.6 Hz, 1H), 7.34 – 7.24 (m, 9H), 7.21 – 7.13 (m, 3H), 7.12 – 6.99 (m, 9H), 6.64 (s, 1H), 5.44 (d, *J* = 6.9 Hz, 1H), 4.78 – 4.63 (m, 2H), 4.27 – 4.12 (m, 1H), 3.55 (s, 3H), 3.09 – 2.96 (m, 3H), 2.89 (dd, *J* = 14.8, 5.9 Hz, 1H), 1.36 (s, 9H), 1.28 (d, *J* = 7.1 Hz, 3H).

¹³C-NMR (75 MHz, CDCl₃) δ 172.7, 171.4, 170.6, 155.4, 142.2, 138.2, 136.8, 136.1, 129.7, 129.2, 128.4, 128.0, 128.0, 126.9, 119.7, 79.6, 75.3, 53.7, 53.1, 52.0, 50.4, 37.9, 29.6, 28.3, 18.7.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₄₃H₄₇N₅O₆: 730.3599, found 730.3605.

Methyl *N*^α-((*tert*-butoxycarbonyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-methioninate (**4d**)



The general procedure was followed starting from Boc-Ala–His(Trt)–OH (170 mg, 0.3 mmol), *L*-H-Met–OMe•HCl (60 mg, 0.3 mmol), HCTU (186 mg, 0.45 mmol) and DiPEA (105 μ L, 0.6 mmol) in DMF (2 mL). The resulting crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to

97:3) to afford **4d** as a white solid (95 mg, 44%). **Mp** 118 – 120 °C; **R_f** = 0.14 (DCM–MeOH, 95:5).

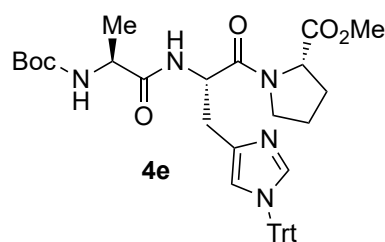
IR (ATR)/ $\bar{\nu}$: 3265, 2969, 2915, 1744, 1715, 1662, 1645, 1518, 1446, 1234, 1215, 1160, 1134, 1036, 1014, 837, 748, 700, 657 cm⁻¹.

¹H-NMR (300 MHz, CDCl₃) δ 8.39 (d, *J* = 7.4 Hz, 1H), 7.61 (d, *J* = 6.6 Hz, 1H), 7.36 – 7.27 (m, 10H), 7.13 – 7.00 (m, 6H), 6.65 (s, 1H), 5.22 (d, *J* = 5.9 Hz, 1H), 4.67 (dt, *J* = 7.5, 5.2 Hz, 1H), 4.58 – 4.45 (m, 1H), 4.10 (p, *J* = 6.9 Hz, 1H), 3.63 (s, 3H), 3.22 – 3.09 (m, 1H), 2.89 – 2.80 (m, 1H), 2.38 (t, *J* = 7.5 Hz, 2H), 2.06 – 2.01 (m, 1H), 1.99 (s, 3H), 1.95 – 1.89 (m, 1H), 1.38 (d, *J* = 6.9 Hz, 3H), 1.32 (s, 9H).

¹³C-NMR (75 MHz, CDCl₃) δ 172.8, 171.9, 171.1, 155.8, 142.1, 138.3, 136.6, 129.8, 128.2, 128.1, 119.9, 80.1, 75.5, 53.3, 53.2, 52.4, 51.7, 31.3, 30.0, 28.4, 18.4, 15.4, 15.0.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₃₉H₄₇N₅O₆S: 714.3320, found 714.3323.

Methyl *N*^α-((*tert*-butoxycarbonyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-prolinate (4e**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-OH (200 mg, 0.35 mmol), *L*-H-Pro-OMe•HCl (58 mg, 0.35 mmol), HATU (201 mg, 0.53 mmol) and DiPEA (120 μ L, 0.7 mmol) in DMF (2 mL). The resulting crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to

97:3) to afford **4e** as a white solid (119 mg, 50%). **Mp** 92 – 94 °C; **R_f** = 0.25 (DCM-MeOH, 95:5).

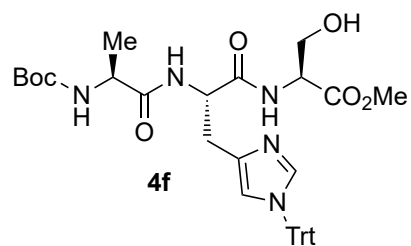
IR (ATR)/ $\bar{\nu}$: 3284, 2959, 1743, 1709, 1643, 1493, 1445, 1365, 1245, 1161, 1131, 1022, 1001, 837, 747, 701, 661, 638 cm^{-1} .

¹H-NMR (600 MHz, CDCl₃) δ 7.66 (s, 1H), 7.37 – 7.28 (m, 10H), 7.14 – 7.10 (m, 6H), 6.86 (s, 1H), 5.36 (d, *J* = 6.2 Hz, 1H), 4.99 – 4.91 (m, 1H), 4.50 (dd, *J* = 8.5, 3.7 Hz, 1H), 4.14 – 4.05 (m, 1H), 3.72 – 3.65 (m, 1H), 3.62 – 3.56 (m, 1H), 3.43 (s, 3H), 3.14 – 3.08 (m, 1H), 2.92 (dd, *J* = 14.8, 5.3 Hz, 1H), 2.20 – 2.12 (m, 1H), 1.99 – 1.87 (m, 3H), 1.35 (s, 9H), 1.26 (d, *J* = 7.0 Hz, 3H).

Presence of conformers: **¹³C-NMR (150 MHz, CDCl₃)** δ 172.5, 172.5, 169.1, 155.6, 141.4, 137.1, 129.9, 129.7, 128.4, 128.3, 128.3, 128.2, 127.8, 121.4, 79.9, 76.8, 58.9, 55.8, 52.2, 50.8, 50.3, 47.0, 28.9, 28.4, 24.8, 18.4.

HRMS-ESI: *m/z* [*M* + *H*]⁺ calcd. for C₃₉H₄₅N₅O₆: 680.3443, found 680.3443.

Methyl *N*^α-((*tert*-butoxycarbonyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-serinate (4f**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-OH (170 mg, 0.3 mmol), *L*-H-Ser-OMe•HCl (47 mg, 0.3 mmol), HCTU (186 mg, 0.53 mmol) and DiPEA (105 μ L, 0.6 mmol) in DMF (2 mL). The resulting crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to

95:5) to afford **4f** as a white solid (107 mg, 53%). **Mp** 100 – 102 °C; **R_f** = 0.29 (DCM-MeOH, 95:5).

IR (ATR)/ $\bar{\nu}$: 3292, 2977, 1743, 1655, 1492, 1445, 1365, 1239, 1205, 1161, 1132, 1068, 1036, 1001, 841, 748, 700, 660, 638 cm^{-1} .

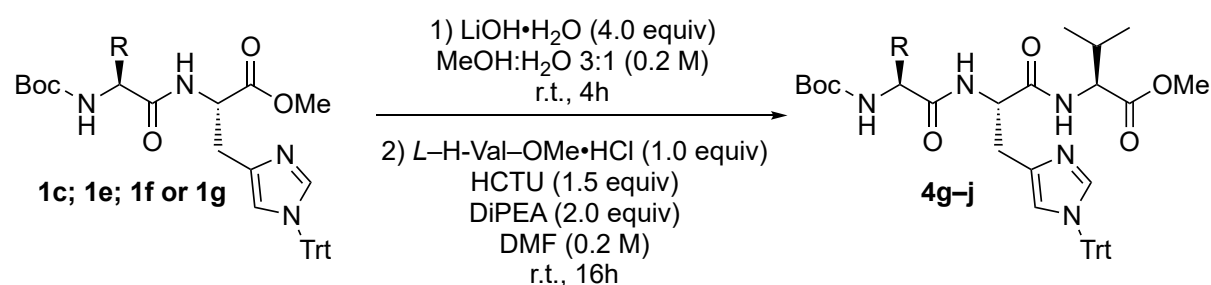
¹H-NMR (300 MHz, CDCl₃) δ 7.88 (d, *J* = 6.1 Hz, 1H), 7.55 (d, *J* = 7.4 Hz, 1H), 7.37 – 7.34 (m, 1H), 7.33 – 7.27 (m, 9H), 7.14 – 7.00 (m, 6H), 6.65 (s, 1H), 5.31 (d, *J* = 6.0 Hz, 1H), 4.80 – 4.68 (m, 1H), 4.49 – 4.40 (m, 1H), 4.13 – 3.99 (m, 1H), 3.91 (dd, *J* = 11.7, 4.5 Hz, 1H), 3.84 (dd, *J* =

11.6, 3.5 Hz, 1H), 3.63 (s, 3H), 3.13 (dd, $J = 14.5, 4.4$ Hz, 1H), 2.94 (dd, $J = 15.2, 4.6$ Hz, 1H), 1.38 – 1.32 (m, 3H), 1.31 (s, 9H).

$^{13}\text{C-NMR}$ (75 MHz, CDCl_3) δ 172.8, 171.0, 170.6, 155.9, 142.2, 138.5, 136.1, 129.8, 128.2, 128.2, 120.5, 80.3, 75.5, 62.0, 55.3, 52.9, 52.4, 51.0, 29.5, 28.4, 18.3.

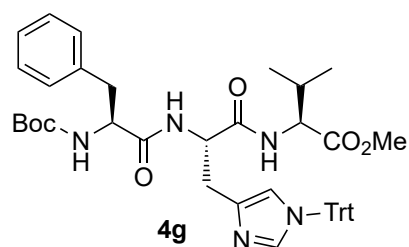
HRMS-ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{37}\text{H}_{43}\text{N}_5\text{O}_7$: 670.3235, found 670.3244.

c. Synthesis of tripeptides Boc-A.A-His(Trt)-Val-OMe (4g-j)



Manuscript, Scheme 5.: In a 50 mL flask, lithium hydroxide monohydrate (4.0 equiv) was added to a solution of methyl N^α -((*tert*-butoxycarbonyl)-*L*-Amino acid)- N^ϵ -trityl-*L*-histidinate (**1c**; **1e**; **1f** or **1g**) (1.0 equiv) in methanol:water 3:1 (9 mL, 0.2 M). The mixture was stirred for 4h at room temperature before concentrated to water under reduce pressure. The residue was diluted with a small amount of water before being acidify to pH 3 with a solution of HCl 1 M, a white precipitate appears. The suspension was extract twice with EtOAc (2x20 mL). The combined organic phases were washed with brine (20 mL), dried over Na_2SO_4 , filtered and concentrated under reduce pressure. The white solid residue was used with no further purification step. In a 50 mL flask, DIPEA (2.0 equiv) was added to Boc-A.A-His(Trt)-OH (1.0 equiv), *L*-H-Val-OMe·HCl (1.0 equiv) and HCTU (1.5 equiv) in DMF (0.2 M). The mixture was stirred for 16h at room temperature before being extracted twice with dichloromethane. The combined organic phases were successively washed with sat. aq. NaHCO_3 (2x20 mL), brine (2x20 mL), dried over Na_2SO_4 , filtered and concentrated under reduce pressure. The resulting crude product was purified by column chromatography using the indicated eluent system to afford the desired pure product.

Methyl N^α -((*tert*-butoxycarbonyl)-*L*-phenylalanyl)- N^ϵ -trityl-*L*-histidyl-*L*-valinate (4g)



The general procedure was followed starting from Boc-Phe-His(Trt)-OH (228 mg, 0.35 mmol), *L*-H-Val-OMe•HCl (60 mg, 0.35 mmol), HCTU (219 mg, 0.53 mmol) and DiPEA (120 μ L, 0.71 mmol) in DMF (2 mL). The resulting crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to

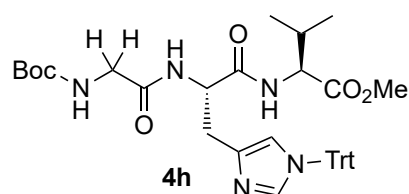
95:5) to afford **4g** as a white solid (150 mg, 56%). **Mp** 83 – 85 °C; **R_f** = 0.43 (DCM-MeOH, 95:5). **IR (ATR)/ $\bar{\nu}$** : 3290, 2966, 1741, 1651, 1492, 1445, 1365, 1246, 1205, 1157, 1086, 1021, 1001, 870, 842, 747, 699, 660, 638 cm^{-1} .

$^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 8.34 (d, J = 6.0 Hz, 1H), 7.46 (d, J = 7.9 Hz, 1H), 7.35 – 7.25 (m, 10H), 7.23 – 7.12 (m, 5H), 7.11 – 7.02 (m, 6H), 6.64 (s, 1H), 5.18 (d, J = 6.8 Hz, 1H), 4.72 (q, J = 6.3 Hz, 1H), 4.46 – 4.33 (m, 2H), 3.63 (s, 3H), 3.17 (dd, J = 14.0, 5.0 Hz, 1H), 3.07 (dd, J = 14.9, 3.7 Hz, 1H), 3.03 – 2.90 (m, 1H), 2.87 – 2.78 (m, 1H), 2.16 – 2.03 (m, 1H), 1.28 (s, 9H), 0.83 (d, J = 3.3 Hz, 3H), 0.81 (d, J = 3.4 Hz, 3H).

$^{13}\text{C-NMR}$ (75 MHz, CDCl_3) δ 172.0, 171.6, 171.0, 155.6, 142.2, 138.4, 136.8, 136.7, 129.7, 129.3, 128.6, 128.1, 128.1, 126.9, 119.6, 79.9, 75.4, 57.7, 56.0, 53.4, 52.0, 30.8, 29.5, 28.2, 19.0, 18.0.

HRMS-ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{45}\text{H}_{51}\text{N}_5\text{O}_6$: 758.3912, found 758.3913.

Methyl *N* $^{\alpha}$ -((*tert*-butoxycarbonyl)-*L*-glycyl)-*N* $^{\epsilon}$ -trityl-*L*-histidyl-*L*-valinate (**4h**)



The general procedure was followed starting from Boc-Gly-His(Trt)-OH (276 mg, 0.5 mmol), *L*-H-Val-OMe•HCl (83 mg, 0.5 mmol), HCTU (309 mg, 0.75 mmol) and DiPEA (170 μ L, 1.0 mmol) in DMF (2.5 mL). The resulting crude product was

purified on silica gel (DCM-MeOH, gradient from 100:0 to 95:5) to afford **4h** as a white solid (64 mg, 19%). **Mp** 78 – 80 °C; **R_f** = 0.64 (DCM-MeOH, 90:10).

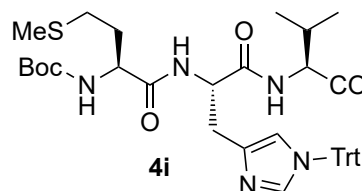
IR (ATR)/ $\bar{\nu}$: 3297, 2966, 2930, 1745, 1715, 1658, 1494, 1445, 1366, 1247, 1206, 1157, 1033, 1001, 870, 749, 702, 661, 639 cm^{-1} .

$^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 8.14 (d, J = 6.7 Hz, 1H), 7.67 (d, J = 8.3 Hz, 1H), 7.35 – 7.28 (m, 10H), 7.13 – 7.05 (m, 6H), 6.65 (s, 1H), 5.37 – 5.27 (m, 1H), 4.77 (q, J = 6.1 Hz, 1H), 4.41 (dd, J = 8.4, 5.1 Hz, 1H), 3.91 – 3.78 (m, 2H), 3.64 (s, 3H), 3.07 (dd, J = 15.0, 4.7 Hz, 1H), 2.95 – 2.85 (m, 1H), 2.18 – 2.06 (m, 1H), 1.38 (s, 9H), 0.84 (d, J = 1.5 Hz, 3H), 0.82 (d, J = 1.6 Hz, 3H).

¹³C-NMR (75 MHz, CDCl₃) δ 172.0, 171.1, 169.5, 156.0, 142.4, 138.3, 137.0, 129.8, 128.2, 128.1, 119.7, 79.9, 75.4, 57.7, 53.1, 52.1, 38.7, 31.0, 29.9, 28.4, 19.2, 18.0.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₃₈H₄₅N₅O₆: 668.3443, found 668.3457.

Methyl N^α-((*tert*-butoxycarbonyl)-*L*-methionyl)-N^ε-trityl-*L*-histidyl-*L*-valinate (4i)



The general procedure was followed starting from Boc–Met–His(Trt)–OH (226 mg, 0.36 mmol), *L*–H–Val–OMe•HCl (60 mg, 0.36 mmol), HCTU (223 mg, 0.54 mmol) and DiPEA (125 μL, 0.72 mmol) in DMF (2 mL). The resulting crude

product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 95:5) to afford **4i** as a white solid (64 mg, 19%). **Mp** 86 – 88 °C; **R_f** = 0.43 (DCM–MeOH, 95:5).

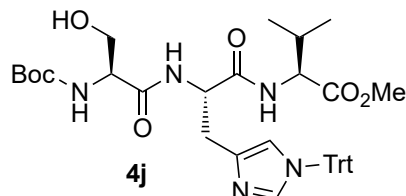
IR (ATR)/ν̄: 3285, 2967, 1740, 1657, 1493, 1445, 1390, 1366, 1246, 1208, 1158, 1024, 1001, 840, 748, 701, 660, 639 cm^{–1}.

¹H-NMR (300 MHz, CDCl₃) δ 8.32 (d, *J* = 6.3 Hz, 1H), 7.42 (d, *J* = 7.9 Hz, 1H), 7.36 – 7.27 (m, 10H), 7.13 – 7.01 (m, 6H), 6.65 (s, 1H), 5.47 (d, *J* = 7.1 Hz, 1H), 4.71 (q, *J* = 6.0 Hz, 1H), 4.39 (dd, *J* = 8.4, 5.2 Hz, 1H), 4.32 – 4.22 (m, 1H), 3.62 (s, 3H), 3.06 (dd, *J* = 14.8, 4.7 Hz, 1H), 2.92 – 2.85 (m, 1H), 2.54 (t, *J* = 7.4 Hz, 2H), 2.18 – 2.06 (m, 2H), 2.05 (s, 3H), 1.95 – 1.82 (m, 1H), 1.35 (s, 9H), 0.82 (d, *J* = 1.2 Hz, 3H), 0.80 (d, *J* = 1.3 Hz, 3H).

¹³C-NMR (75 MHz, CDCl₃) δ 172.1, 171.9, 171.0, 155.7, 142.3, 138.4, 136.7, 129.8, 128.2, 128.1, 119.8, 80.0, 75.5, 57.6, 54.2, 53.4, 52.1, 32.1, 30.9, 30.2, 28.4, 19.1, 18.0, 15.3.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₄₁H₅₁N₅O₆S: 742.3633, found 742.3647.

Methyl N^α-((*tert*-butoxycarbonyl)-*L*-seryl)-N^ε-trityl-*L*-histidyl-*L*-valinate (4j)



The general procedure was followed starting from Boc–Ser–His(Trt)–OH (160 mg, 0.27 mmol), *L*–H–Val–OMe•HCl (46 mg, 0.27 mmol), HCTU (170 mg, 0.41 mmol) and DiPEA (95 μL, 0.55 mmol) in DMF (1.5 mL). The resulting crude product

was purified on silica gel (DCM–MeOH, gradient from 100:0 to 95:5) to afford **4j** as a white solid (45 mg, 24%). **Mp** 104 – 106 °C; **R_f** = 0.34 (DCM–MeOH, 95:5).

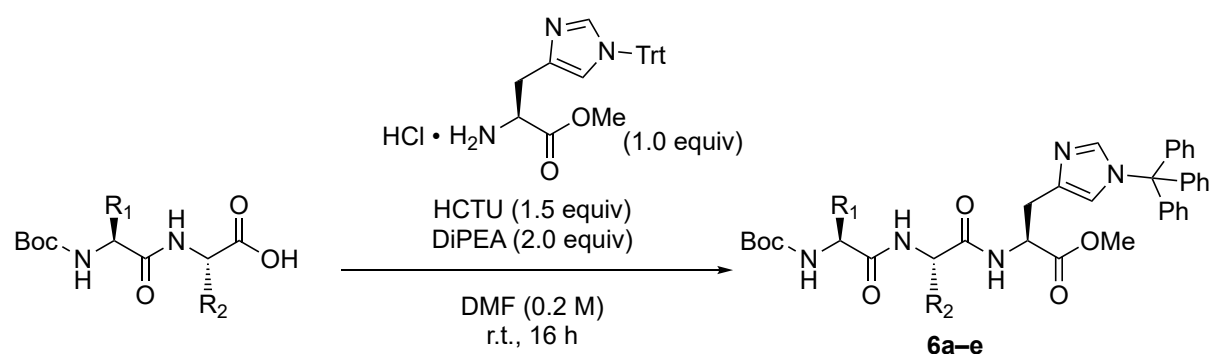
IR (ATR)/ν̄: 3285, 2959, 1739, 1655, 1492, 1445, 1366, 1208, 1156, 1057, 1037, 1001, 841, 748, 700, 660 cm^{–1}.

¹H-NMR (300 MHz, CDCl₃) δ 7.82 (d, *J* = 6.6 Hz, 1H), 7.37 – 7.27 (m, 11H), 7.14 – 7.01 (m, 7H), 6.62 (s, 1H), 5.69 (d, *J* = 6.1 Hz, 1H), 4.77 (q, *J* = 5.9 Hz, 1H), 4.44 (dd, *J* = 8.6, 5.0 Hz, 1H), 4.15 (s(br), 1H), 4.01 (dd, *J* = 11.3, 2.0 Hz, 1H), 3.68 (dd, *J* = 11.3, 5.6 Hz, 1H), 3.56 (s, 3H), 3.11 (dd, *J* = 14.9, 4.2 Hz, 1H), 2.93 (dd, *J* = 15.2, 6.0 Hz, 1H), 2.20 – 2.02 (m, 1H), 1.40 (s, 9H), 0.84 (d, *J* = 4.3 Hz, 3H), 0.82 (d, *J* = 4.4 Hz, 3H).

¹³C-NMR (150 MHz, CDCl₃) δ 172.4, 171.5, 170.7, 155.6, 142.2, 138.6, 136.4, 129.9, 128.2, 128.2, 120.1, 80.1, 75.6, 63.1, 57.4, 57.0, 53.9, 52.2, 31.1, 29.5, 28.4, 19.1, 17.9.

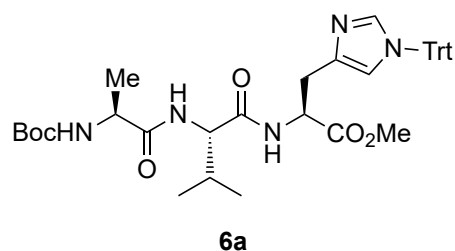
HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₃₉H₄₇N₅O₇: 698.3548, found 698.3556.

d. Synthesis of tripeptides Boc–A.A₁–A.A₂–His(Trt)–OMe (6a–e)



Manuscript, Scheme 6.: In a 50 mL flask, DIPEA (2.0 equiv) was added to Boc–A.A₁–A.A₂–OH (1.0 equiv), L–H–His(Trt)–OMe•HCl (1.0 equiv) and HCTU (1.5 equiv) in DMF (0.2 M). The mixture was stirred for 16h at room temperature before being extracted twice with dichloromethane. The combined organic phases were successively washed with sat. aq. NaHCO₃ (2x20 mL), brine (2x20 mL), dried over Na₂SO₄, filtered and concentrated under reduce pressure. The resulting crude product was purified by column chromatography using the indicated eluent system to afford the desired pure product.

Methyl *N*^α-(*tert*-butoxycarbonyl)-*L*-alanyl-*L*-valyl-*N*^ε-trityl-*L*-histidinate (6a)



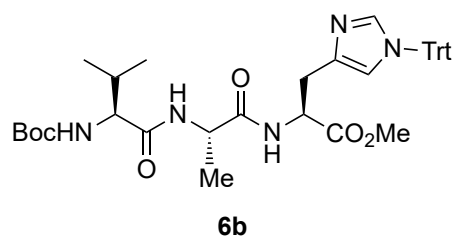
This compound has been previously reported ¹: **Mp** 87 – 89 °C; **R_f** = 0.34 (DCM–MeOH, 95:5).

IR (ATR)/ $\bar{\nu}$: 3292, 2972, 1650, 1493, 1445, 1162, 838, 748, 701, 660 cm⁻¹.

¹H-NMR (300 MHz, CDCl₃) δ 7.82 (d, *J* = 7.4 Hz, 1H), 7.43

(s, 1H), 7.35 – 7.25 (m, 9H), 7.15 – 7.09 (m, 1H), 7.09 – 7.01 (m, 6H), 6.58 (s, 1H), 5.44 (s, 1H), 4.79 – 4.63 (m, 1H), 4.31 (dd, *J* = 8.2, 5.7 Hz, 1H), 4.23 – 4.09 (m, 1H), 3.53 (s, 3H), 3.01 (dd, *J* = 14.6, 5.1 Hz, 1H), 2.94 (dd, *J* = 14.9, 5.0 Hz, 1H), 2.20 – 2.05 (m, 1H), 1.38 (s, 9H), 1.26 (d, *J* = 7.0 Hz, 3H), 0.90 (t, *J* = 6.1 Hz, 6H).

Methyl *N*^α-(*tert*-butoxycarbonyl)-*L*-valyl-*L*-alanyl-*N*^ε-trityl-*L*-histidinate (6b)

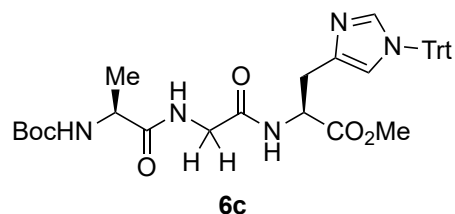


This compound has been previously reported ¹: **Mp** 92 – 94 °C; **R_f** = 0.36 (DCM–MeOH, 95:5).

IR (ATR)/ $\bar{\nu}$: 3275, 2972, 1745, 1651, 1493, 1445, 1366, 1160, 1132, 838, 748, 701, 660 cm⁻¹.

¹H-NMR (300 MHz, CDCl₃) δ 7.70 (d, *J* = 7.8 Hz, 1H), 7.45 (s, 1H), 7.34 – 7.27 (m, 9H), 7.13 – 7.00 (m, 7H), 6.56 (d, *J* = 1.0 Hz, 1H), 5.25 (d, *J* = 8.9 Hz, 1H), 4.80 – 4.70 (m, 1H), 4.48 (p, *J* = 6.9 Hz, 1H), 4.01 – 3.91 (m, 1H), 3.55 (s, 3H), 3.04 (dd, *J* = 14.9, 5.1 Hz, 1H), 2.94 (dd, *J* = 10.2, 5.6 Hz, 1H), 2.14 – 1.99 (m, 1H), 1.39 (s, 9H), 1.36 (d, *J* = 7.1 Hz, 3H), 0.85 (dd, *J* = 13.5, 6.8 Hz, 6H).

Methyl *N*^α-(*tert*-butoxycarbonyl)-*L*-alanylglycyl-*N*^ε-trityl-*L*-histidinate (6c)



The general procedure was followed starting from Boc–Ala–Gly–OH (100 mg, 0.41 mmol), *L*–H–His(Trt)–OMe•HCl (182 mg, 0.41 mmol), HCTU (252 mg, 0.61 mmol) and DiPEA (140 μ L, 0.81 mmol) in DMF (2 mL). The

resulting crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 97:3) to afford **6c** as a white solid (146 mg, 56%). **Mp** 86 – 89 °C; **R_f** = 0.20 (DCM–MeOH, 95:5).

¹ Chan, H.-C.; Bueno, B.; Le Roch, A.; Gagnon, *Chem.Eur. J.* **2021**, 27, 13330–13336

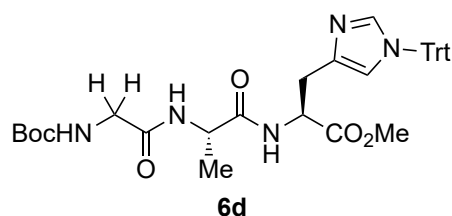
IR (ATR)/ $\bar{\nu}$: 3298, 2978, 1743, 1662, 1493, 1445, 1365, 1241, 1210, 1162, 1132, 1025, 1001, 870, 842, 747, 700, 660, 638 cm^{-1} .

$^1\text{H-NMR}$ (600 MHz, CDCl_3) δ 7.72 (d, J = 6.5 Hz, 1H), 7.39 (s, 1H), 7.33 – 7.27 (m, 9H), 7.12 (s, 1H), 7.10 – 7.04 (m, 6H), 6.53 (s, 1H), 5.54 (d, J = 7.8 Hz, 1H), 4.74 (dt, J = 7.6, 5.0 Hz, 1H), 4.24 (s(br), 1H), 4.00 (dd, J = 16.6, 3.4 Hz, 1H), 3.89 (dd, J = 17.0, 5.0 Hz, 1H), 3.54 (s, 3H), 3.01 (dd, J = 14.6, 5.4 Hz, 1H), 2.93 (dd, J = 14.6, 4.6 Hz, 1H), 1.36 (s, 9H), 1.31 (d, J = 7.1 Hz, 3H).

$^{13}\text{C-NMR}$ (150 MHz, CDCl_3) δ 173.1, 171.4, 168.6, 155.5, 142.2, 138.8, 136.1, 129.7, 128.2, 128.1, 119.7, 79.9, 75.4, 52.7, 52.2, 50.1, 42.8, 29.6, 28.3, 18.7.

HRMS–ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{36}\text{H}_{41}\text{N}_5\text{O}_6$: 640.3130, found 640.3139.

Methyl N^α -(*tert*-butoxycarbonyl)glycyl-*L*-alanyl- N^ϵ -trityl-*L*-histidinate (**6d**)



The general procedure was followed starting from Boc–Gly–Ala–OH (100 mg, 0.41 mmol), *L*–H–His(Trt)–OMe•HCl (182 mg, 0.41 mmol), HCTU (252 mg, 0.61 mmol) and DiPEA (140 μL , 0.81 mmol) in DMF (2 mL). The

resulting crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 97:3) to afford **6d** as a white solid (120 mg, 46%). **Mp** 89 – 91 $^\circ\text{C}$; **R_f** = 0.18 (DCM–MeOH, 95:5).

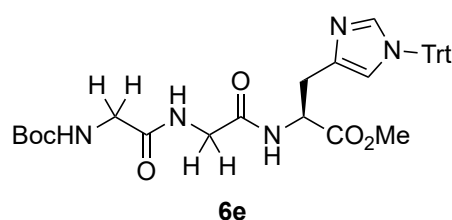
IR (ATR)/ $\bar{\nu}$: 3291, 2978, 1652, 1494, 1445, 1366, 1241, 1209, 1159, 1088, 1034, 1001, 942, 842, 747, 700, 660, 638 cm^{-1} .

$^1\text{H-NMR}$ (600 MHz, CDCl_3) δ ^1H 7.76 (d, J = 7.6 Hz, 1H), 7.37 (s, 1H), 7.34 – 7.28 (m, 9H), 7.11 – 7.04 (m, 7H), 6.54 (s, 1H), 5.54 (t, J = 5.1 Hz, 1H), 4.73 (dt, J = 7.6, 4.9 Hz, 1H), 4.59 – 4.49 (m, 1H), 3.82 (dd, J = 16.6, 4.3 Hz, 1H), 3.75 (dd, J = 16.2, 3.7 Hz, 1H), 3.55 (s, 3H), 3.02 (dd, J = 14.7, 5.4 Hz, 1H), 2.95 (dd, J = 14.7, 4.3 Hz, 1H), 1.39 (s, 9H), 1.36 (d, J = 7.0 Hz, 3H).

$^{13}\text{C-NMR}$ (150 MHz, CDCl_3) δ 172.0, 171.4, 169.2, 156.0, 142.2, 138.7, 136.2, 129.7, 128.2, 128.2, 119.7, 78.0, 75.4, 52.7, 52.2, 48.8, 44.2, 29.4, 28.3, 18.6.

HRMS–ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{36}\text{H}_{41}\text{N}_5\text{O}_6$: 640.3130, found 640.3150.

Methyl N^α -(*tert*-butoxycarbonyl)glycylglycyl- N^ϵ -trityl-*L*-histidinate (**6e**)



The general procedure was followed starting from Boc–Gly–Gly–OH (229 mg, 0.99 mmol), *L*–H–His(Trt)–OMe•HCl (442 mg, 0.99 mmol), HCTU (612 mg, 1.48 mmol) and DiPEA (335 μL , 1.97 mmol) in DMF (5 mL). The

resulting crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 97:3) to afford **6e** as a white solid (150 mg, 24%). **Mp** 85 – 87 °C; **R_f** = 0.17 (DCM–MeOH, 95:5).

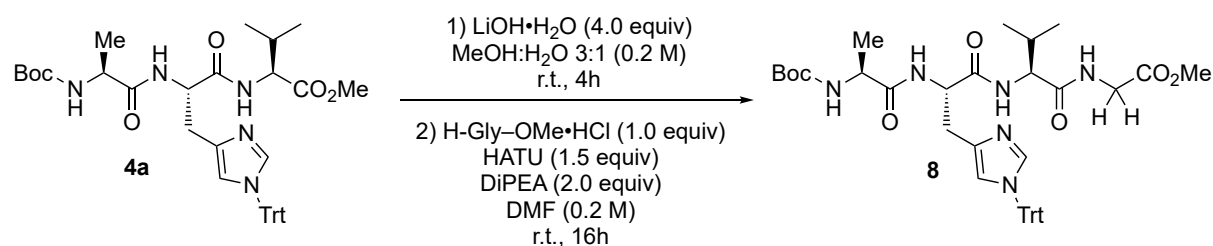
IR (ATR)/ $\bar{\nu}$: 3315, 2971, 1744, 1668, 1516, 1495, 1445, 1281, 1248, 1213, 1163, 1132, 1110, 950, 843, 749, 702, 661 cm^{-1} .

$^1\text{H-NMR}$ (600 MHz, CDCl_3) δ 7.78 (d, J = 7.6 Hz, 1H), 7.45 (s, 1H), 7.32 – 7.27 (m, 10H), 7.11 – 7.04 (m, 6H), 6.57 (s, 1H), 5.80 (s, 1H), 4.74 – 4.69 (m, 1H), 3.95 (dd, J = 16.9, 5.3 Hz, 1H), 3.90 (dd, J = 16.9, 5.2 Hz, 1H), 3.79 (s(br), 2H), 3.53 (s, 3H), 2.99 (dd, J = 14.8, 5.7 Hz, 1H), 2.95 (dd, J = 14.9, 4.9 Hz, 1H), 1.36 (s, 9H).

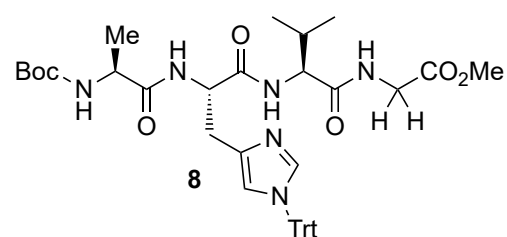
$^{13}\text{C-NMR}$ (150 MHz, CDCl_3) δ 171.5, 170.6, 168.9, 156.2, 142.0, 138.5, 135.7, 129.7, 128.2, 128.1, 119.9, 80.0, 75.6, 52.7, 52.2, 44.1, 42.7, 29.4, 28.3.

HRMS–ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{35}\text{H}_{39}\text{N}_5\text{O}_6$: 626.2973, found 626.2994.

e. Synthesis of tetrapeptide Boc–Ala–His(Trt)–Val–Gly–OMe (**8**)



Methyl N^α -((*tert*-butoxycarbonyl)-*L*-alanyl)- N^ϵ -trityl-*L*-histidyl-*L*-valylglycinate (**8**)



Manuscript, Scheme 7.: In a 50 mL flask, lithium hydroxide monohydrate (62 mg, 1.5 mmol, 4.0 equiv) was added to a solution of methyl N^α -((*tert*-butoxycarbonyl)-*L*-alanyl)- N^ϵ -trityl-*L*-histidyl-*L*-

valinate (**4a**) (250 mg, 0.37 mmol, 1.0 equiv) in methanol:water 3:1 (3 mL, 0.1 M). The mixture was stirred for 4h at room temperature before concentrated to water under reduce pressure. The residue was diluted with a small amount of water before being acidify to pH 3 with a solution of HCl 1 M, a white precipitate appears. The suspension was extract twice with EtOAc (2x20 mL). The combined organic phases were washed with brine (20 mL), dried over Na_2SO_4 , filtered and concentrated under reduce pressure. The white solid residue was used with no further purification step. In a 50 mL flask, DIPEA (115 μL , 0.67 mmol, 2.0 equiv) was added to

Boc-Ala-His(Trt)-Val-OH (223 mg, 0.33 mmol, 1.0 equiv), H-Gly-OMe•HCl (42 mg, 0.33 mmol, 1.0 equiv) and HATU (190 mg, 0.50 mmol, 1.5 equiv) in DMF (2 mL, 0.2 M). The mixture was stirred for 16h at room temperature before being extracted twice with dichloromethane. The combined organic phases were successively washed with sat. aq. NaHCO₃ (2x20 mL), brine (2x20 mL), dried over Na₂SO₄, filtered and concentrated under reduce pressure. The resulting crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to 97:3) to afford **8** as a white solid (143 mg, 58%). **Mp** 104 – 106 °C; **R_f** = 0.38 (DCM-MeOH, 95:5).

IR (ATR)/ $\bar{\nu}$: 3285, 2970, 1751, 1639, 1493, 1445, 1366, 1206, 1161, 1132, 1036, 1001, 841, 747, 701, 659, 638 cm⁻¹.

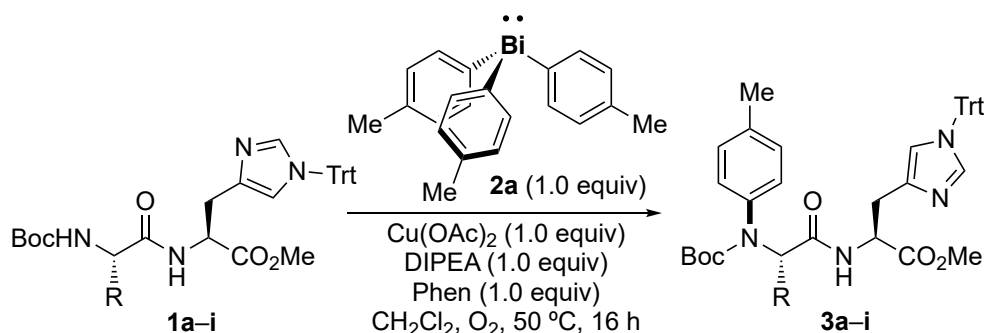
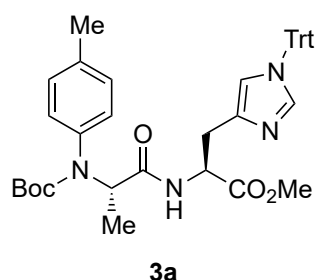
¹H-NMR (600 MHz, CDCl₃) δ 8.71 (d, *J* = 4.9 Hz, 1H), 7.73 (s, 1H), 7.33 – 7.28 (m, 10H), 7.10 – 7.03 (m, 6H), 7.01 (d, *J* = 8.8 Hz, 1H), 6.68 (s, 1H), 5.12 (d, *J* = 4.4 Hz, 1H), 4.56 (q, *J* = 5.5 Hz, 1H), 4.36 (dd, *J* = 8.7, 5.0 Hz, 1H), 4.12 – 4.04 (m, 2H), 3.81 (dd, *J* = 17.7, 5.3 Hz, 1H), 3.63 (s, 3H), 3.13 – 3.07 (m, 1H), 2.95 (dd, *J* = 15.0, 5.9 Hz, 1H), 2.39 – 2.28 (m, 1H), 1.38 – 1.35 (m, 3H), 1.33 (s, 9H), 0.86 (d, *J* = 6.8 Hz, 3H), 0.83 (d, *J* = 6.8 Hz, 3H).

¹³C-NMR (150 MHz, CDCl₃) δ 173.9, 171.8, 171.3, 170.4, 155.8, 142.2, 138.7, 136.6, 129.7, 128.3, 128.2, 120.0, 80.3, 75.5, 58.8, 54.5, 52.1, 51.4, 41.1, 29.6, 28.9, 28.4, 19.4, 17.4.

HRMS-ESI: *m/z* [M + H]⁺ calcd. for C₄₁H₅₀N₆O₇: 739.3814, found 739.3818.

4. Procedure for the coupling between methyl ester N-protected histidine-containing peptides (**1**, **4**, **6** and **8**) and triarylismuthines (**2**)

General method: In a sealed tube, *N,N*-diisopropylethylamine (1.0 equiv) was added to the *N*^T-trityl-histidine-containing peptide (1.0 equiv), the corresponding triarylismuthine (**2**) (1.0 equiv), copper(II) acetate (1.0 equiv) and phenanthroline (1.0 equiv) in CH₂Cl₂ (0.1 M). Oxygen was bubbled in the reaction mixture before stirred in an oil bath set at 50 °C for 16 hours. After cooled down at room temperature, the mixture was directly purified on silica gel chromatography using the corresponding eluent system to afford desired pure product.

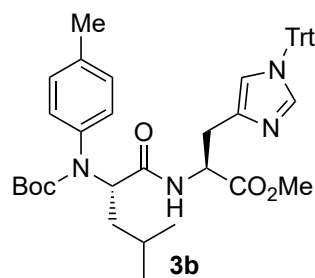
a. Arylated compounds (**3a–i**) from dipeptides (**1a–i**) using tritolylbismuth (**2a**)Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidinate (**3a**)

This compound has been previously reported ¹: **Mp** 64 – 66 °C; **R_f** = 0.47 (DCM–MeOH, 95:5).

IR (ATR)/ $\bar{\nu}$: 3288, 2972, 2928, 1746, 1687, 1514, 1494, 1445, 1366, 1323, 1159, 1132, 748, 701, 660, 637 cm^{-1} .

¹H-NMR (300 MHz, CDCl_3) δ 8.07 (s, 1H), 7.38 – 7.28 (m, 10H), 7.17

(d, J = 8.1 Hz, 2H), 7.12 – 7.00 (m, 8H), 6.58 (s, 1H), 4.85 – 4.75 (m, 1H), 4.67 – 4.57 (m, 1H), 3.60 (s, 3H), 3.12 (dd, J = 14.6, 5.1 Hz, 1H), 3.00 (dd, J = 14.6, 4.8 Hz, 1H), 2.29 (s, 3H), 1.36 (s, 9H), 1.30 (d, J = 7.1 Hz, 3H).

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-leucyl)-*N*^ε-trityl-*L*-histidinate (**3b**)

The general procedure was followed starting from Boc–Leu–His(Trt)–OMe **1b** (65 mg, 0.10 mmol), tritolylbismuth **2a** (50 mg, 0.10 mmol), copper(II) acetate (19 mg, 0.10 mmol), phenanthroline (19 mg, 0.1 mmol) and DIPEA (18 μL , 0.1 mmol) in CH_2Cl_2 (1 mL).

The resulting crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 98:2) to afford **3b** as a white solid (41 mg, 55%). **Mp** 45 – 47 °C; **R_f** = 0.55 (DCM–MeOH, 95:5).

IR (ATR)/ $\bar{\nu}$: 3319, 2953, 1747, 1687, 1513, 1493, 1445, 1366, 1321, 1238, 1163, 1132, 1036, 1002, 826, 747, 701, 660, 638 cm^{-1} .

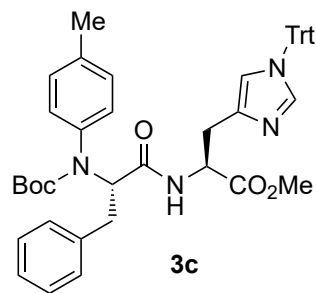
¹H-NMR (600 MHz, CDCl_3) δ 7.96 (s(br), 1H), 7.24 – 7.17 (m, 10H), 7.04 – 6.95 (m, 8H), 6.92 (d, J = 8.1 Hz, 2H), 6.49 (s, 1H), 4.70 (q, J = 5.8 Hz, 1H), 4.53 (s(br), 1H), 3.51 (s, 3H), 3.01 (dd, J

= 14.7, 5.2 Hz, 1H), 2.90 (dd, J = 14.6, 5.0 Hz, 1H), 2.18 (s, 3H), 1.61 – 1.52 (m, 2H), 1.51 – 1.44 (m, 1H), 1.27 (s(br), 9H), 0.74 (d, J = 3.4 Hz, 3H), 0.73 (d, J = 3.5 Hz, 3H).

^{13}C -NMR (150 MHz, CDCl_3) δ 172.1, 171.9, 155.2, 142.4, 138.8, 136.5, 136.2, 129.9, 129.8, 129.2, 128.5, 128.1, 128.1, 119.4, 80.8, 75.4, 53.0, 52.2, 38.1, 30.0, 28.4, 24.9, 23.1, 22.1, 21.1.

HRMS–ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{44}\text{H}_{50}\text{N}_4\text{O}_5$: 715.3854, found 715.3852.

Methyl N^α -(N -(*tert*-butoxycarbonyl)- N -(*p*-tolyl)- L -phenylalanyl)- N^ϵ -trityl- L -histidinate (**3c**)



The general procedure was followed starting from Boc–Phe–His(Trt)–OMe **1c** (62 mg, 94 μmol), tritolylbismuth **2a** (45 mg, 94 μmol), copper(II) acetate (17 mg, 94 μmol), phenanthroline (17 mg, 94 μmol) and DiPEA (16 μL , 94 μmol) in CH_2Cl_2 (1 mL). The resulting crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 98:2) to afford **3c** as a white solid (35 mg, 50%). **Mp** 70 – 72 $^\circ\text{C}$; **R_f** = 0.42 (Hexane–EtOAc, 50:50).

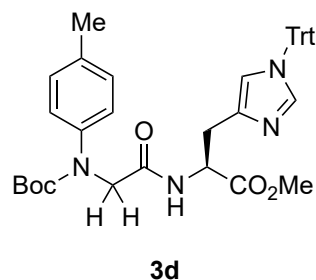
IR (ATR)/ $\bar{\nu}$: 3304, 2912, 1746, 1685, 1513, 1494, 1445, 1366, 1321, 1302, 1239, 1158, 1133, 1034, 828, 747, 700, 660 cm^{-1} .

^1H -NMR (600 MHz, CDCl_3) δ 8.26 (s, 1H), 7.37 – 7.25 (m, 9H), 7.24 – 7.19 (m, 3H), 7.19 – 7.15 (m, 1H), 7.12 (d, J = 7.4 Hz, 2H), 7.10 – 7.02 (m, 6H), 6.81 (d, J = 7.9 Hz, 2H), 6.69 – 6.46 (m, 3H), 4.85 – 4.79 (m, 1H), 4.63 – 4.55 (m, 1H), 3.59 (s, 3H), 3.34 – 3.14 (m, 2H), 3.10 (dd, J = 14.7, 5.1 Hz, 1H), 3.04 – 2.97 (m, 1H), 2.19 (s, 3H), 1.33 (s(br), 9H).

^{13}C -NMR (150 MHz, CDCl_3) δ 171.9, 171.3, 154.9, 142.5, 139.5, 138.8, 138.6, 136.5, 136.0, 129.9, 129.5, 129.0, 128.4, 128.1, 127.6, 126.5, 119.5, 80.9, 75.4, 65.6, 53.1, 52.2, 35.1, 30.0, 28.4, 21.0.

HRMS–ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{47}\text{H}_{48}\text{N}_4\text{O}_5$: 749.3698, found 749.3706.

Methyl N^α -(N -(*tert*-butoxycarbonyl)- N -(*p*-tolyl)glycyl)- N^ϵ -trityl- L -histidinate (**3d**)



The general procedure was followed starting from Boc–Gly–His(Trt)–OMe **1d** (60 mg, 0.11 mmol), tritolylbismuth **2a** (51 mg, 0.11 mmol), copper(II) acetate (19 mg, 0.11 mmol), phenanthroline (19 mg, 0.11 mmol) and DiPEA (18 μL , 0.11 mmol) in CH_2Cl_2 (1 mL). The resulting crude product was purified on silica gel (DCM–MeOH,

gradient from 100:0 to 98:2) to afford **3d** as a white solid (52 mg, 75%). **Mp** 57 – 59 °C; **R_f** = 0.35 (DCM–MeOH, 95:5).

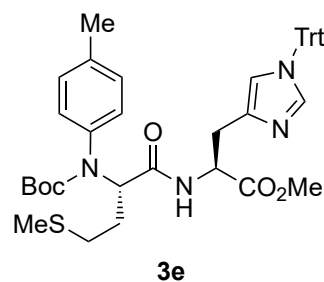
IR (ATR)/ $\bar{\nu}$: 3304, 2912, 1747, 1690, 1514, 1493, 1444, 1366, 1234, 1152, 1034, 1017, 824, 748, 701, 660, 638 cm⁻¹.

¹H-NMR (300 MHz, CDCl₃) δ 8.19 (s, 1H), 7.38 – 7.28 (m, 10H), 7.27 (s, 1H), 7.25 (s, 1H), 7.12 – 7.00 (m, 8H), 6.53 (s, 1H), 4.87 – 4.77 (m, 1H), 4.33 – 4.17 (m, 2H), 3.58 (s, 3H), 3.08 (dd, *J* = 14.6, 4.5 Hz, 1H), 2.95 (dd, *J* = 14.5, 4.4 Hz, 1H), 2.27 (s, 3H), 1.40 (s, 9H).

¹³C-NMR (75 MHz, CDCl₃) δ 171.6, 169.4, 154.7, 142.4, 140.4, 138.8, 136.6, 135.8, 129.9, 129.4, 128.1, 126.3, 119.6, 81.1, 75.4, 54.4, 52.6, 52.1, 29.7, 28.3, 21.1.

HRMS–ESI: *m/z* [*M* + *H*]⁺ calcd. for C₄₀H₄₂N₄O₅: 659.3228, found 659.3232.

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-methionyl)-*N*^ε-trityl-*L*-histidinate (**3e**)



The general procedure was followed starting from Boc–Met–His(Trt)–OMe **1e** (100 mg, 0.16 mmol), tritolylbismuth **2a** (75 mg, 0.16 mmol), copper(II) acetate (28 mg, 0.16 mmol), phenanthroline (28 mg, 0.16 mmol) and DiPEA (27 μ L, 0.16 mmol) in CH₂Cl₂ (1.6 mL). The resulting crude product was purified on

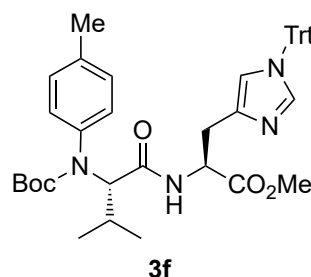
silica gel (Hexane–EtOAc, gradient from 100:0 to 60:40) to afford **3e** as a yellow solid (70 mg, 61%). **Mp** 51 – 53 °C; **R_f** = 0.70 (Hexane–EtOAc, 50:50).

IR (ATR)/ $\bar{\nu}$: 3308, 2919, 1746, 1687, 1513, 1493, 1445, 1366, 1322, 1302, 1158, 1132, 1035, 828, 748, 700, 659, 638 cm⁻¹.

¹H-NMR (600 MHz, CDCl₃) δ 8.14 (s, 1H), 7.34 – 7.27 (m, 10H), 7.20 – 7.13 (m, 2H), 7.11 – 7.06 (m, 6H), 7.02 (d, *J* = 8.1 Hz, 2H), 6.58 (s, 1H), 4.79 (dt, *J* = 7.1, 5.2 Hz, 1H), 4.71 (t, *J* = 6.4 Hz, 1H), 3.60 (s, 3H), 3.11 (dd, *J* = 14.5, 5.5 Hz, 1H), 2.99 (dd, *J* = 14.6, 4.9 Hz, 1H), 2.52 (t, *J* = 7.2 Hz, 2H), 2.28 (s, 3H), 2.19 (s(br), 1H), 1.98 (s, 3H), 1.96 – 1.88 (m, 1H), 1.36 (s(br), 9H).

¹³C-NMR (150 MHz, CDCl₃) δ 171.8, 171.4, 155.0, 142.3, 138.8, 138.0, 136.4, 135.0, 129.8, 129.3, 128.1, 128.0, 119.5, 81.1, 75.4, 61.0, 53.0, 52.1, 31.1, 29.8, 29.2, 28.4, 21.1, 15.3.

HRMS–ESI: *m/z* [*M* + *H*]⁺ calcd. for C₄₃H₄₈N₄O₅S: 733.3418, found 733.3429.

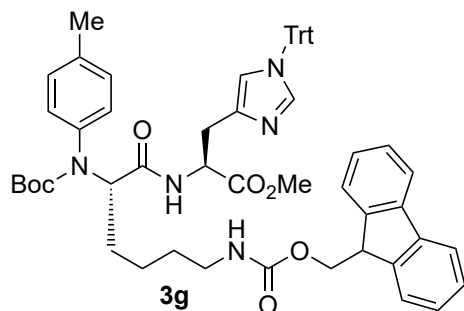
Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-valyl)-*N*^ε-trityl-*L*-histidinate (3f)

The general procedure was followed starting from Boc-Val-His(Trt)-OMe **1f** (60 mg, 98 μmol), tritolylbismuth **2a** (47 mg, 98 μmol), copper(II) acetate (18 mg, 98 μmol), phenanthroline (18 mg, 98 μmol) and DiPEA (17 μL, 98 μmol) in CH₂Cl₂ (1 mL). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 100:0 to 60:40) to afford **3f** as a white solid (9 mg, 13%). **Mp** 67 – 69 °C; **R_f** = 0.48 (Hexane–EtOAc, 30:70).

IR (ATR)/ $\bar{\nu}$: 3298, 2967, 2927, 1747, 1686, 1513, 1493, 1445, 1366, 1325, 1303, 1242, 1167, 1088, 1035, 1001, 870, 827, 748, 701, 660, 638 cm⁻¹.

¹H-NMR (300 MHz, CDCl₃) δ 8.12 (s(br), 1H), 7.38 – 7.28 (m, 10H), 7.17 – 7.10 (m, 6H), 7.10 – 7.01 (m, 4H), 6.63 (s, 1H), 4.83 – 4.75 (m, 1H), 3.86 – 3.70 (m, 1H), 3.64 (s, 3H), 3.62 – 3.57 (m, 1H), 3.11 (dd, *J* = 14.5, 5.7 Hz, 1H), 2.99 (dd, *J* = 15.5, 6.2 Hz, 1H), 2.29 (s, 3H), 1.33 (s, 9H), 1.00 (d, *J* = 6.6 Hz, 3H), 0.85 (d, *J* = 6.7 Hz, 3H).

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₄₃H₄₈N₄O₅: 701.3698, found 701.3700.

Methyl *N*^α-(*N*⁶-(((9*H*-fluoren-9-yl)methoxy)carbonyl)-*N*²-(*tert*-butoxycarbonyl)-*N*²-(*p*-tolyl)-*L*-lysyl)-*N*^ε-trityl-*L*-histidinate (3g)

The general procedure was followed starting from Boc-Lys-His(Trt)-OMe **1g** (50 mg, 58 μmol), tritolylbismuth **2a** (28 mg, 58 μmol), copper(II) acetate (11 mg, 58 μmol), phenanthroline (11 mg, 58 μmol) and DiPEA (10 μL, 58 μmol) in CH₂Cl₂ (0.6 mL). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 100:0 to 60:40) to afford **3g** as an white solid (12 mg, 22%). **Mp** 67 – 69 °C; **R_f** = 0.78 (Hexane–EtOAc, 30:70).

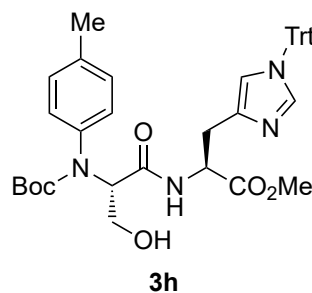
IR (ATR)/ $\bar{\nu}$: 3269, 2924, 2853, 1687, 1513, 1447, 1366, 1325, 1244, 1162, 1133, 1001, 829, 758, 742, 701, 661, 638 cm⁻¹.

¹H-NMR (600 MHz, CDCl₃) δ 8.05 – 8.00 (m, 1H), 7.69 (d, *J* = 7.5 Hz, 2H), 7.51 (d, *J* = 7.6 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 7.27 – 7.21 (m, 12H), 7.04 (s(br), 2H), 7.03 – 6.99 (m, 6H), 6.94 (d, *J* = 8.3 Hz, 2H), 6.51 (s, 1H), 4.84 (s(br), 1H), 4.73 (dt, *J* = 7.4, 5.2 Hz, 1H), 4.61 (s, 2H), 4.45

(s(br), 1H), 4.32 – 4.26 (m, 2H), 4.12 (t, $J = 7.1$ Hz, 1H), 3.52 (s, 3H), 3.08 – 3.00 (m, 3H), 2.93 (dd, $J = 14.4, 5.0$ Hz, 1H), 2.20 (s, 3H), 1.41 – 1.32 (m, 4H), 1.18 (s, 9H).

HRMS–ESI: m/z $[M + H]^+$ calcd. for $C_{59}H_{61}N_5O_7$: 952.4644, found 952.4643.

Methyl N^α -(N -(*tert*-butoxycarbonyl)- N,O -di-*p*-tolyl-*L*-seryl)- N^ϵ -trityl-*L*-histidinate (3h)



The general procedure was followed starting from Boc–Ser–His(Trt)–OMe **1h** (52 mg, 87 μ mol), tritolylbismuth **2a** (42 mg, 87 μ mol), copper(II) acetate (16 mg, 87 μ mol), phenanthroline (16 mg, 87 μ mol) and DiPEA (15 μ L, 87 μ mol) in CH_2Cl_2 (1 mL). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 100:0 to 60:40) to afford **3h** as a white solid (33 mg, 49%). **Mp** 65 – 67 $^\circ$ C; **R_f** = 0.78 (Hexane–EtOAc, 50:50).

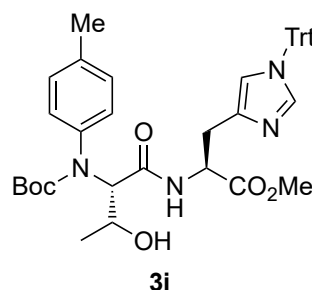
IR (ATR)/ $\bar{\nu}$: 3304, 2919, 1747, 1688, 1510, 1445, 1366, 1324, 1239, 1205, 1156, 1133, 1036, 817, 747, 700, 660, 638 cm^{-1} .

1H -NMR (300 MHz, $CDCl_3$) δ 8.36 (s(br), 1H), 7.38 – 7.26 (m, 12H), 7.13 – 7.05 (m, 6H), 7.02 (d, $J = 7.5$ Hz, 4H), 6.79 (d, $J = 8.5$ Hz, 2H), 6.58 (s, 1H), 4.84 (dt, $J = 7.3, 4.9$ Hz, 2H), 4.59 – 4.44 (m, 1H), 4.26 (t, $J = 9.5$ Hz, 1H), 3.59 (s, 3H), 3.14 (dd, $J = 14.7, 5.1$ Hz, 1H), 3.01 (dd, $J = 14.7, 4.8$ Hz, 1H), 2.28 (s, 3H), 2.26 (s, 3H), 1.36 (s(br), 9H).

^{13}C -NMR (75 MHz, $CDCl_3$) δ 171.7, 169.6, 156.3, 142.4, 138.8, 136.5, 130.3, 129.9, 129.9, 129.4, 128.3, 128.2, 119.5, 114.8, 81.0, 75.4, 66.0, 52.9, 52.2, 29.8, 29.7, 28.4, 21.1, 20.6.

HRMS–ESI: m/z $[M + H]^+$ calcd. for $C_{48}H_{50}N_4O_6$: 779.3803, found 779.3811.

Methyl N^α -(N -(*tert*-butoxycarbonyl)- O -(*p*-tolyl)-*L*-threonyl)- N^ϵ -trityl-*L*-histidinate (3i)



The general procedure was followed starting from Boc–Thr–His(Trt)–OMe **1i** (60 mg, 98 μ mol), tritolylbismuth **2a** (47 mg, 98 μ mol), copper(II) acetate (18 mg, 98 μ mol), phenanthroline (18 mg, 98 μ mol) and DiPEA (17 μ L, 98 μ mol) in CH_2Cl_2 (1 mL). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 100:0 to 60:40) to afford **3i** as a white solid (30 mg, 44%). **Mp** 66 – 68 $^\circ$ C; **R_f** = 0.79 (Hexane–EtOAc, 30:70).

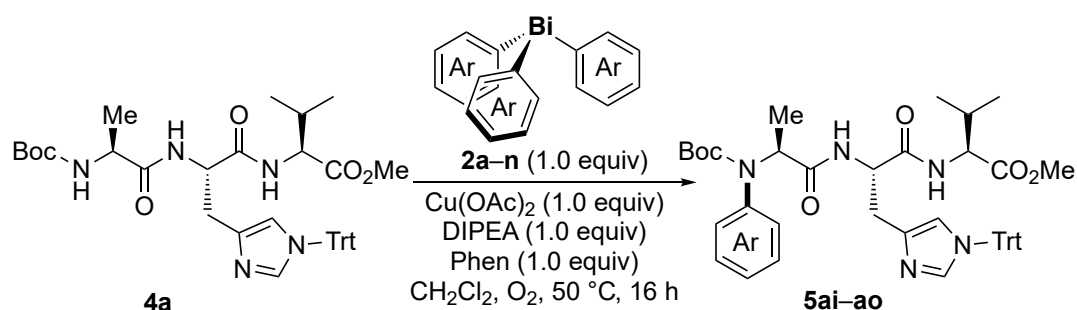
IR (ATR)/ $\bar{\nu}$: 3354, 2973, 2919, 1746, 1715, 1675, 1508, 1492, 1445, 1366, 1229, 1166, 1132, 1034, 749, 702, 661 cm^{-1} .

¹H-NMR (600 MHz, CDCl₃) δ 7.89 (d, *J* = 7.6 Hz, 1H), 7.37 – 7.27 (m, 10H), 7.09 – 7.03 (m, 6H), 6.93 (d, *J* = 8.3 Hz, 2H), 6.83 (d, *J* = 8.2 Hz, 2H), 6.48 (s, 1H), 5.67 (d, *J* = 6.5 Hz, 1H), 4.92 – 4.84 (m, 1H), 4.83 – 4.77 (m, 1H), 4.41 (dd, *J* = 7.3, 2.3 Hz, 1H), 3.61 (s, 3H), 3.08 (dd, *J* = 14.6, 4.7 Hz, 1H), 2.94 (dd, *J* = 14.5, 4.2 Hz, 1H), 2.22 (s, 3H), 1.44 (s, 9H), 1.24 (d, *J* = 6.3 Hz, 3H).

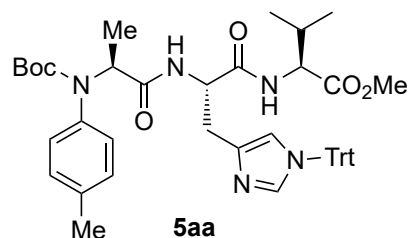
¹³C-NMR (150 MHz, CDCl₃) δ 171.6, 169.4, 155.7, 154.9, 142.4, 138.7, 136.2, 130.9, 130.0, 129.9, 129.9, 128.2, 119.8, 116.9, 80.0, 75.4, 74.0, 57.6, 52.7, 52.2, 30.2, 28.5, 20.6, 15.1.

HRMS-ESI: *m/z* [M + H]⁺ calcd. for C₄₂H₄₆N₄O₆: 703.3490, found 703.3516.

b. Arylated compounds (5aa –an) from tripeptides (4a) using triarylbi-muth (2)



Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (5aa)

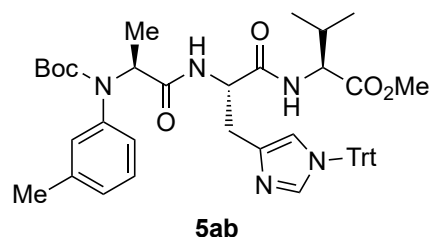


This compound has been previously reported¹: **Mp** 134 – 136 °C; **R_f** = 0.46 (DCM–MeOH, 95:5)

IR (ATR)/ $\bar{\nu}$: 3155, 2970, 1740, 1668, 1514, 1495, 1447, 1367, 1155, 1129, 1019, 1001, 839, 747, 701, 662, 640 cm⁻¹.

¹H-NMR (300 MHz, CDCl₃) δ 8.35 (d, *J* = 7.2 Hz, 1H), 8.08 (s, 1H), 7.74 (d, *J* = 5.9 Hz, 1H), 7.41 – 7.31 (m, 10H), 7.10 (dd, *J* = 5.9, 1.7 Hz, 8H), 7.04 (d, *J* = 8.2 Hz, 1H), 6.87 (s, 1H), 5.27 – 5.11 (m, 1H), 4.50 – 4.29 (m, 2H), 3.67 (s, 3H), 3.54 (d, *J* = 15.1 Hz, 1H), 2.96 – 2.81 (m, 1H), 2.27 (s, 3H), 2.26 – 2.20 (m, 1H), 1.31 (d, *J* = 6.6 Hz, 3H), 1.22 (s, 9H), 1.02 (dd, *J* = 15.7, 6.8 Hz, 6H).

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*m*-tolyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (5ab**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-Val-OMe **4a** (50 mg, 73 μmol), tri-*m*-tolylbismuthane **2b** (35 mg, 73 μmol), copper(II) acetate (13 mg, 73 μmol), phenanthroline (13 mg, 73 μmol) and DiPEA (13 μL, 73 μmol) in CH₂Cl₂ (1 mL). The resulting crude

product was purified on silica gel (DCM-MeOH, gradient from 100:0 to 98:2) to afford **5ab** as a white solid (35 mg, 61%). **Mp** 75 – 77 °C; **R_f** = 0.39 (DCM-MeOH, 95:5).

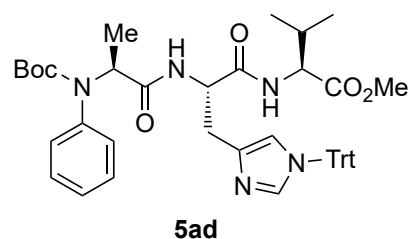
IR (ATR)/ $\bar{\nu}$: 3315, 2972, 2923, 1741, 1674, 1491, 1445, 1367, 1317, 1253, 1202, 1154, 1088, 1036, 1001, 841, 748, 701, 660, 637 cm⁻¹.

¹H-NMR (600 MHz, CDCl₃) δ 8.64 (d, *J* = 4.6 Hz, 1H), 7.71 (d, *J* = 7.9 Hz, 1H), 7.34 – 7.28 (m, 10H), 7.19 – 7.14 (m, 2H), 7.12 – 7.07 (m, 7H), 7.02 (d, *J* = 7.5 Hz, 1H), 6.70 (s, 1H), 4.88 – 4.81 (m, 1H), 4.54 – 4.47 (m, 1H), 4.41 (dd, *J* = 7.9, 5.6 Hz, 1H), 3.64 (s, 3H), 3.25 – 3.17 (m, 1H), 2.92 (dd, *J* = 14.5, 5.4 Hz, 1H), 2.27 (s, 3H), 2.16 – 2.09 (m, 1H), 1.41 (d, *J* = 6.9 Hz, 3H), 1.34 (s, 9H), 0.83 (d, *J* = 6.8 Hz, 6H).

¹³C-NMR (150 MHz, CDCl₃) δ 172.1, 172.1, 171.4, 154.7, 142.4, 141.3, 138.6, 138.2, 137.2, 129.9, 129.0, 128.6, 128.2, 128.2, 127.6, 125.2, 119.6, 80.9, 75.5, 57.7, 53.3, 52.0, 31.0, 29.6, 28.4, 21.4, 19.2, 18.1, 15.7.

HRMS-ESI: *m/z* [M + H]⁺ calcd. for C₄₆H₅₃N₅O₆: 772.4069, found 772.4063.

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-phenyl-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (5ad**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-Val-OMe **4a** (70 mg, 0.10 mmol), triphenylbismuthane **2d** (45 mg, 0.10 mmol), copper(II) acetate (19 mg, 0.10 mmol), phenanthroline (18 mg, 0.10 mmol) and DiPEA (18 μL, 0.10 mmol) in CH₂Cl₂ (1 mL). The

resulting crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to 98:2) to afford **5ad** as a white solid (38 mg, 49%). **Mp** 76 – 78 °C; **R_f** = 0.50 (DCM-MeOH, 95:5).

IR (ATR)/ $\bar{\nu}$: 3315, 2972, 1741, 1674, 1494, 1446, 1368, 1310, 1256, 1156, 1002, 838, 749, 701, 660, 637 cm⁻¹.

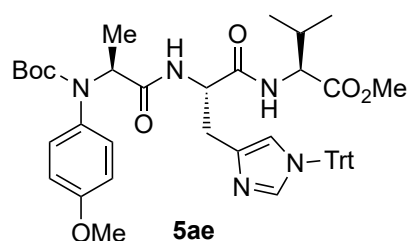
¹H-NMR (300 MHz, CDCl₃) δ 8.56 (d, *J* = 6.6 Hz, 1H), 7.67 (d, *J* = 8.3 Hz, 1H), 7.35 – 7.27 (m, 14H), 7.25 – 7.18 (m, 1H), 7.12 – 7.05 (m, 6H), 6.71 (s, 1H), 4.81 (q, *J* = 5.6 Hz, 1H), 4.56 (q, *J* =

7.1 Hz, 1H), 4.39 (dd, $J = 8.3, 5.4$ Hz, 1H), 3.64 (s, 3H), 3.22 – 3.17 (m, 1H), 2.93 (dd, $J = 14.5, 6.0$ Hz, 1H), 2.18 – 2.06 (m, 1H), 1.36 (d, $J = 7.1$ Hz, 3H), 1.32 (s, 9H), 0.84 (d, $J = 6.8$ Hz, 6H).

Presence of conformers: $^{13}\text{C-NMR}$ (150 MHz, CDCl_3) δ 172.2, 172.1, 171.4, 154.9, 142.3, 141.0, 138.2, 136.9, 129.9, 128.8, 128.5, 128.2, 128.2, 127.0, 119.8, 81.0, 75.6, 57.8, 55.5, 53.4, 52.1, 47.3, 43.5, 30.9, 29.6, 28.4, 19.8, 18.1.

HRMS–ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{45}\text{H}_{51}\text{N}_5\text{O}_6$: 758.3912, found 758.3909.

Methyl N^α -(N -(tert-butoxycarbonyl)- N -(4-methoxyphenyl)- L -alanyl)- N^ϵ -trityl- L -histidyl- L -valinate (5ae**)**



The general procedure was followed starting from Boc–Ala–His(Trt)–Val–OMe **4a** (60 mg, 88 μmol), tris(4-methoxyphenyl)bismuthane **2e** (47 mg, 88 μmol), copper(II) acetate (16 mg, 88 μmol), phenanthroline (16 mg, 88 μmol) and DiPEA (15 μL , 88 μmol) in CH_2Cl_2 (1 mL). The resulting

crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 98:2) to afford **5ae** as a white solid (35 mg, 50%). **Mp** 89 – 91 $^\circ\text{C}$; **R_f** = 0.46 (Hexane–EtOAc, 20:80).

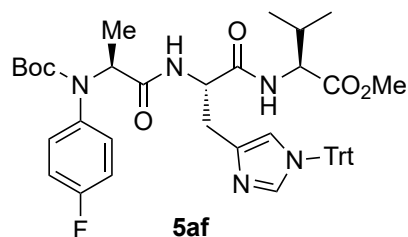
IR (ATR)/ $\bar{\nu}$: 3315, 2972, 2928, 1741, 1673, 1511, 1445, 1366, 1321, 1294, 1244, 1205, 1155, 1033, 1001, 839, 748, 701, 660 cm^{-1} .

$^1\text{H-NMR}$ (600 MHz, CDCl_3) δ 8.49 (d, $J = 6.1$ Hz, 1H), 7.68 (d, $J = 7.2$ Hz, 1H), 7.35 – 7.27 (m, 10H), 7.23 – 7.14 (m, 2H), 7.13 – 7.05 (m, 6H), 6.79 (d, $J = 8.8$ Hz, 2H), 6.70 (s, 1H), 4.85 – 4.77 (m, 1H), 4.58 (s(br), 1H), 4.40 (dd, $J = 8.2, 5.5$ Hz, 1H), 3.76 (s, 3H), 3.64 (s, 3H), 3.18 (dd, $J = 14.9, 4.3$ Hz, 1H), 2.92 (dd, $J = 14.9, 6.4$ Hz, 1H), 2.15 – 2.09 (m, 1H), 1.31 (s(br), 12H), 0.84 (d, $J = 6.8$ Hz, 6H).

$^{13}\text{C-NMR}$ (150 MHz, CDCl_3) δ 172.4, 172.1, 171.4, 158.3, 155.3, 142.4, 138.3, 137.1, 129.9, 129.8, 128.2, 128.2, 119.7, 113.9, 80.8, 75.5, 57.8, 55.5, 53.4, 52.1, 30.9, 29.6, 28.4, 19.2, 18.1, 15.8, 14.3.

HRMS–ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{46}\text{H}_{53}\text{N}_5\text{O}_7$: 788.4018, found 788.4020.

Methyl N^{α} -(*N*-(tert-butoxycarbonyl)-*N*-(4-fluorophenyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (5af)



The general procedure was followed starting from Boc-Ala-His(Trt)-Val-OMe **4a** (50 mg, 73 μmol), tris(4-fluorophenyl)bismuthane **2f** (36 mg, 73 μmol), copper(II) acetate (13 mg, 73 μmol), phenanthroline (13 mg, 73 μmol) and DiPEA (13 μL, 73 μmol) in CH₂Cl₂ (1 mL). The resulting

crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to 98:2) to afford **5af** as a white solid (33 mg, 58%). **Mp** 81 – 83 °C; **R_f** = 0.65 (Hexane-EtOAc, 30:70).

IR (ATR)/ $\bar{\nu}$: 3301, 2959, 2914, 1741, 1682, 1508, 1445, 1367, 1318, 1215, 1152, 1036, 1001, 843, 748, 701, 660 cm⁻¹.

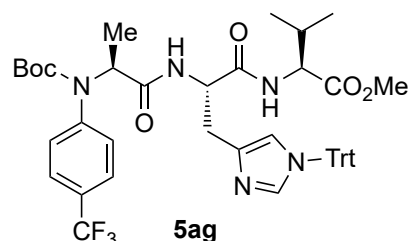
¹H-NMR (600 MHz, CDCl₃) δ 8.47 (s, 1H), 7.69 (d, *J* = 6.7 Hz, 1H), 7.37 – 7.23 (m, 12H), 7.16 – 7.04 (m, 6H), 6.96 (t, *J* = 8.6 Hz, 2H), 6.71 (s, 1H), 4.81 (q, *J* = 5.7 Hz, 1H), 4.64 – 4.53 (m, 1H), 4.41 (dd, *J* = 7.9, 5.5 Hz, 1H), 3.65 (s, 3H), 3.18 (dd, *J* = 14.8, 3.2 Hz, 1H), 2.93 (dd, *J* = 14.4, 5.5 Hz, 1H), 2.17 – 2.08 (m, 1H), 1.32 (s(br), 12H), 0.84 (d, *J* = 6.8 Hz, 6H).

¹³C-NMR (150 MHz, CDCl₃) δ 172.1, 171.29, 161.41 (d, *J* = 244.7 Hz), 154.89, 142.33, 138.18, 136.94, 136.70, 130.52 (d, *J* = 8.4 Hz), 129.87, 128.25, 128.20, 119.78, 115.5 (d, *J* = 22.4 Hz), 81.11, 75.62, 57.77, 53.39, 52.05, 30.94, 29.61, 28.34, 19.16, 18.09, 15.89.

¹⁹F NMR (564 MHz, CDCl₃) δ -115.24.

HRMS-ESI: *m/z* [M + H]⁺ calcd. for C₄₅H₅₀FN₅O₆: 776.3818, found 776.3818.

Methyl N^{α} -(*N*-(tert-butoxycarbonyl)-*N*-(4-(trifluoromethyl)phenyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (5ag)



The general procedure was followed starting from Boc-Ala-His(Trt)-Val-OMe **4a** (43 mg, 63 μmol), tris(4-(trifluoromethyl)phenyl)bismuthane **2g** (41 mg, 63 μmol), copper(II) acetate (12 mg, 63 μmol), phenanthroline (11 mg, 63 μmol) and DiPEA (11 μL, 63 μmol) in CH₂Cl₂ (700 μL). The

resulting crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to 98:2) to afford **5ag** as a white solid (11 mg, 21%). **Mp** 82 – 84 °C; **R_f** = 0.46 (DCM-MeOH, 95:5).

IR (ATR)/ $\bar{\nu}$: 3315, 2963, 2928, 1743, 1689, 1519, 1494, 1446, 1369, 1324, 1254, 1162, 1123, 1068, 1018, 1002, 854, 748, 701, 660 cm⁻¹.

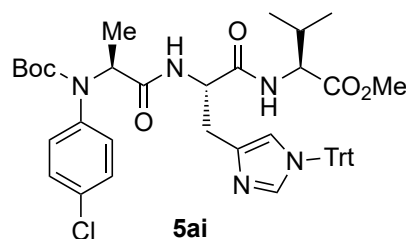
¹H-NMR (600 MHz, CDCl₃) δ 8.52 (d, *J* = 5.2 Hz, 1H), 7.70 (d, *J* = 7.5 Hz, 1H), 7.54 (d, *J* = 8.4 Hz, 2H), 7.45 (d, *J* = 8.3 Hz, 2H), 7.37 – 7.29 (m, 10H), 7.12 – 7.06 (m, 6H), 6.71 (s, 1H), 4.83 (q, *J* = 5.6 Hz, 1H), 4.62 (q, *J* = 7.1 Hz, 1H), 4.41 (dd, *J* = 8.3, 5.3 Hz, 1H), 3.65 (s, 3H), 3.17 (dd, *J* = 15.0, 4.4 Hz, 1H), 2.98 – 2.89 (m, 1H), 2.16 – 2.09 (m, 1H), 1.39 (d, *J* = 7.1 Hz, 3H), 1.34 (s, 9H), 0.84 (d, *J* = 6.8 Hz, 6H).

¹³C-NMR (150 MHz, CDCl₃) δ 172.1, 171.7, 171.2, 154.2, 144.3, 142.3, 138.1, 129.9, 128.8, 128.3, 128.2, 125.9 (q, *J* = 3.5 Hz), 124.1 (d, *J* = 270.6 Hz), 119.8, 81.7, 58.2, 57.8, 53.5, 52.1, 31.0, 29.7, 28.3, 19.2, 18.1, 15.9.

¹⁹F NMR (282 MHz, CDCl₃) δ -62.40.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₄₆H₅₀F₃N₅O₆: 826.3786, found 826.3797.

Methyl *N*^α-(*N*-(tert-butoxycarbonyl)-*N*-(4-chlorophenyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (5ai)



The general procedure was followed starting from Boc-Ala-His(Trt)-Val-OMe **4a** (50 mg, 73 μmol), tris(4-chlorophenyl)bismuthane **2i** (40 mg, 73 μmol), copper(II) acetate (13 mg, 73 μmol), phenanthroline (13 mg, 73 μmol) and DiPEA (13 μL, 73 μmol) in CH₂Cl₂ (740 μL). The resulting

crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 98:2) to afford **5ai** as a white solid (27 mg, 47%). **Mp** 74 – 76 °C; **R_f** = 0.68 (Hexane–EtOAc, 30:70).

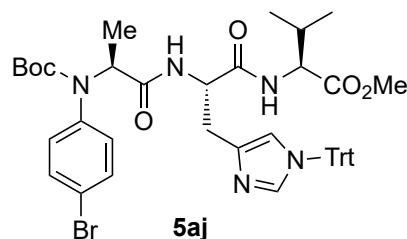
IR (ATR)/ $\bar{\nu}$: 3310, 2959, 2928, 1741, 1683, 1493, 1446, 1367, 1317, 1255, 1156, 1091, 1015, 840, 748, 701, 660 cm⁻¹.

¹H-NMR (600 MHz, CDCl₃) δ 8.46 (d, *J* = 5.5 Hz, 1H), 7.68 (d, *J* = 8.2 Hz, 1H), 7.42 – 7.27 (m, 10H), 7.25 (s, 4H), 7.10 – 7.04 (m, 6H), 6.71 (s, 1H), 4.81 (q, *J* = 5.9 Hz, 1H), 4.59 (q, *J* = 6.7 Hz, 1H), 4.41 (dd, *J* = 8.3, 5.3 Hz, 1H), 3.65 (s, 3H), 3.17 (dd, *J* = 15.0, 4.4 Hz, 1H), 2.94 (dd, *J* = 14.9, 5.9 Hz, 1H), 2.17 – 2.09 (m, 1H), 1.34 (d, *J* = 7.8 Hz, 3H), 1.33 (s, 9H), 0.84 (d, *J* = 6.8 Hz, 6H).

¹³C-NMR (150 MHz, CDCl₃) δ 172.1, 172.0, 171.2, 154.6, 142.3, 139.4, 138.1, 136.8, 132.6, 130.1, 129.9, 128.9, 128.3, 128.2, 119.8, 81.3, 75.7, 57.8, 53.4, 52.1, 30.9, 29.5, 28.3, 19.2, 18.1, 15.9.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₄₅H₅₀ClN₅O₆: 792.3522, found 792.3516.

Methyl *N*^α-(*N*-(tert-butoxycarbonyl)-*N*-(4-bromophenyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (5aj**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-Val-OMe **4a** (50 mg, 73 μmol), tris(4-bromophenyl)bismuthane **2j** (50 mg, 73 μmol), copper(II) acetate (13 mg, 73 μmol), phenanthroline (13 mg, 73 μmol) and DiPEA (13 μL, 73 μmol) in CH₂Cl₂ (740 μL). The resulting

crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to 98:2) to afford **5aj** as a white solid (29 mg, 48%). **Mp** 85 – 87 °C; **R_f** = 0.44 (DCM-MeOH, 95:5).

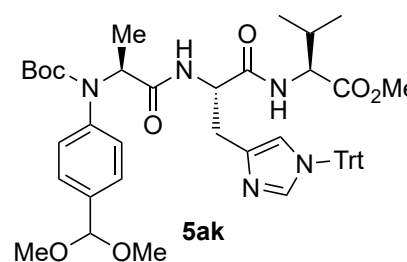
IR (ATR)/ $\bar{\nu}$: 3319, 2963, 2919, 1742, 1683, 1489, 1445, 1367, 1318, 1254, 1156, 1070, 1012, 837, 748, 701, 661 cm⁻¹.

¹H-NMR (600 MHz, CDCl₃) δ 8.45 (d, *J* = 5.7 Hz, 1H), 7.64 (d, *J* = 8.0 Hz, 1H), 7.33 (d, *J* = 8.5 Hz, 2H), 7.28 – 7.22 (m, 10H), 7.12 (d, *J* = 8.3 Hz, 2H), 7.06 – 6.98 (m, 6H), 6.63 (s, 1H), 4.78 – 4.72 (m, 1H), 4.52 (q, *J* = 6.8 Hz, 1H), 4.34 (dd, *J* = 8.2, 5.4 Hz, 1H), 3.58 (s, 3H), 3.10 (dd, *J* = 14.8, 3.5 Hz, 1H), 2.91 – 2.80 (m, 1H), 2.14 – 1.99 (m, 1H), 1.26 (s, 12H), 0.77 (d, *J* = 6.8 Hz, 6H).

¹³C-NMR (150 MHz, CDCl₃) δ 172.1, 171.9, 171.3, 154.5, 142.3, 140.0, 138.2, 137.1, 131.9, 130.4, 129.9, 128.2, 128.2, 120.6, 119.7, 81.3, 75.6, 57.7, 53.4, 52.1, 30.9, 29.6, 28.3, 19.2, 18.1, 15.8.

HRMS-ESI: *m/z* [M + H]⁺ calcd. for C₄₅H₅₀BrN₅O₆: 836.3018, found 836.3026.

Methyl *N*^α-(*N*-(tert-butoxycarbonyl)-*N*-(4-(dimethoxymethyl)phenyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (5ak**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-Val-OMe **4a** (50 mg, 73 μmol), tris(4-(dimethoxymethyl)phenyl)bismuthane **2k** (49 mg, 73 μmol), copper(II) acetate (13 mg, 73 μmol), phenanthroline (13 mg, 73 μmol) and DiPEA (13 μL, 73 μmol) in CH₂Cl₂ (740 μL). The

resulting crude product was purified on basified silica gel (DCM-MeOH, gradient from 100:0 to 98:2) to afford **5ak** as a white solid (39 mg, 64%). **Mp** 64 – 66 °C; **R_f** = 0.29 (DCM-MeOH, 95:5).

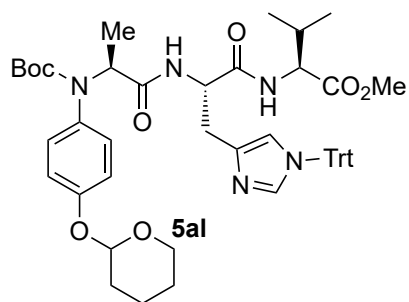
IR (ATR)/ $\bar{\nu}$: 3315, 2924, 2852, 1740, 1682, 1512, 1494, 1445, 1366, 1319, 1254, 1156, 1099, 1052, 1020, 984, 828, 748, 701, 660 cm⁻¹.

¹H-NMR (300 MHz, CDCl₃) δ 8.58 (d, *J* = 7.0 Hz, 1H), 7.76 (d, *J* = 8.5 Hz, 1H), 7.37 (d, *J* = 8.4 Hz, 2H), 7.34 – 7.27 (m, 12H), 7.15 – 7.02 (m, 6H), 6.69 (s, 1H), 5.37 (s, 1H), 4.82 (dt, *J* = 6.6, 4.4 Hz, 1H), 4.56 (q, *J* = 7.1 Hz, 1H), 4.41 (dd, *J* = 8.4, 5.4 Hz, 1H), 3.64 (s, 3H), 3.29 (s, 6H), 3.19 (dd, *J* = 15.0, 4.2 Hz, 1H), 2.90 (dd, *J* = 14.9, 6.2 Hz, 1H), 2.23 – 2.04 (m, 1H), 1.37 (d, *J* = 9.1 Hz, 3H), 1.32 (s, 9H), 0.83 (d, *J* = 6.8 Hz, 6H).

¹³C-NMR (75 MHz, CDCl₃) δ 172.1, 172.0, 171.4, 154.7, 142.4, 141.2, 138.2, 137.2, 136.6, 129.9, 128.2, 128.2, 127.2, 119.6, 102.8, 81.1, 75.5, 57.8, 53.4, 52.7, 52.0, 45.9, 30.9, 29.8, 28.3, 19.2, 18.1, 15.7.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₄₈H₅₇N₅O₈: 832.4280, found 832.4291.

Methyl *N*^α-(*N*-(tert-butoxycarbonyl)-*N*-(4-((tetrahydro-2*H*-pyran-2-yl)oxy)phenyl))-L-alanyl)-*N*^γ-trityl-L-histidyl-L-valinate (5al**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-Val-OMe **4a** (50 mg, 73 μmol), tris(4-((tetrahydro-2*H*-pyran-2-yl)oxy)phenyl)bismuthane **2l** (54 mg, 73 μmol), copper(II) acetate (13 mg, 73 μmol), phenanthroline (13 mg, 73 μmol) and DiPEA (13 μL, 73 μmol) in CH₂Cl₂ (740 μL). The resulting crude product was purified

on basified silica gel (DCM–MeOH, gradient from 100:0 to 98:2) to afford **5al** as a white solid (33 mg, 52%). **Mp** 69 – 71 °C; **R_f** = 0.31 (DCM–MeOH, 95:5).

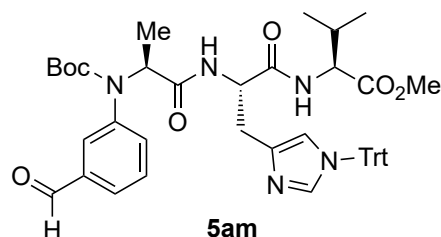
IR (ATR)/ $\bar{\nu}$: 3310, 2929, 1740, 1681, 1509, 1445, 1367, 1321, 1233, 1202, 1156, 1112, 1075, 1035, 960, 920, 841, 748, 701, 660 cm⁻¹.

¹H-NMR (600 MHz, CDCl₃) δ 8.55 (d, *J* = 6.7 Hz, 1H), 7.79 – 7.71 (m, 1H), 7.35 – 7.28 (m, 10H), 7.22 – 7.13 (m, 2H), 7.12 – 7.05 (m, 6H), 6.94 (d, *J* = 8.8 Hz, 2H), 6.69 (s, 1H), 5.34 (q, *J* = 2.9 Hz, 1H), 4.84 – 4.77 (m, 1H), 4.57 (s(br), 1H), 4.41 (dd, *J* = 8.2, 5.5 Hz, 1H), 3.90 – 3.84 (m, 1H), 3.64 (s, 3H), 3.59 – 3.54 (m, 1H), 3.19 (dd, *J* = 15.0, 4.2 Hz, 1H), 2.90 (dd, *J* = 15.0, 6.5 Hz, 1H), 2.85 (q, *J* = 7.1 Hz, 1H), 2.16 – 2.09 (m, 1H), 2.02 – 1.95 (m, 1H), 1.89 – 1.79 (m, 2H), 1.70 – 1.62 (m, 1H), 1.60 – 1.55 (m, 1H), 1.32 (s(br), 12H), 0.83 (d, *J* = 6.8 Hz, 6H).

¹³C-NMR (150 MHz, CDCl₃) δ 172.2, 172.2, 172.1, 171.4, 171.4, 155.9, 142.5, 138.3, 137.3, 129.9, 129.6, 128.1, 119.6, 116.5, 96.6, 80.8, 75.4, 62.2, 57.8, 53.4, 52.0, 45.9, 31.0, 30.5, 29.8, 29.6, 28.4, 25.3, 19.2, 18.9, 18.1, 15.7.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₅₀H₅₉N₅O₈: 858.4436, found 858.4443.

Methyl *N*^α-(*N*-(tert-butoxycarbonyl)-*N*-(3-formylphenyl))-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (5am**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-Val-OMe **4a** (50 mg, 73 μmol), 3,3',3''-bismuthanetriyltribenzaldehyde **2m** (38 mg, 73 μmol), copper(II) acetate (13 mg, 73 μmol), phenanthroline (13 mg, 73 μmol) and DiPEA (13 μL, 73 μmol) in CH₂Cl₂ (740

μL). The resulting crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to 98:2) to afford **5am** as a yellow solid (27 mg, 47%). **Mp** 63 – 65 °C; **R_f** = 0.26 (DCM-MeOH, 95:5).

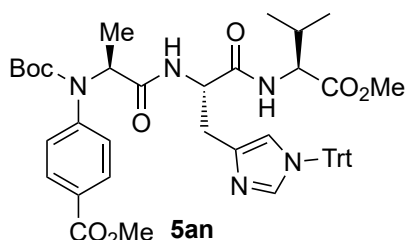
IR (ATR)/ $\bar{\nu}$: 3301, 2968, 2923, 1740, 1690, 1519, 1490, 1445, 1368, 1315, 1254, 1151, 1035, 1001, 839, 748, 700, 660 cm⁻¹.

¹H-NMR (600 MHz, CDCl₃) δ 9.95 (s, 1H), 8.69 – 8.55 (m, 1H), 7.89 (s, 1H), 7.75 (d, *J* = 7.6 Hz, 1H), 7.63 (d, *J* = 7.9 Hz, 1H), 7.59 (d, *J* = 8.0 Hz, 1H), 7.47 (t, *J* = 7.8 Hz, 1H), 7.35 – 7.29 (m, 10H), 7.12 – 7.05 (m, 6H), 6.72 (s, 1H), 4.83 (q, *J* = 5.5 Hz, 1H), 4.61 (q, *J* = 6.9 Hz, 1H), 4.40 (dd, *J* = 8.2, 5.4 Hz, 1H), 3.63 (s, 3H), 3.21 (dd, *J* = 14.9, 3.5 Hz, 1H), 2.94 (dd, *J* = 14.8, 5.8 Hz, 1H), 2.17 – 2.08 (m, 1H), 1.40 (d, *J* = 7.2 Hz, 3H), 1.33 (s, 9H), 0.82 (d, *J* = 6.7 Hz, 6H).

¹³C-NMR (150 MHz, CDCl₃) δ 191.8, 172.1, 171.9, 171.2, 154.4, 142.3, 138.2, 137.2, 134.7, 130.0, 129.9, 129.5, 128.3, 128.2, 128.0, 127.9, 127.3, 119.9, 81.6, 75.7, 57.7, 55.6, 53.4, 52.1, 43.6, 30.9, 28.3, 19.2, 18.1, 16.1.

HRMS-ESI: *m/z* [M + H]⁺ calcd. for C₄₆H₅₁N₅O₇: 786.3861, found 786.3878.

Methyl (6*S*,9*S*,12*S*)-12-isopropyl-5-(4-(methoxycarbonyl)phenyl)-2,2,6-trimethyl-4,7,10-trioxo-9-((1-trityl-1*H*-imidazol-4-yl)methyl)-3-oxa-5,8,11-triazatridecan-13-oate (5an**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-Val-OMe **4a** (50 mg, 73 μmol), trimethyl 4,4',4''-bismuthanetriyltribenzoate **2n** (45 mg, 73 μmol), copper(II) acetate (13 mg, 73 μmol), phenanthroline (13 mg, 73 μmol) and DiPEA (13 μL, 73 μmol) in CH₂Cl₂ (740 μL). The resulting

crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to 98:2) to afford **5an** as a white solid (22 mg, 37%). **Mp** 84 – 86 °C; **R_f** = 0.24 (DCM-MeOH, 95:5).

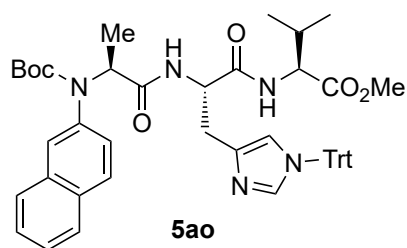
IR (ATR)/ $\bar{\nu}$: 3199, 2963, 1683, 1511, 1493, 1445, 1392, 1369, 1321, 1277, 1156, 1113, 1019, 836, 749, 701, 660 cm^{-1} .

$^1\text{H-NMR}$ (600 MHz, CDCl_3) δ 8.57 (s, 1H), 7.97 (d, J = 8.5 Hz, 2H), 7.61 (d, J = 7.6 Hz, 1H), 7.39 (d, J = 8.4 Hz, 2H), 7.35 – 7.29 (m, 10H), 7.14 – 7.02 (m, 6H), 6.71 (s, 1H), 4.85 – 4.77 (m, 1H), 4.60 (q, J = 7.1 Hz, 1H), 4.39 (dd, J = 8.0, 5.5 Hz, 1H), 3.90 (s, 3H), 3.64 (s, 3H), 3.18 (q, J = 7.4 Hz, 1H), 2.92 (dd, J = 13.7, 4.4 Hz, 1H), 2.16 – 2.07 (m, 1H), 1.39 (d, J = 7.1 Hz, 3H), 1.33 (s, 9H), 0.83 (d, J = 6.7 Hz, 6H).

Presence of conformers: **$^{13}\text{C-NMR}$ (150 MHz, CDCl_3)** δ 172.1, 171.9, 166.7, 154.2, 145.4, 142.3, 138.1, 130.2, 129.9, 128.4, 128.3, 128.2, 128.1, 128.0, 127.4, 119.9, 81.6, 75.7, 58.4, 57.8, 55.9, 53.4, 52.3, 52.1, 43.8, 30.9, 28.3, 19.8, 18.1, 15.8, 12.7.

HRMS–ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{47}\text{H}_{53}\text{N}_5\text{O}_8$: 816.3967, found 816.3978.

Methyl N^α -(N -(*tert*-butoxycarbonyl)- N -(naphthalen-2-yl)- L -alanyl)- N^ϵ -trityl- L -histidyl- L -valinate (5ao**)**



The general procedure was followed starting from Boc–Ala–His(Trt)–Val–OMe **4a** (50 mg, 73 μmol), tri(naphthalen-2-yl)bismuthane **2o** (43 mg, 73 μmol), copper(II) acetate (13 mg, 73 μmol), phenanthroline (13 mg, 73 μmol) and DiPEA (13 μL , 73 μmol) in CH_2Cl_2 (740 μL). The resulting crude

product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 98:2) to afford **5ao** as a white solid (30 mg, 51%). **Mp** 67 – 69 $^\circ\text{C}$; **R_f** = 0.47 (DCM–MeOH, 95:5).

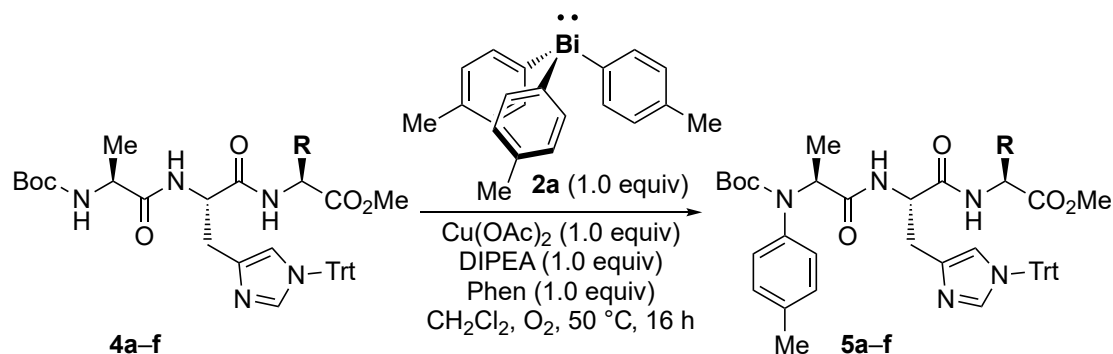
IR (ATR)/ $\bar{\nu}$: 3310, 2963, 2928, 1741, 1682, 1493, 1445, 1391, 1367, 1312, 1254, 1202, 1155, 1129, 1036, 1001, 858, 818, 747, 700, 660 cm^{-1} .

$^1\text{H-NMR}$ (600 MHz, CDCl_3) δ 8.71 (d, J = 5.7 Hz, 1H), 7.84 – 7.78 (m, 2H), 7.75 (d, J = 8.7 Hz, 1H), 7.72 (d, J = 7.8 Hz, 2H), 7.47 – 7.39 (m, 3H), 7.35 – 7.27 (m, 10H), 7.12 – 7.05 (m, 6H), 6.71 (s, 1H), 4.88 (q, J = 5.4 Hz, 1H), 4.67 (q, J = 6.8 Hz, 1H), 4.42 (dd, J = 8.3, 5.4 Hz, 1H), 3.64 (s, 3H), 3.23 (dd, J = 14.9, 3.5 Hz, 1H), 2.96 (dd, J = 14.6, 5.4 Hz, 1H), 2.17 – 2.09 (m, 1H), 1.43 (d, J = 7.0 Hz, 3H), 1.34 (s, 9H), 0.84 (d, J = 4.3 Hz, 3H), 0.83 (d, J = 4.1 Hz, 3H).

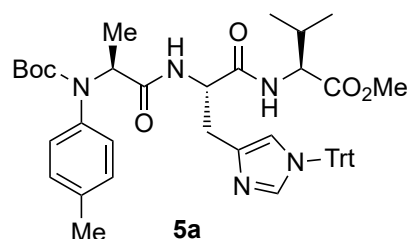
$^{13}\text{C-NMR}$ (150 MHz, CDCl_3) δ 172.1, 172.1, 171.4, 154.8, 153.2, 142.4, 138.7, 138.2, 133.6, 132.1, 129.9, 128.4, 128.2, 128.2, 128.1, 127.6, 127.3, 126.3, 126.2, 126.1, 119.7, 81.1, 75.5, 58.7, 57.8, 53.4, 52.0, 31.0, 29.6, 28.4, 19.2, 18.1, 15.9.

HRMS–ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{49}\text{H}_{53}\text{N}_5\text{O}_6$: 808.4069, found 808.4071.

c. Arylated compounds (**5a–f**) from tripeptides (**4a–f**) using tritolylbismuth (**2a**)



Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (5a**)**

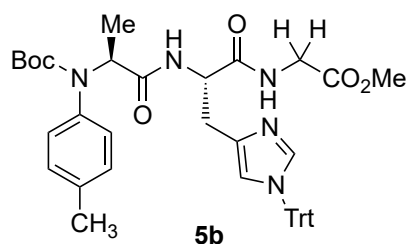


This compound has been previously reported¹: **Mp** 134 – 136 °C; R_f = 0.46 (DCM–MeOH, 95:5)

IR (ATR)/ $\bar{\nu}$: 3155, 2970, 1740, 1668, 1514, 1495, 1447, 1367, 1155, 1129, 1019, 1001, 839, 747, 701, 662, 640 cm^{-1} .

¹H-NMR (300 MHz, CDCl_3) δ 8.35 (d, J = 7.2 Hz, 1H), 8.08 (s, 1H), 7.74 (d, J = 5.9 Hz, 1H), 7.41 – 7.31 (m, 10H), 7.10 (dd, J = 5.9, 1.7 Hz, 8H), 7.04 (d, J = 8.2 Hz, 1H), 6.87 (s, 1H), 5.27 – 5.11 (m, 1H), 4.50 – 4.29 (m, 2H), 3.67 (s, 3H), 3.54 (d, J = 15.1 Hz, 1H), 2.96 – 2.81 (m, 1H), 2.27 (s, 3H), 2.26 – 2.20 (m, 1H), 1.31 (d, J = 6.6 Hz, 3H), 1.22 (s, 9H), 1.02 (dd, J = 15.7, 6.8 Hz, 6H).

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidylglycinate (5b**)**



The general procedure was followed starting from Boc-Ala-His(Trt)-Gly-OMe **4b** (70 mg, 0.11 mmol), tri-*p*-tolylbismuthane **2a** (53 mg, 0.11 mmol), copper(II) acetate (20 mg, 0.11 mmol), phenanthroline (20 mg, 0.11 mmol) and DIPEA (19 μL , 0.11 mmol) in CH_2Cl_2 (1 mL). The resulting

crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 98:2) to afford **5b** as a yellowish solid (59 mg, 74%). **Mp** 75 – 77 °C; R_f = 0.26 (DCM–MeOH, 95:5).

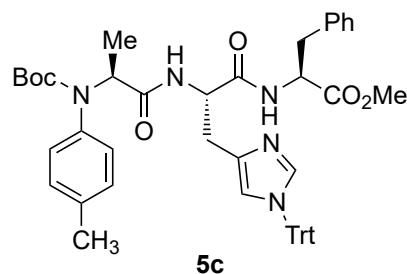
IR (ATR)/ $\bar{\nu}$: 3323, 2977, 2919, 1754, 1668, 1513, 1494, 1445, 1366, 1322, 1237, 1201, 1132, 1036, 1001, 851, 829, 747, 700, 660 cm^{-1} .

¹H-NMR (600 MHz, CDCl₃) δ 8.72 (s, 1H), 7.83 (s, 1H), 7.38 – 7.28 (m, 10H), 7.20 (d, *J* = 8.0 Hz, 2H), 7.14 – 7.07 (m, 8H), 6.70 (s, 1H), 4.77 (s, 1H), 4.40 (q, *J* = 7.2 Hz, 1H), 3.99 (dd, *J* = 17.7, 5.7 Hz, 1H), 3.87 (dd, *J* = 17.7, 5.3 Hz, 1H), 3.65 (s, 3H), 3.25 (d, *J* = 14.5 Hz, 1H), 2.92 (d, *J* = 14.6 Hz, 1H), 2.31 (s, 3H), 1.36 (d, *J* = 7.3 Hz, 3H), 1.27 (s, 9H).

¹³C-NMR (150 MHz, CDCl₃) δ 172.2, 171.7, 169.9, 155.4, 142.4, 138.5, 138.3, 136.9, 136.7, 129.9, 129.4, 128.3, 128.1, 120.0, 80.9, 75.5, 59.2, 53.6, 52.1, 41.3, 29.2, 28.4, 21.1, 15.8.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₄₃H₄₇N₅O₆: 730.3599, found 730.3617.

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-phenylalaninate (5c)



The general procedure was followed starting from Boc–Ala–His(Trt)–Phe–OMe **4c** (70 mg, 96 μmol), tri-*p*-tolylbismuthane **2a** (46 mg, 96 μmol), copper(II) acetate (17 mg, 96 μmol), phenanthroline (17 mg, 96 μmol) and DiPEA (16 μL, 96 μmol) in CH₂Cl₂ (1 mL). The resulting crude

product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 98:2) to afford **5c** as a white solid (56 mg, 72%). **Mp** 85 – 87 °C; **R_f** = 0.24 (DCM–MeOH, 95:5).

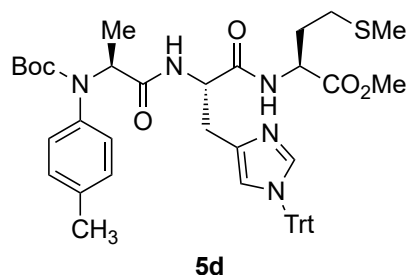
IR (ATR)/ $\bar{\nu}$: 3292, 2923, 1744, 1669, 1514, 1494, 1445, 1366, 1322, 1250, 1203, 1158, 1131, 1031, 854, 830, 747, 700, 660 cm^{–1}.

¹H-NMR (300 MHz, CDCl₃) δ 8.40 (d, *J* = 6.9 Hz, 1H), 7.84 (d, *J* = 7.7 Hz, 1H), 7.38 – 7.27 (m, 10H), 7.21 – 7.02 (m, 15H), 6.66 (s, 1H), 4.86 – 4.77 (m, 1H), 4.72 (q, *J* = 6.9 Hz, 1H), 4.45 (q, *J* = 7.0 Hz, 1H), 3.60 (s, 3H), 3.24 – 3.10 (m, 1H), 3.05 (d, *J* = 6.4 Hz, 2H), 2.88 (dd, *J* = 15.0, 5.3 Hz, 1H), 2.31 (s, 3H), 1.38 – 1.32 (m, 12H).

¹³C-NMR (75 MHz, CDCl₃) δ 172.0, 171.7, 171.2, 154.9, 142.4, 138.6, 136.5, 129.9, 129.4, 129.3, 128.5, 128.1, 126.9, 119.7, 80.8, 75.4, 58.6, 54.0, 53.2, 52.2, 37.8, 29.5, 28.4, 21.1, 15.5.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₅₀H₅₃N₅O₆: 820.4069, found 820.4069.

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-methioninate (5d)



The general procedure was followed starting from Boc-Ala-His(Trt)-Met-OMe **4d** (88 mg, 0.12 mmol), tri-*p*-tolylbismuthane **2a** (60 mg, 0.12 mmol), copper(II) acetate (22 mg, 0.12 mmol), phenanthroline (22 mg, 0.12 mmol) and DiPEA (21 μ L, 0.12 mmol) in CH₂Cl₂ (1.2 mL). The resulting

crude product was purified on silica gel (DCM-MeOH, gradient from 100:0 to 98:2) to afford **5d** as a yellow solid (65 mg, 65%). **Mp** 80 – 82 °C; **R_f** = 0.29 (DCM-MeOH, 95:5).

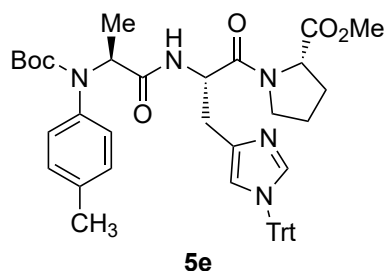
IR (ATR)/ $\bar{\nu}$: 3331, 2977, 2915, 1742, 1668, 1514, 1494, 1445, 1158, 1131, 1001, 839, 748, 701, 660 cm⁻¹.

¹H-NMR (600 MHz, CDCl₃) δ 8.58 (s, 1H), 7.83 (d, *J* = 7.7 Hz, 1H), 7.36 – 7.28 (m, 10H), 7.15 (d, *J* = 8.2 Hz, 2H), 7.12 – 7.06 (m, 8H), 6.70 (s, 1H), 4.78 (q, *J* = 5.9 Hz, 1H), 4.54 (dt, *J* = 8.0, 5.1 Hz, 1H), 4.45 (q, *J* = 7.2 Hz, 1H), 3.65 (s, 3H), 3.21 (dd, *J* = 14.6, 4.1 Hz, 1H), 2.91 (dd, *J* = 15.2, 6.0 Hz, 1H), 2.41 (t, *J* = 7.6 Hz, 2H), 2.31 (s, 3H), 2.15 – 2.07 (m, 1H), 2.03 – 1.97 (m, 1H), 2.00 (s, 3H), 1.35 (d, *J* = 7.3 Hz, 3H), 1.30 (s, 9H).

Presence of conformers: **¹³C-NMR (150 MHz, CDCl₃)** δ 172.1, 172.1, 171.4, 155.1, 142.3, 138.5, 138.2, 137.0, 136.6, 129.9, 129.4, 128.2, 128.2, 119.9, 80.9, 75.6, 58.8, 55.6, 53.3, 52.3, 51.9, 31.3, 30.2, 29.2, 28.4, 21.1, 15.6, 15.4.

HRMS-ESI: *m/z* [M + H]⁺ calcd. for C₄₆H₅₃N₅O₆S: 804.3789, found 804.3795.

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-prolinate (5e)



The general procedure was followed starting from Boc-Ala-His(Trt)-Pro-OMe **4e** (70 mg, 0.10 mmol), tri-*p*-tolylbismuthane **2a** (50 mg, 0.10 mmol), copper(II) acetate (19 mg, 0.10 mmol), phenanthroline (19 mg, 0.10 mmol) and DiPEA (18 μ L, 0.10 mmol) in CH₂Cl₂ (1.2 mL). The resulting

crude product was purified preparative TLC (DCM-MeOH, 96:4) to afford **5e** as a white solid (52 mg, 66%). **Mp** 66 – 68 °C; **R_f** = 0.29 (DCM-MeOH, 95:5).

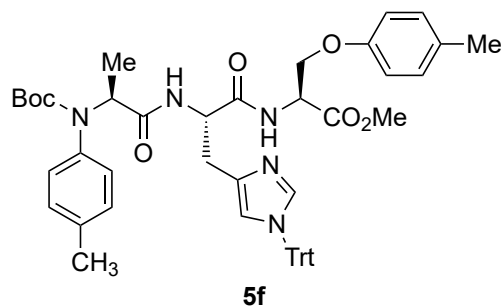
IR (ATR)/ $\bar{\nu}$: 2925, 1743, 1689, 1651, 1514, 1494, 1444, 1366, 1323, 1248, 1159, 1131, 1036, 1020, 1001, 841, 747, 701, 660 cm⁻¹.

¹H-NMR (600 MHz, CDCl₃) δ 7.57 (s, 1H), 7.31 – 7.27 (m, 10H), 7.15 – 7.10 (m, 6H), 7.08 (d, *J* = 10.9 Hz, 2H), 7.01 (d, *J* = 8.0 Hz, 2H), 6.76 (s, 1H), 4.99 (q, *J* = 6.1 Hz, 1H), 4.60 – 4.47 (m, 2H), 3.83 (q, *J* = 7.9 Hz, 1H), 3.62 (dt, *J* = 9.4, 6.3 Hz, 1H), 3.47 (s, 3H), 3.07 (dd, *J* = 14.6, 5.6 Hz, 1H), 2.91 (dd, *J* = 14.6, 6.2 Hz, 1H), 2.28 (s, 3H), 2.19 – 2.11 (m, 1H), 2.00 – 1.93 (m, 2H), 1.93 – 1.86 (m, 1H), 1.42 – 1.35 (m, 3H), 1.34 (s, 9H).

Presence of conformers: **¹³C-NMR (150 MHz, CDCl₃)** δ 172.6, 172.0, 170.3, 154.9, 142.6, 138.5, 136.4, 130.0, 129.8, 129.3, 128.3, 128.2, 128.1, 128.0, 120.2, 80.9, 75.5, 58.7, 52.1, 51.3, 47.0, 30.5, 29.8, 29.1, 28.4, 25.0, 21.1, 15.5.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₄₆H₅₁N₅O₆: 770.3912, found 770.3910.

Methyl *N*-(*N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl)-*O*-(*p*-tolyl)-*L*-serinate (5f)



The general procedure was followed starting from Boc-Ala-His(Trt)-Ser-OMe **4f** (55 mg, 82 μmol), tri-*p*-tolylbismuthane **2a** (40 mg, 82 μmol), copper(II) acetate (15 mg, 82 μmol), phenanthroline (15 mg, 82 μmol) and DiPEA (14 μL, 82 μmol) in CH₂Cl₂ (1 mL).

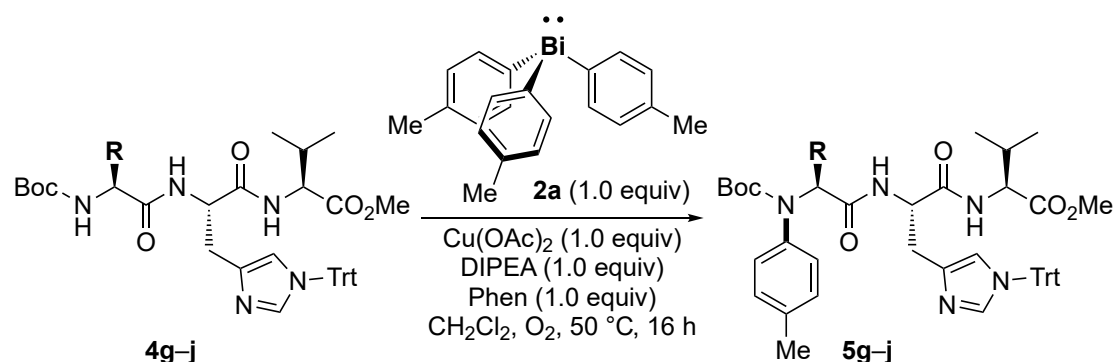
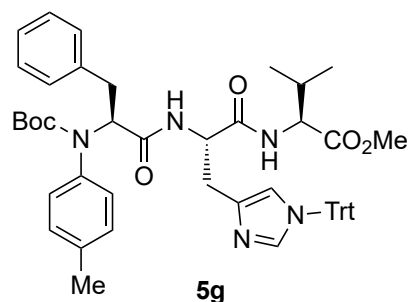
The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 80:20 to 30:70) to afford **5f** as a white solid (32 mg, 45%). **Mp** 80 – 82 °C; **R_f** = 0.63 (Hexane–EtOAc, 20:80).

IR (ATR)/ $\bar{\nu}$: 3299, 2921, 1748, 1671, 1511, 1445, 1366, 1323, 1237, 1204, 1157, 1132, 1087, 1036, 1001, 816, 748, 701, 660 cm⁻¹.

¹H-NMR (600 MHz, CDCl₃) δ 8.25 (s, 1H), 8.17 – 8.09 (m, 1H), 7.38 – 7.26 (m, 10H), 7.13 (d, *J* = 6.7 Hz, 2H), 7.10 – 7.06 (m, 6H), 7.05 (d, *J* = 8.1 Hz, 2H), 6.97 (d, *J* = 8.3 Hz, 2H), 6.70 (s, 1H), 6.68 (d, *J* = 8.3 Hz, 2H), 4.91 – 4.78 (m, 2H), 4.54 (s, 1H), 4.31 (dd, *J* = 9.3, 3.6 Hz, 1H), 4.13 (dd, *J* = 9.3, 3.6 Hz, 1H), 3.66 (s, 3H), 3.20 – 3.13 (m, 1H), 3.02 – 2.93 (m, 1H), 2.30 (s, 3H), 2.24 (s, 3H), 1.31 (s(br), 12H).

¹³C-NMR (150 MHz, CDCl₃) δ 172.2, 171.6, 170.1, 156.3, 154.5, 142.4, 138.3, 138.2, 136.6, 130.6, 129.9, 129.9, 129.4, 128.3, 128.2, 119.8, 114.9, 80.9, 75.6, 68.0, 53.9, 53.4, 52.7, 52.6, 30.0, 28.4, 21.1, 20.6, 15.7.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₅₁H₅₅N₅O₇: 850.4174, found 850.4187.

d. Arylated compounds (**5g–j**) from tripeptides (**4g–j**) using tritolylbismuth (**2a**)Methyl *N^α-(N-(tert-butoxycarbonyl)-N-(p-tolyl)-L-phenylalanyl)-N^ε-trityl-L-histidyl-L-valinate* (**5g**)

The general procedure was followed starting from Boc-Phe-His(Trt)-Val-OMe **4g** (70 mg, 92 μmol), tri-*p*-tolylbismuthane **2a** (45 mg, 92 μmol), copper(II) acetate (17 mg, 92 μmol), phenanthroline (17 mg, 92 μmol) and DIPEA (16 μL , 92 μmol) in CH_2Cl_2 (1 mL). The resulting crude product was purified on silica gel (DCM–MeOH, gradient

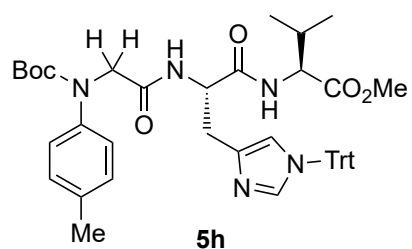
from 100:0 to 98:2) to afford **5g** as a yellow solid (28 mg, 36%). **Mp** $74 - 76\text{ }^\circ\text{C}$; **R_f** = 0.36 (DCM–MeOH, 95:5).

IR (ATR)/ $\bar{\nu}$: 3295, 2965, 2928, 1740, 1674, 1513, 1495, 1445, 1367, 1319, 1303, 1253, 1203, 1154, 1033, 1001, 827, 747, 700, 660 cm^{-1} .

^1H -NMR (600 MHz, CDCl_3) δ 8.61 (s, 1H), 7.73 (d, $J = 8.2\text{ Hz}$, 1H), 7.33 – 7.28 (m, 10H), 7.24 (d, $J = 7.4\text{ Hz}$, 2H), 7.21 (d, $J = 7.1\text{ Hz}$, 1H), 7.15 (d, $J = 7.2\text{ Hz}$, 2H), 7.11 – 7.06 (m, 6H), 6.87 (d, $J = 8.0\text{ Hz}$, 2H), 6.71 (s, 1H), 6.59 (s(br), 2H), 4.90 – 4.84 (m, 1H), 4.63 – 4.56 (m, 1H), 4.47 – 4.40 (m, 1H), 3.64 (s, 3H), 3.37 (s(br), 1H), 3.27 (s(br), 1H), 3.21 – 3.14 (m, 1H), 2.98 – 2.90 (m, 1H), 2.22 (s, 3H), 2.19 – 2.09 (m, 1H), 1.33 (s(br), 9H), 0.84 (d, $J = 6.7\text{ Hz}$, 6H).

^{13}C -NMR (150 MHz, CDCl_3) δ 172.1, 171.4, 171.2, 165.9, 154.7, 142.5, 141.1, 139.6, 139.0, 138.5, 136.1, 129.9, 129.6, 129.1, 128.5, 128.1, 127.5, 126.6, 119.6, 80.9, 75.5, 65.6, 57.7, 53.4, 52.0, 31.0, 29.8, 28.4, 21.0, 19.2, 18.0.

HRMS–ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{52}\text{H}_{57}\text{N}_5\text{O}_6$: 848.4382, found 848.4387.

Methyl N^α -(N -(*tert*-butoxycarbonyl)- N -(*p*-tolyl)glycyl)- N^ϵ -trityl- L -histidyl- L -valinate (5h**)**

The general procedure was followed starting from Boc-Gly-His(Trt)-Val-OMe **4h** (54 mg, 81 μ mol), tri-*p*-tolylbismuthane **2a** (39 mg, 81 μ mol), copper(II) acetate (15 mg, 81 μ mol), phenanthroline (15 mg, 81 μ mol) and DiPEA (14 μ L, 81 μ mol) in CH_2Cl_2 (1 mL). The resulting crude

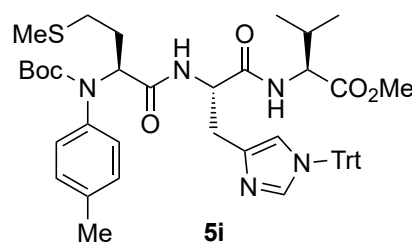
product was purified on silica gel (DCM-MeOH, gradient from 100:0 to 98:2) to afford **5h** as a white solid (37 mg, 61%). **Mp** 83 – 85 $^\circ\text{C}$; **R_f** = 0.36 (Hexane-EtOAc, 30:70).

IR (ATR)/ $\bar{\nu}$: 3292, 2965, 2931, 1742, 1683, 1514, 1494, 1445, 1367, 1206, 1152, 1035, 1017, 1001, 868, 824, 748, 701, 659 cm^{-1} .

$^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 8.54 (d, J = 7.1 Hz, 1H), 7.60 (d, J = 8.4 Hz, 1H), 7.36 – 7.28 (m, 10H), 7.23 (d, J = 8.2 Hz, 2H), 7.13 – 7.03 (m, 8H), 6.67 (s, 1H), 4.85 – 4.76 (m, 1H), 4.41 (dd, J = 8.5, 5.3 Hz, 1H), 4.31 (d, J = 17.0 Hz, 1H), 4.24 (d, J = 17.0 Hz, 1H), 3.64 (s, 3H), 3.15 (dd, J = 15.0, 4.2 Hz, 1H), 2.87 (dd, J = 15.0, 6.1 Hz, 1H), 2.29 (s, 3H), 2.18 – 2.04 (m, 1H), 1.36 (s, 9H), 0.83 (d, J = 3.0 Hz, 3H), 0.81 (d, J = 3.0 Hz, 3H).

$^{13}\text{C-NMR}$ (75 MHz, CDCl_3) δ 172.0, 171.1, 169.5, 154.9, 142.4, 140.3, 138.3, 137.1, 136.0, 129.9, 129.6, 128.2, 128.2, 126.3, 119.7, 81.2, 75.5, 57.7, 54.6, 53.2, 52.0, 31.0, 29.7, 28.3, 21.1, 19.2, 18.0.

HRMS-ESI: m/z $[\text{M} + \text{H}]^+$ calcd. for $\text{C}_{45}\text{H}_{51}\text{N}_5\text{O}_6$: 758.3912, found 758.3920.

Methyl N^α -(N -(*tert*-butoxycarbonyl)- N -(*p*-tolyl)- L -methionyl)- N^ϵ -trityl- L -histidyl- L -valinate (5i**)**

The general procedure was followed starting from Boc-Met-His(Trt)-Val-OMe **4i** (84 mg, 0.11 mmol), tri-*p*-tolylbismuthane **2a** (55 mg, 0.11 mmol), copper(II) acetate (21 mg, 0.11 mmol), phenanthroline (20 mg, 0.11 mmol) and DiPEA (19 μ L, 0.11 mmol) in CH_2Cl_2 (1.1 mL). The

resulting crude product was purified on silica gel (Hexane-EtOAc, gradient from 80:20 to 30:70) to afford **5i** as a yellow solid (48 mg, 51%). **Mp** 71 – 73 $^\circ\text{C}$; **R_f** = 0.35 (DCM-MeOH, 95:5).

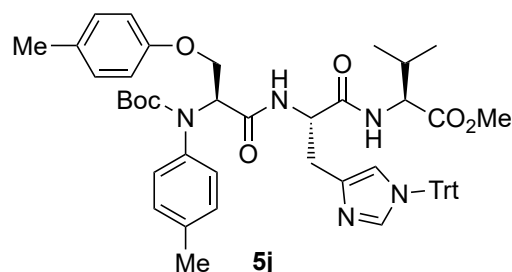
IR (ATR)/ $\bar{\nu}$: 3306, 2965, 2925, 1741, 1683, 1513, 1494, 1445, 1367, 1323, 1303, 1255, 1204, 1155, 1037, 1001, 870, 828, 748, 701, 660 cm^{-1} .

¹H-NMR (600 MHz, CDCl₃) δ 8.45 (s, 1H), 7.70 (d, *J* = 7.2 Hz, 1H), 7.36 – 7.27 (m, 10H), 7.17 – 7.06 (m, 8H), 7.04 (d, *J* = 7.9 Hz, 2H), 6.70 (s(br), 1H), 4.82 (s, 1H), 4.76 – 4.68 (m, 1H), 4.48 – 4.40 (m, 1H), 3.63 (s, 3H), 3.17 – 3.08 (m, 1H), 2.96 – 2.89 (m, 1H), 2.54 (t, *J* = 7.1 Hz, 2H), 2.28 (s, 4H), 2.16 – 2.08 (m, 1H), 1.99 (s, 3H), 1.98 – 1.91 (m, 1H), 1.35 (s, 9H), 0.85 (d, *J* = 2.9 Hz, 3H), 0.84 (d, *J* = 2.9 Hz, 3H).

¹³C-NMR (150 MHz, CDCl₃) δ 172.0, 171.3, 171.3, 165.8, 154.9, 142.4, 138.1, 136.5, 129.8, 129.4, 128.1, 127.8, 119.4, 81.2, 75.5, 61.2, 57.6, 53.4, 52.0, 31.2, 31.0, 29.8, 29.0, 28.3, 21.1, 19.1, 18.0, 15.3.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₄₈H₅₇N₅O₆S: 832.4102, found 832.4113.

Methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*,*O*-di-*p*-tolyl-*L*-seryl)-*N*^ε-trityl-*L*-histidyl-*L*-valinate (5j**)**



The general procedure was followed starting from Boc–Ser–His(Trt)–Val–OMe **4j** (45 mg, 65 μmol), tri-*p*-tolylbismuthane **2a** (31 mg, 65 μmol), copper(II) acetate (12 mg, 65 μmol), phenanthroline (12 mg, 65 μmol) and DiPEA (11 μL, 65 μmol) in CH₂Cl₂ (650

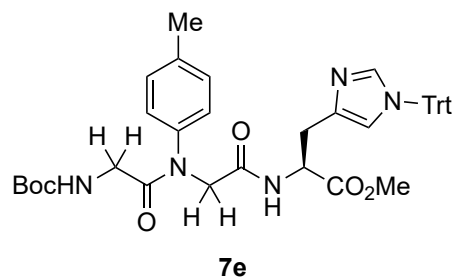
μL). The resulting crude product was purified on silica gel (Hexane–EtOAc, gradient from 80:20 to 30:70) to afford **5j** as a white solid (19 mg, 33%). **Mp** 72 – 74 °C; **R_f** = 0.57 (Hexane–EtOAc, 20:80).

IR (ATR)/ $\bar{\nu}$: 3334, 2965, 2929, 1742, 1683, 1511, 1445, 1368, 1324, 1241, 1209, 1156, 1037, 818, 749, 702, 660 cm^{–1}.

¹H-NMR (600 MHz, CDCl₃) δ 7.34 – 7.28 (m, 12H), 7.12 – 7.07 (m, 8H), 7.05 (d, *J* = 7.9 Hz, 2H), 7.02 (d, *J* = 7.9 Hz, 2H), 6.78 (d, *J* = 7.3 Hz, 2H), 6.71 (s, 1H), 4.91 – 4.84 (m, 1H), 4.85 – 4.81 (m, 1H), 4.57 – 4.51 (m, 1H), 4.44 (dd, *J* = 8.0, 5.2 Hz, 1H), 4.28 (t, *J* = 9.5 Hz, 1H), 3.64 (s, 3H), 3.21 – 3.15 (m, 1H), 3.00 – 2.91 (m, 1H), 2.29 (s, 3H), 2.25 (s, 3H), 2.15 – 2.06 (m, 1H), 1.33 (s, 9H), 0.85 – 0.79 (m, 6H).

¹³C-NMR (150 MHz, CDCl₃) δ 172.1, 171.2, 169.6, 156.3, 142.4, 136.7, 130.3, 130.3, 130.0, 129.9, 129.8, 129.5, 128.7, 128.5, 128.2, 126.9, 119.6, 114.7, 80.8, 75.5, 53.3, 52.1, 36.8, 31.0, 29.8, 28.4, 24.8, 23.5, 21.2, 20.6, 19.2, 18.0.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₅₃H₅₉N₅O₇: 878.4487, found 878.4479.

e. Arylated compounds (**7**) from tripeptides (**6**) using tritolylbismuth (**2a**)Methyl *N*^α-(*N*-((*tert*-butoxycarbonyl)glycyl)-*N*-(*p*-tolyl)glycyl)-*N*^ε-trityl-*L*-histidinate (**7e**)

The general procedure was followed starting from Boc–Gly–Gly–His(Trt)–OMe **6e** (60 mg, 96 μmol), tri-*p*-tolylbismuthane **2a** (69 mg, 144 μmol, 1.5 equiv.), copper(II) acetate (26 mg, 144 μmol, 1.5 equiv.), phenanthroline (26 mg, 144 μmol, 1.5 equiv.) and DIPEA

(24.5 μL, 144 μmol, 1.5 equiv.) in CH₂Cl₂ (1 mL). The resulting crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 98:2) to afford **7e** as a white solid (17 mg, 25%).

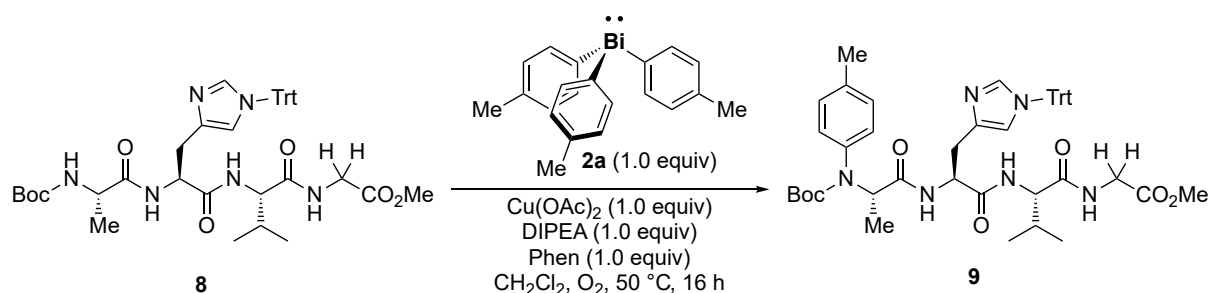
Mp 74 – 76 °C; **R_f** = 0.24 (DCM–MeOH, 95:5).

IR (ATR)/ $\bar{\nu}$: 3322, 2975, 2930, 1746, 1674, 1514, 1494, 1445, 1391, 1366, 1245, 1210, 1169, 1133, 1034, 870, 829, 750, 703, 661 cm^{−1}.

¹H-NMR (600 MHz, CDCl₃) δ 7.93 (s, 1H), 7.37 – 7.29 (m, 10H), 7.25 (d, *J* = 8.3 Hz, 2H), 7.13 (d, *J* = 8.3 Hz, 2H), 7.11 – 7.07 (m, 6H), 6.56 (s, 1H), 5.36 (s(br), 1H), 4.82 – 4.76 (m, 1H), 4.43 (d, *J* = 16.3 Hz, 1H), 4.22 (d, *J* = 16.9 Hz, 1H), 3.76 – 3.67 (m, 2H), 3.59 (s, 3H), 3.07 (dd, *J* = 14.6, 4.9 Hz, 1H), 2.98 (dd, *J* = 14.5, 4.6 Hz, 1H), 2.32 (s, 3H), 1.36 (s, 9H).

¹³C-NMR (150 MHz, CDCl₃) δ 171.4, 169.7, 167.9, 155.6, 142.2, 138.8, 138.3, 136.2, 130.6, 130.0, 129.7, 128.1, 127.6, 119.7, 115.1, 79.4, 75.4, 53.5, 52.6, 52.1, 43.2, 29.6, 28.3, 21.0.

HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₄₂H₄₅N₅O₆: 716.3443, found 716.3471.

f. Arylated compounds (**9**) from tetrapeptide (**8**) using tritolylbismuth (**2a**)Methyl *N*^α-(*N*-((*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl)-*N*^ε-trityl-*L*-histidyl-*L*-valylglycinate (**9**)

The general procedure was followed starting from Boc–Ala–His(Trt)–Val–Gly–OMe **8** (70 mg, 95 μmol), tri-*p*-tolylbismuthane **2a** (46 mg, 95 μmol), copper(II) acetate (17 mg, 95 μmol), phenanthroline (17 mg, 95 μmol) and DIPEA (16 μL, 95 μmol) in CH₂Cl₂ (1 mL). The resulting

crude product was purified on silica gel (DCM–MeOH, gradient from 100:0 to 98:2) to afford **9** as a white solid (50 mg, 64%). **Mp** 95 – 97 °C; **R_f** = 0.36 (DCM–MeOH, 95:5).

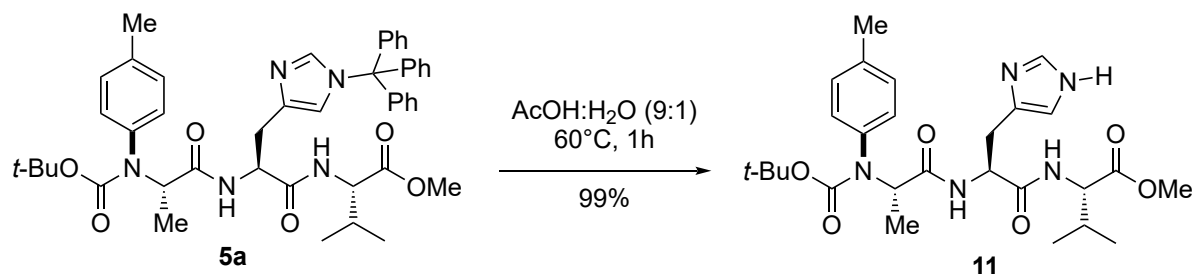
IR (ATR)/ $\bar{\nu}$: 3315, 2968, 2923, 1752, 1667, 1513, 1494, 1445, 1367, 1323, 1203, 1157, 1131, 1036, 1001, 988, 842, 748, 701, 660 cm⁻¹.

¹H-NMR (600 MHz, CDCl₃) δ 8.76 (s(br), 1H), 7.83 (s, 1H), 7.36 – 7.28 (m, 11H), 7.15 (d, *J* = 8.0 Hz, 2H), 7.10 – 7.05 (m, 8H), 6.75 (s, 1H), 4.59 (q, *J* = 5.5 Hz, 1H), 4.46 (q, *J* = 7.2 Hz, 1H), 4.39 (dd, *J* = 8.8, 4.9 Hz, 1H), 4.09 (dd, *J* = 17.7, 6.1 Hz, 1H), 3.83 (dd, *J* = 17.7, 5.3 Hz, 1H), 3.64 (s, 3H), 3.17 (dd, *J* = 14.8, 4.2 Hz, 1H), 3.00 (dd, *J* = 14.8, 6.3 Hz, 1H), 2.43 – 2.38 (m, 1H), 2.32 (s, 3H), 1.28 (s, 12H), 0.91 – 0.86 (m, 6H).

¹³C-NMR (150 MHz, CDCl₃) δ 173.5, 171.8, 171.5, 170.3, 155.2, 142.3, 138.6, 137.9, 136.8, 136.7, 129.8, 129.4, 128.5, 128.3, 128.2, 120.0, 81.0, 75.6, 59.0, 58.5, 54.9, 52.1, 41.2, 29.4, 28.3, 21.2, 19.6, 17.4, 16.0.

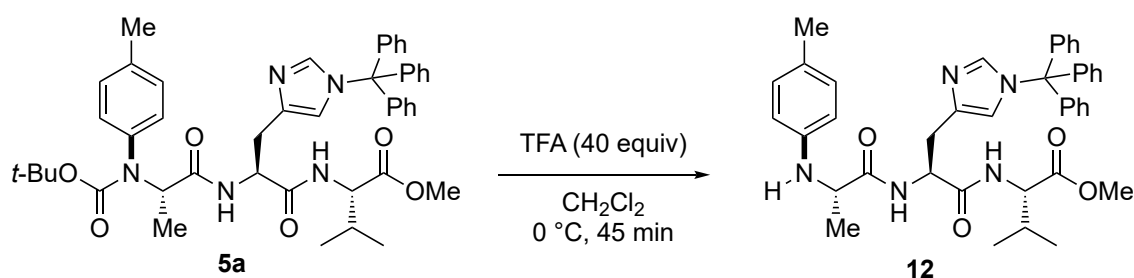
HRMS–ESI: *m/z* [M + H]⁺ calcd. for C₄₈H₅₆N₆O₇: 829.4283, found 829.4301.

5. Procedure for selective deprotection of **5a**



Methyl *N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-*L*-alanyl-*L*-histidyl-*L*-valinate (**11**)

In a flask, Boc-*N*(Tolyl)-Ala-His(trt)-Val-OMe **5a** (50 mg, 0.65 mmol) in AcOH:H₂O (9:1, 0.35 M) was stirred for 1h30 at 60 °C. After cooling down at room temperature, the solvent was co-evaporated with cyclohexane under reduce pressure. The white solid residue was purified on silica gel chromatography (5% MeOH/CH₂Cl₂) to afford **11** as a white solid (35 mg, 99%): This compound has been previously reported¹: **Mp** 154 – 157 °C; **R_f** = 0.46 (6% MeOH/CH₂Cl₂); **¹H-NMR (600 MHz, CDCl₃)** δ 7.86 (s(br), 1H), 7.62 (s(br), 1H), 7.56 (s, 1H), 7.14 – 7.08 (m, 4H), 6.83 (s, 1H), 4.80 – 4.75 (m, 1H), 4.57 – 4.51 (m, 1H), 4.39 (dd, *J* = 8.3, 5.2 Hz, 1H), 3.71 (s, 3H), 3.16 (dd, *J* = 15.1, 4.7 Hz, 1H), 3.09 (dd, *J* = 15.2, 5.9 Hz, 1H), 2.33 (s, 3H), 2.18 – 2.11 (m, 1H), 1.36 (s, 9H), 1.30 (d, *J* = 6.9 Hz, 3H), 0.86 (dd, *J* = 6.8, 3.5 Hz, 6H).



Methyl *N*^α-(*p*-tolyl-L-alanyl)-*N*^ε-trityl-L-histidyl-L-valinate (**12**)

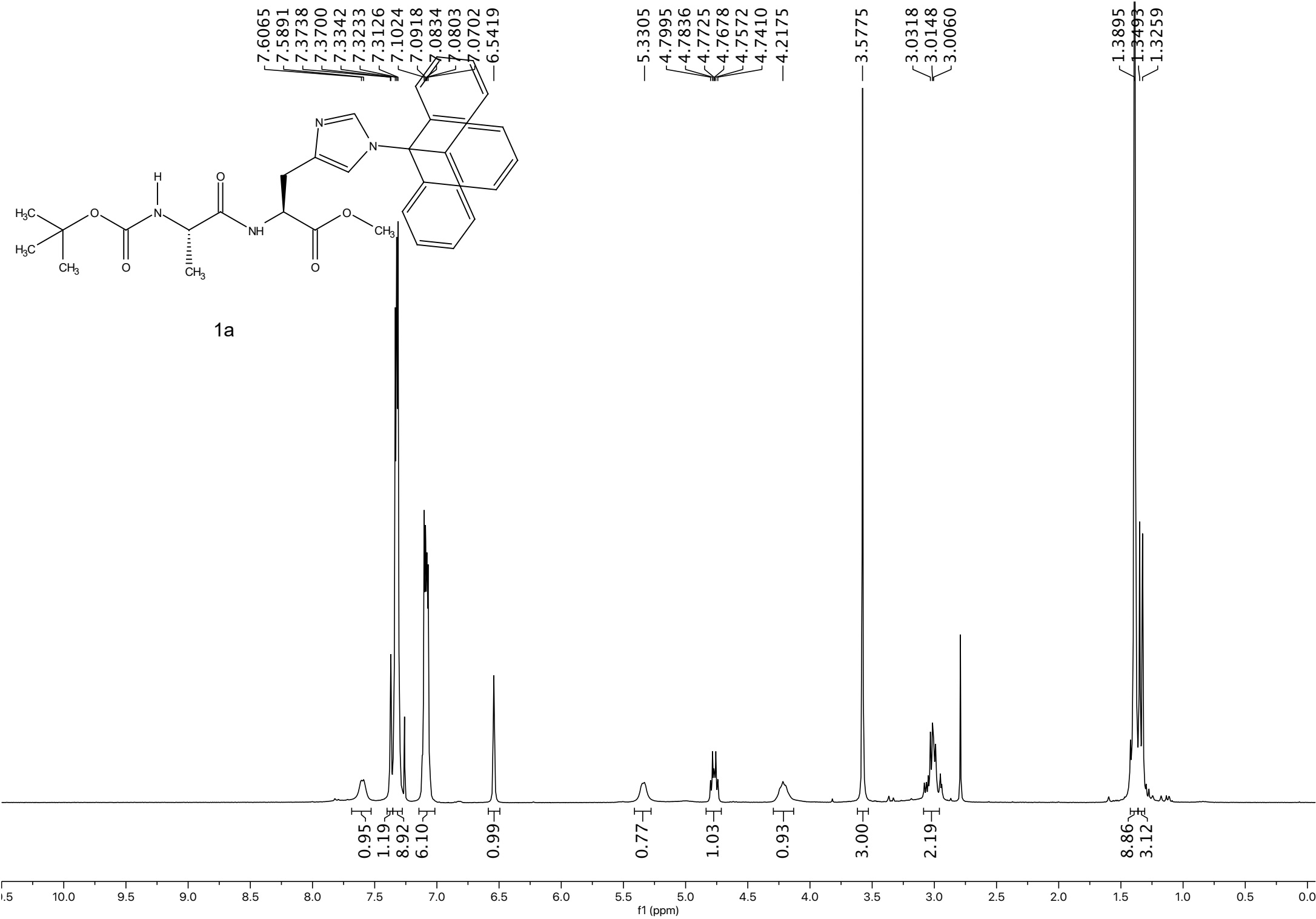
In a flask, trifluoroacetic acid (450 μL , 5.8 mmol, 40 equiv.) was added slowly to a solution of methyl *N*^α-(*N*-(*tert*-butoxycarbonyl)-*N*-(*p*-tolyl)-L-alanyl)-*N*^ε-trityl-L-histidyl-L-valinate (**5a**) (112 mg, 0.15 mmol, 1.0 equiv.) in dichloromethane (3 mL, 0.5 M) at 0 $^\circ\text{C}$. After stirred at 0 $^\circ\text{C}$ for 45 min (followed by TLC), the mixture was poured in a sat. solution of NaHCO_3 . The combined organic phases were washed with brine (20 mL), dried over Na_2SO_4 , filtered and concentrated under reduce pressure. The white solid residue was purified on silica gel chromatography (2% MeOH/ CH_2Cl_2) to afford **12** as a white solid (67 mg, 69%). **Mp** 74 – 76 $^\circ\text{C}$; **R_f** = 0.36 (CH_2Cl_2 –MeOH, 93:7).

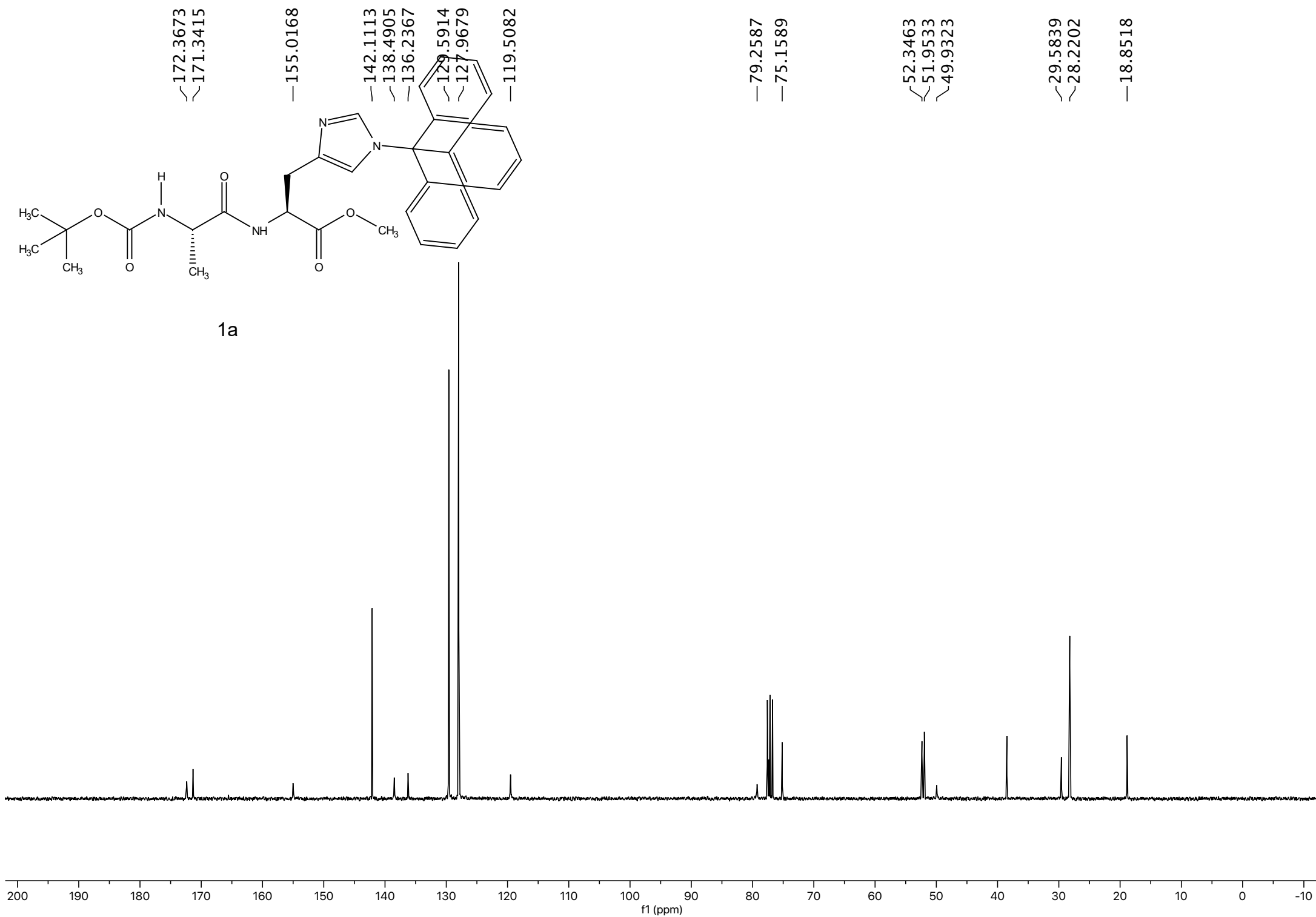
IR (ATR)/ $\bar{\nu}$: 3324, 2965, 1743, 1667, 1521, 1494, 1446, 1318, 1205, 1155, 749, 702, 661 cm^{-1} .

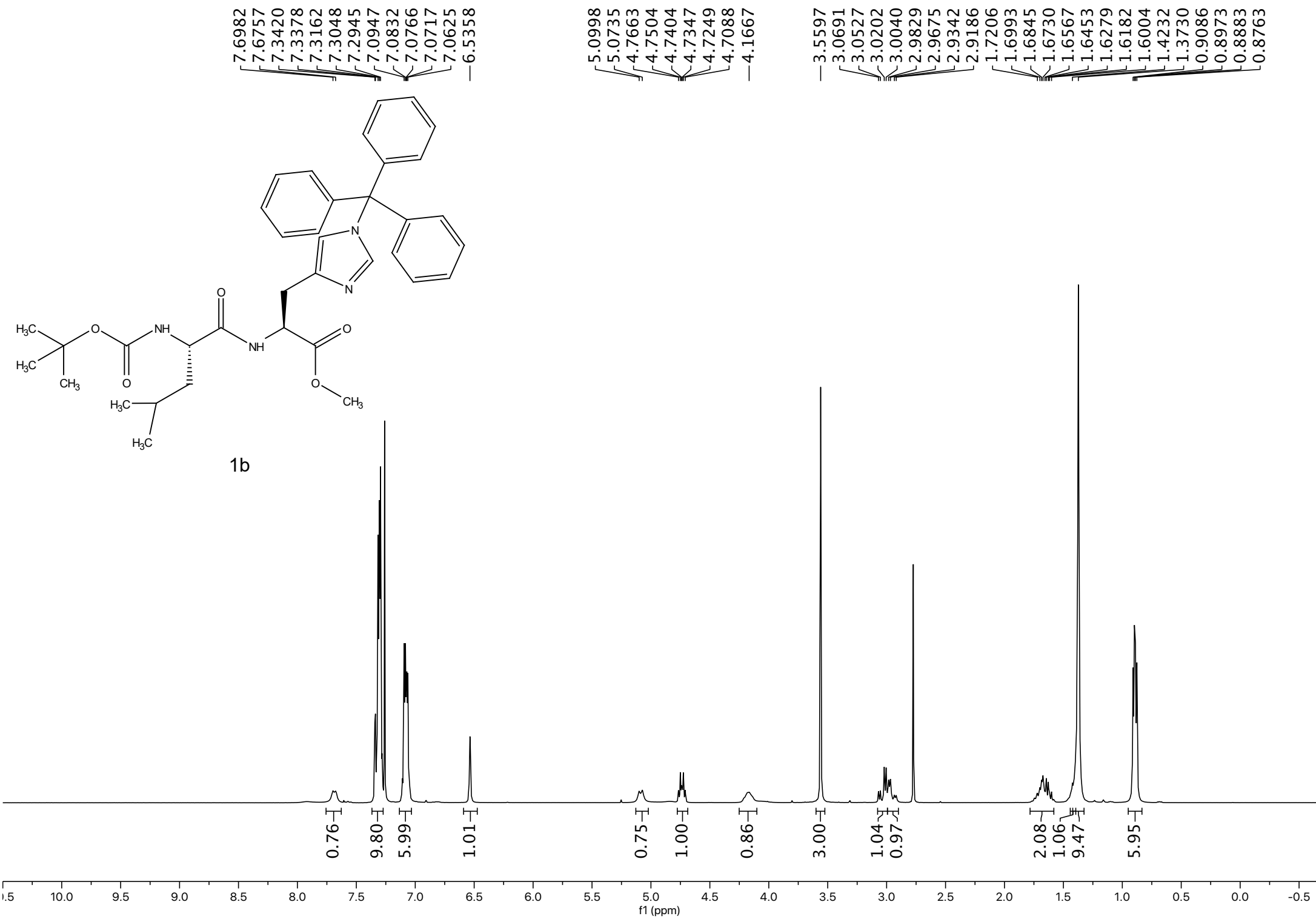
^1H -NMR (600 MHz, CDCl_3) δ 8.70 (d, J = 7.3 Hz, 1H), 7.35 – 7.28 (m, 9H), 7.20 (d, J = 1.0 Hz, 1H), 7.10 (d, J = 8.7 Hz, 1H), 7.06 – 7.00 (m, 6H), 6.91 (d, J = 8.2 Hz, 2H), 6.64 (s, 1H), 6.52 (d, J = 8.3 Hz, 2H), 4.70 – 4.64 (m, 1H), 4.36 (dd, J = 8.7, 5.3 Hz, 1H), 3.95 (s(br), 1H), 3.89 (q, J = 6.8 Hz, 1H), 3.70 (s, 3H), 3.10 (dd, J = 14.6, 5.0 Hz, 1H), 2.88 (dd, J = 14.8, 5.6 Hz, 1H), 2.16 (s, 3H), 1.97 – 1.88 (m, 1H), 1.47 (d, J = 7.0 Hz, 3H), 0.71 (d, J = 6.8 Hz, 3H), 0.60 (d, J = 6.8 Hz, 3H).

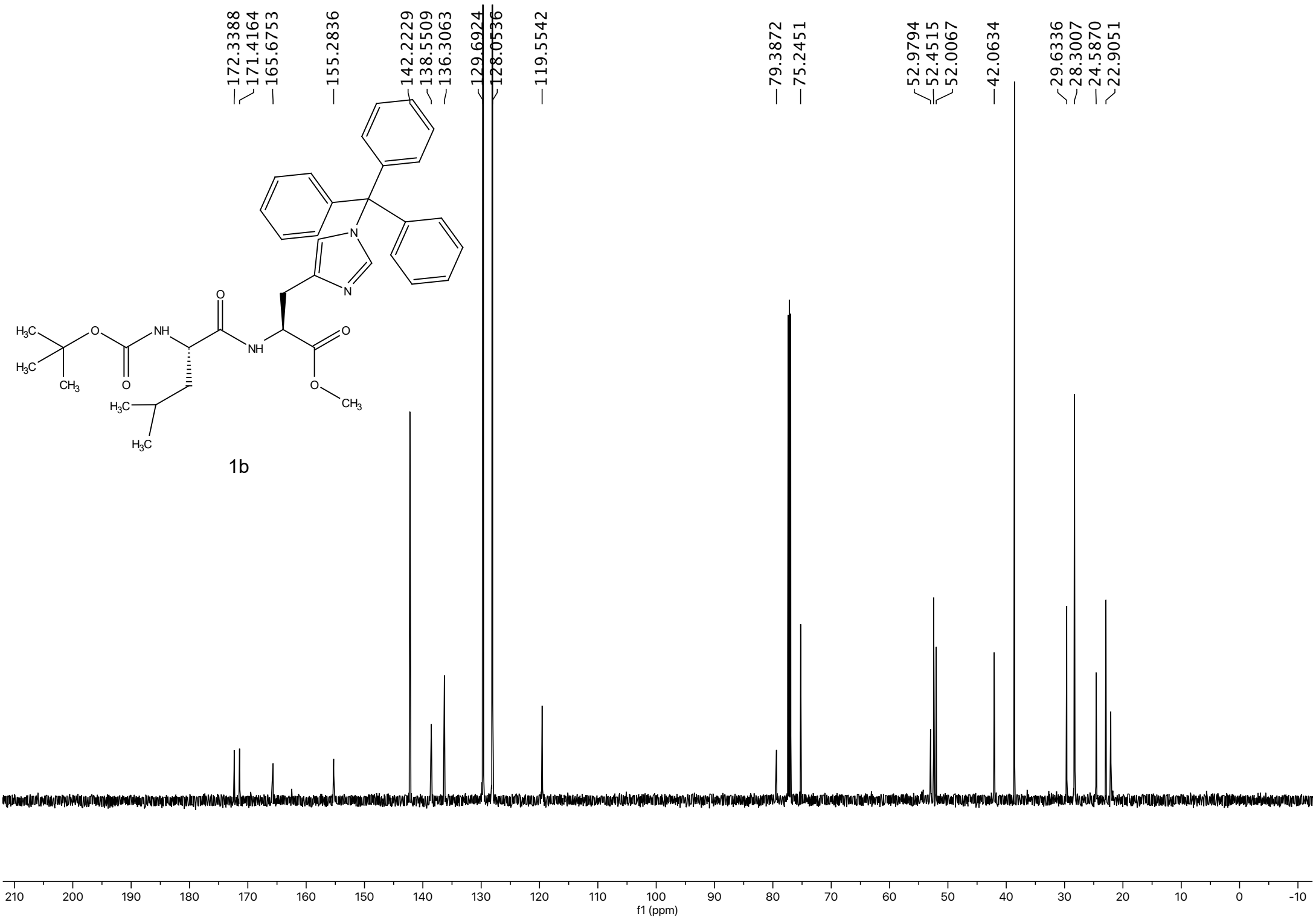
^{13}C -NMR (150 MHz, CDCl_3) δ 175.0, 172.0, 171.0, 144.3, 142.4, 138.4, 136.8, 130.1, 129.8, 128.2, 128.14, 127.6, 119.7, 113.2, 75.4, 57.4, 54.7, 53.4, 52.1, 31.1, 29.3, 20.5, 19.7, 19.2, 17.9.

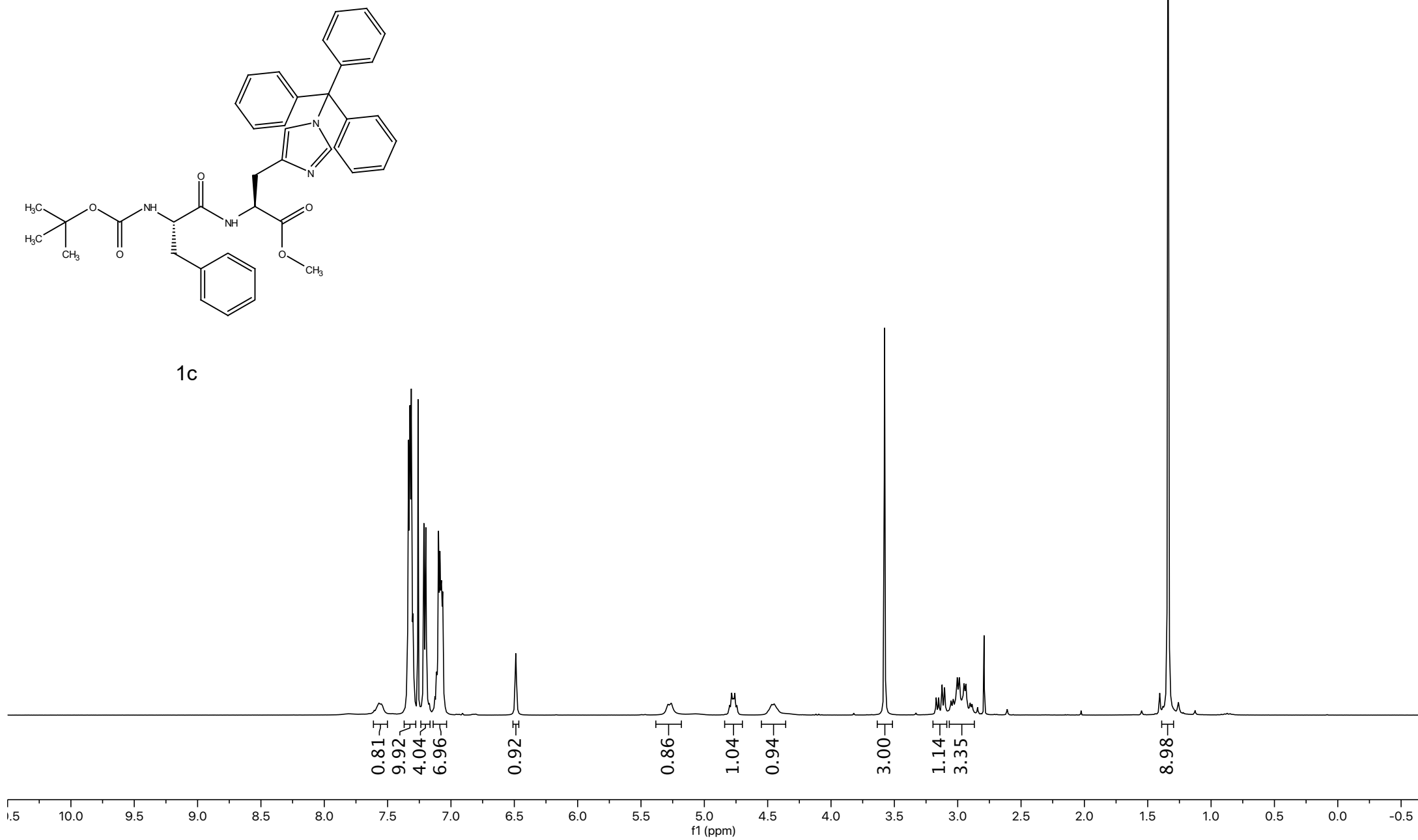
HRMS–ESI: m/z [$\text{M} + \text{H}$]⁺ calcd. for $\text{C}_{41}\text{H}_{45}\text{N}_5\text{O}_4$: 672.3544, found 672.3575.

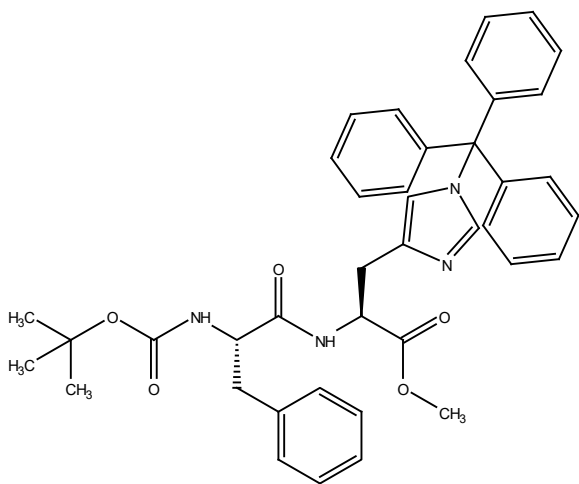




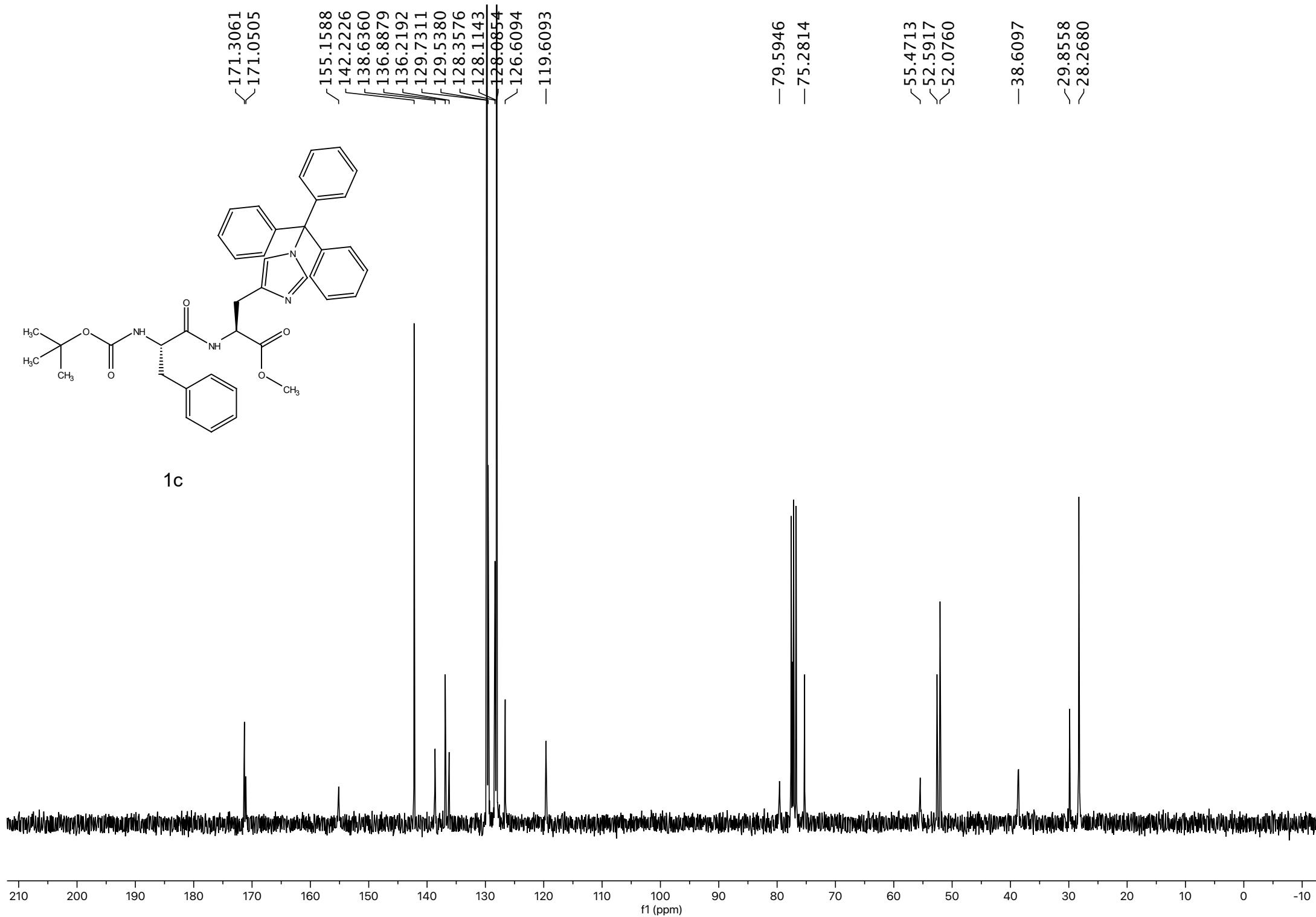


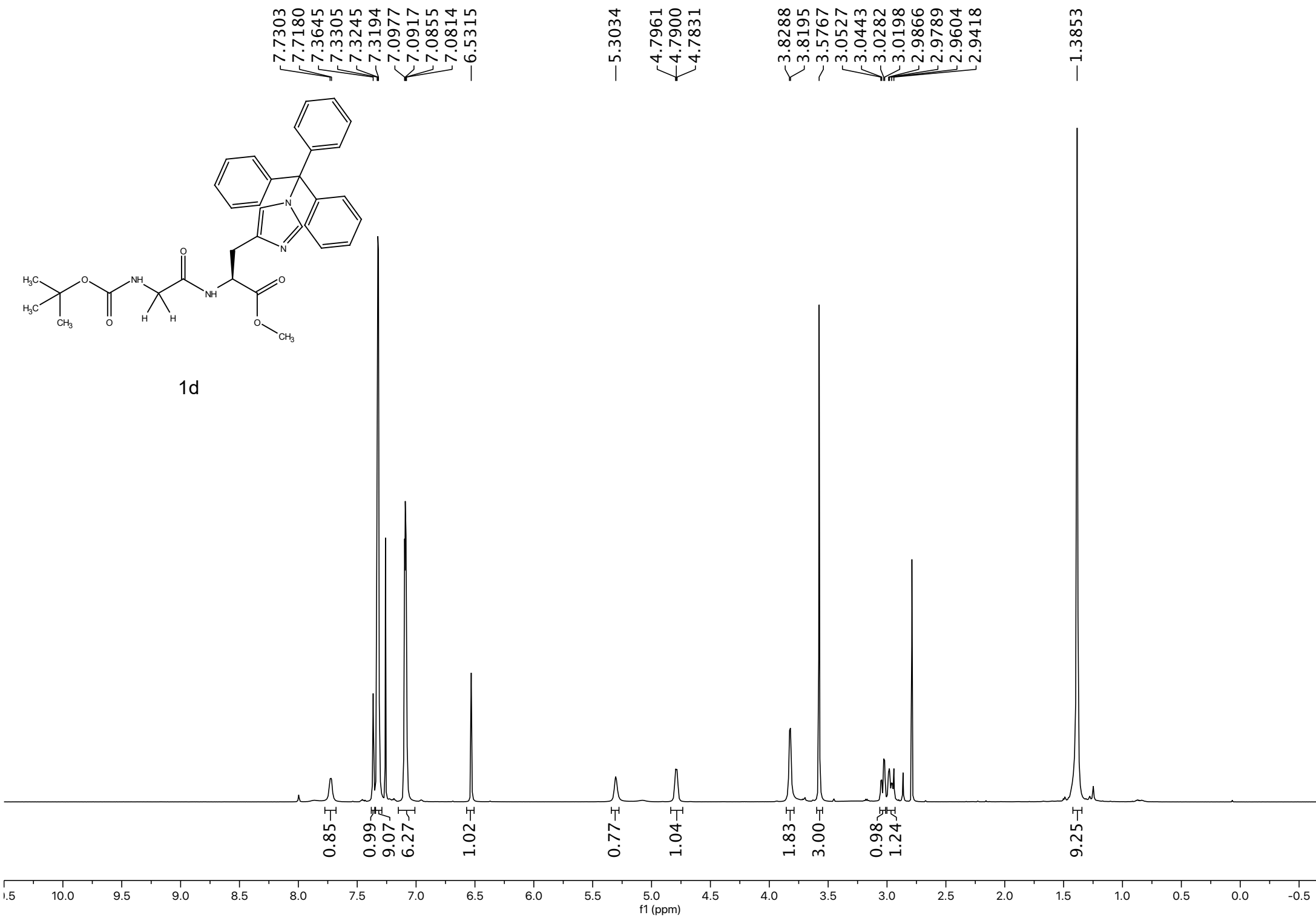


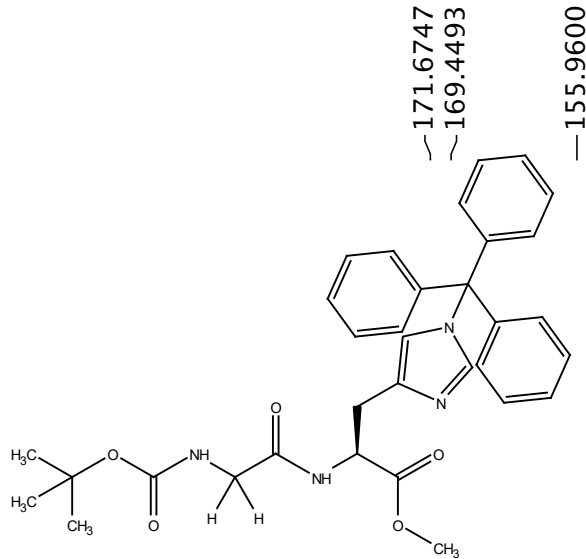




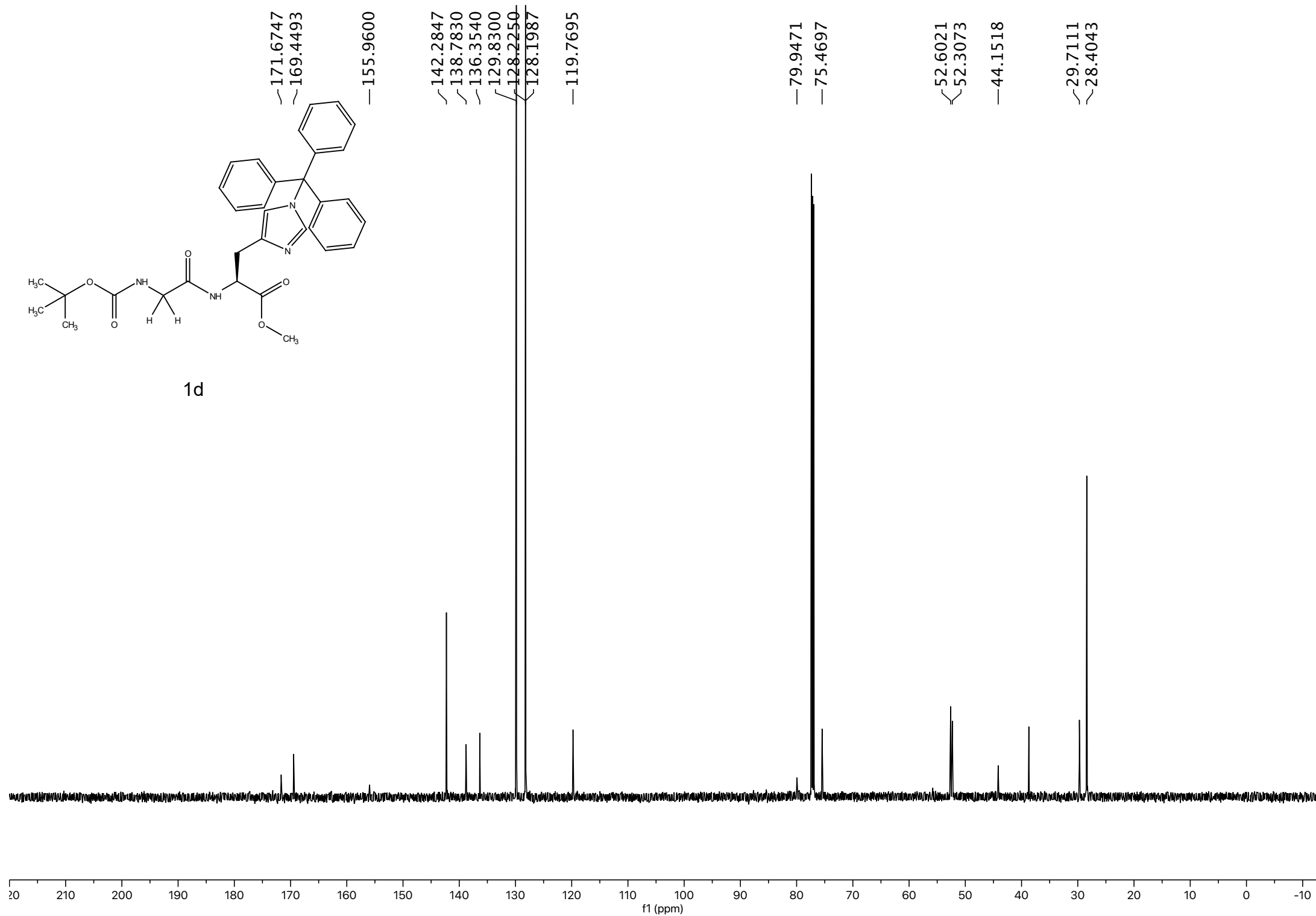
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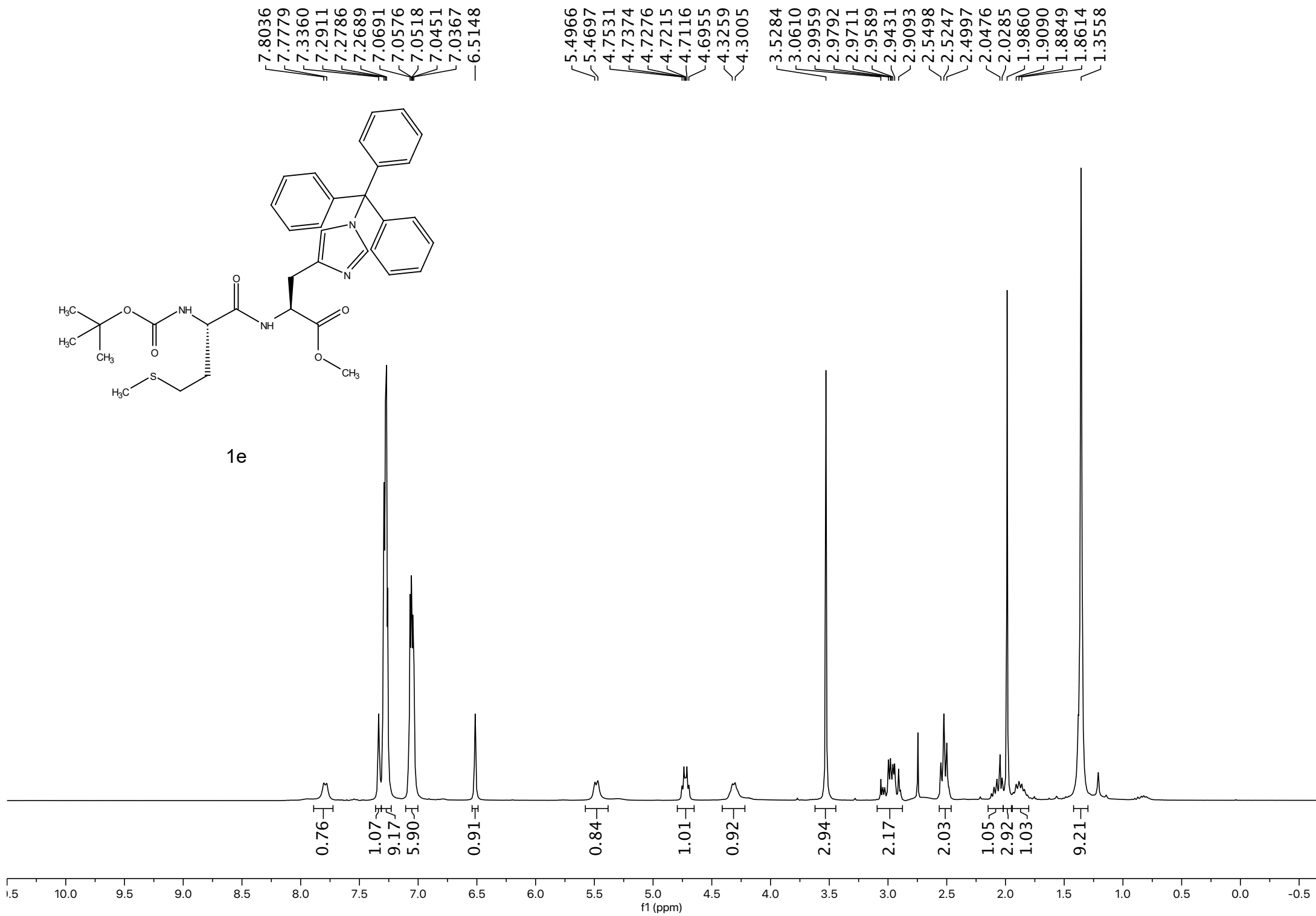


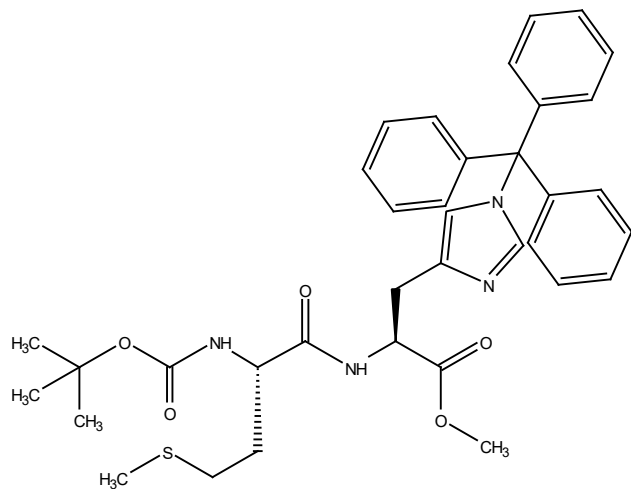




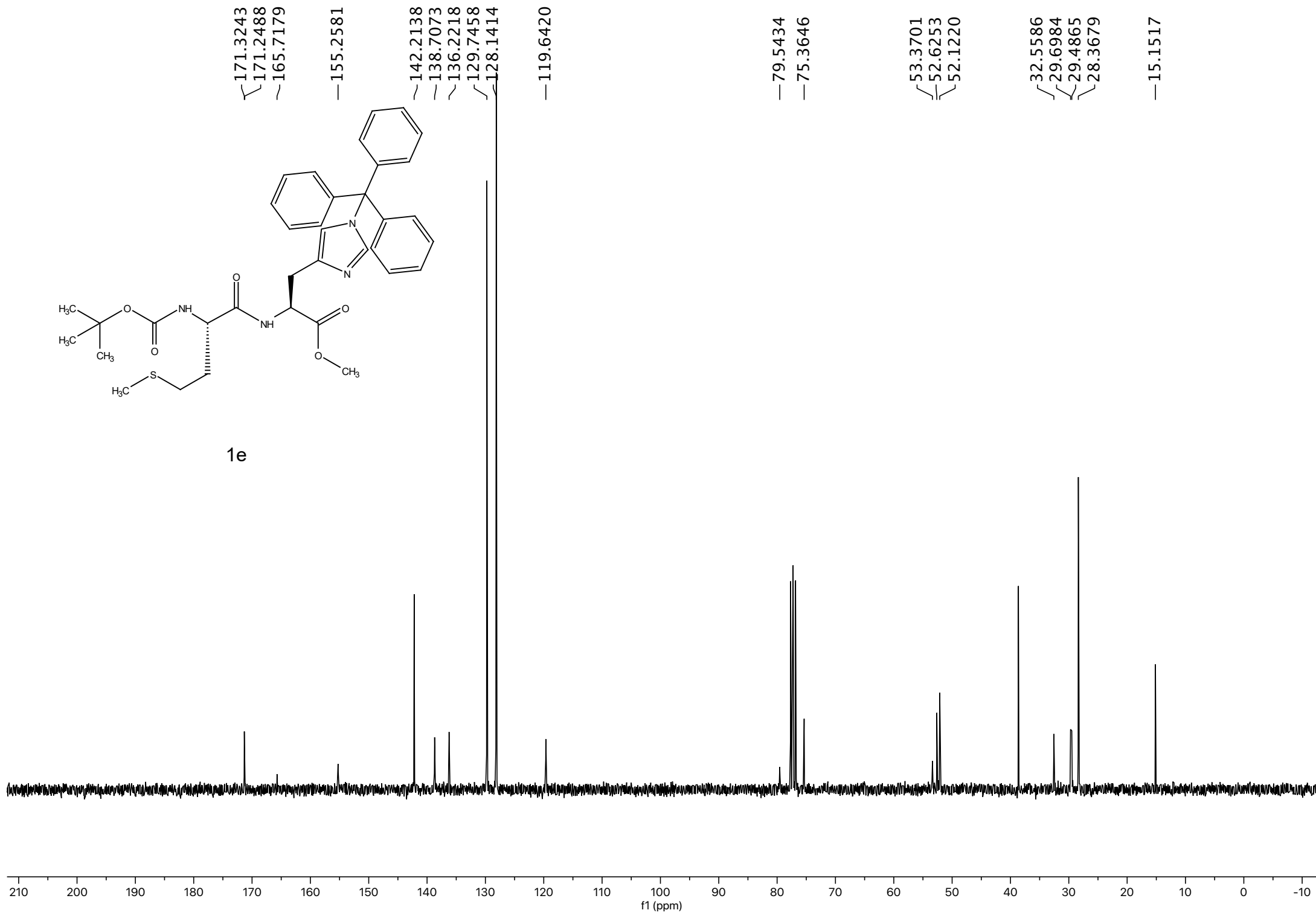
1d

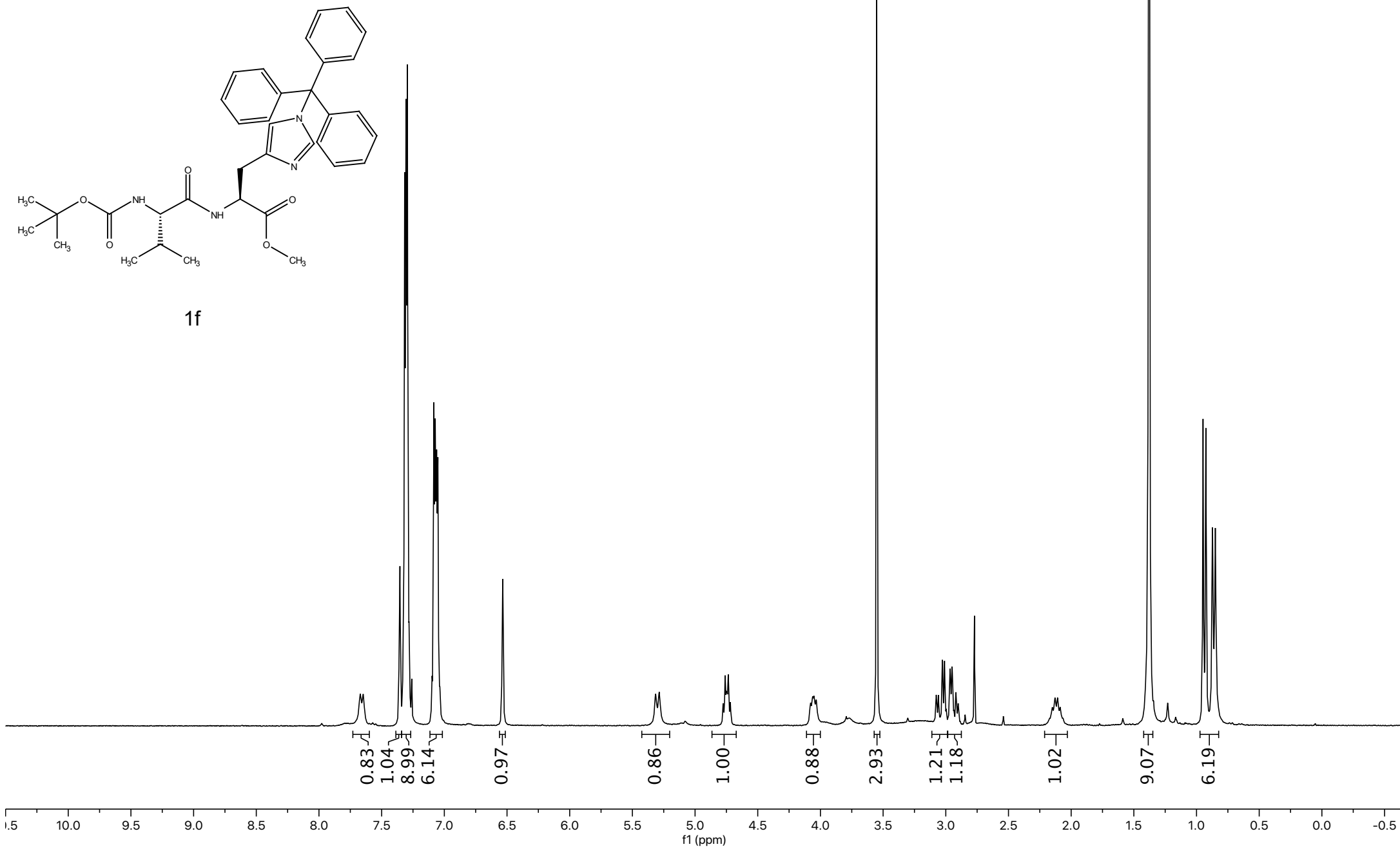


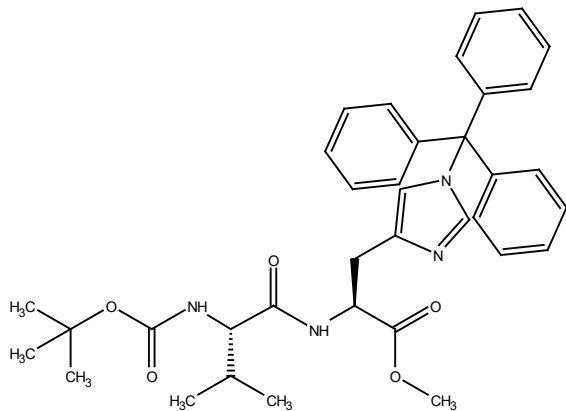




1e







1f

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171.3835

155.7637

142.2034
138.6848
136.2527
129.7509
128.1643
128.1365

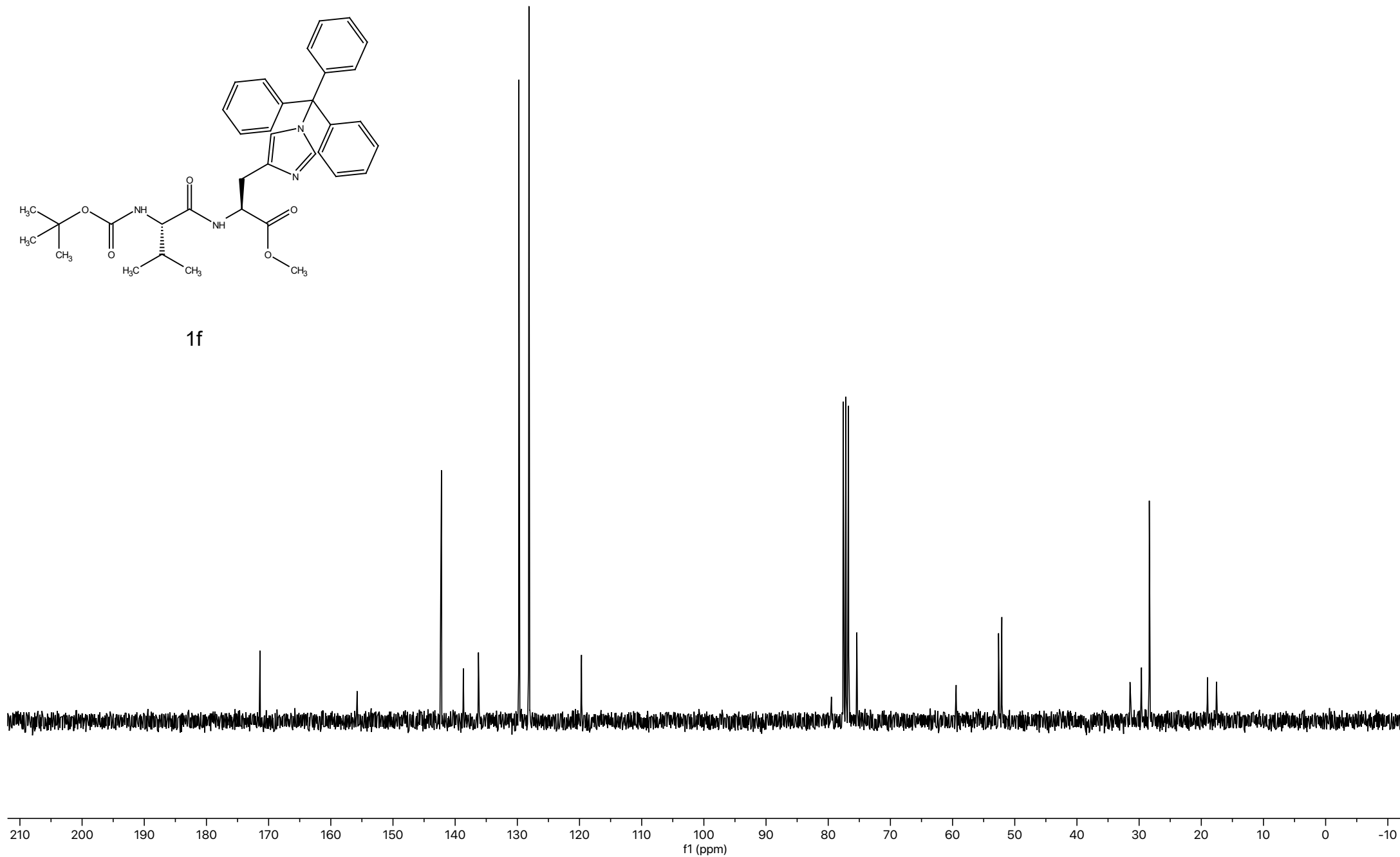
119.6840

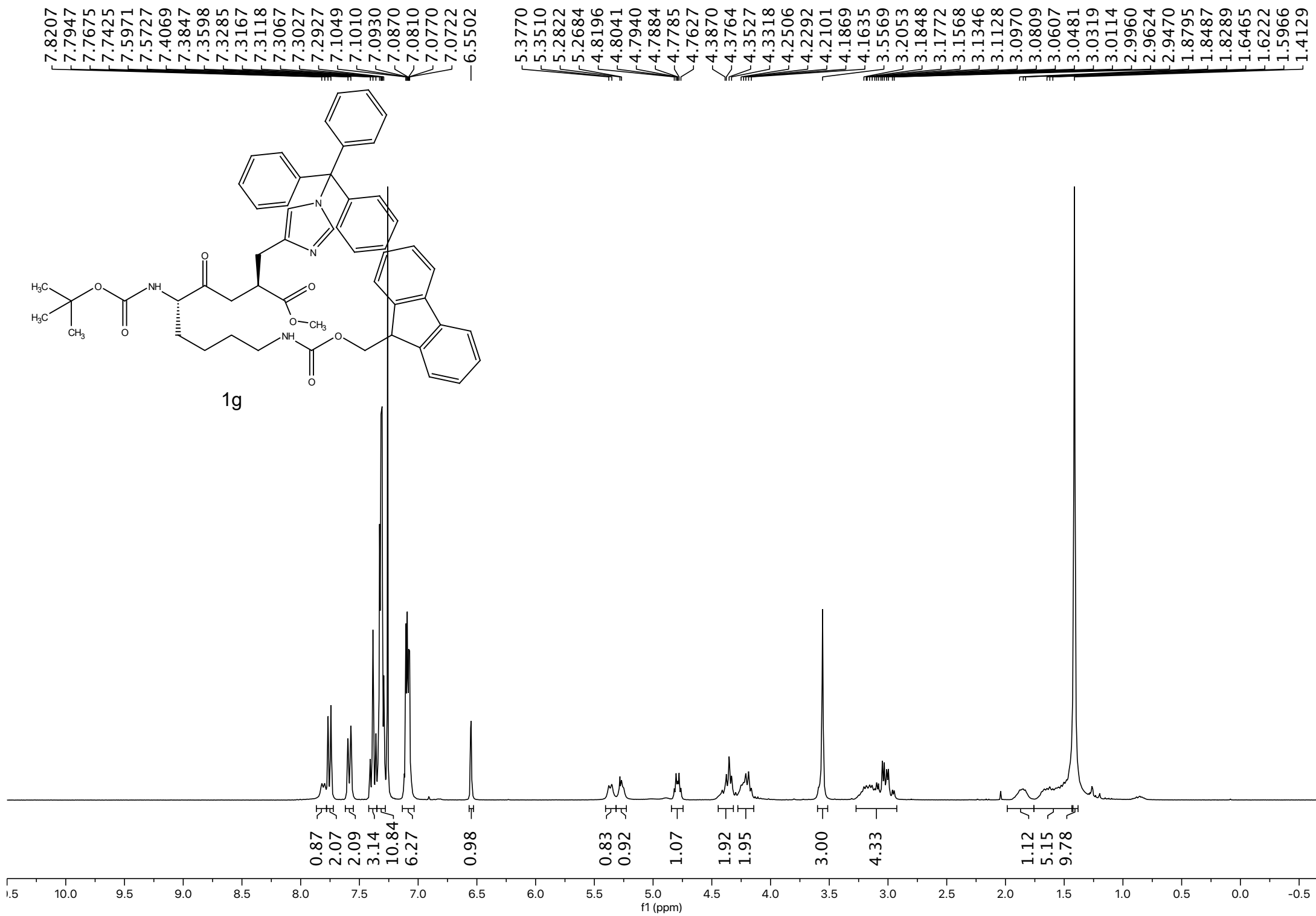
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75.3992

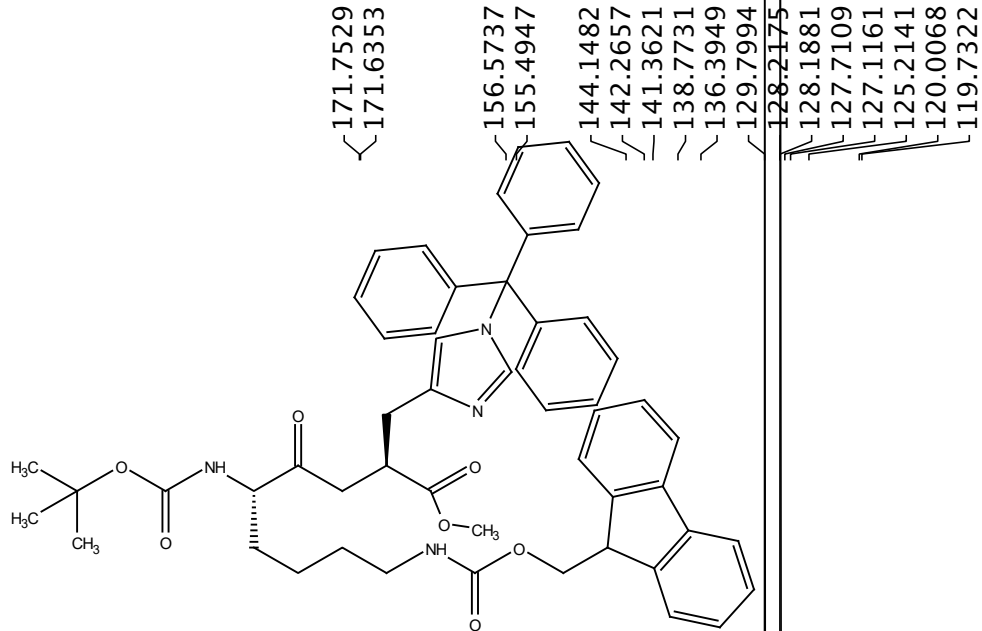
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52.0790

31.4593
29.6256
28.3604

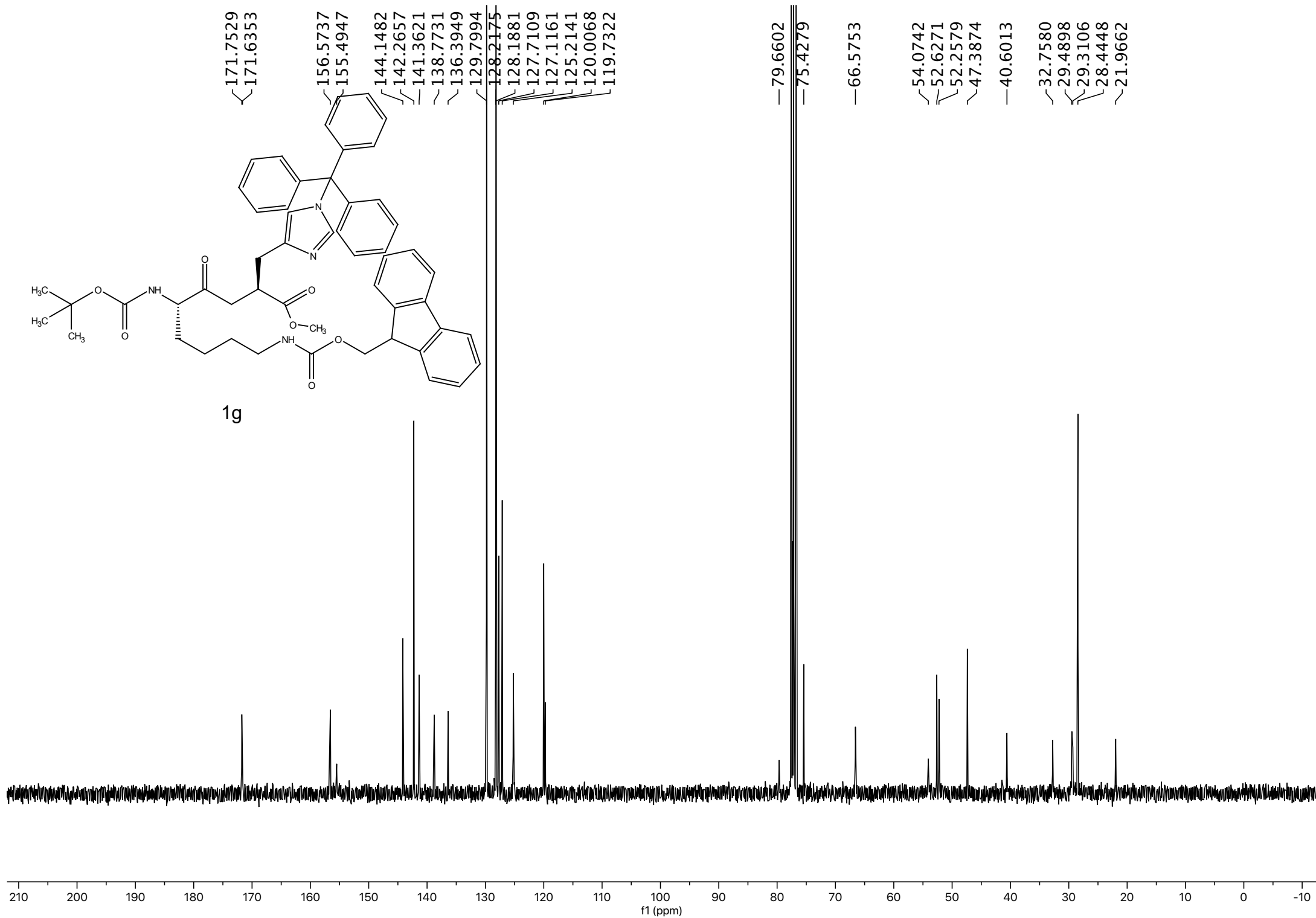
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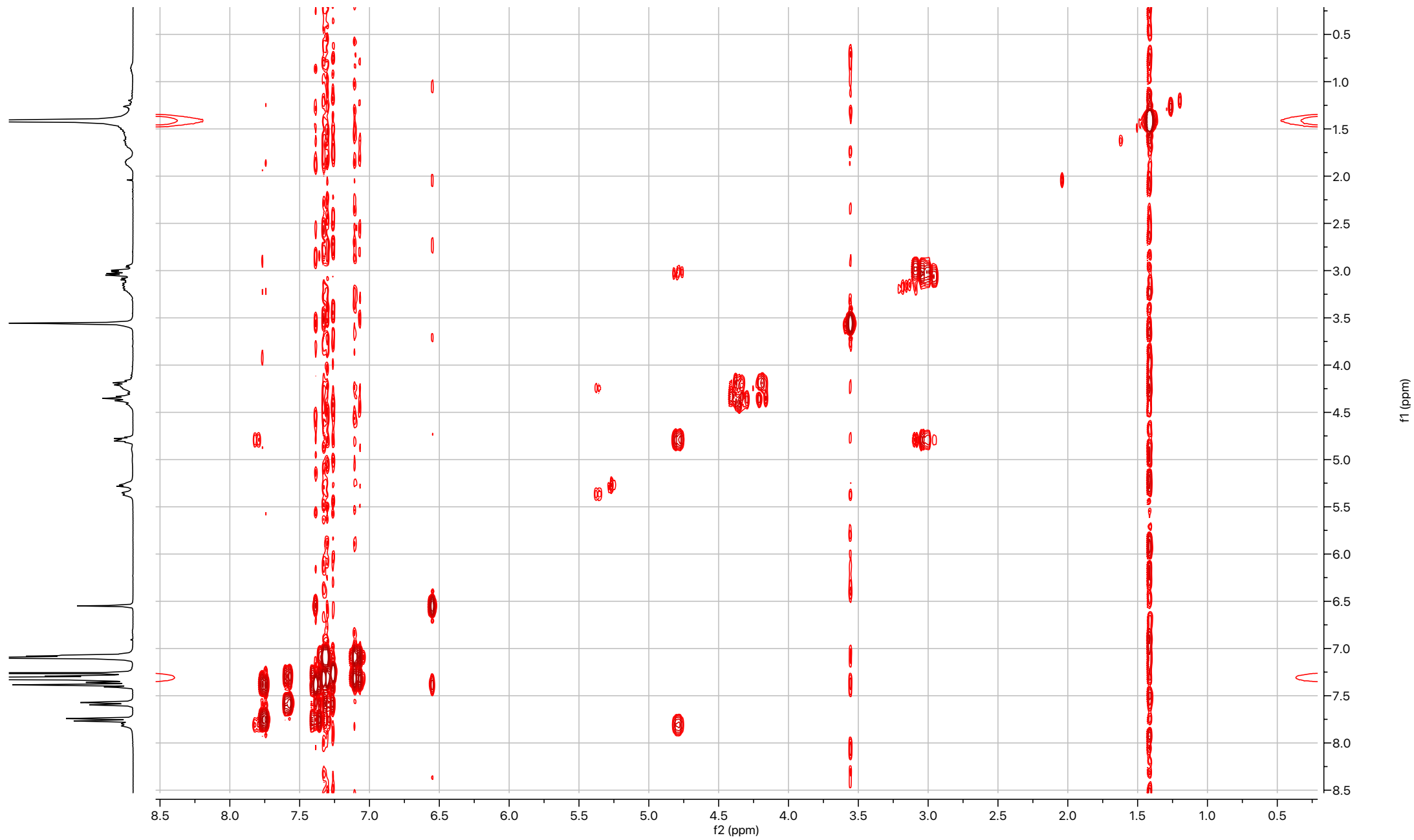


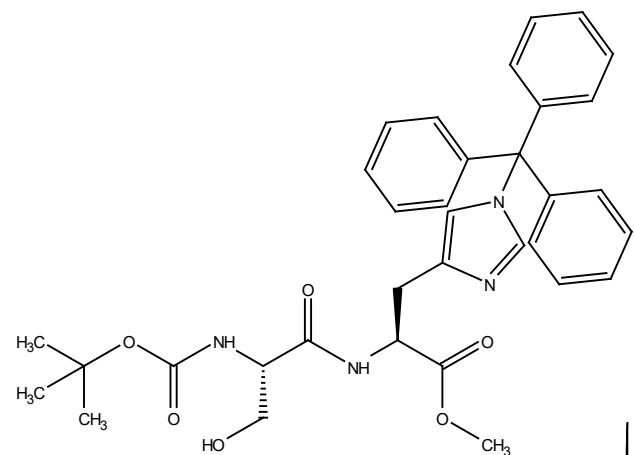


1g

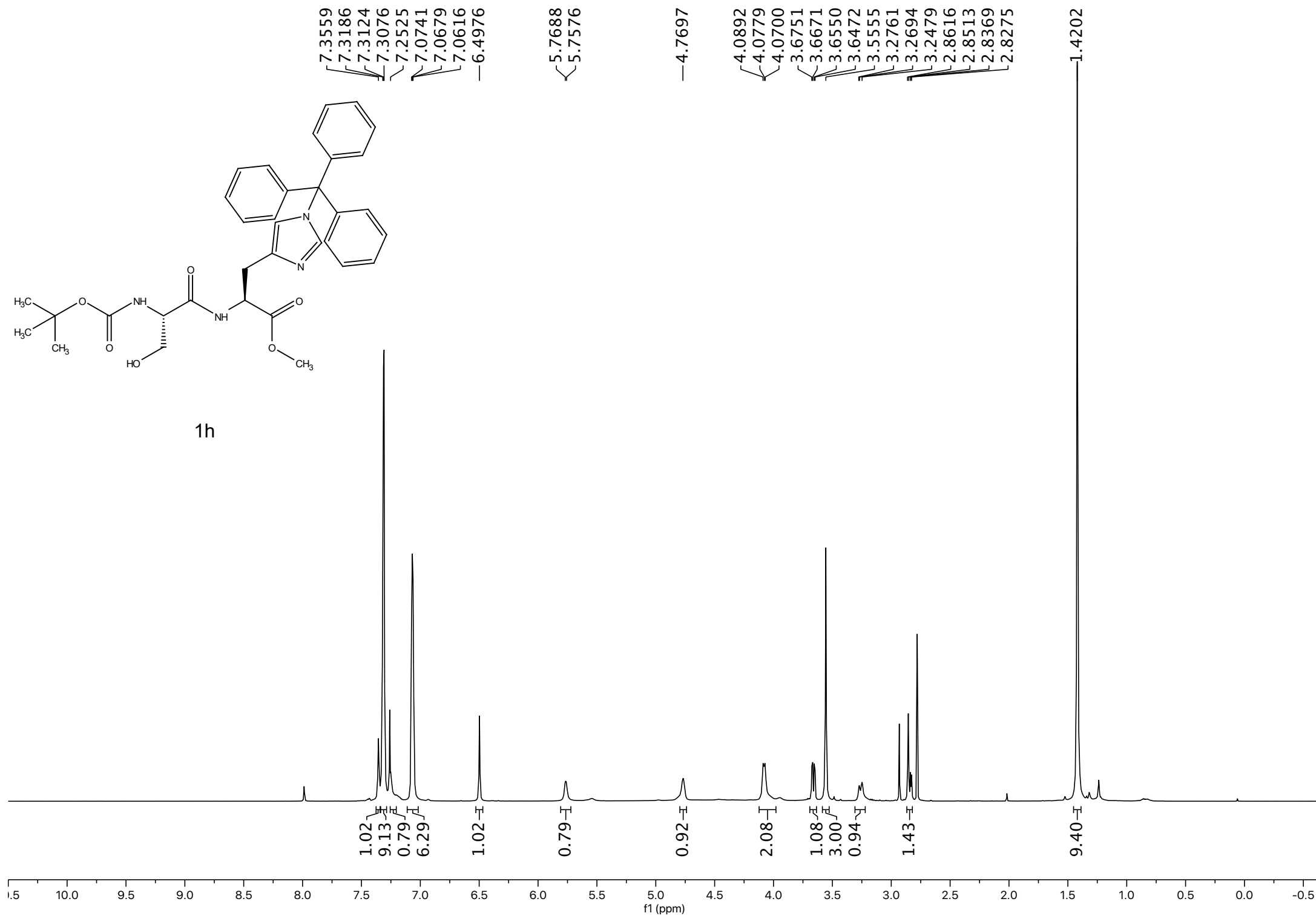


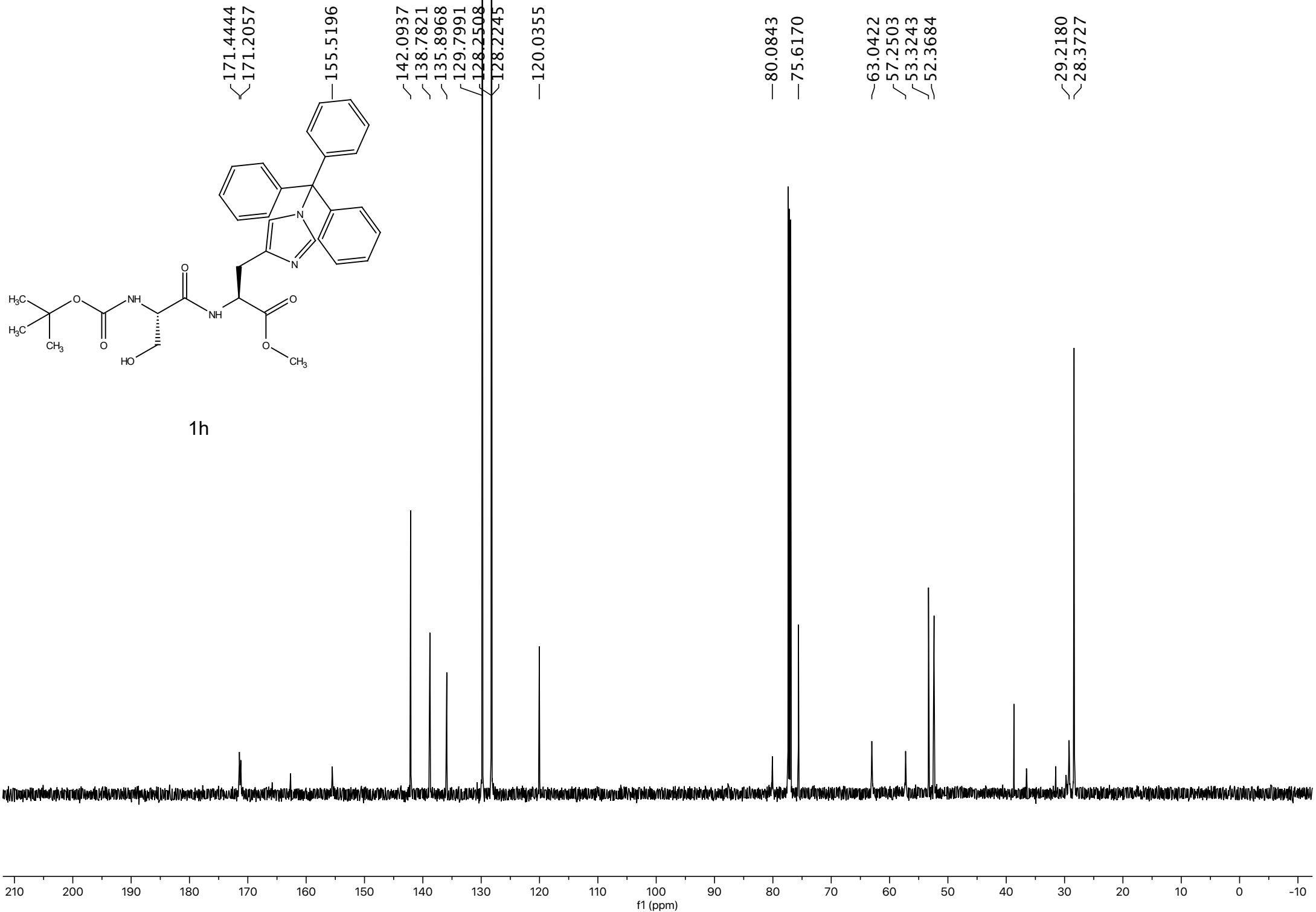
1g COSY

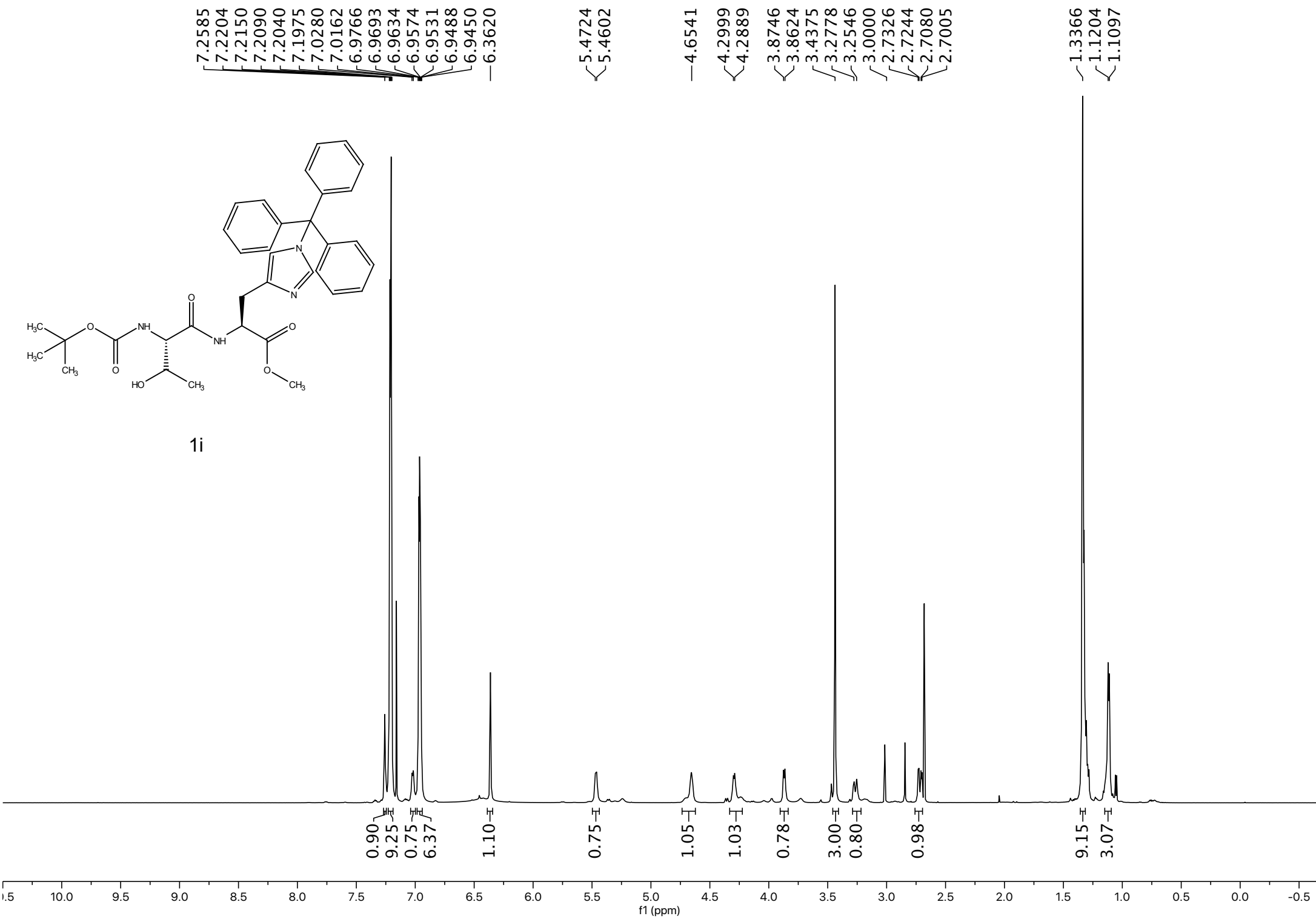


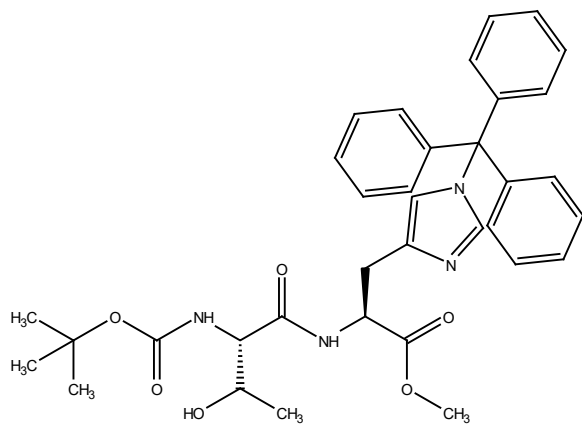


1h









1i

171.4471
171.3037

155.9666

142.0788

138.7923

135.8741

129.7905

128.2152

128.2152

120.0373

80.0237

75.5257

67.2735

61.0205

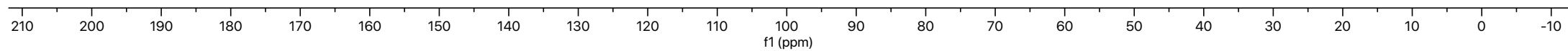
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52.3216

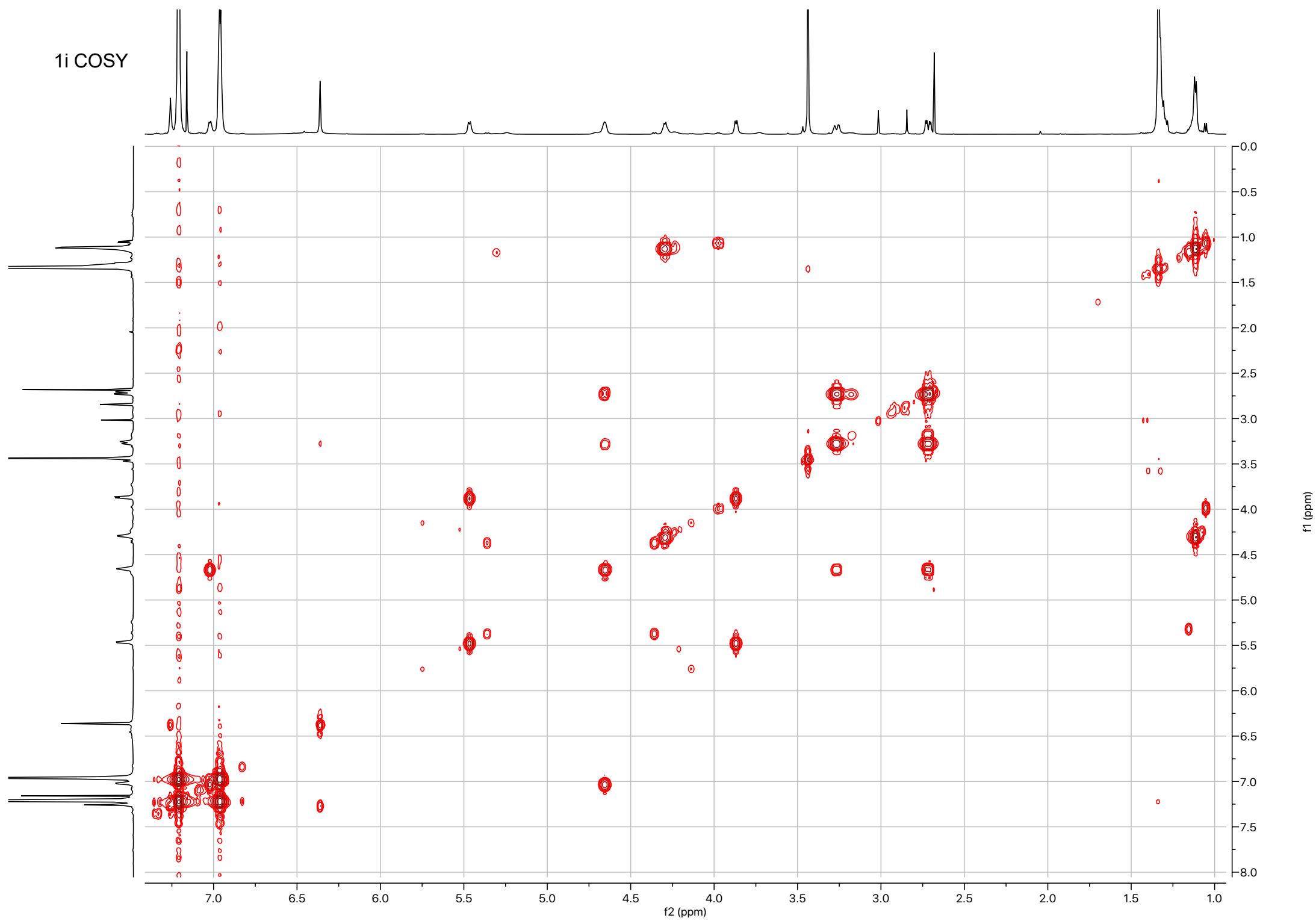
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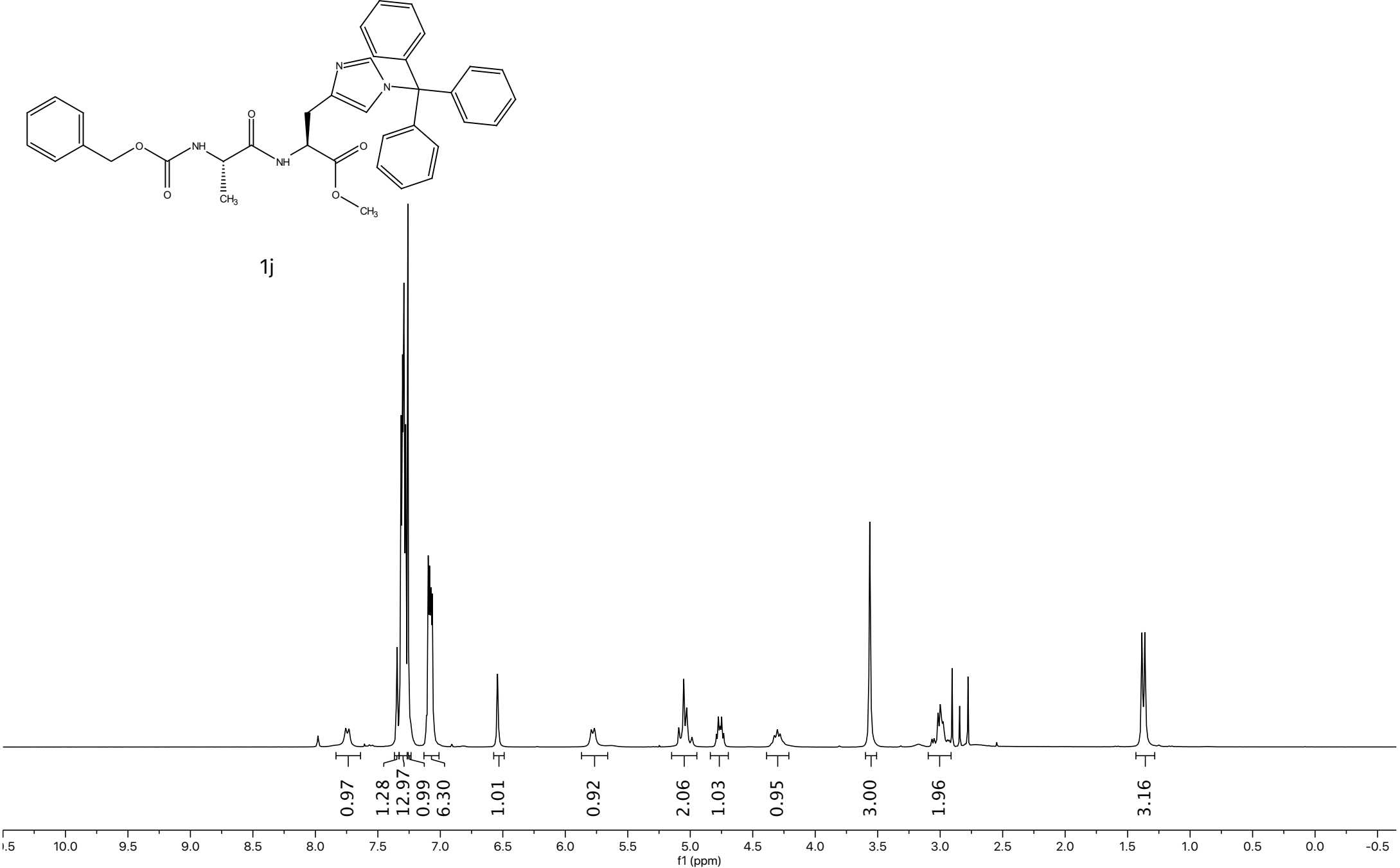
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19.9825

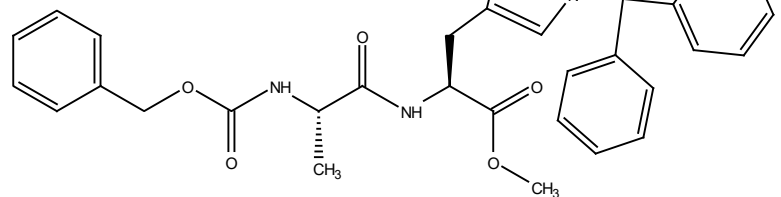


1i COSY





1j



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136.4490
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128.0650
127.9579
127.8757
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66.6201

52.5383

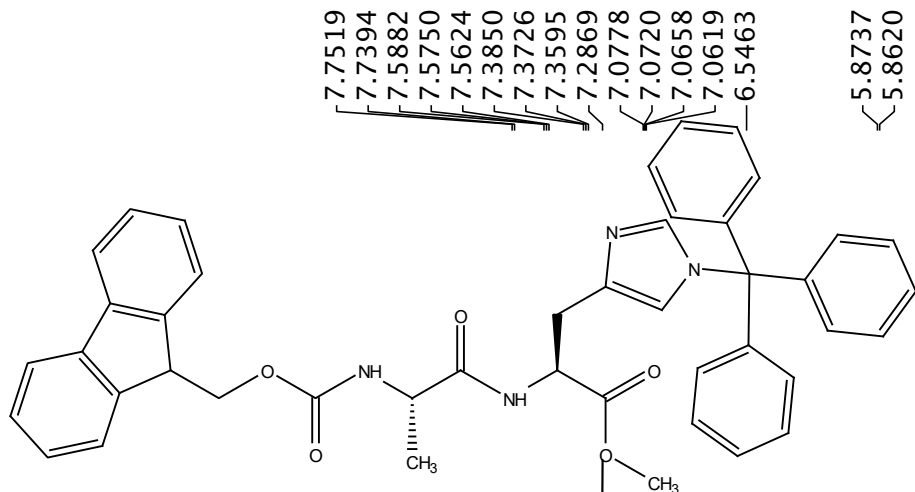
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29.5017

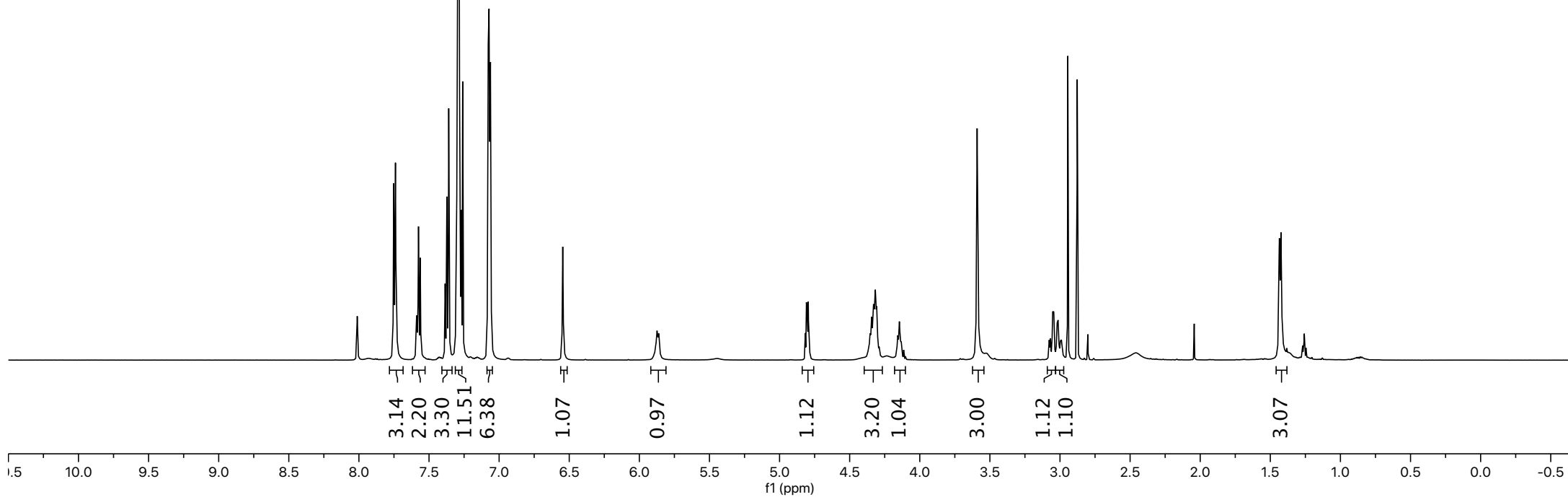
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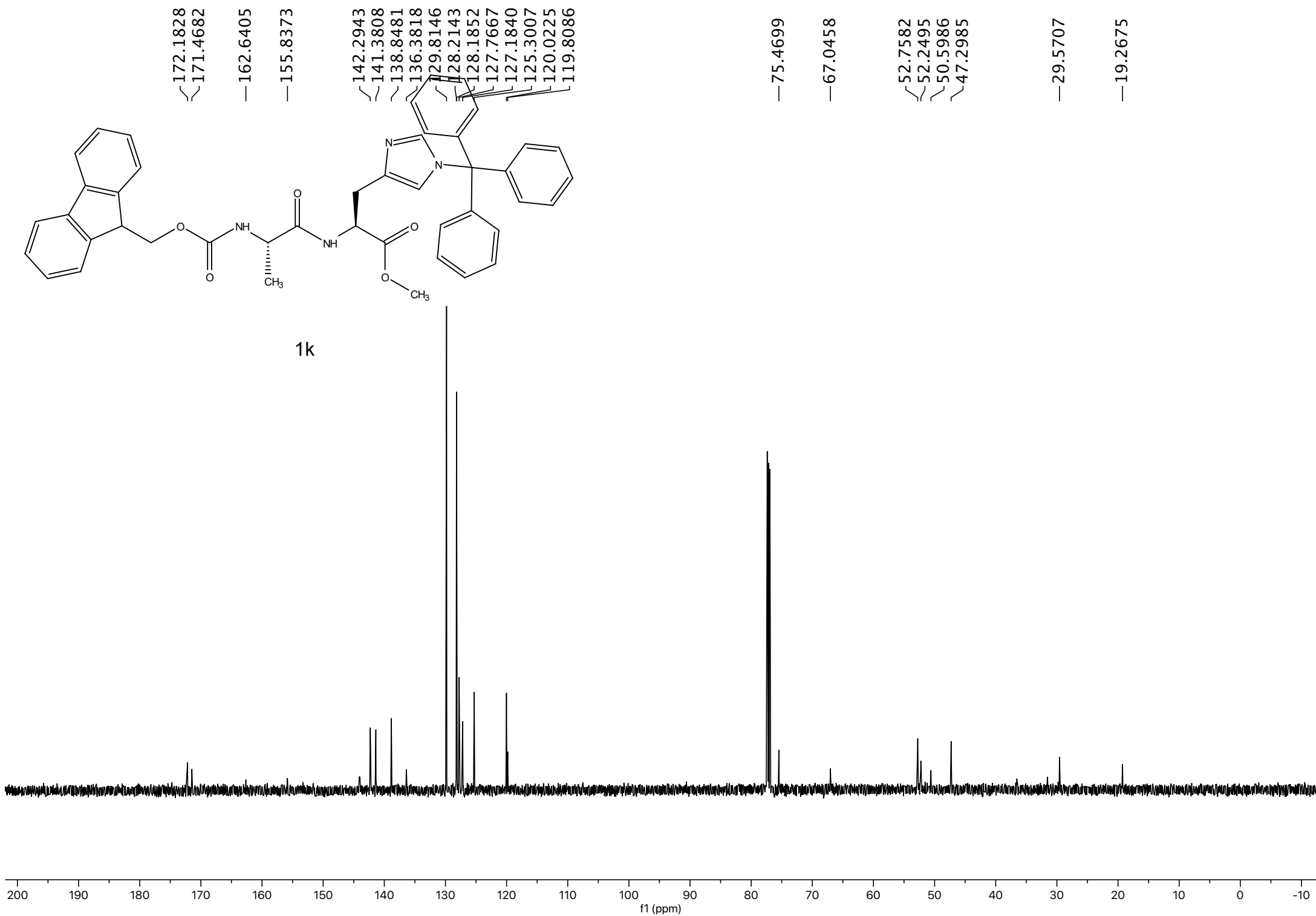
f1 (ppm)

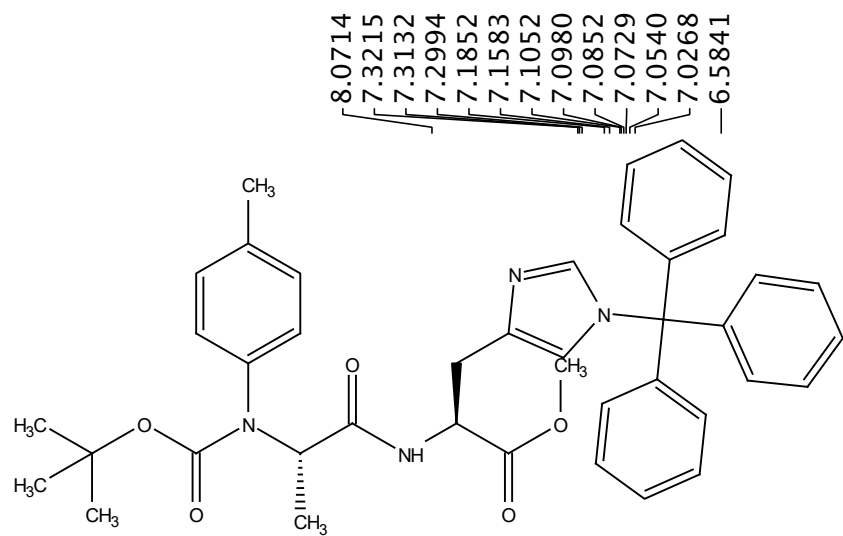


1k

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7.5882
7.5750
7.5624
7.3850
7.3726
7.3595
7.2869
7.0778
7.0720
7.0658
7.0619
6.5463
5.8737
5.8620
4.8161
4.8083
4.8024
4.8008
4.7952
4.7872
4.3170
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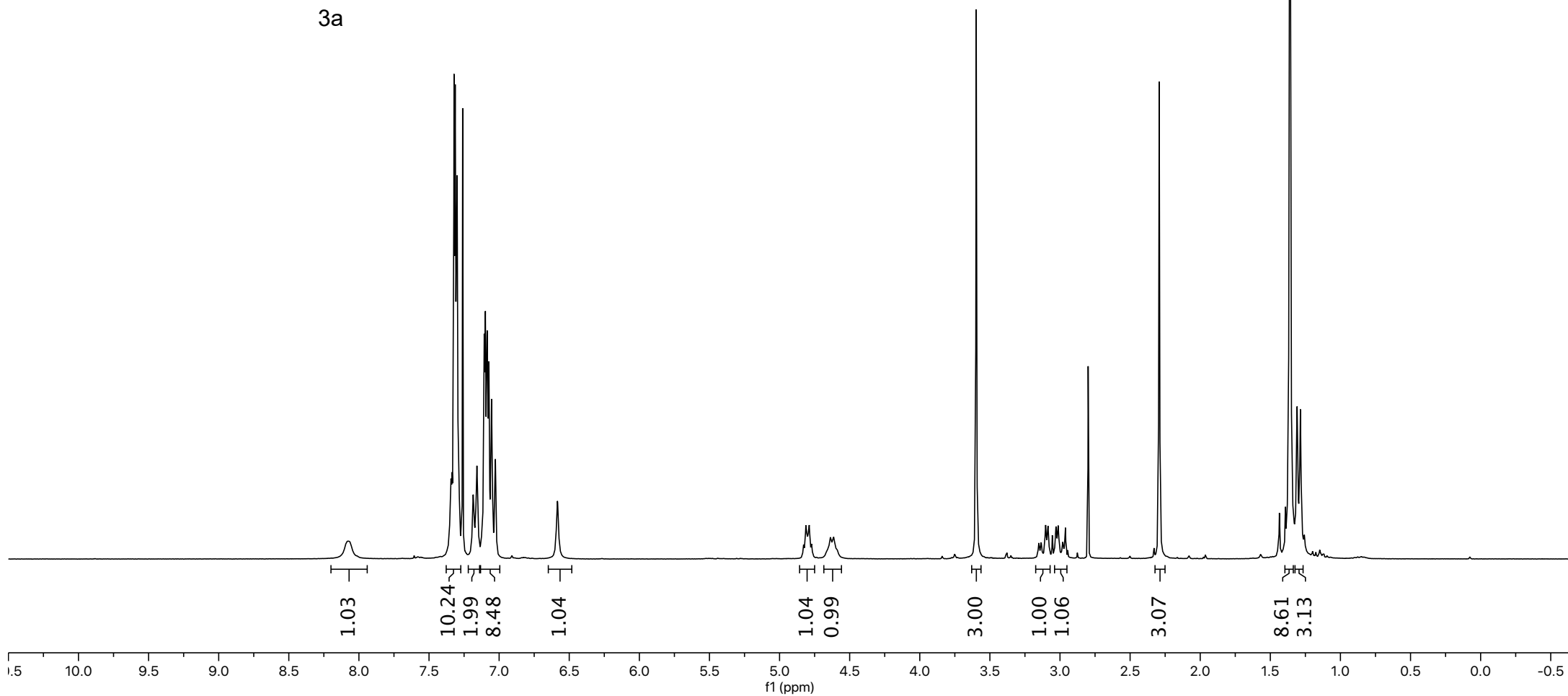
3a

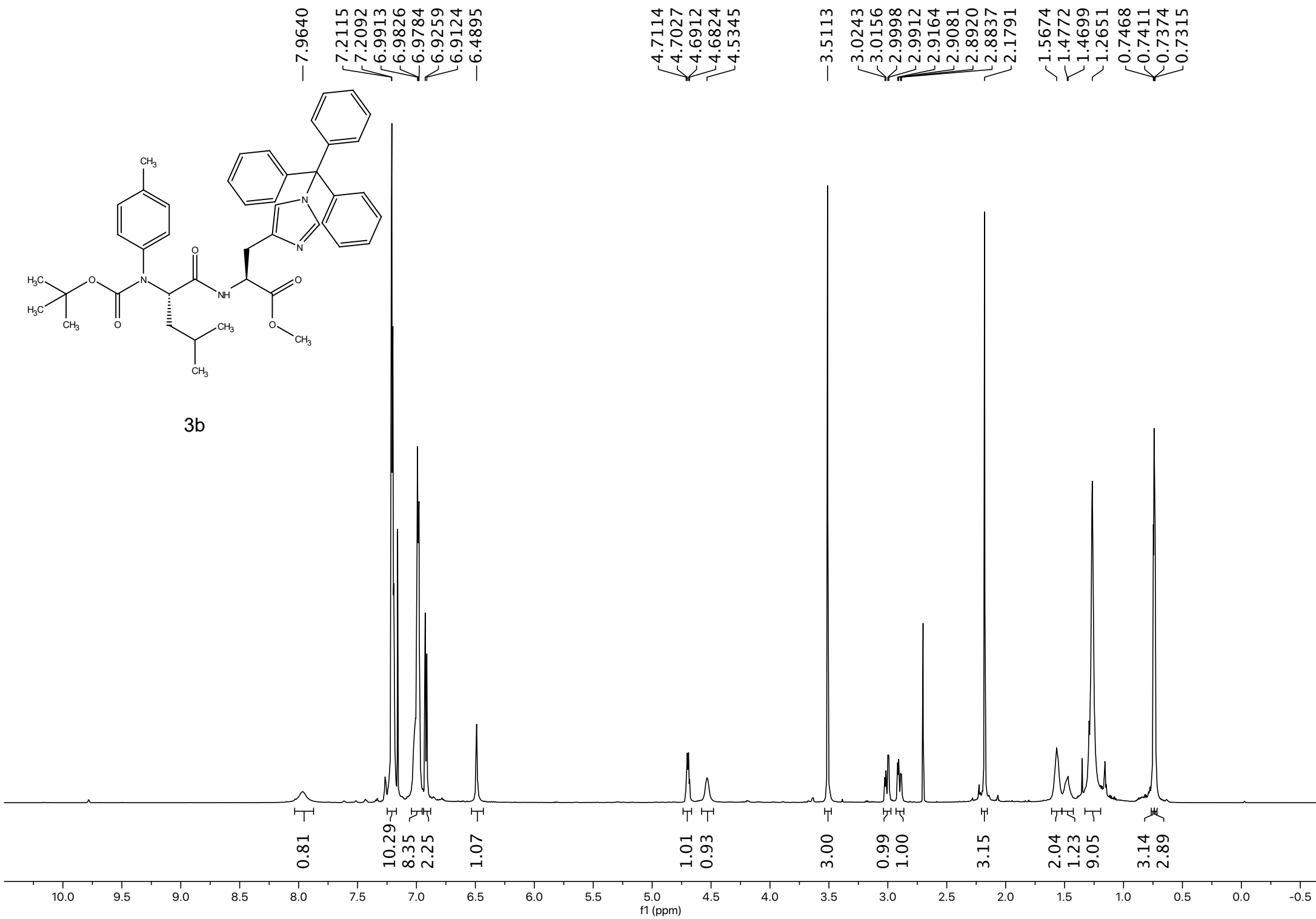
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7.1052
7.0980
7.0852
7.0729
7.0540
7.0268
6.5841

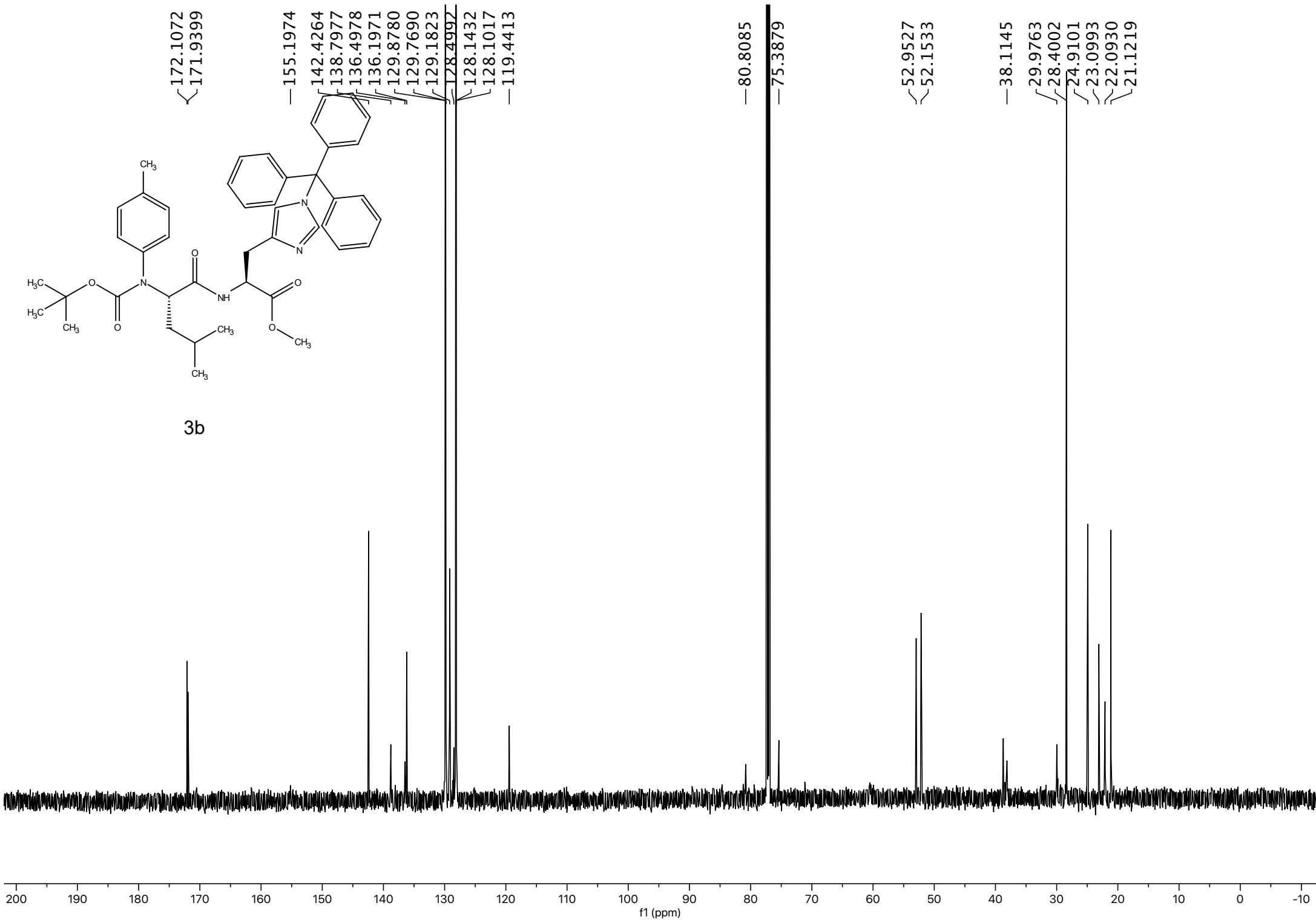
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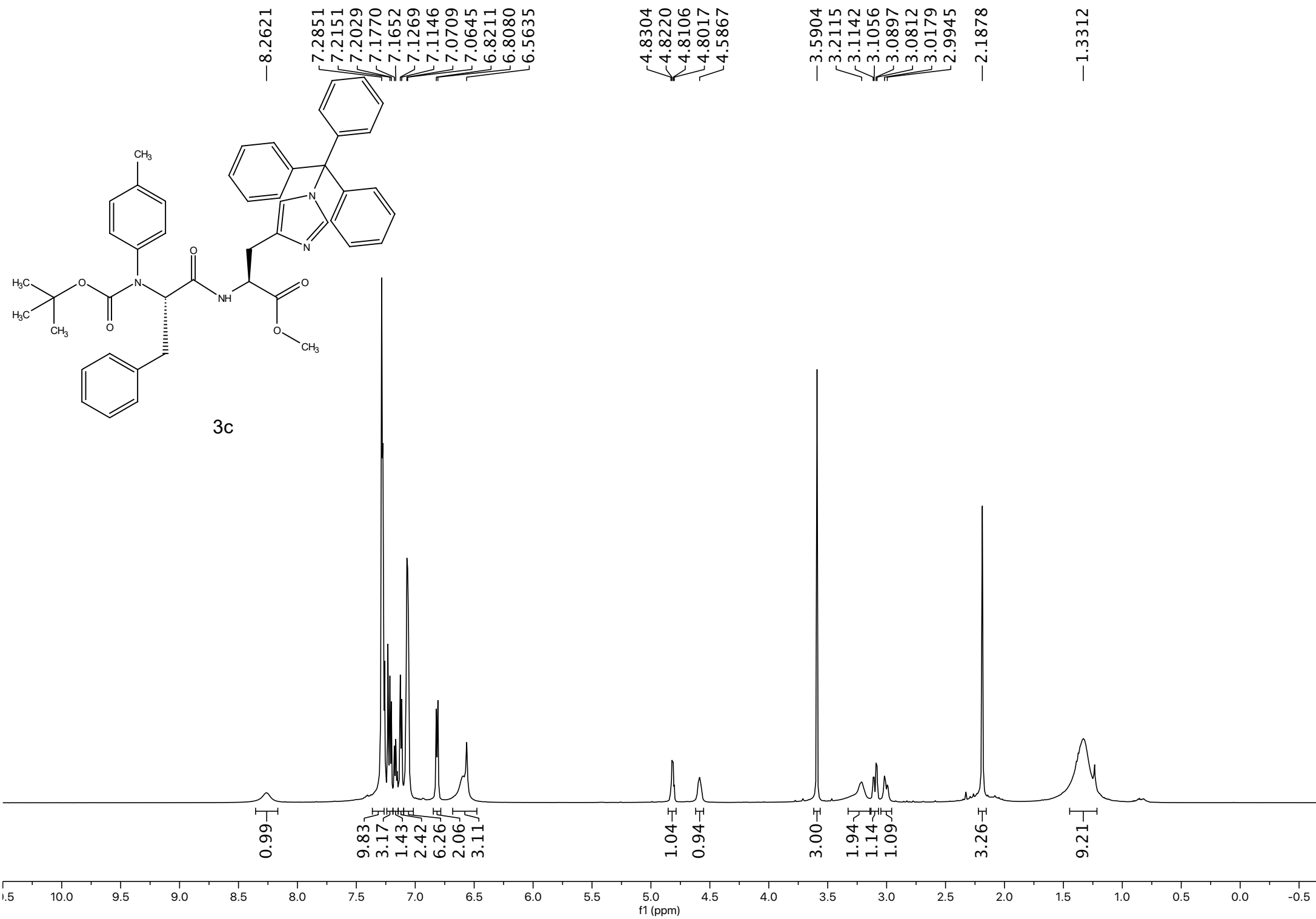
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2.2923

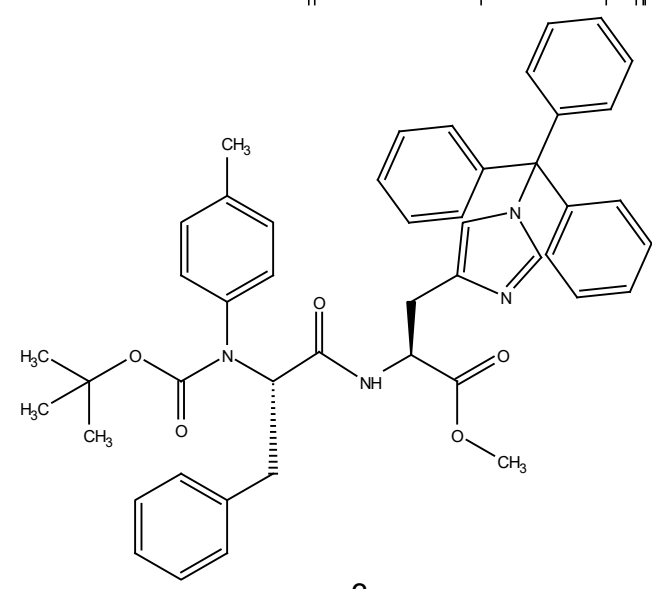
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1.2853





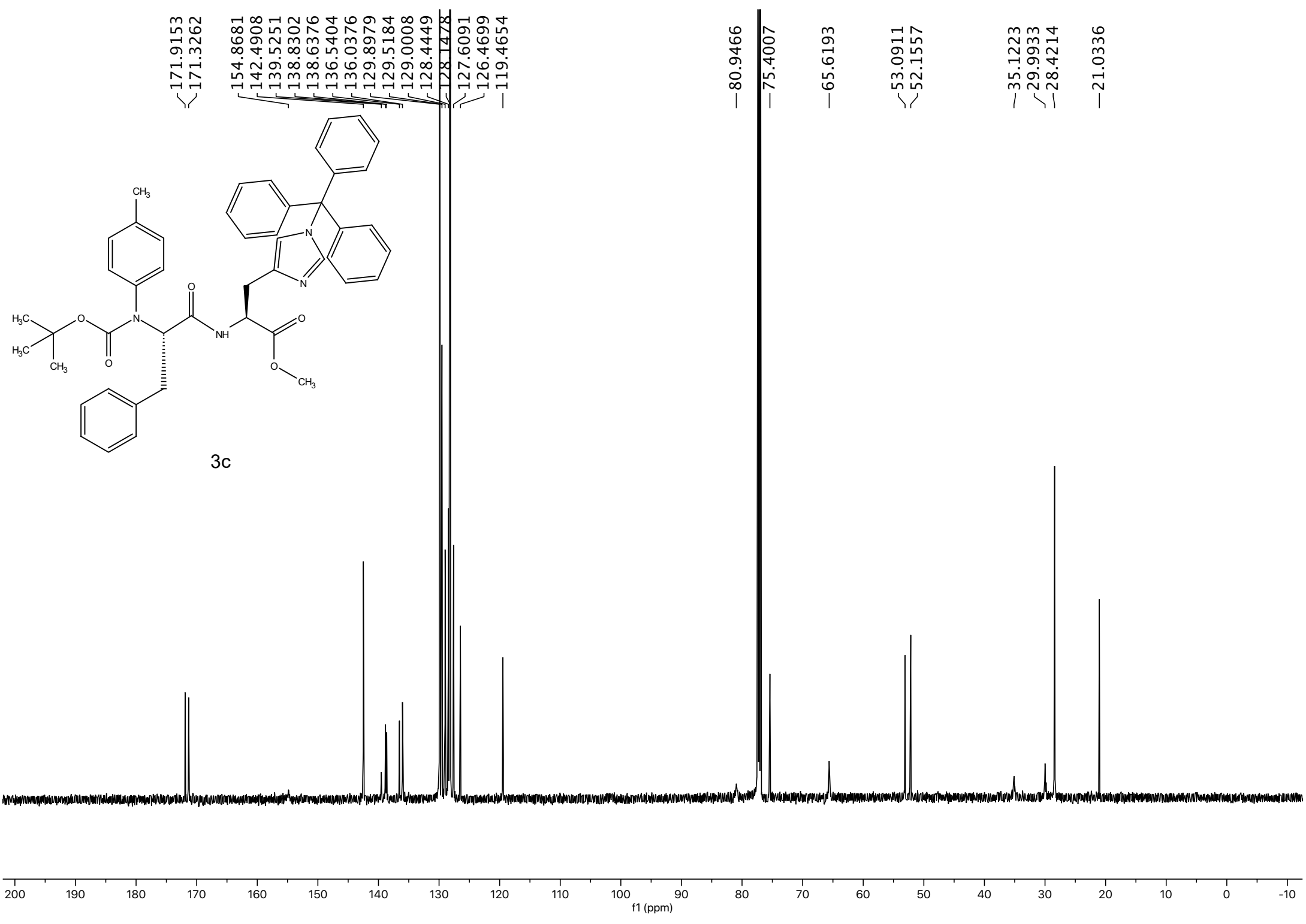


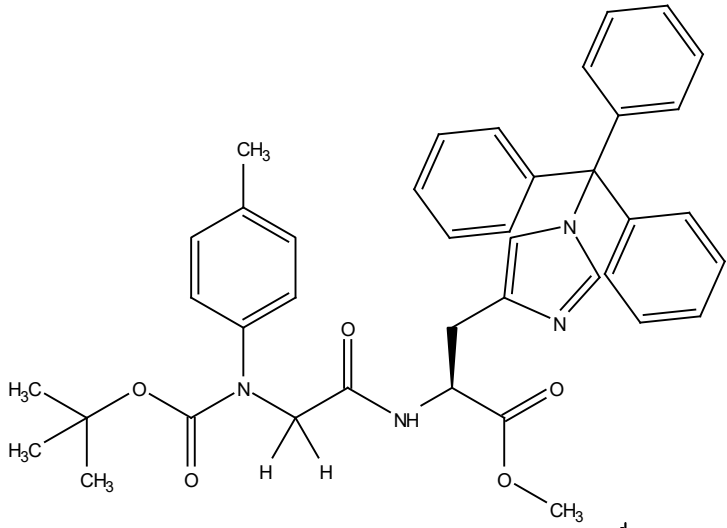




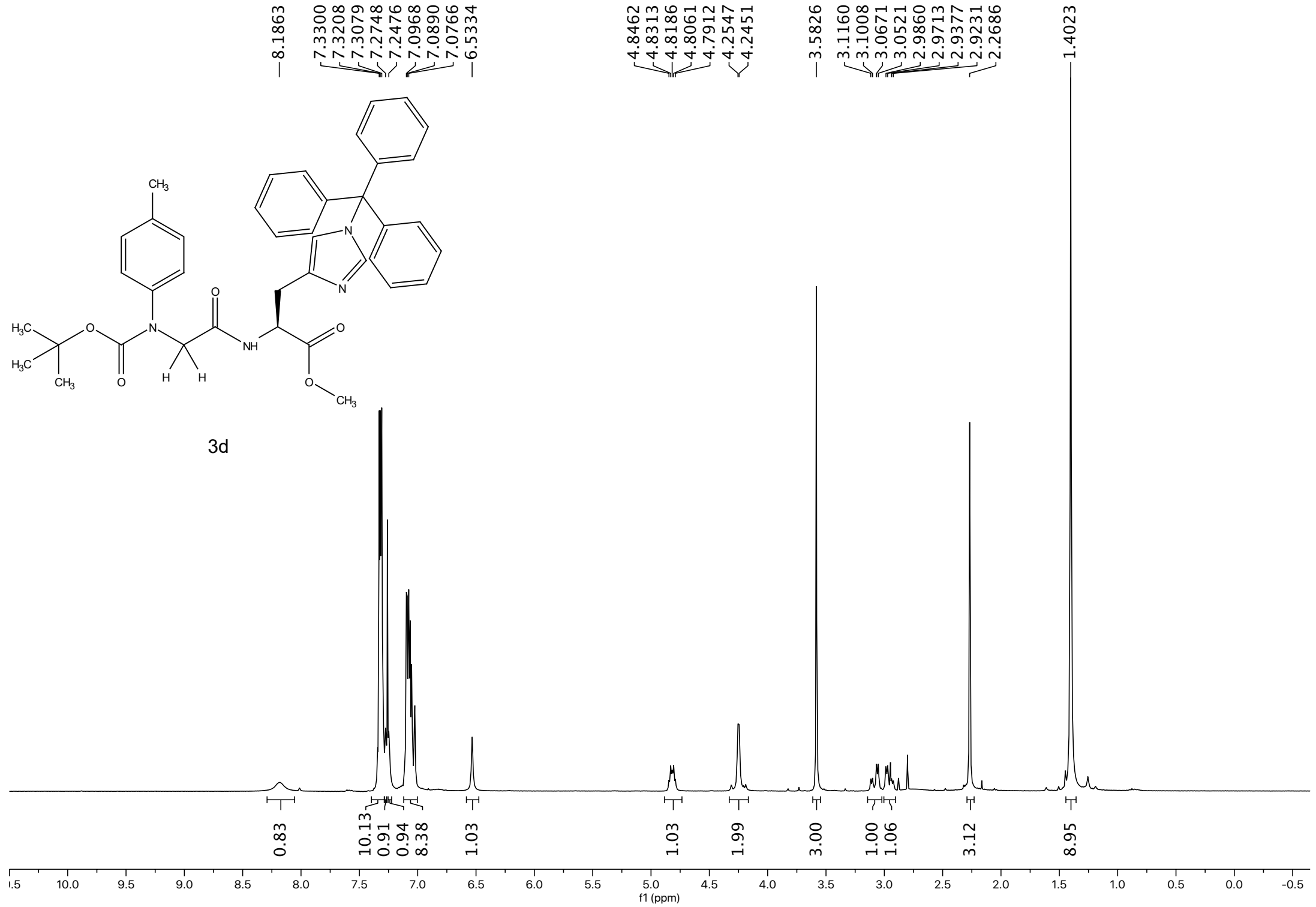
3c

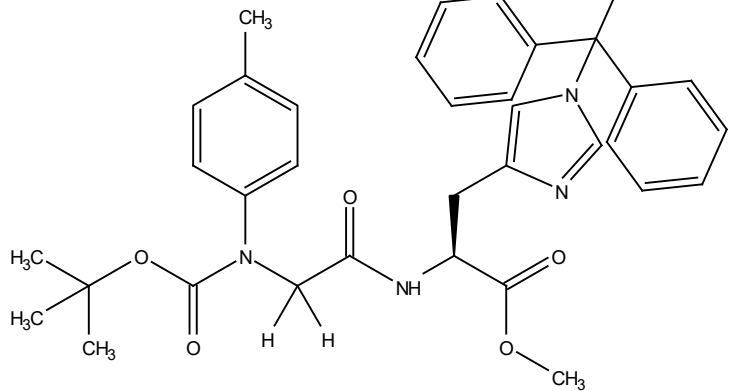
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- 171.3262
- 154.8681
- 142.4908
- 139.5251
- 138.8302
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- 129.8979
- 129.5184
- 129.0008
- 128.4449
- 128.1478
- 127.6091
- 126.4699
- 119.4654
- 80.9466
- 75.4007
- 65.6193
- 53.0911
- 52.1557
- 35.1223
- 29.9933
- 28.4214
- 21.0336



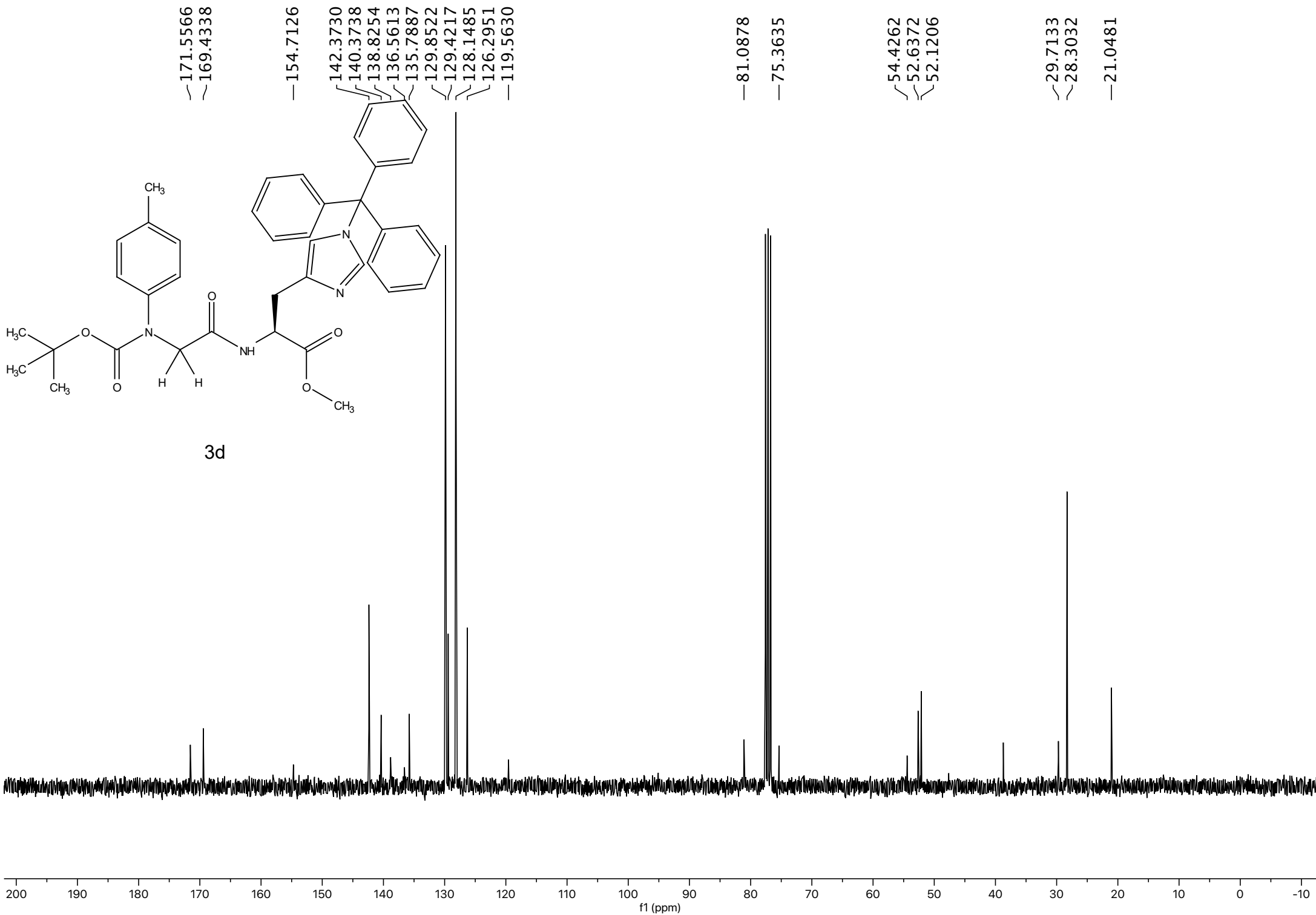


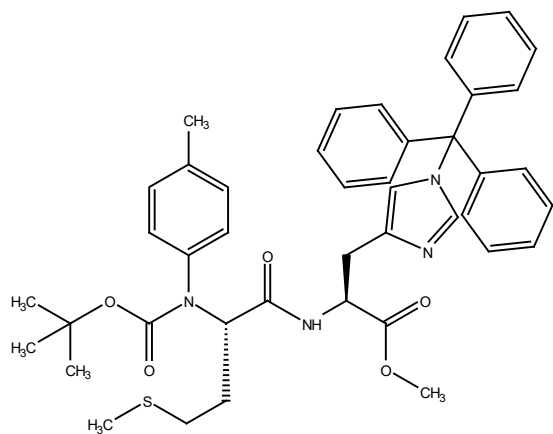
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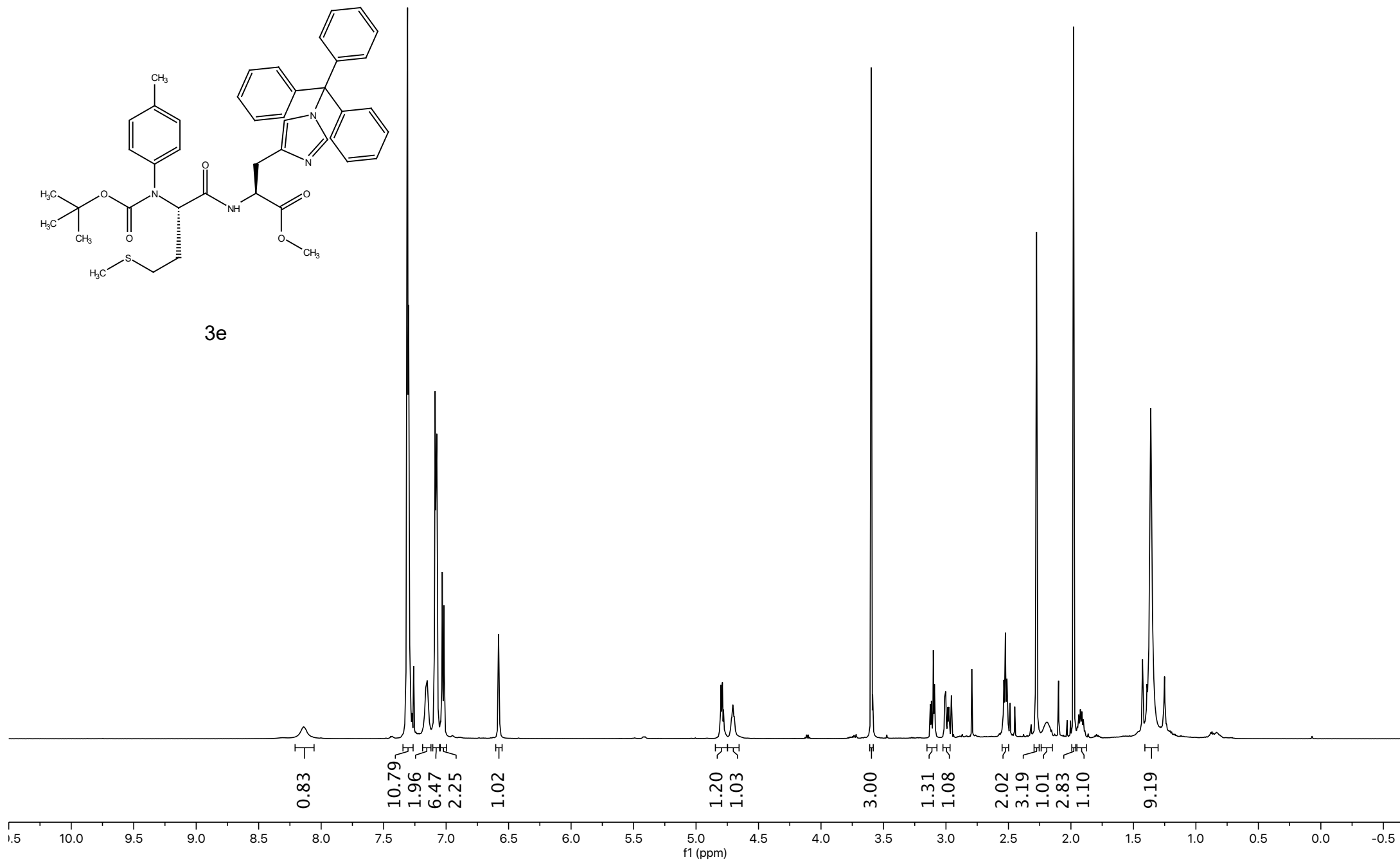


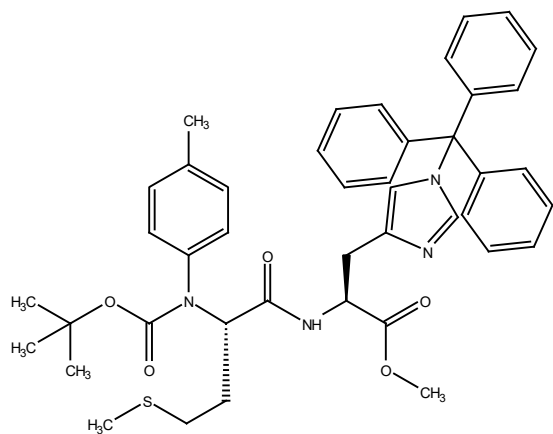
3d





3e





3e

171.7861
171.4463

154.9895

142.3474

138.7540

138.0149

136.3864

134.9852

129.8368

129.2680

128.1316

128.0258

119.4818

81.0522

75.4005

61.0398

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52.1466

31.1156

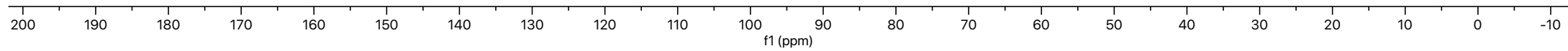
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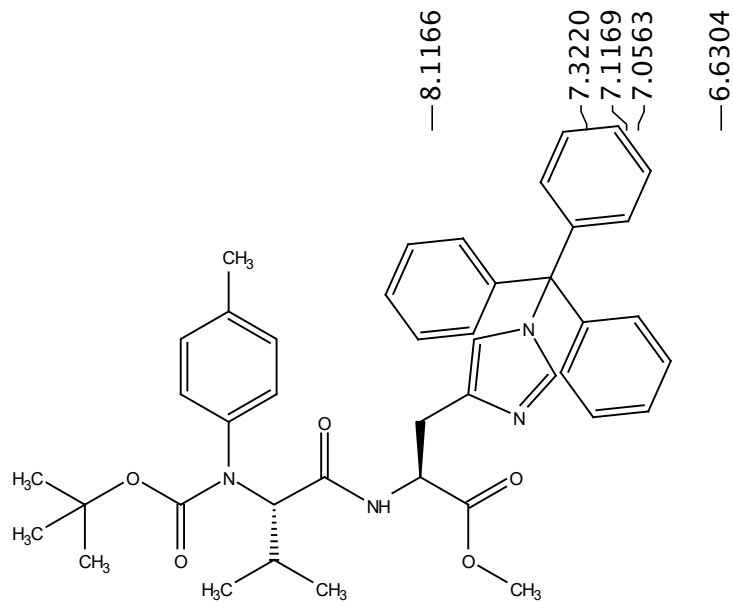
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28.3537

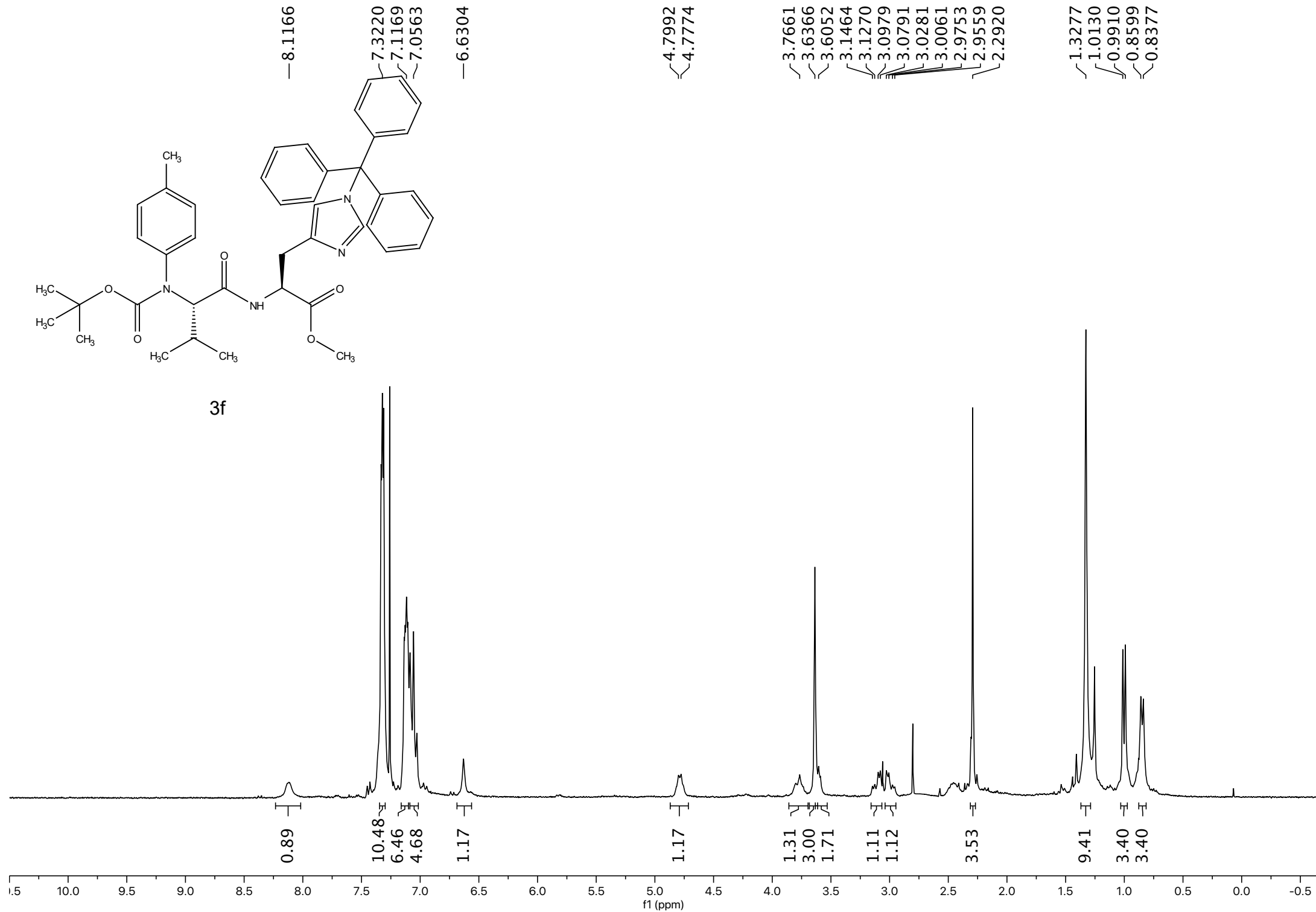
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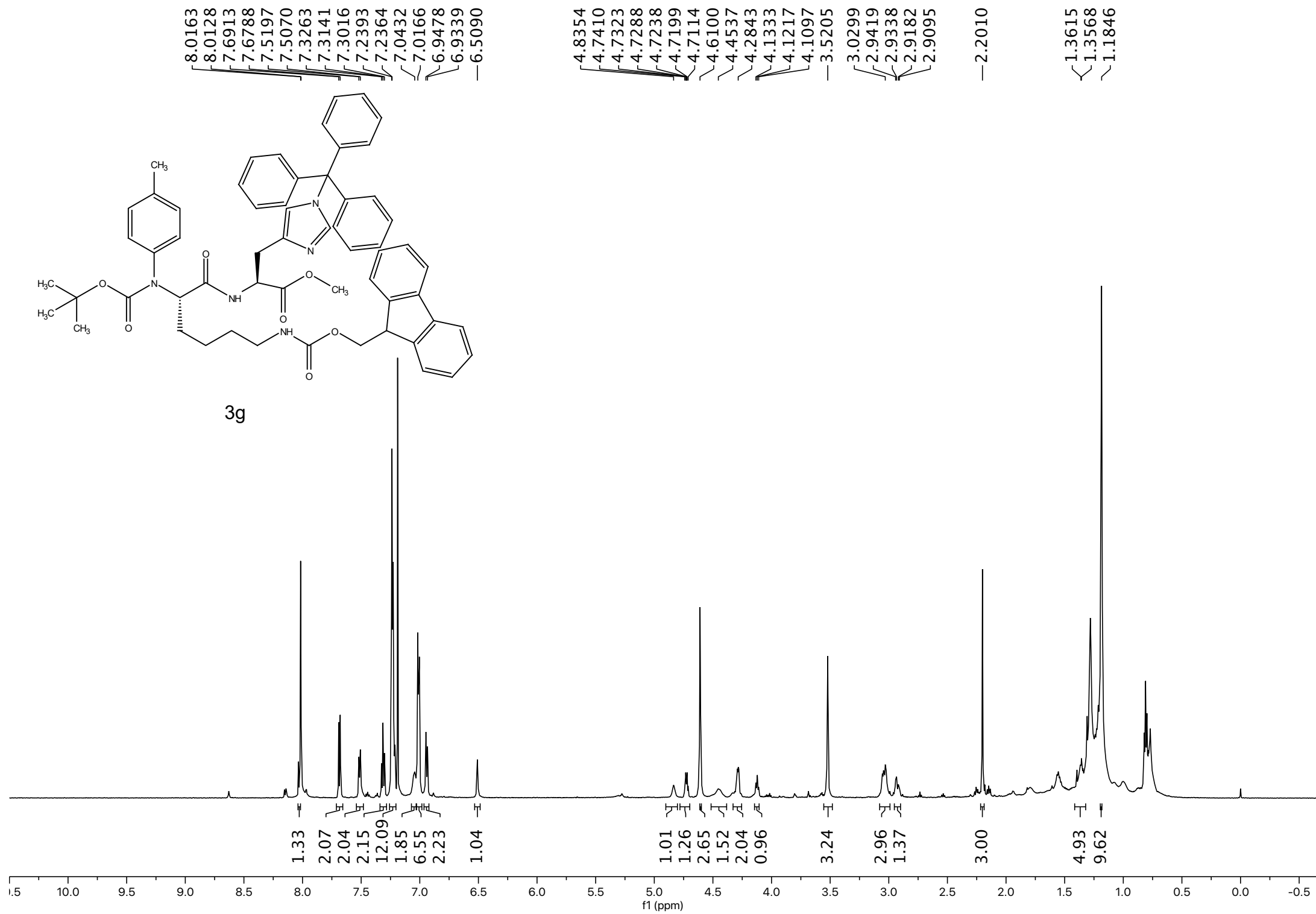
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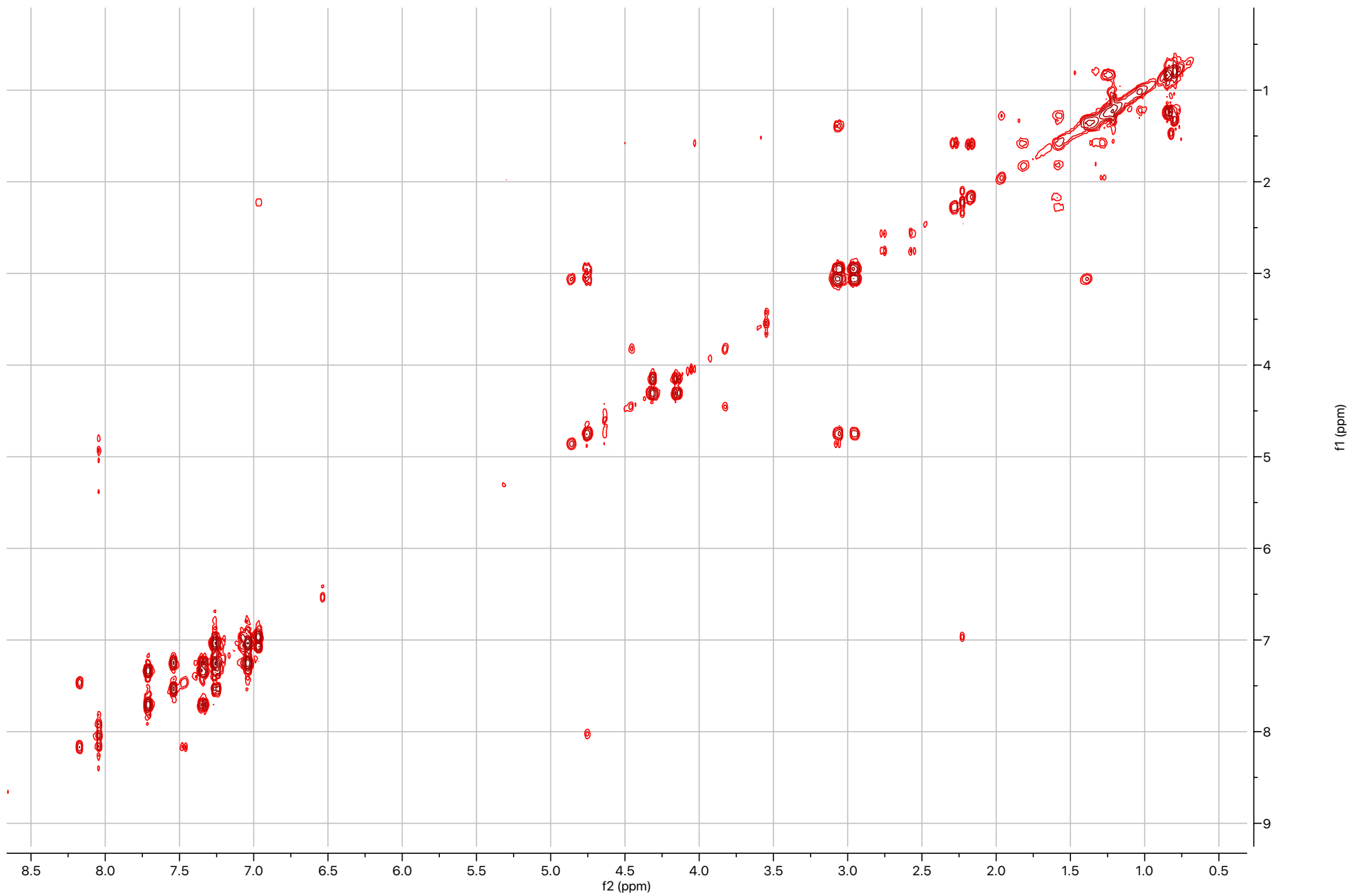
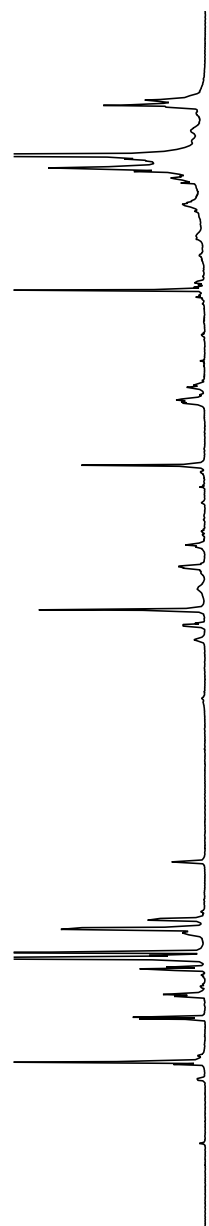
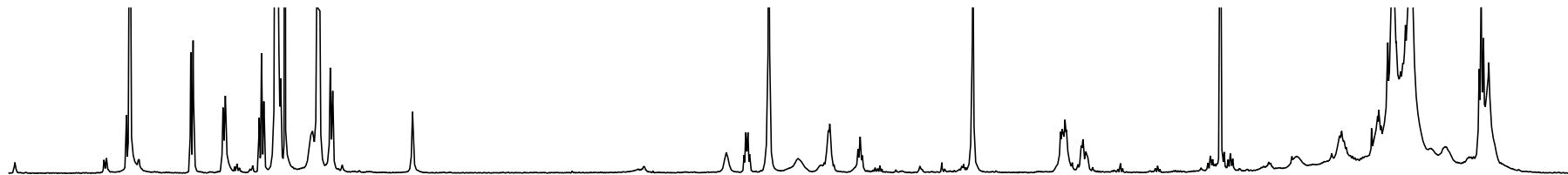


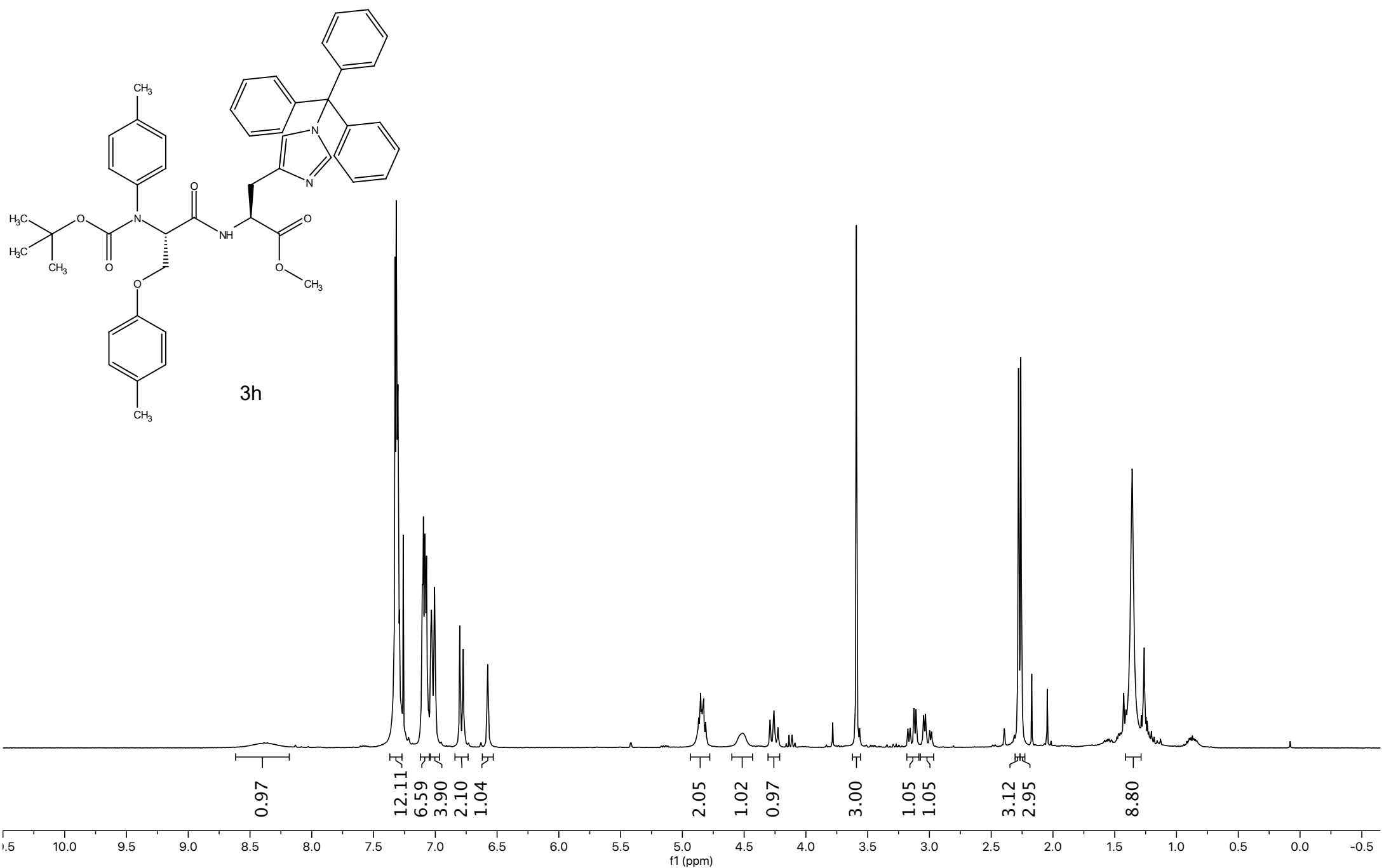
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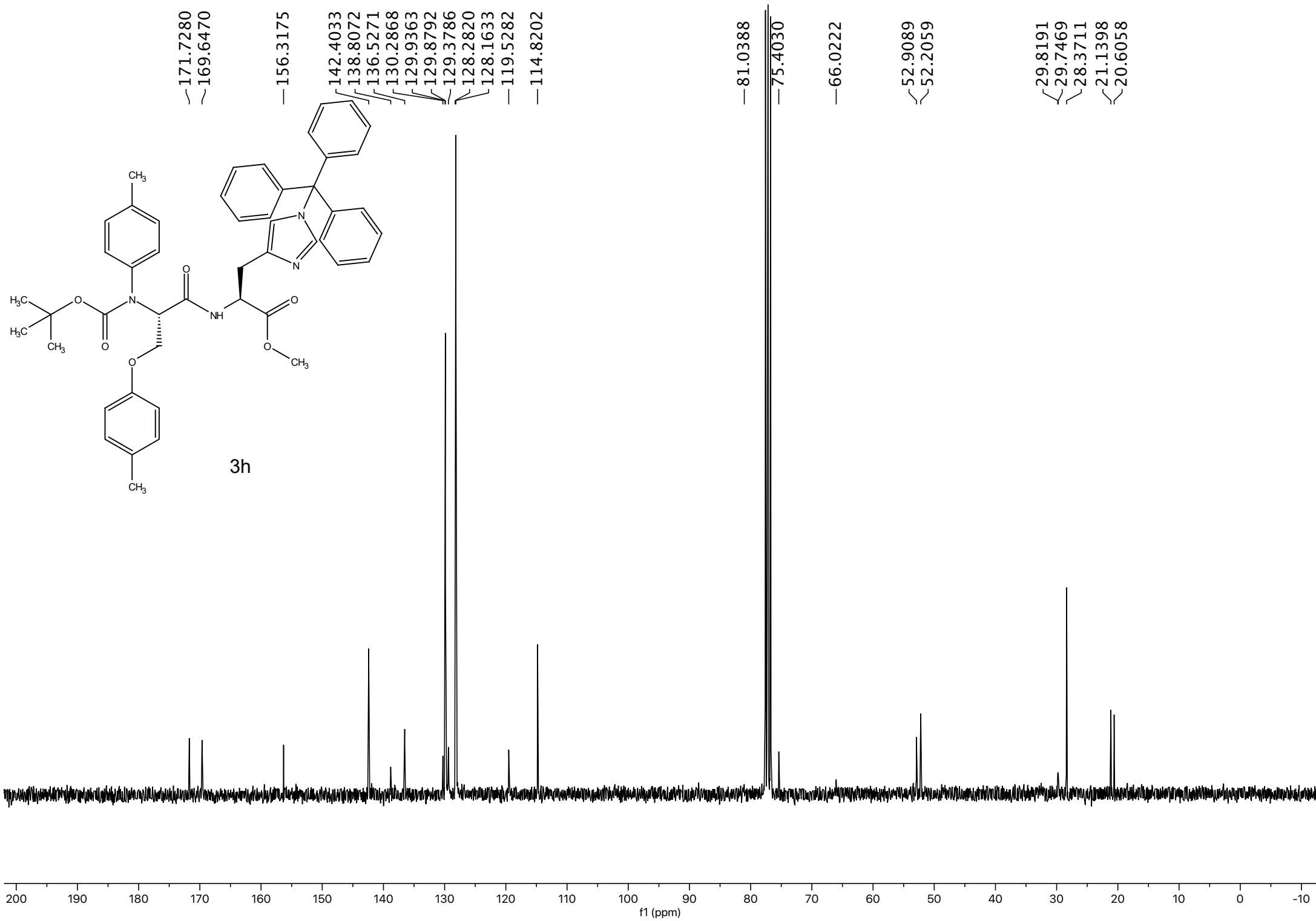


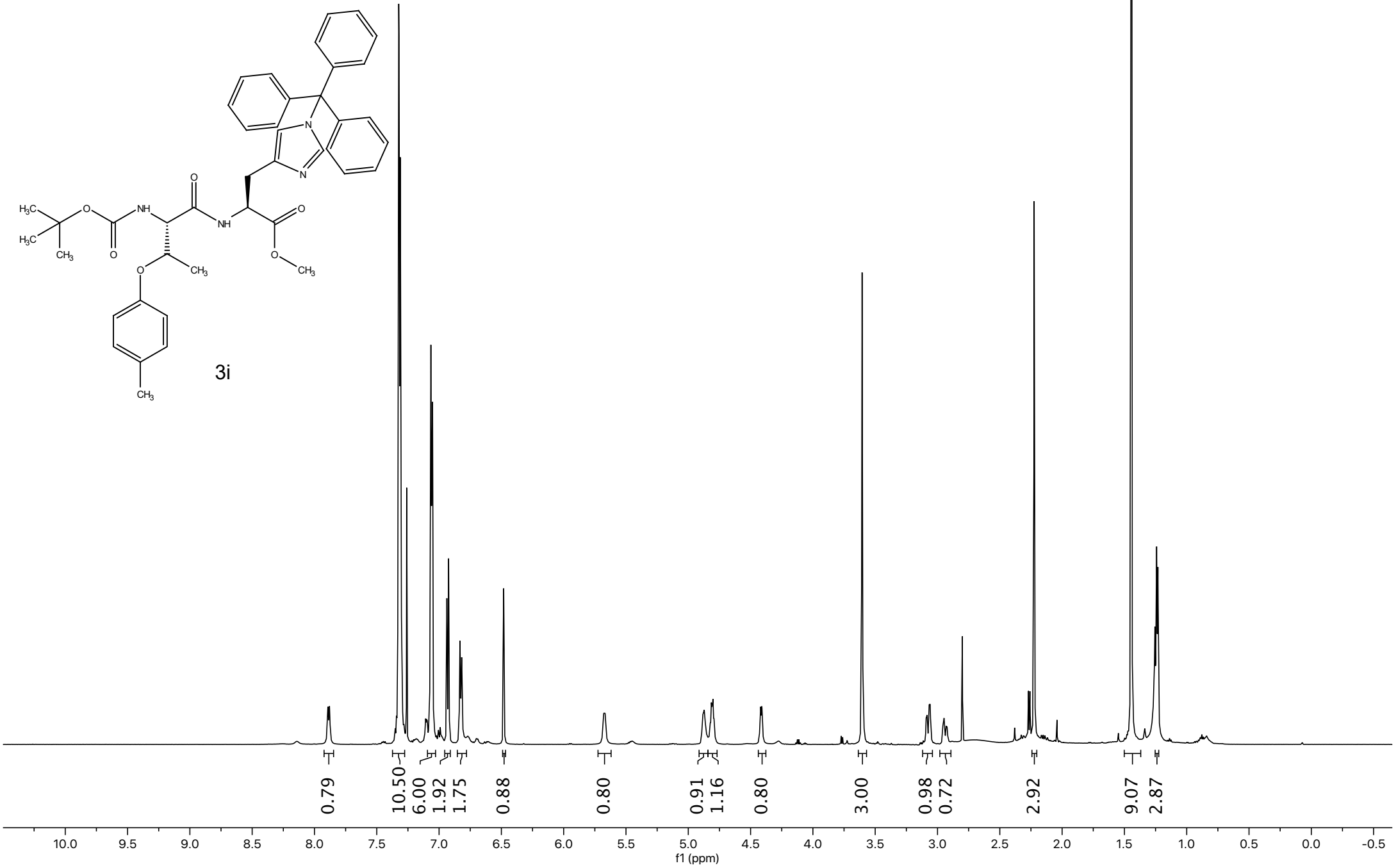


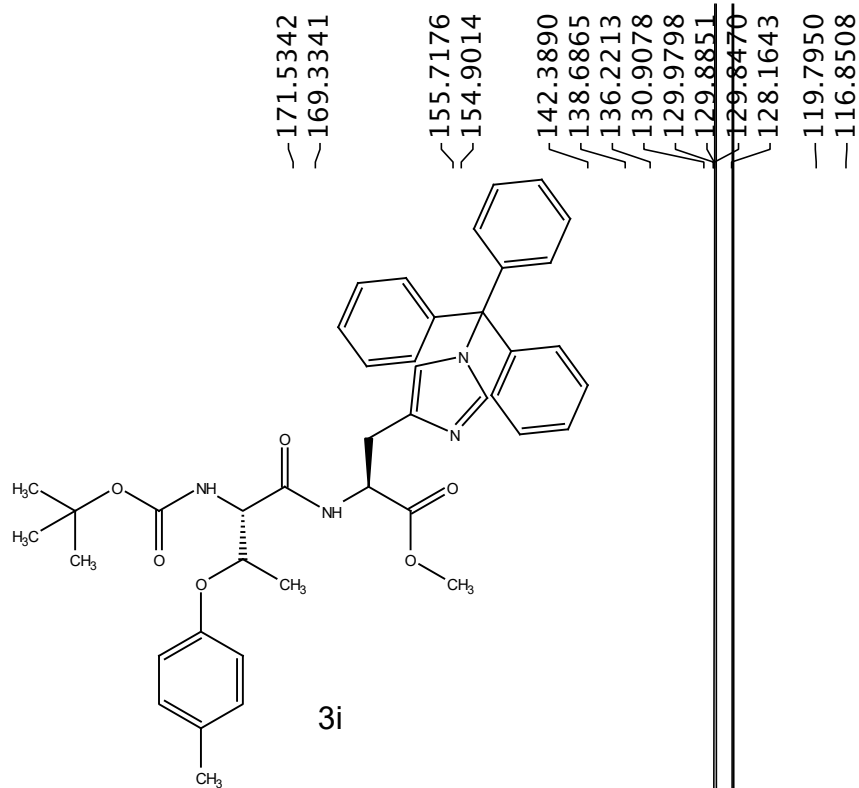
3g COSY



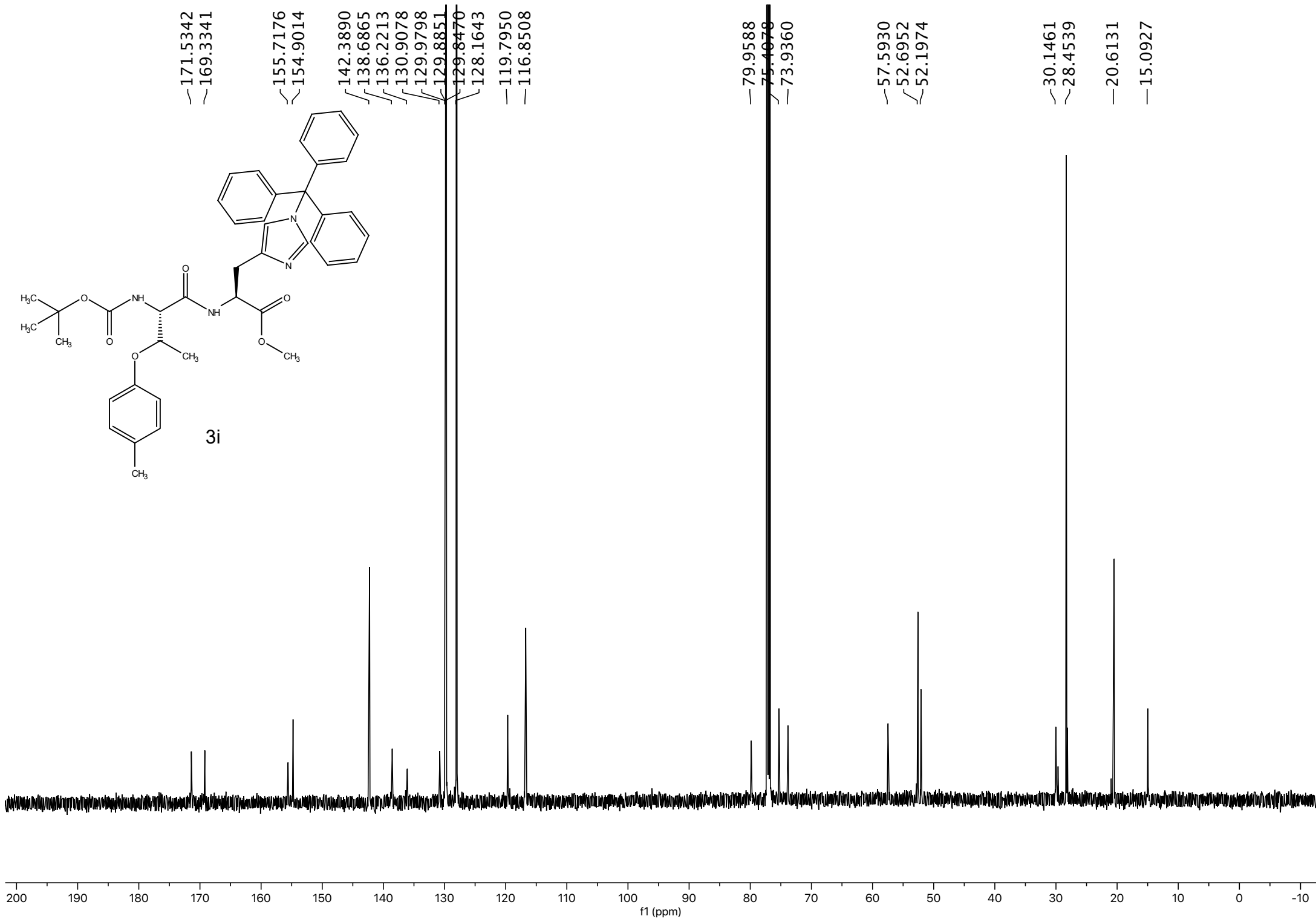




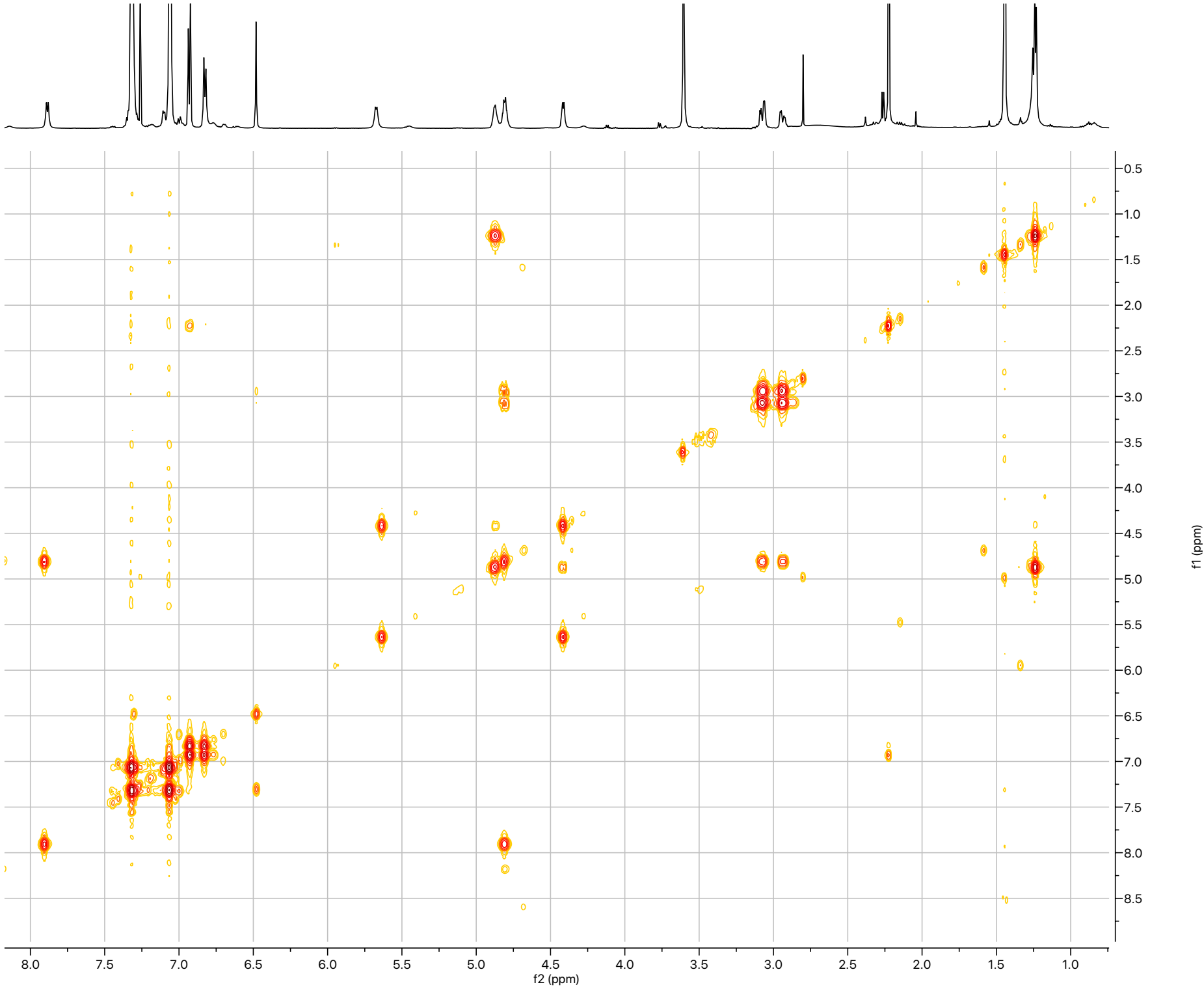


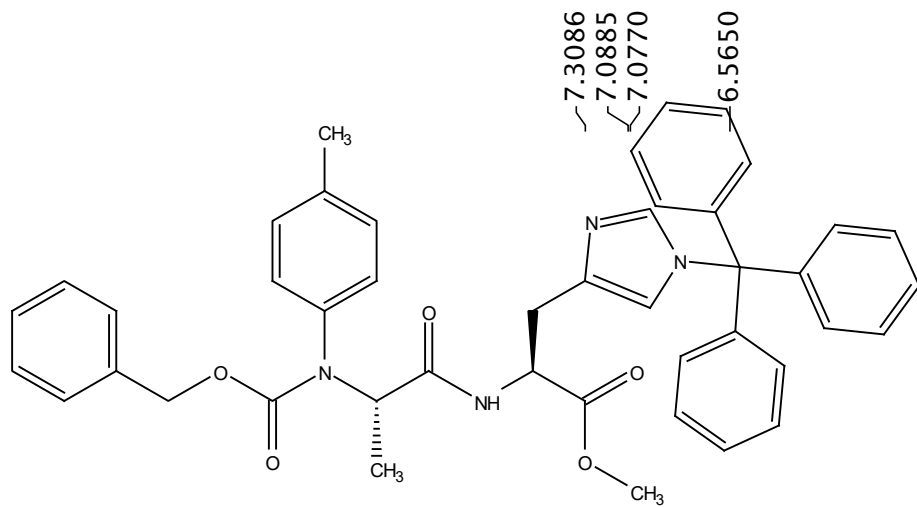


3i

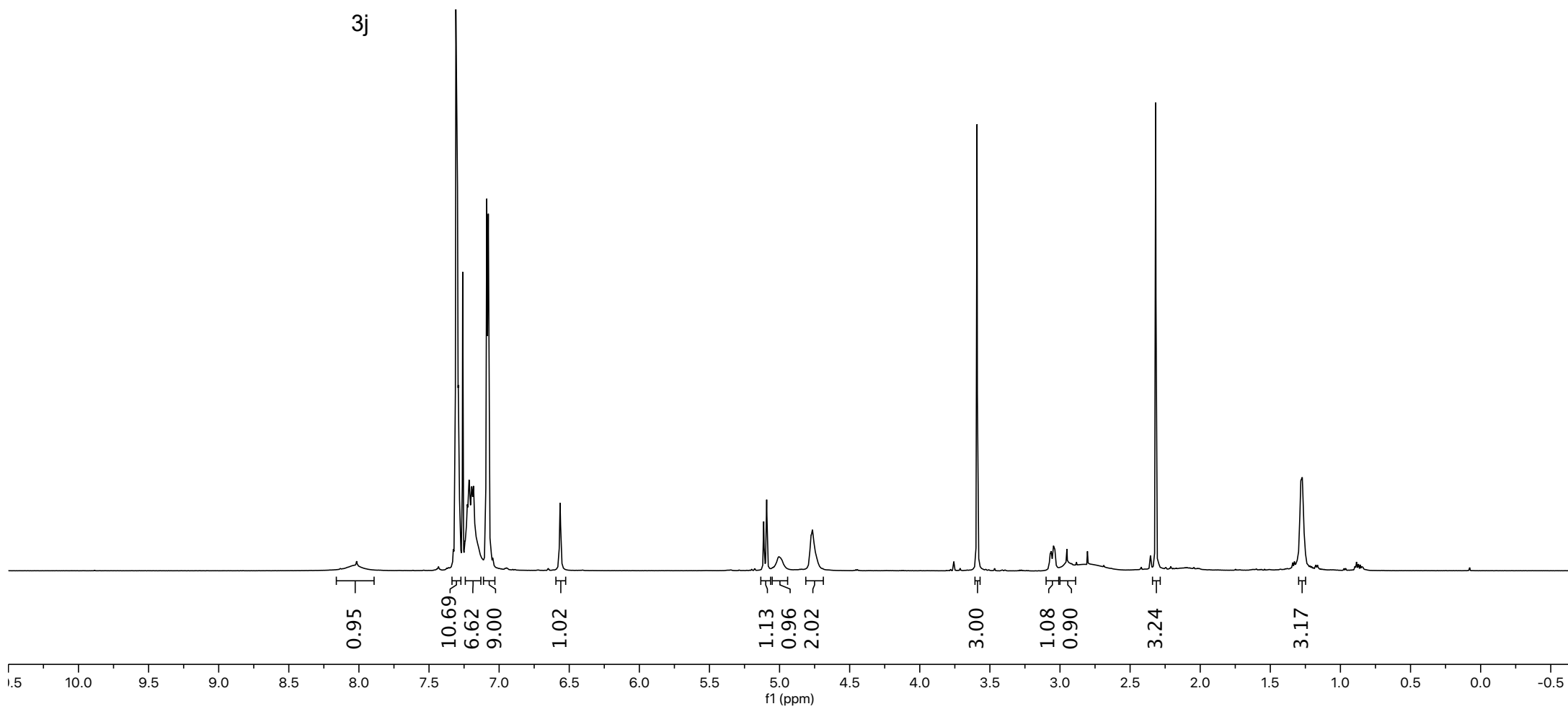


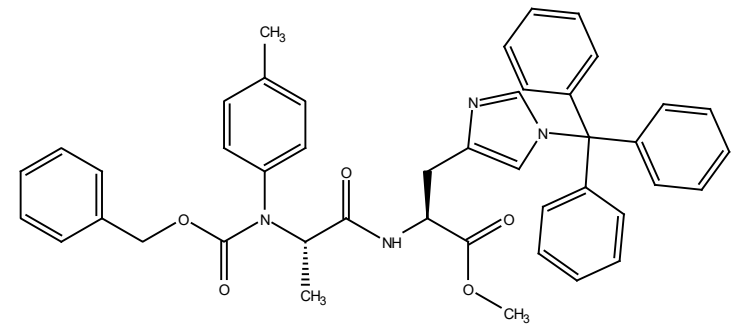
3i COSY



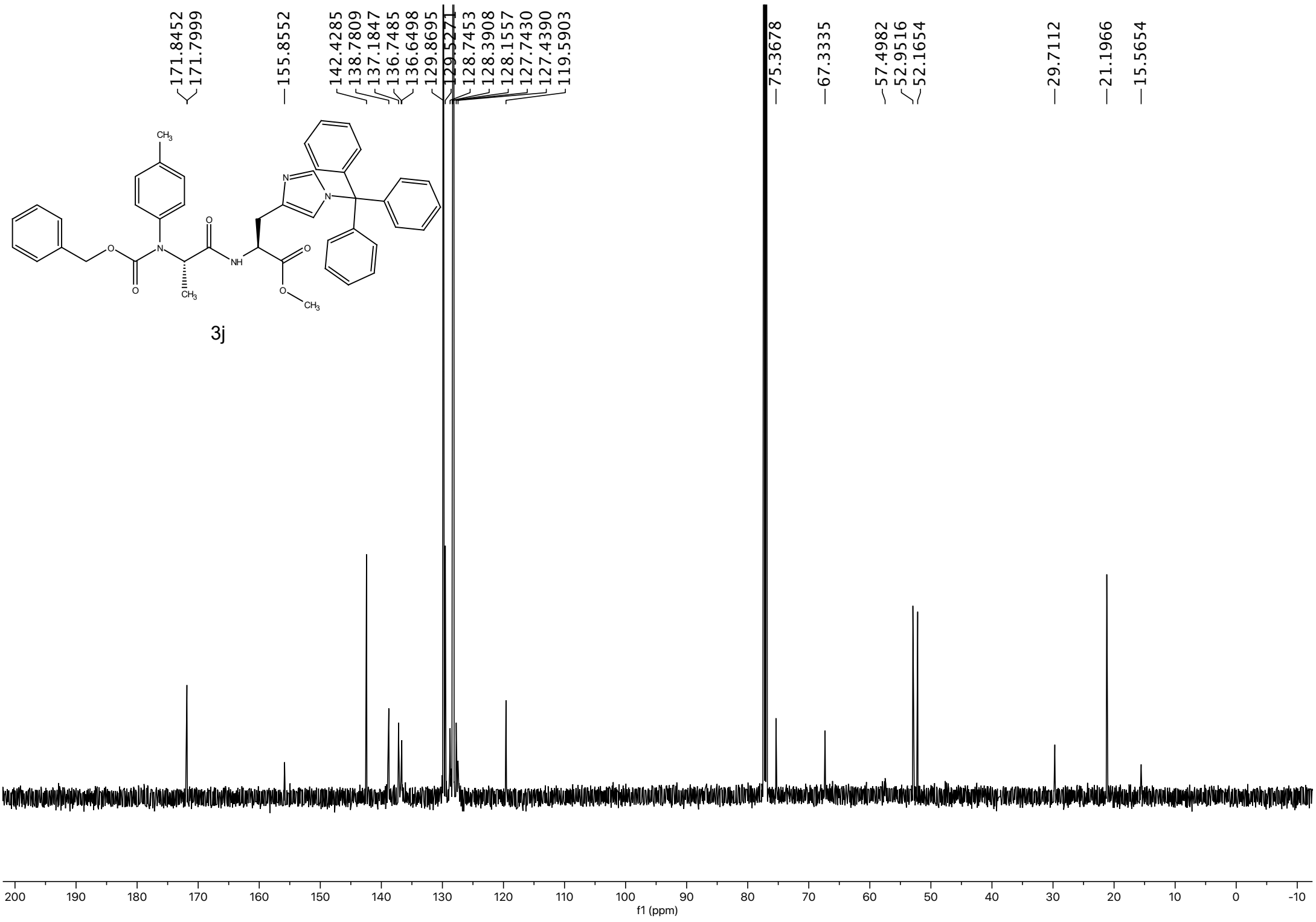


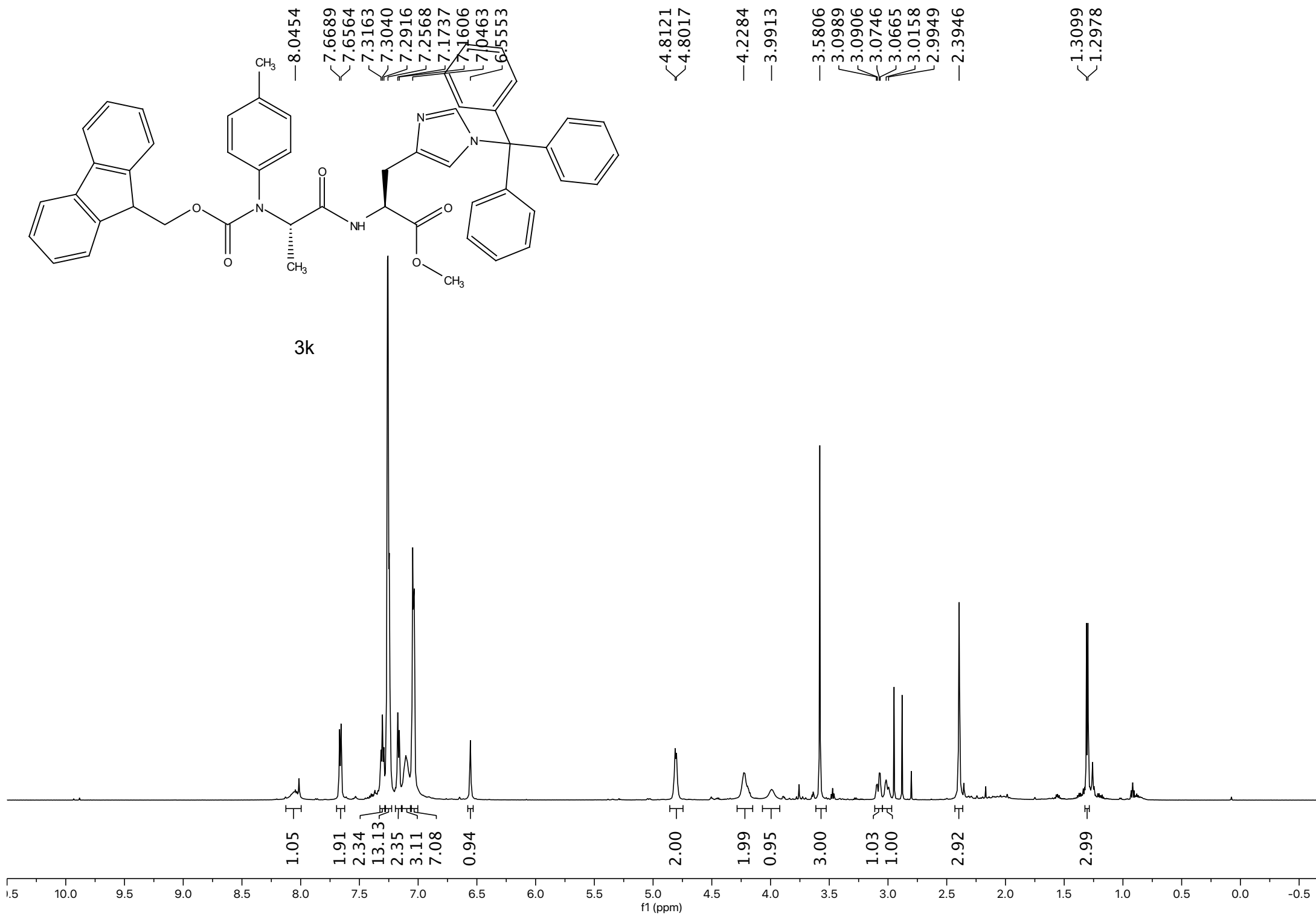
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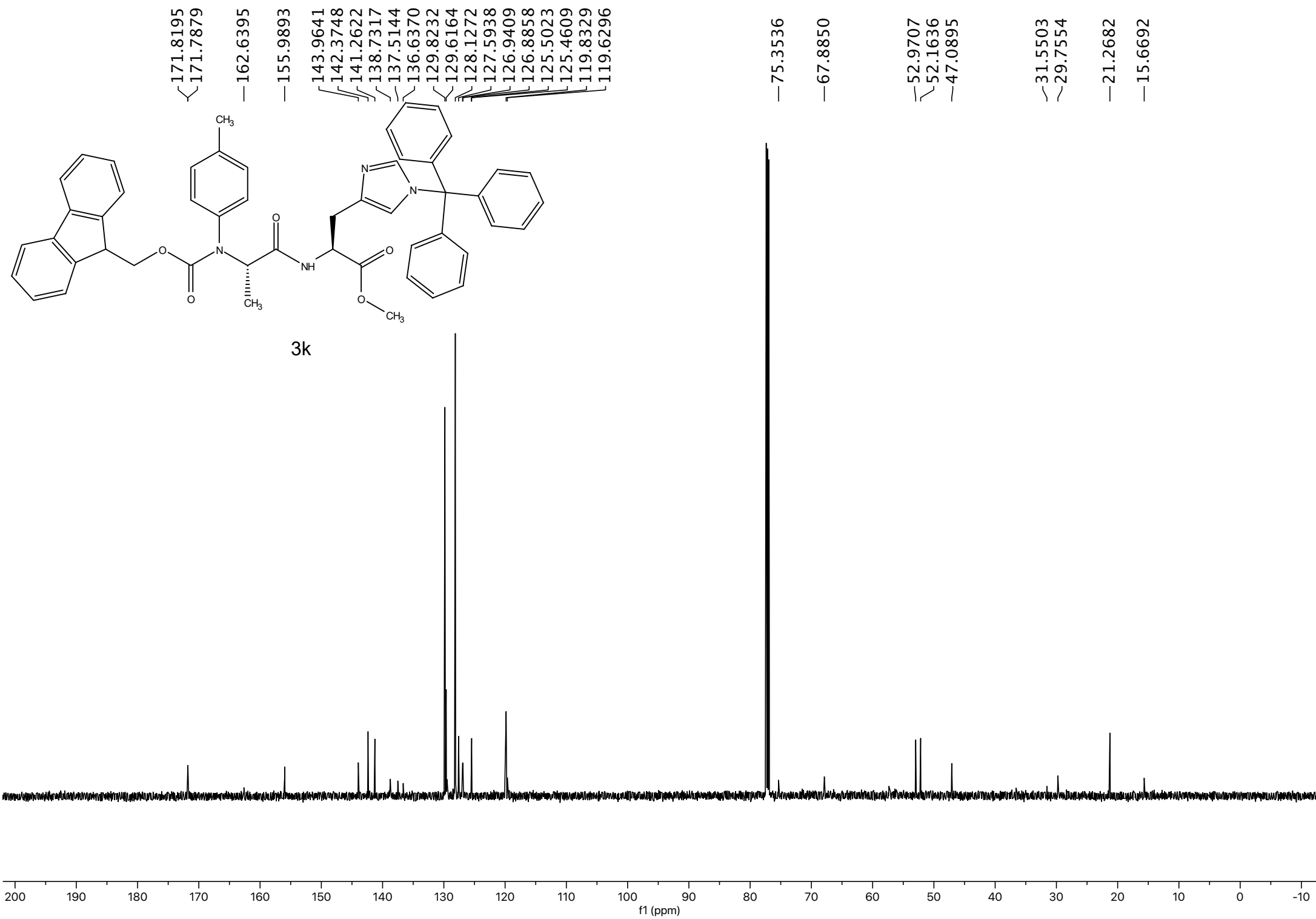


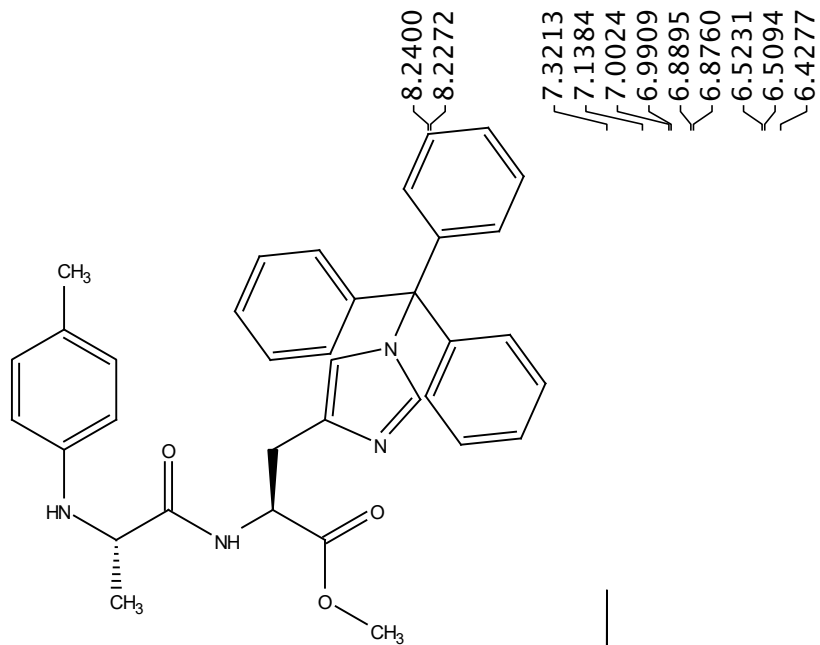


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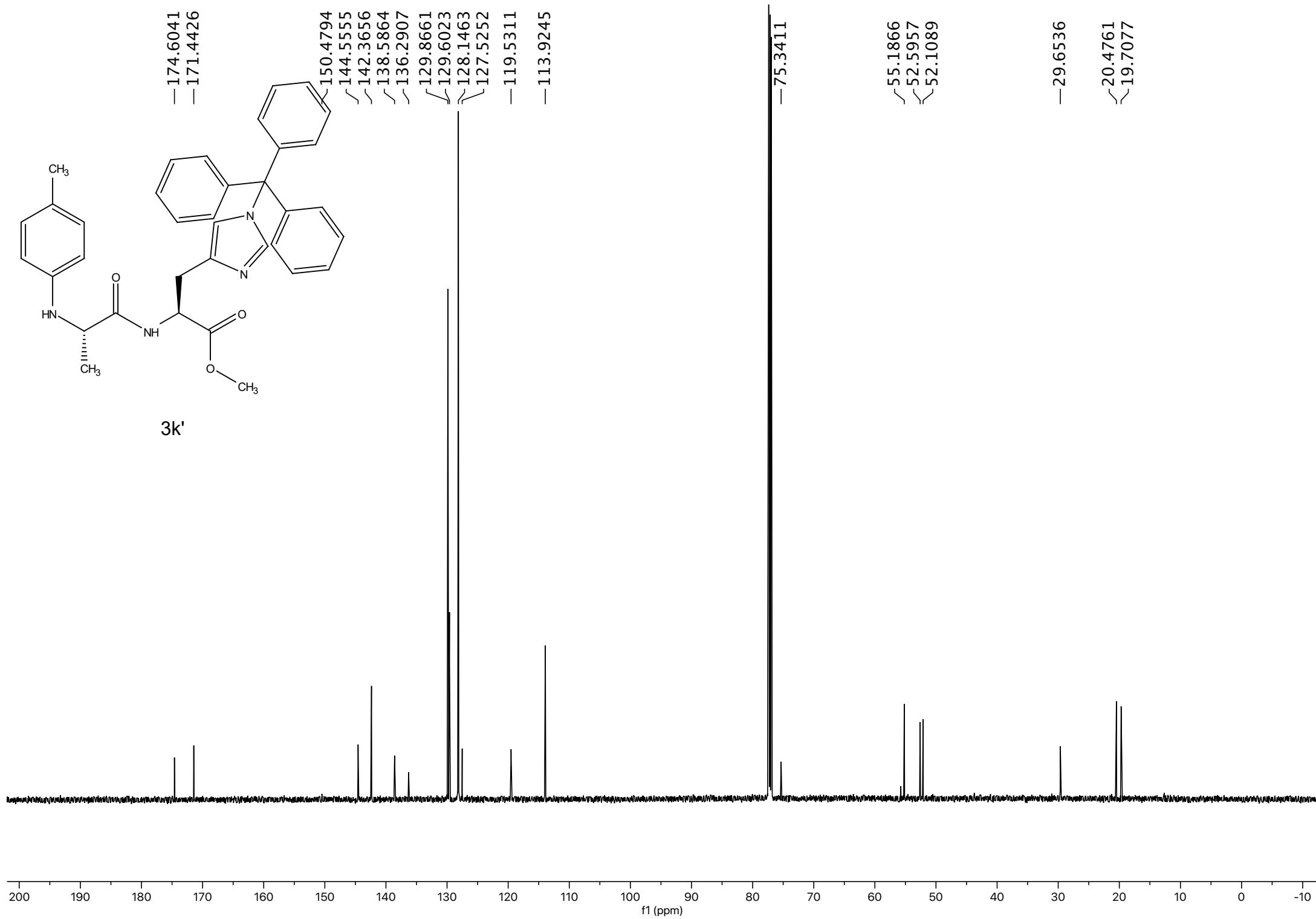


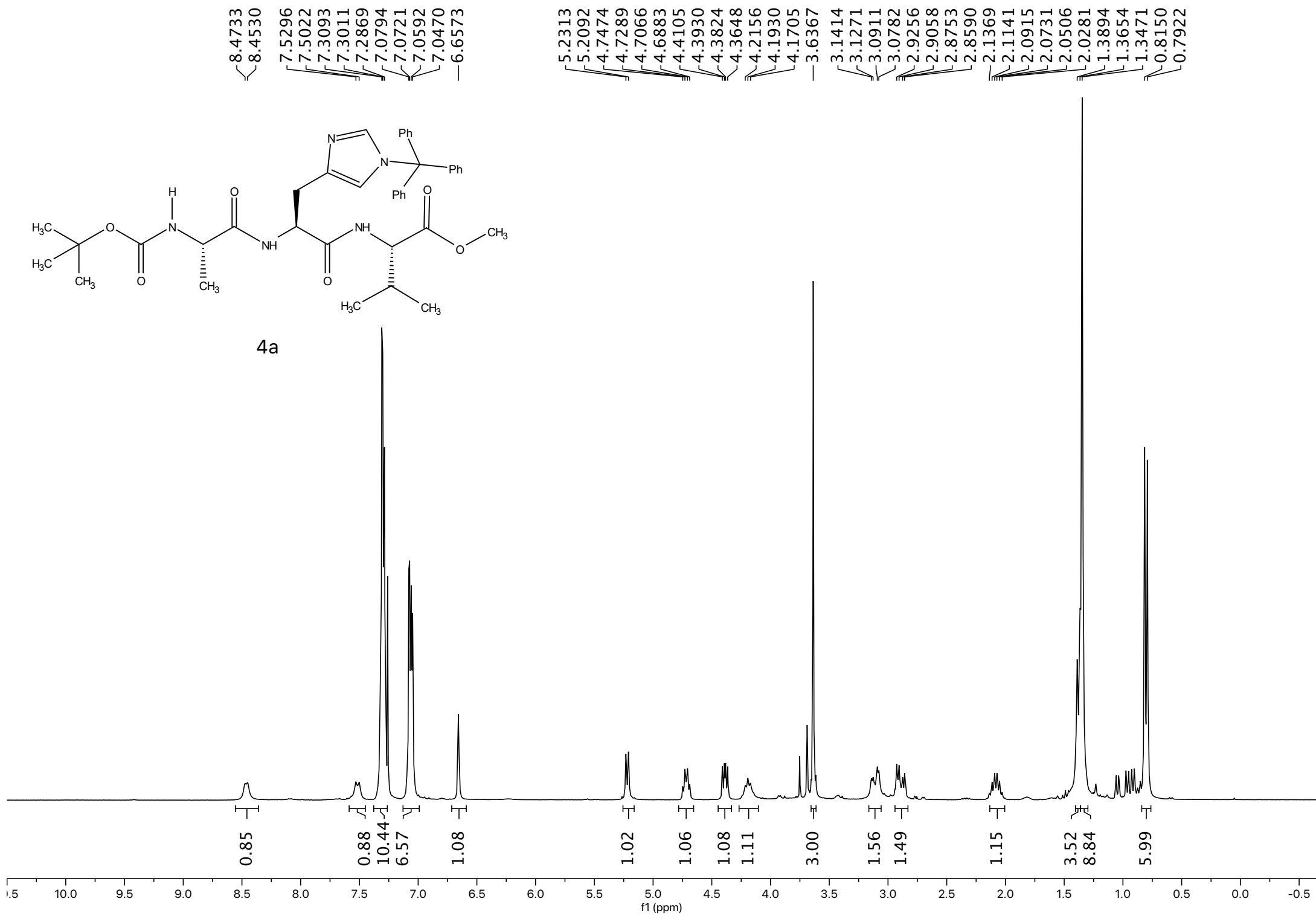


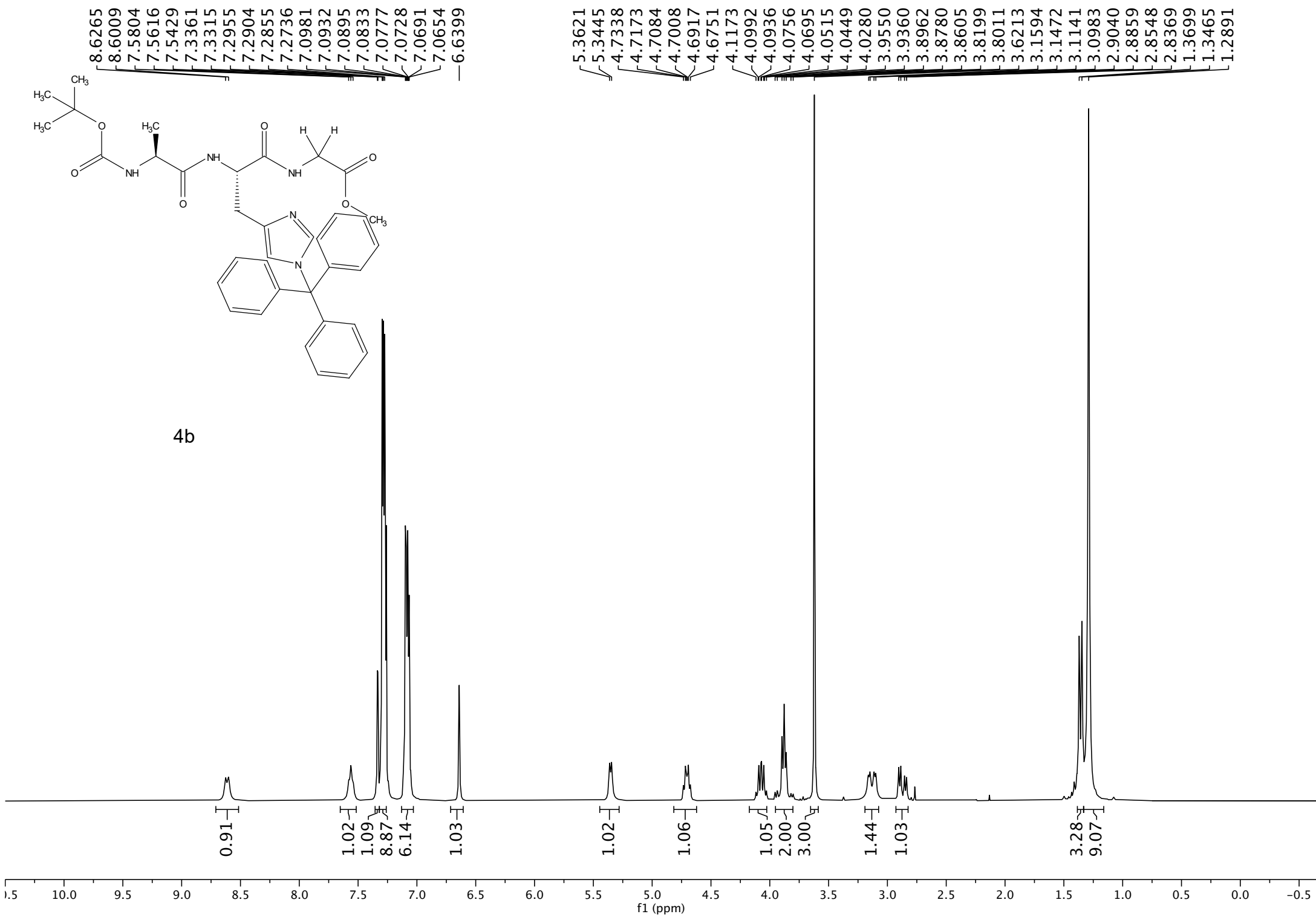


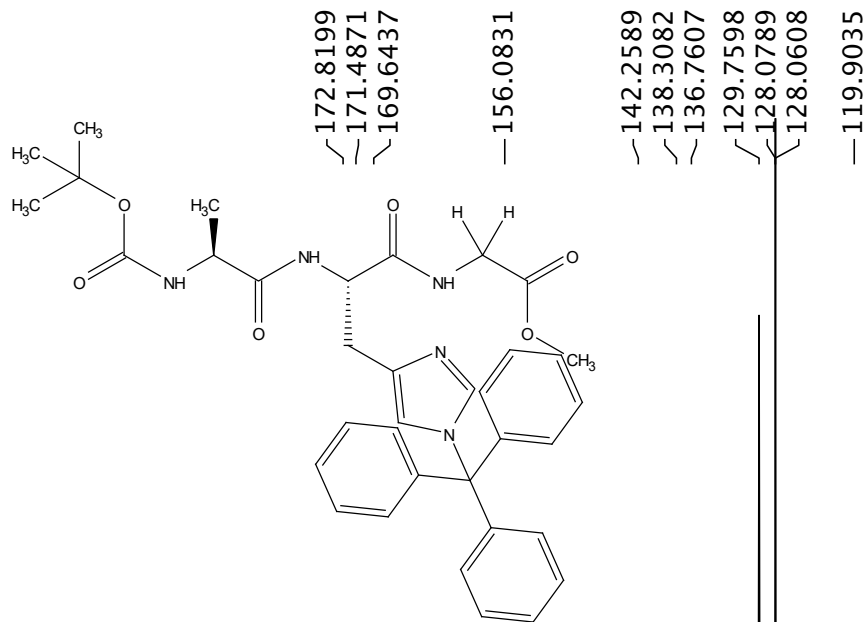
3k'

f1 (ppm)

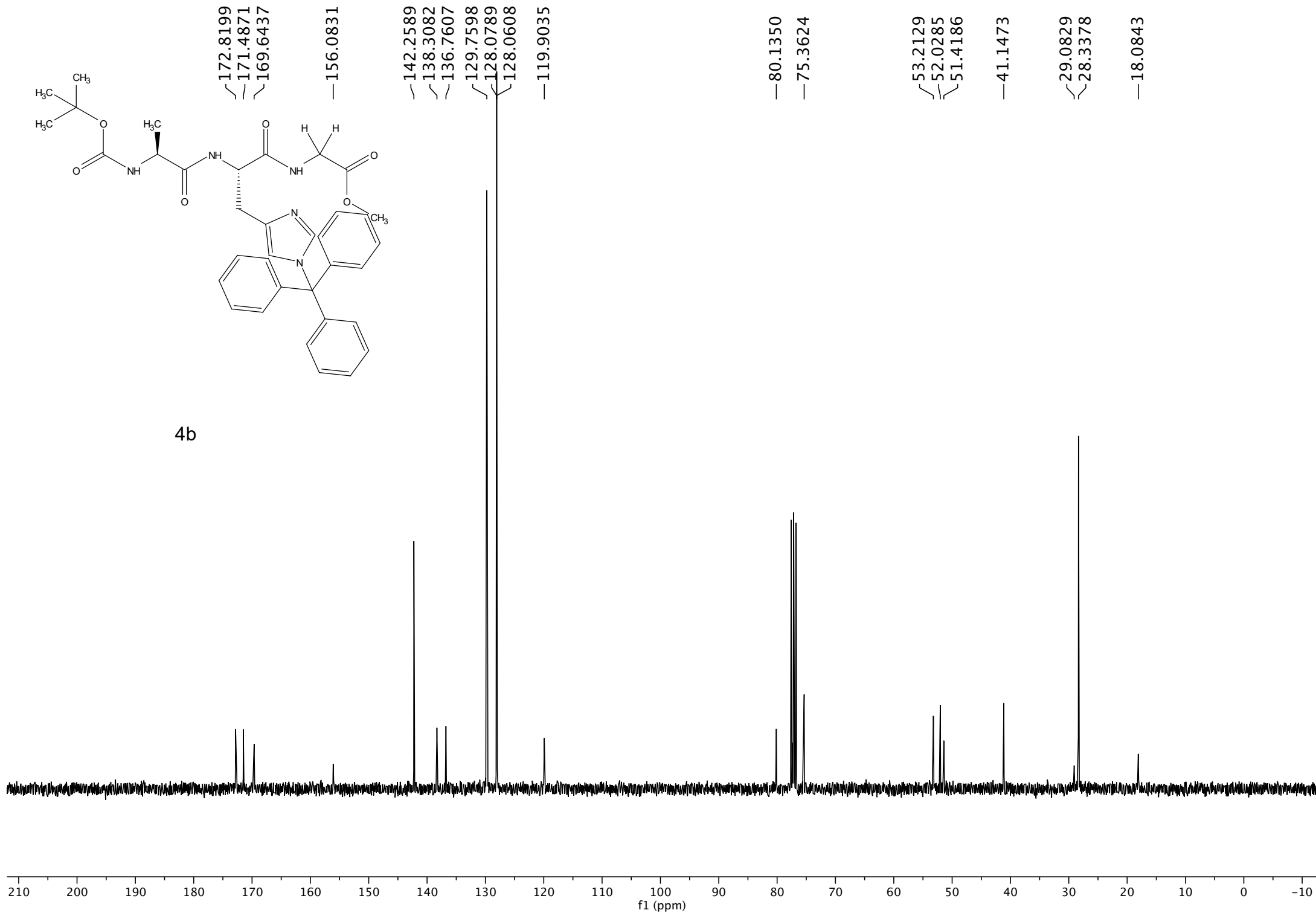


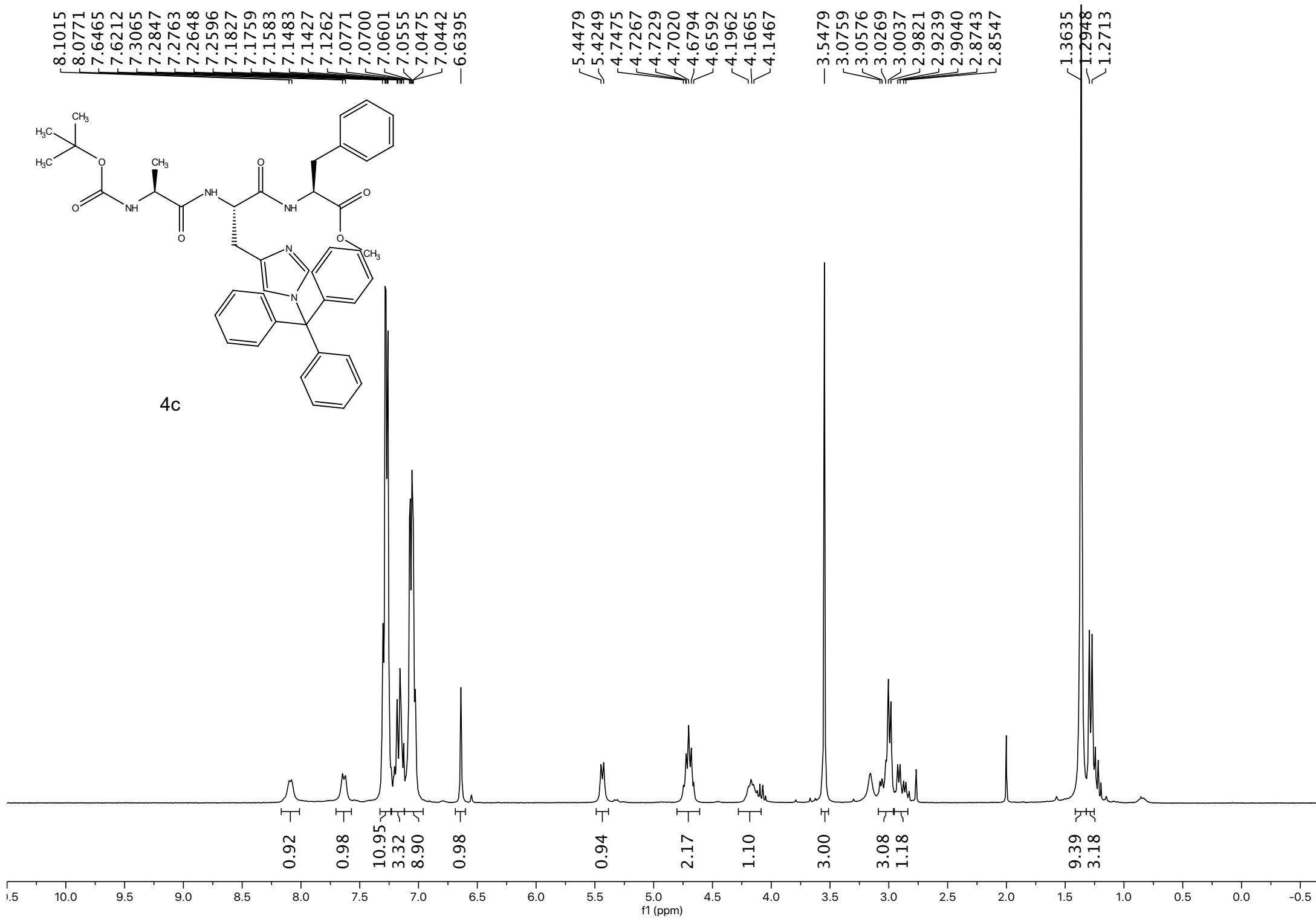


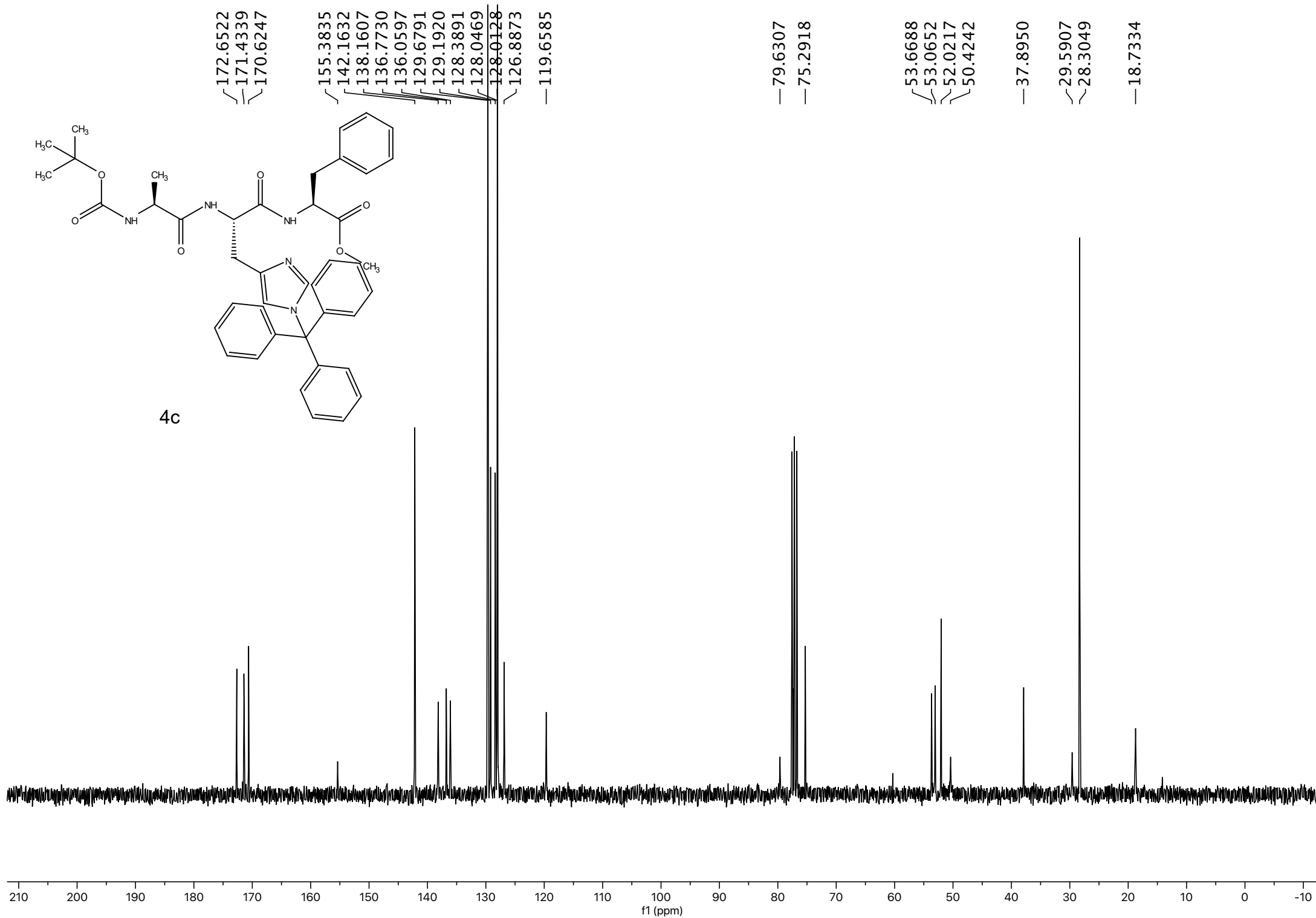


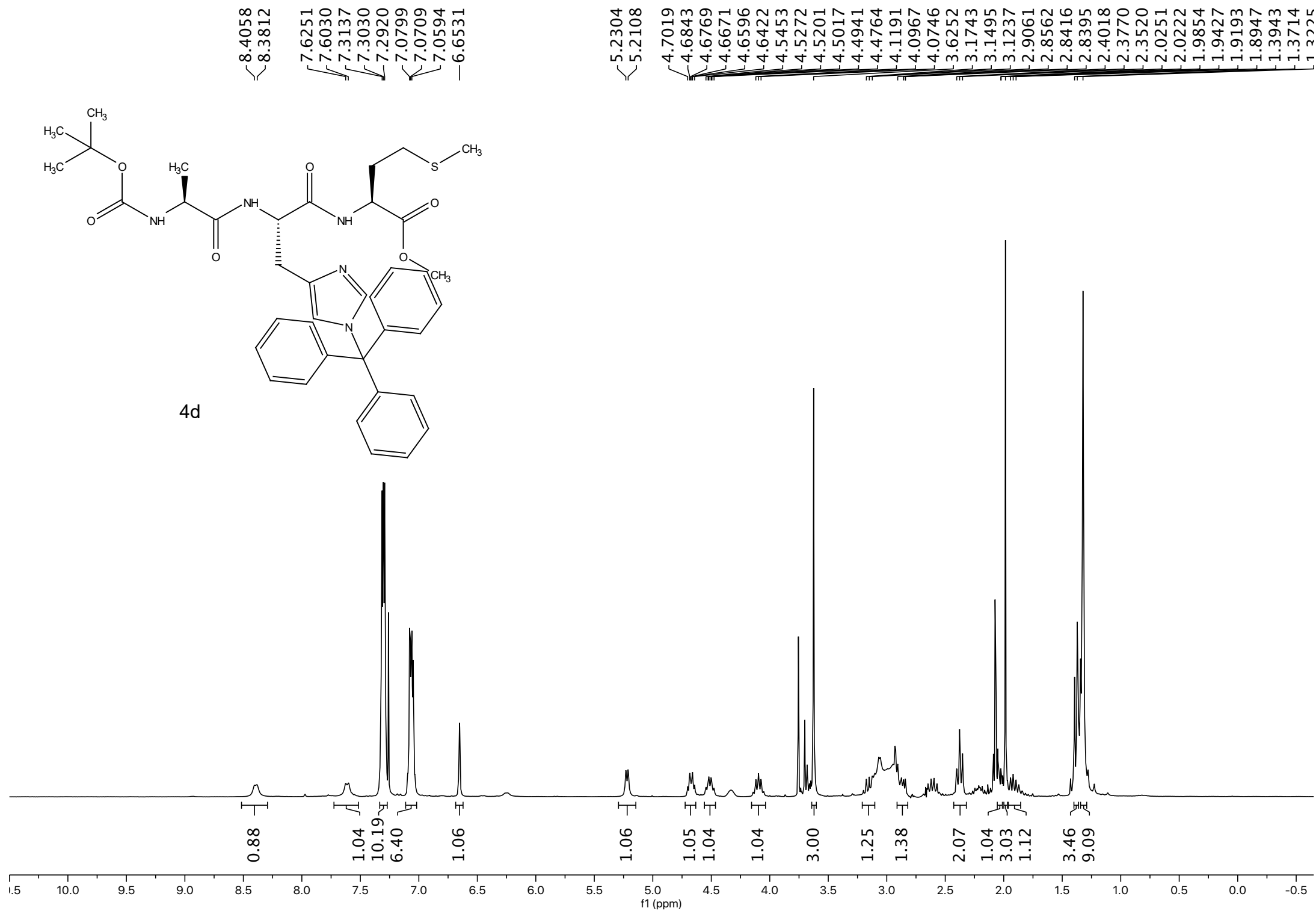
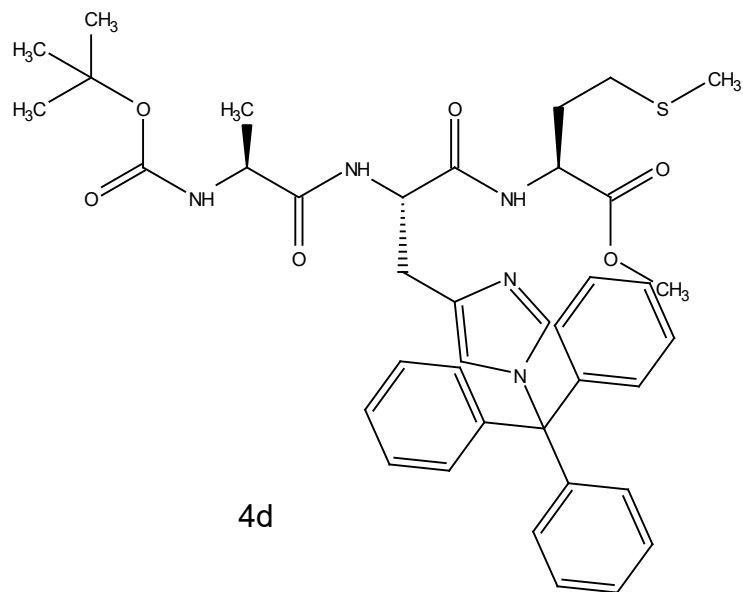


4b

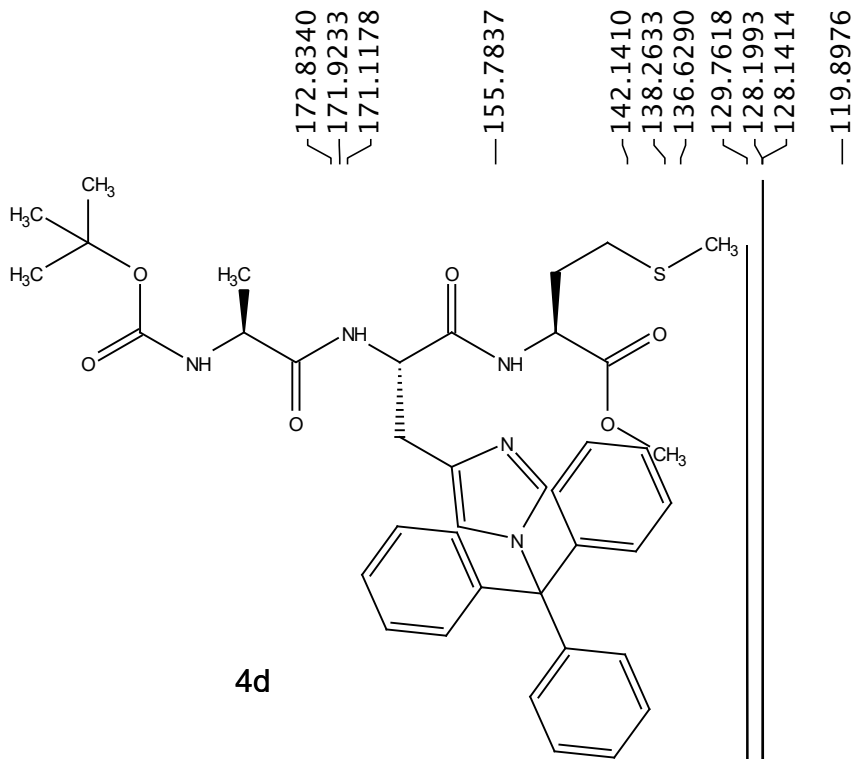








4d



172.8340
171.9233
171.1178

155.7837

142.1410
138.2633
136.6290
129.7618
128.1993
128.1414

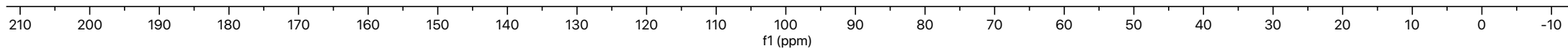
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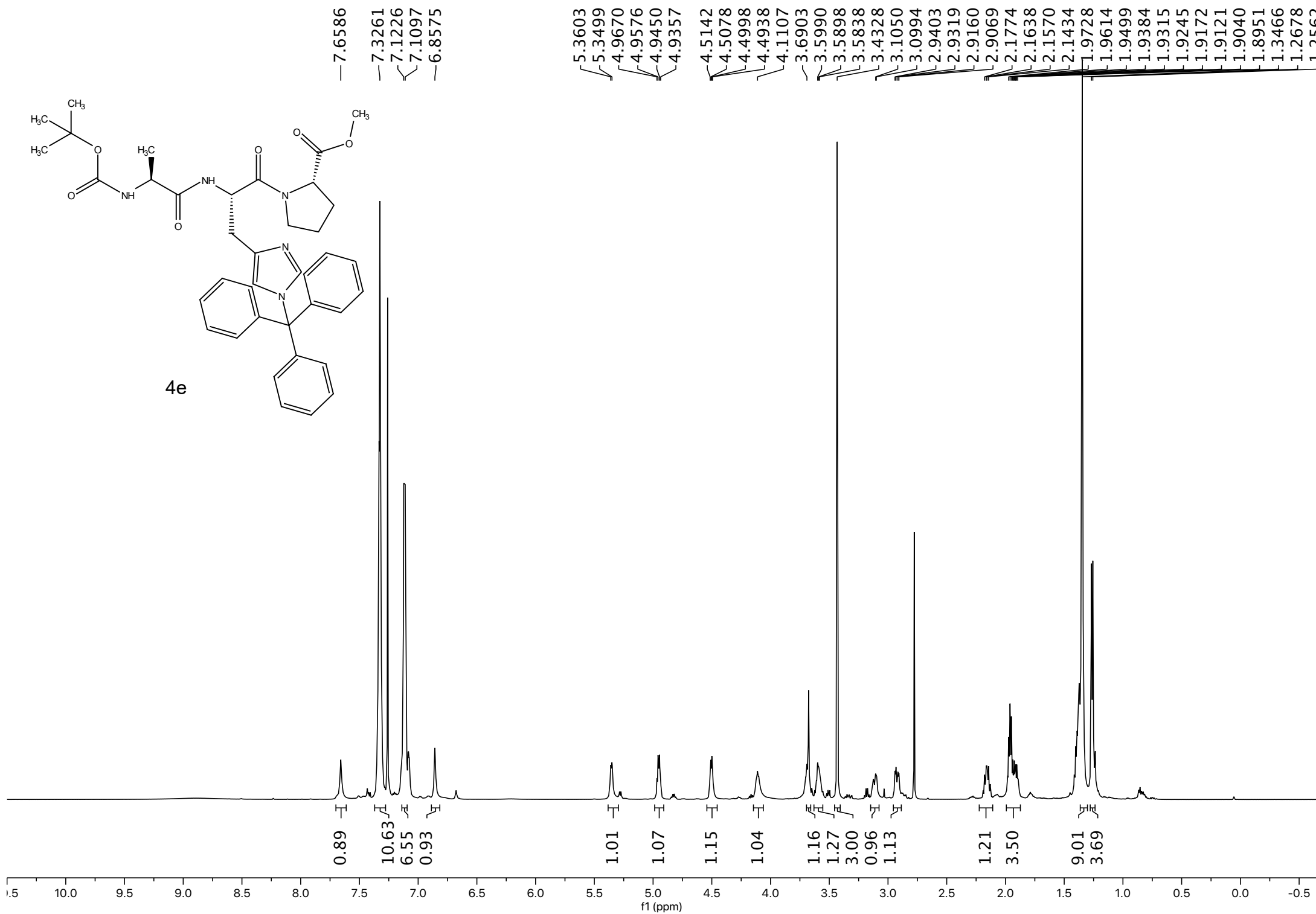
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75.5271

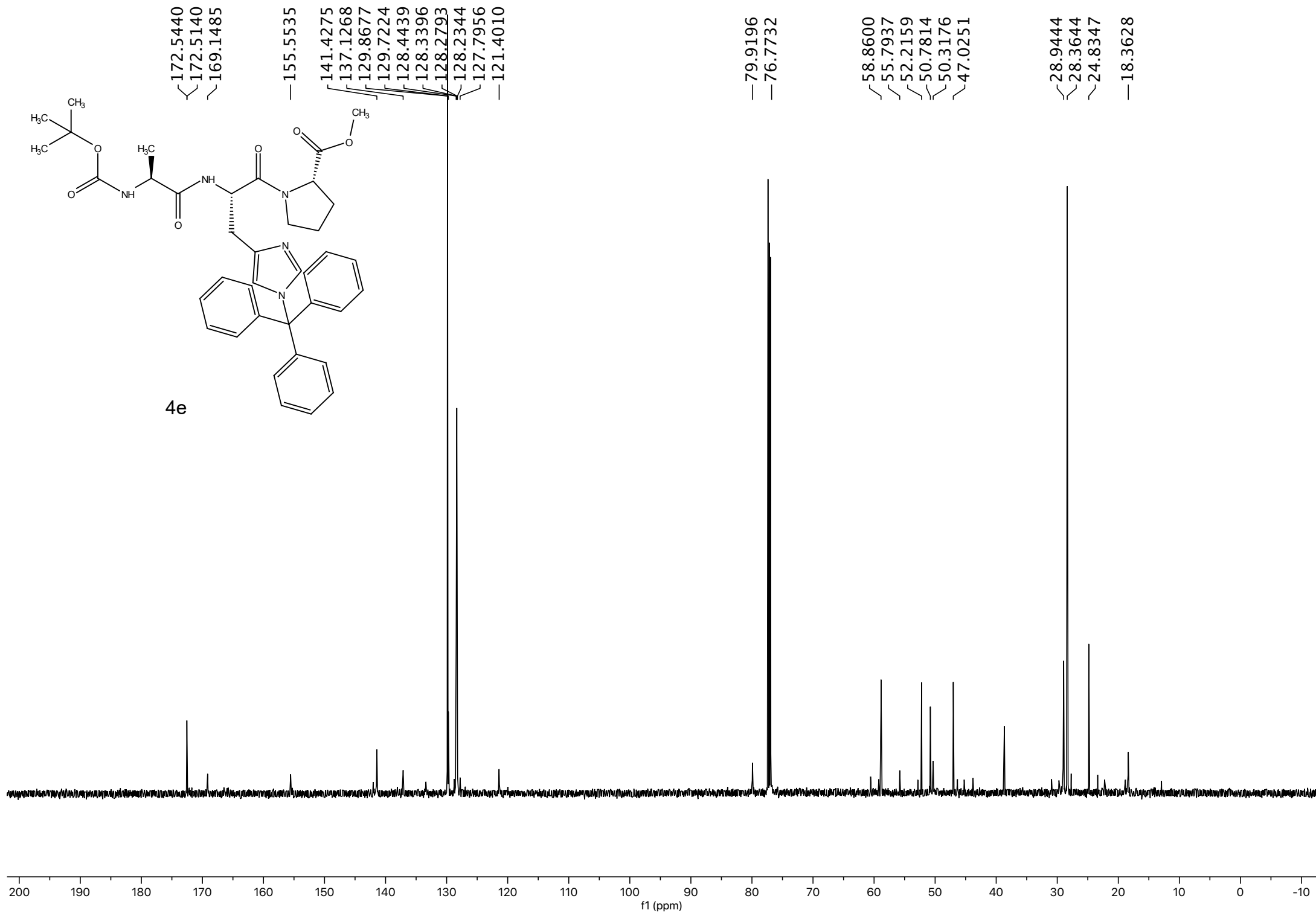
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52.3525
51.7443

31.2683
30.0042
28.3742

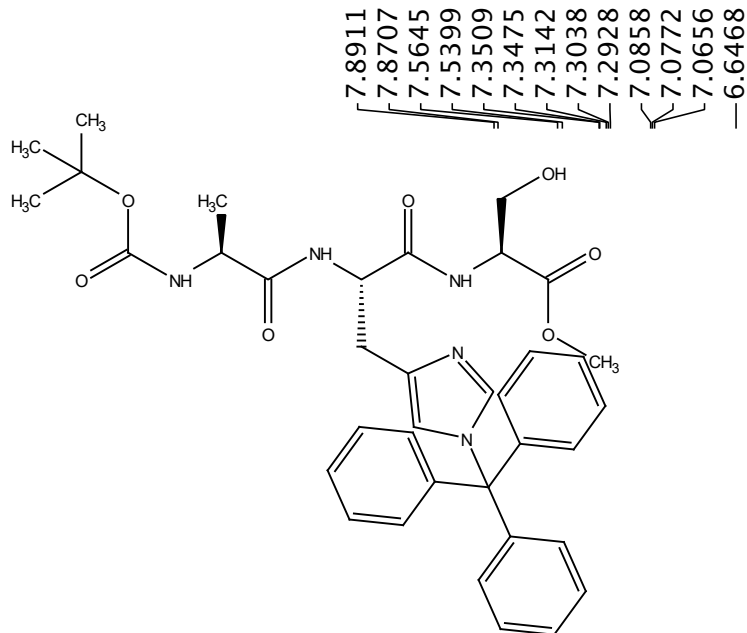
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14.9815





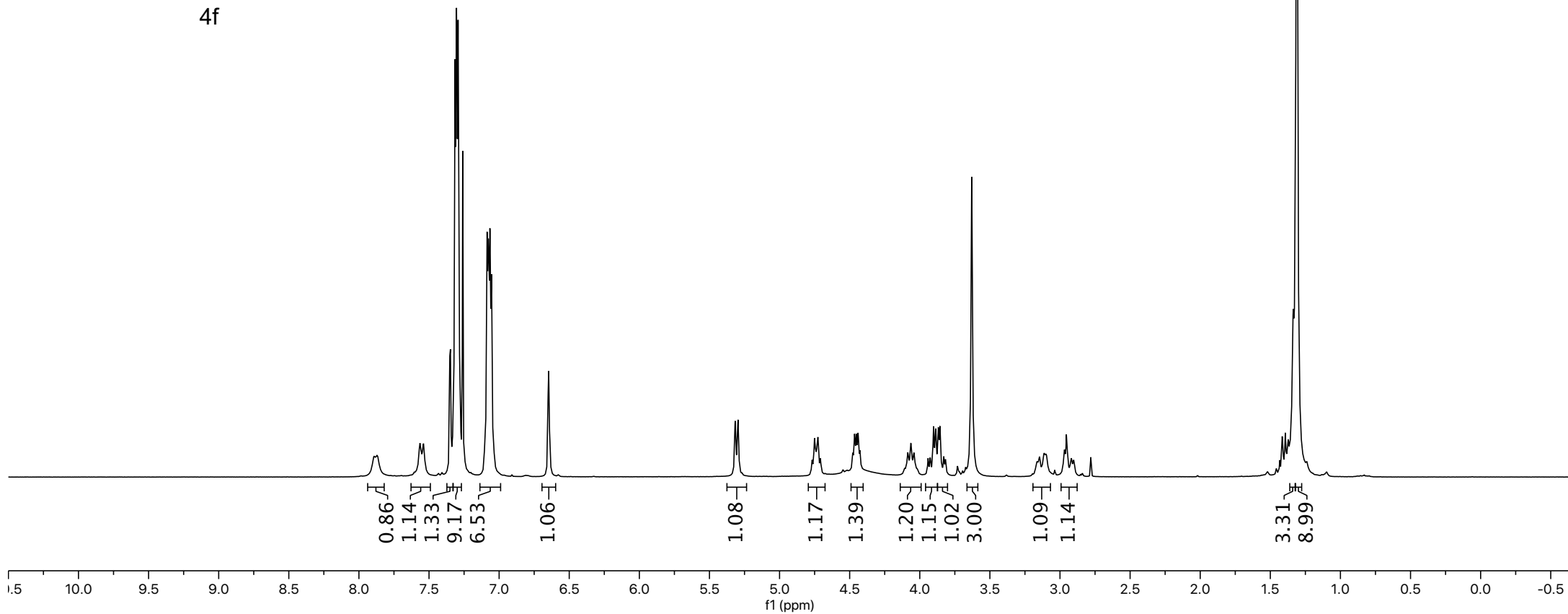


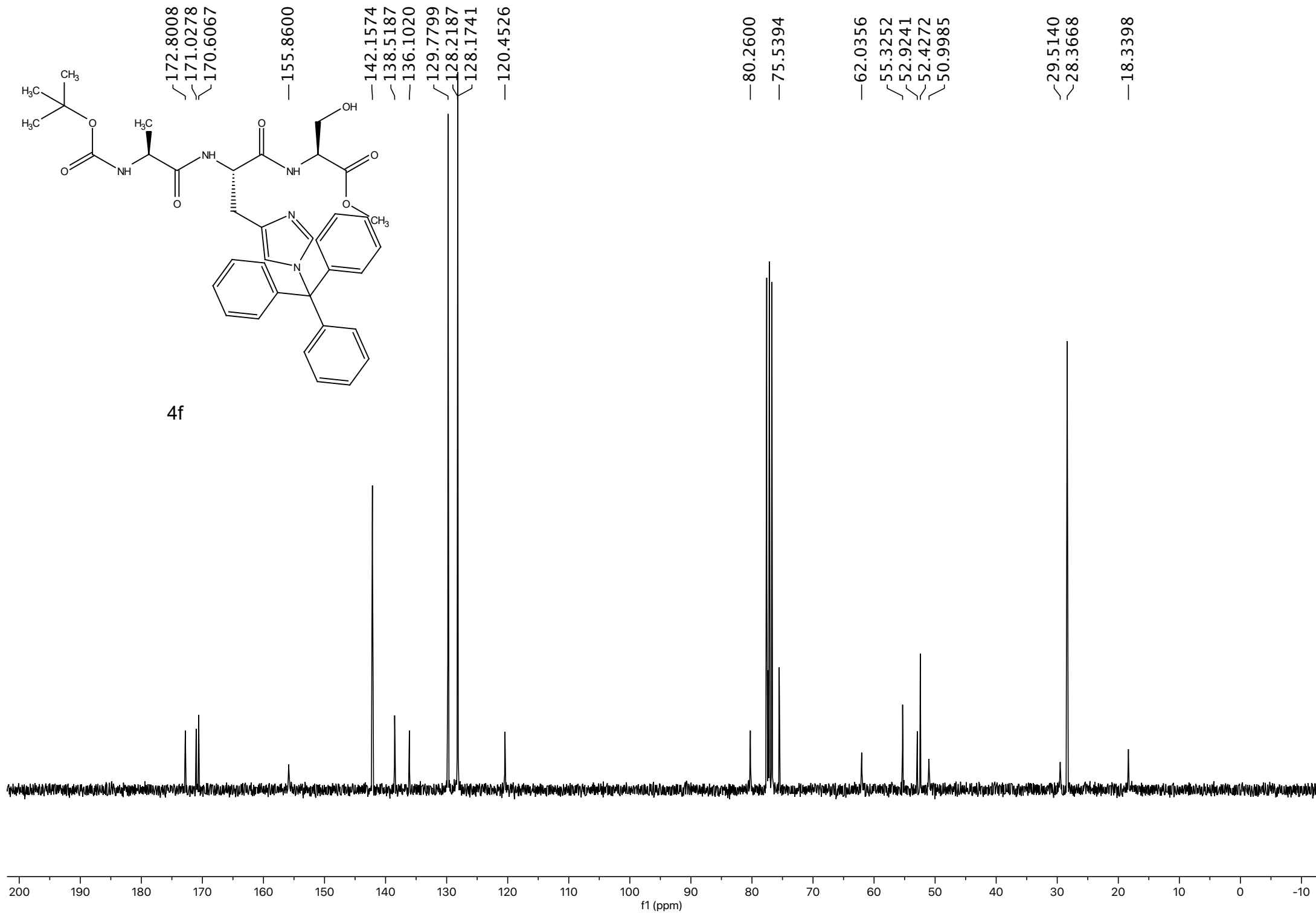
4f

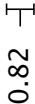


7.8911
7.8707
7.5645
7.5399
7.3509
7.3475
7.3142
7.3038
7.2928
7.0858
7.0772
7.0656
—6.6468

5.3163
5.2962
4.7679
4.7509
4.7264
4.7090
4.4775
4.4651
4.4522
4.4403
4.4267
4.0853
4.0631
4.0413
3.9407
3.9257
3.9017
3.8870
3.8675
3.8558
3.8287
3.8170
3.6299
3.1628
3.1463
3.1127
3.0998
2.9683
2.9544
2.9193
2.9023
1.3359
1.3112





0.94 γ

10.36 正

5.19

6.4/江

0.99
H

1.03 T

1.04 T

2.05 T

3.00

1.09 γ

1.07

0.95
45

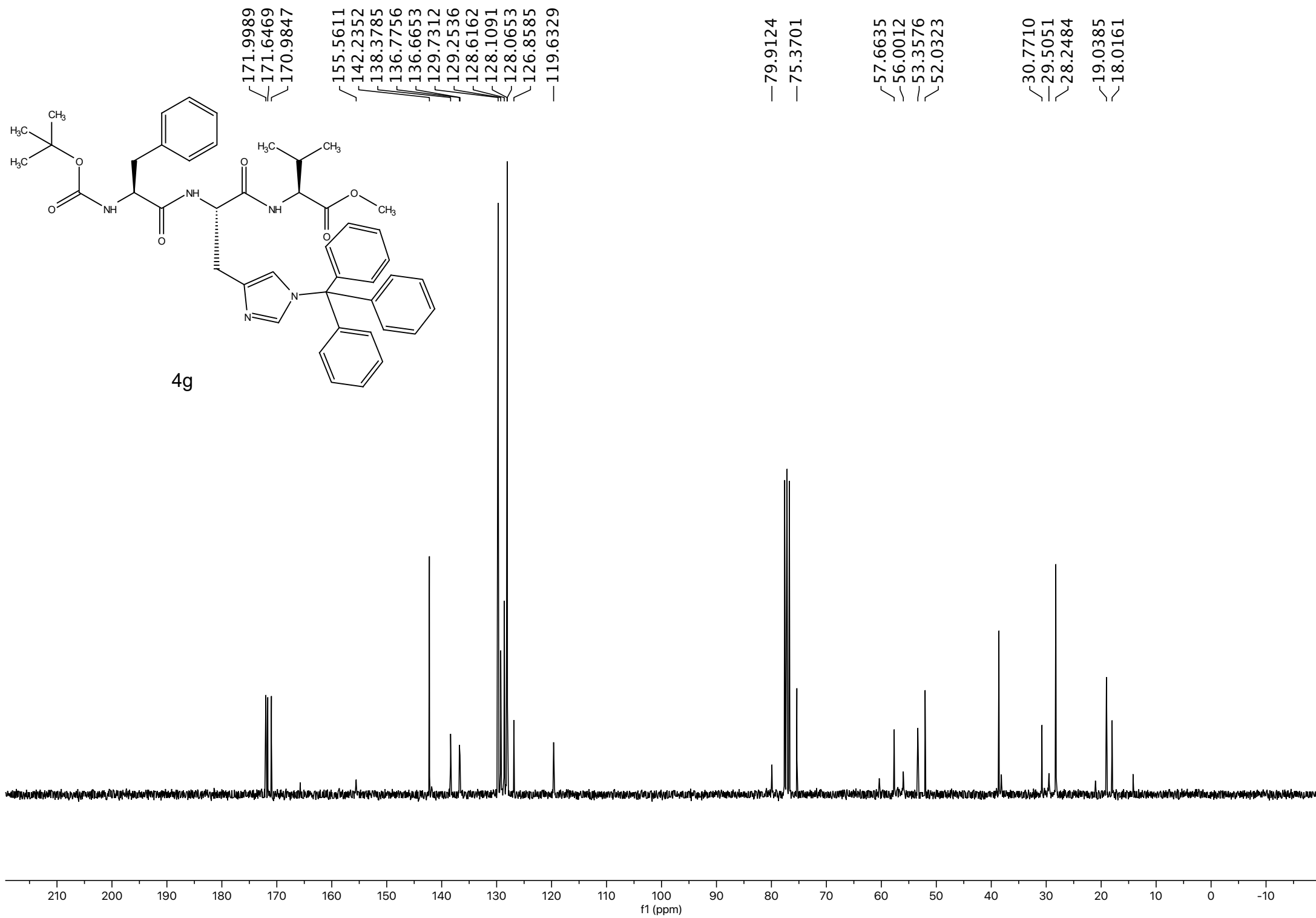
0.62 Л

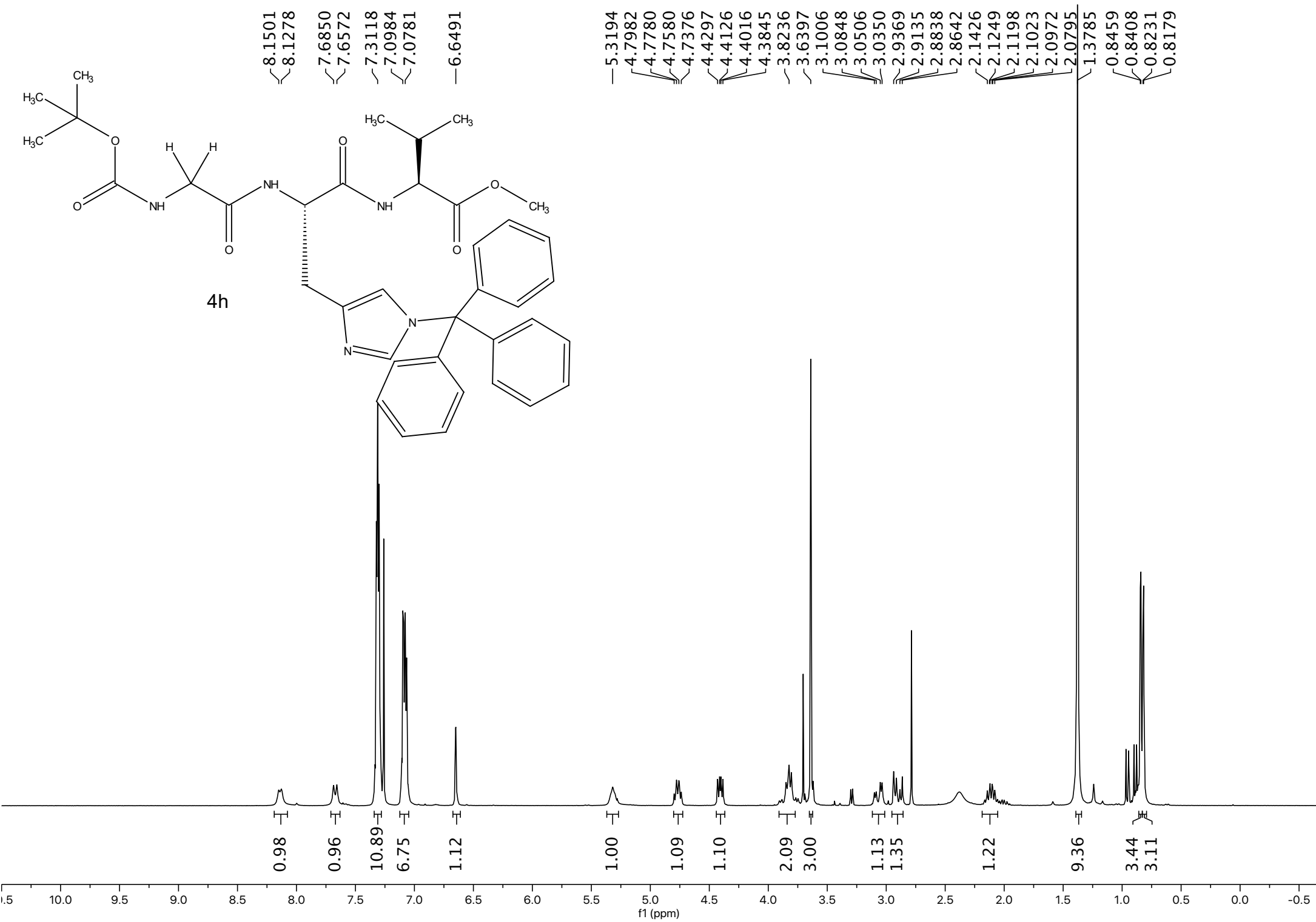
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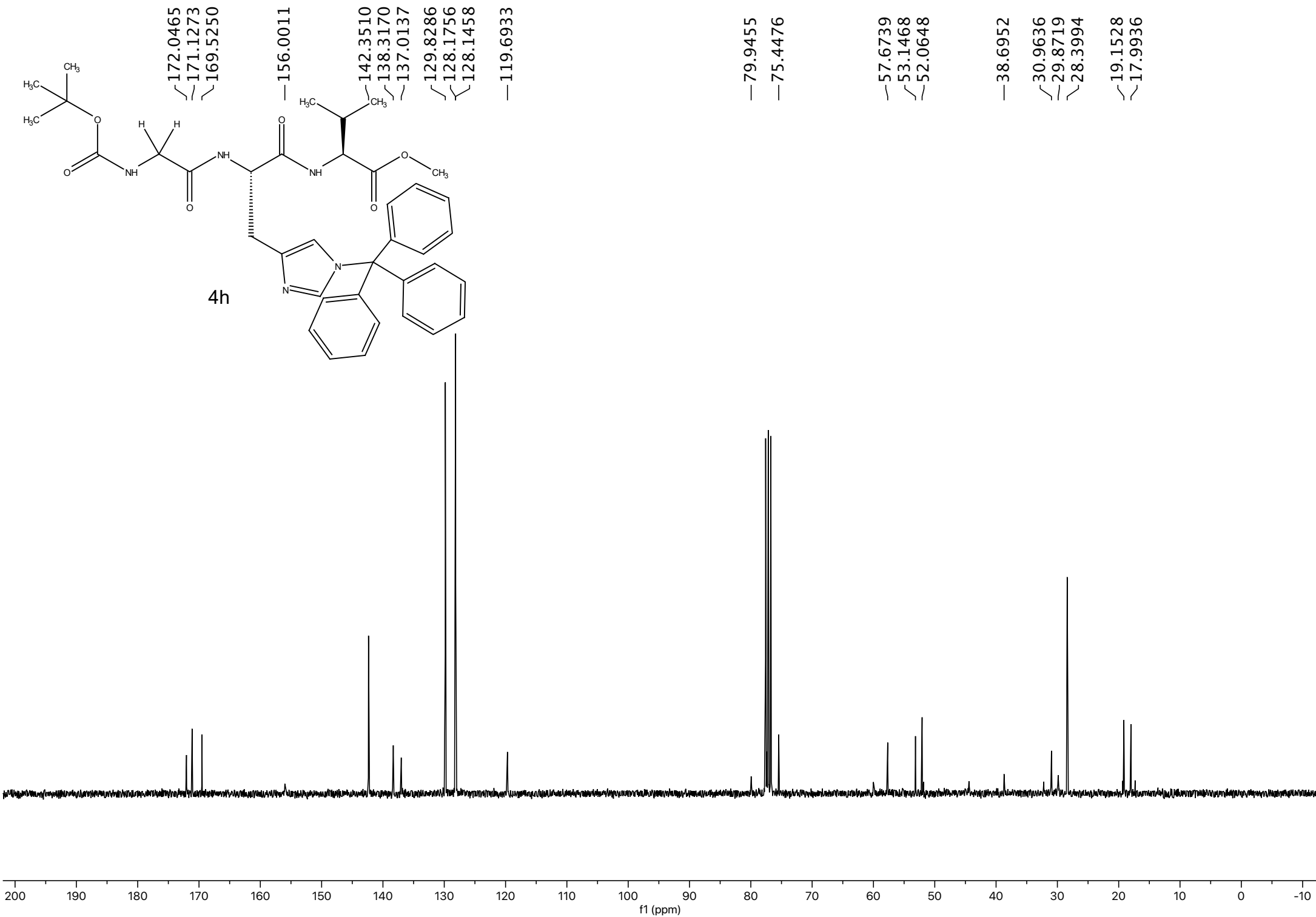
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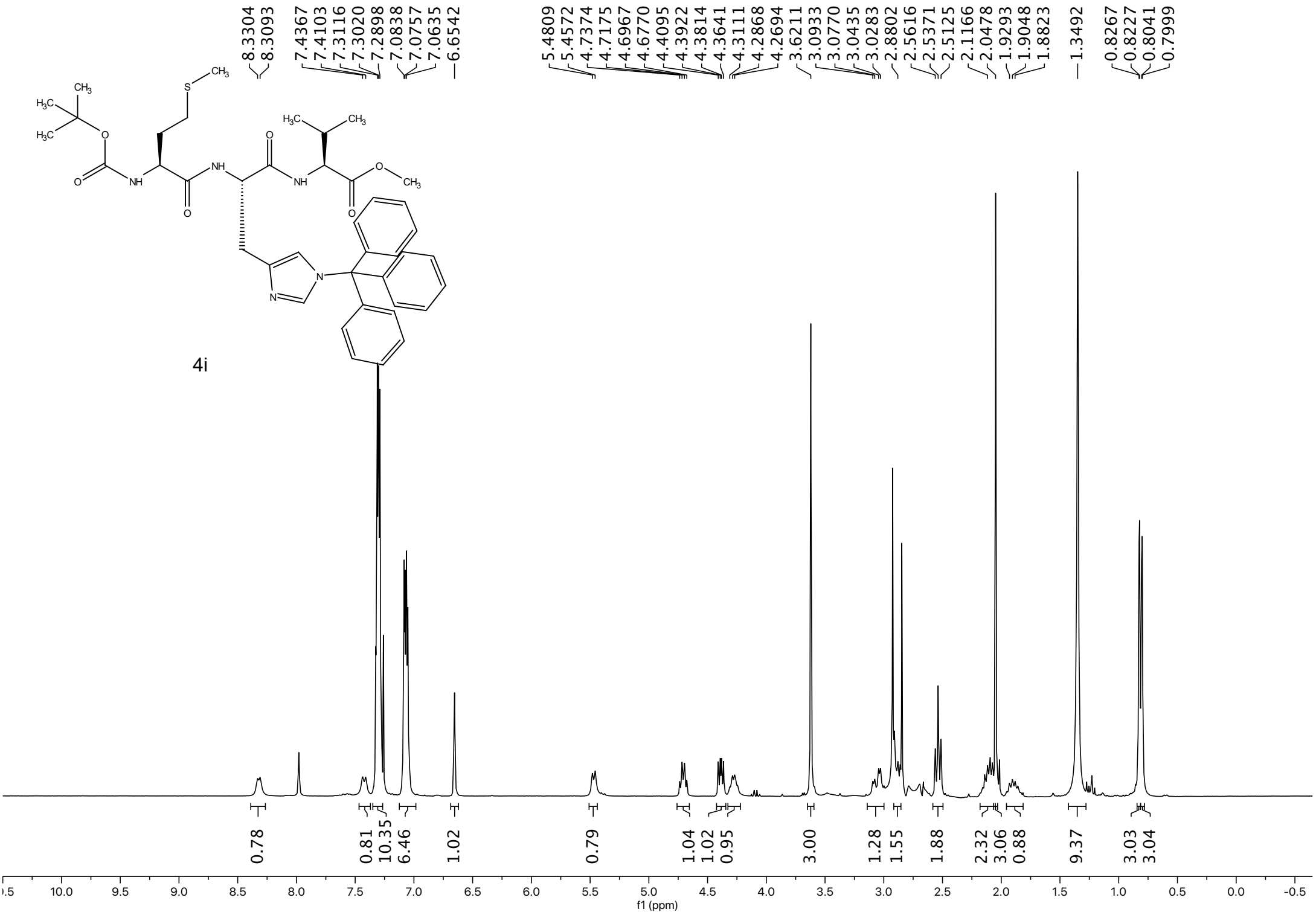
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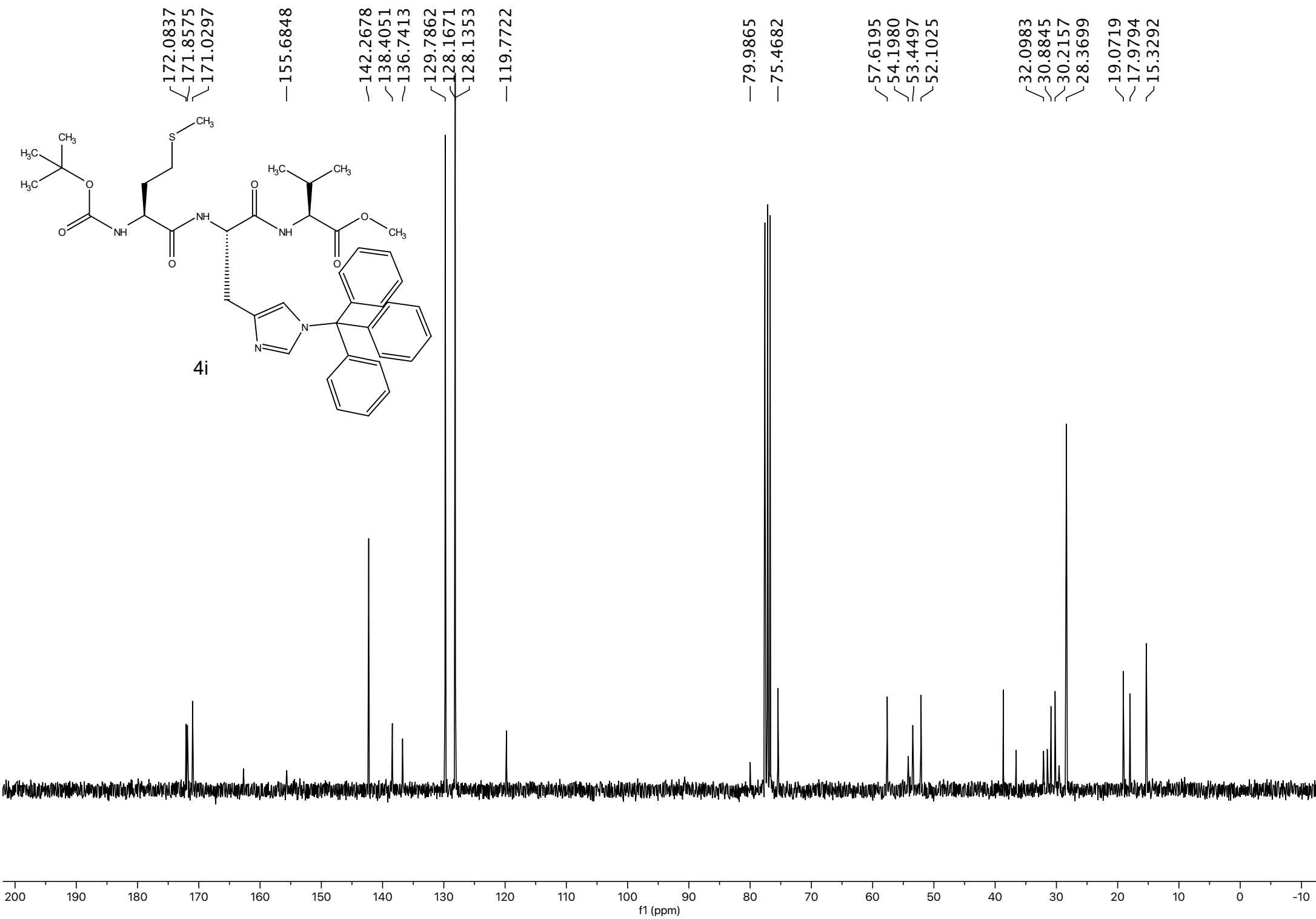
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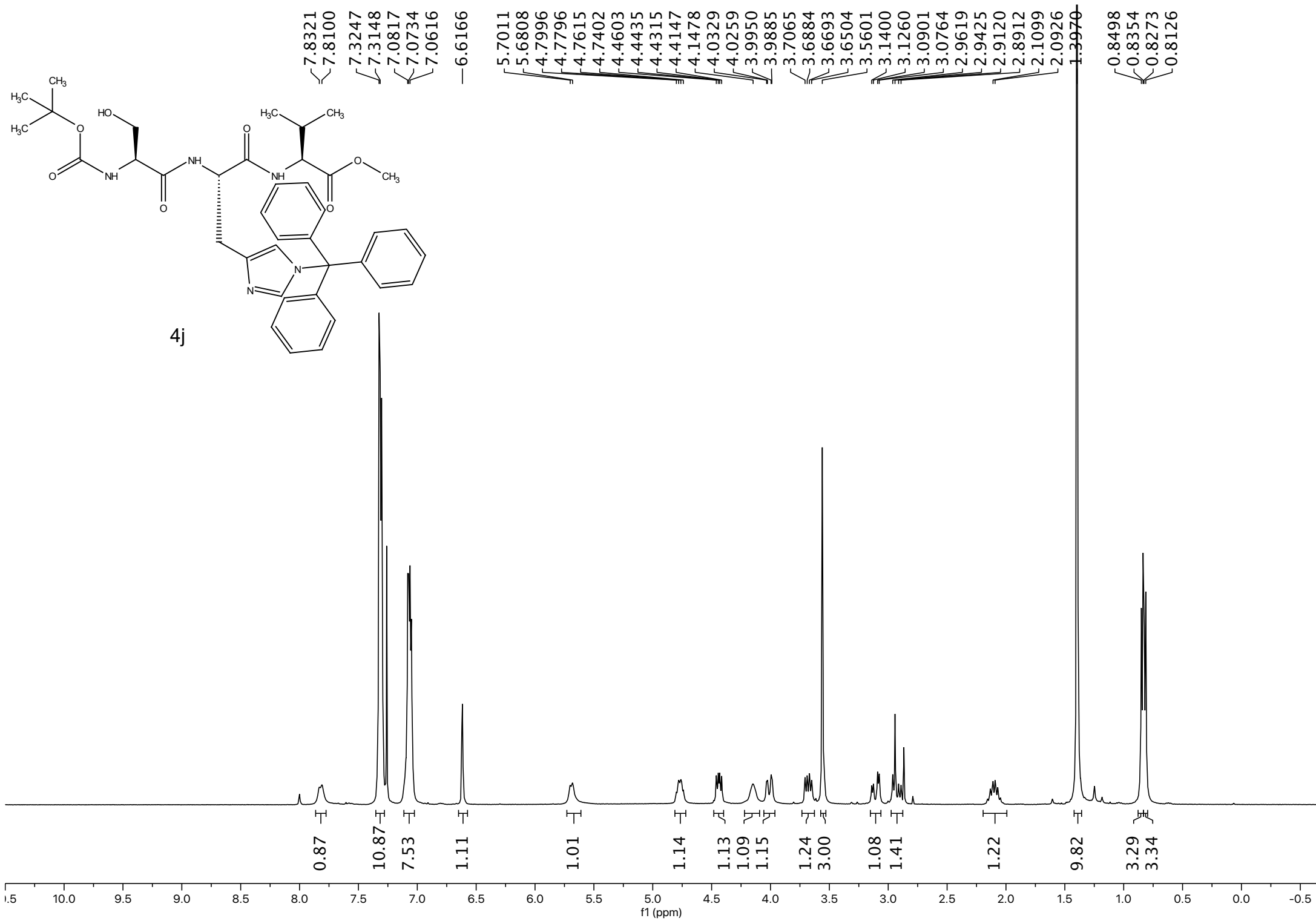


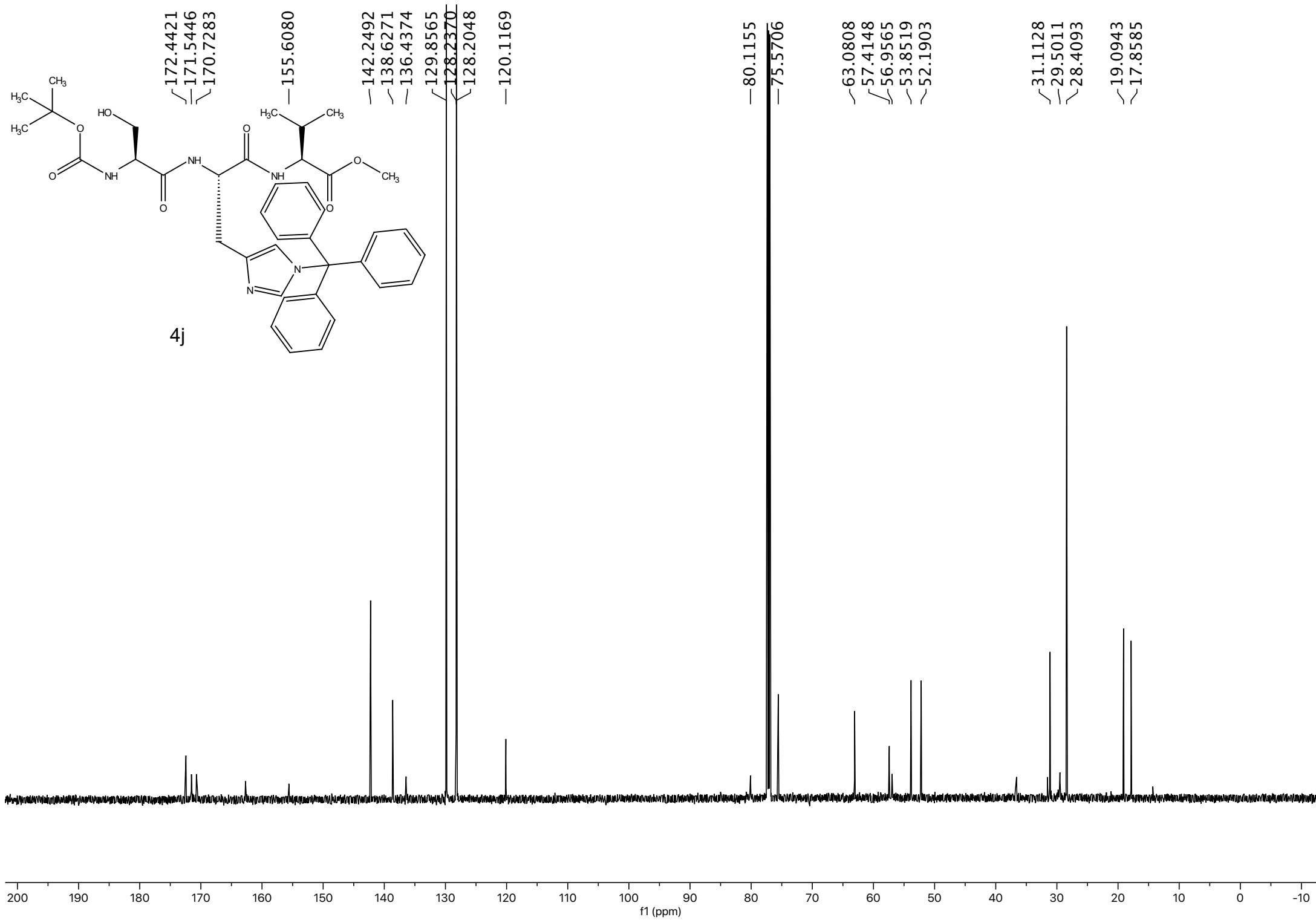


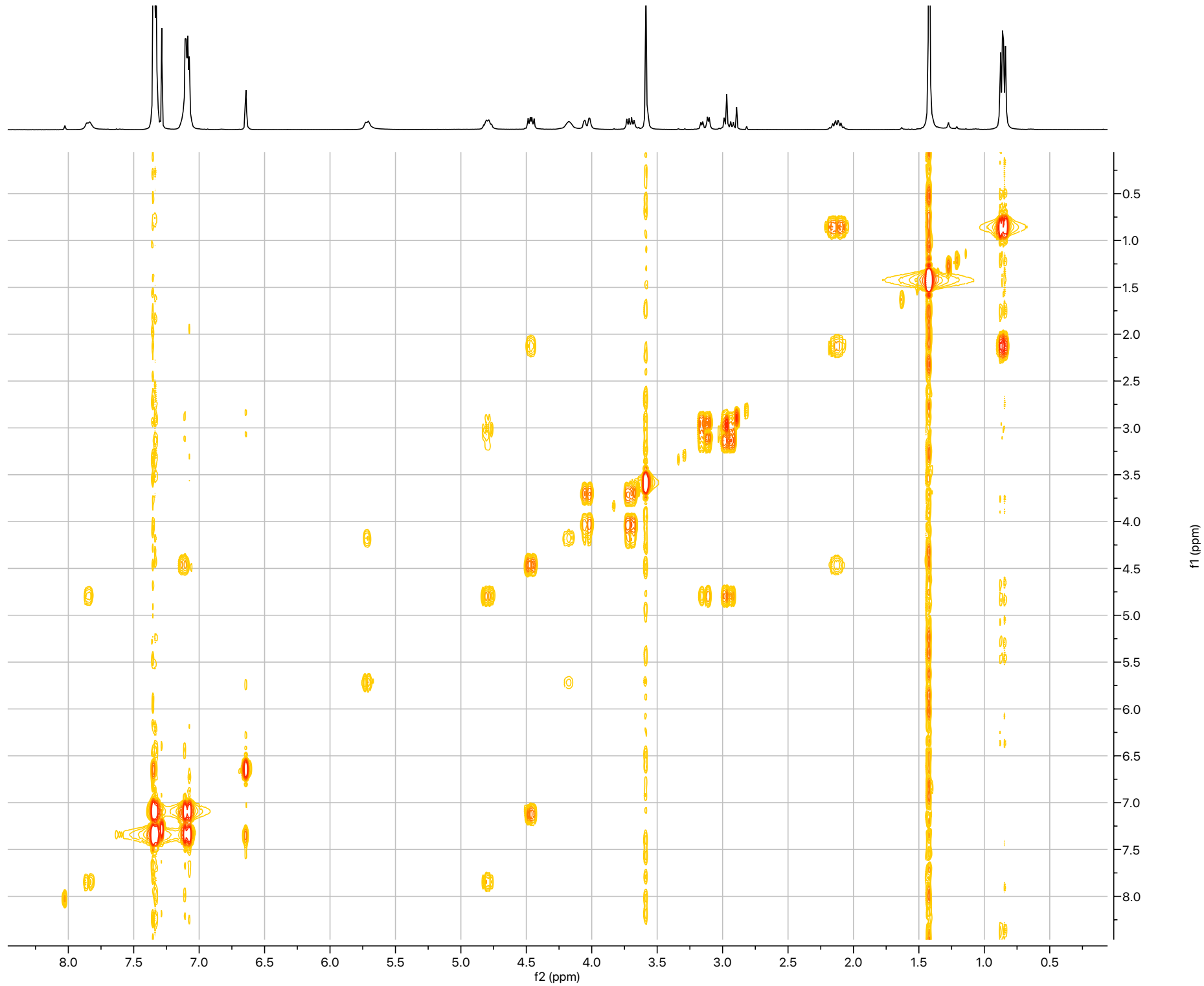


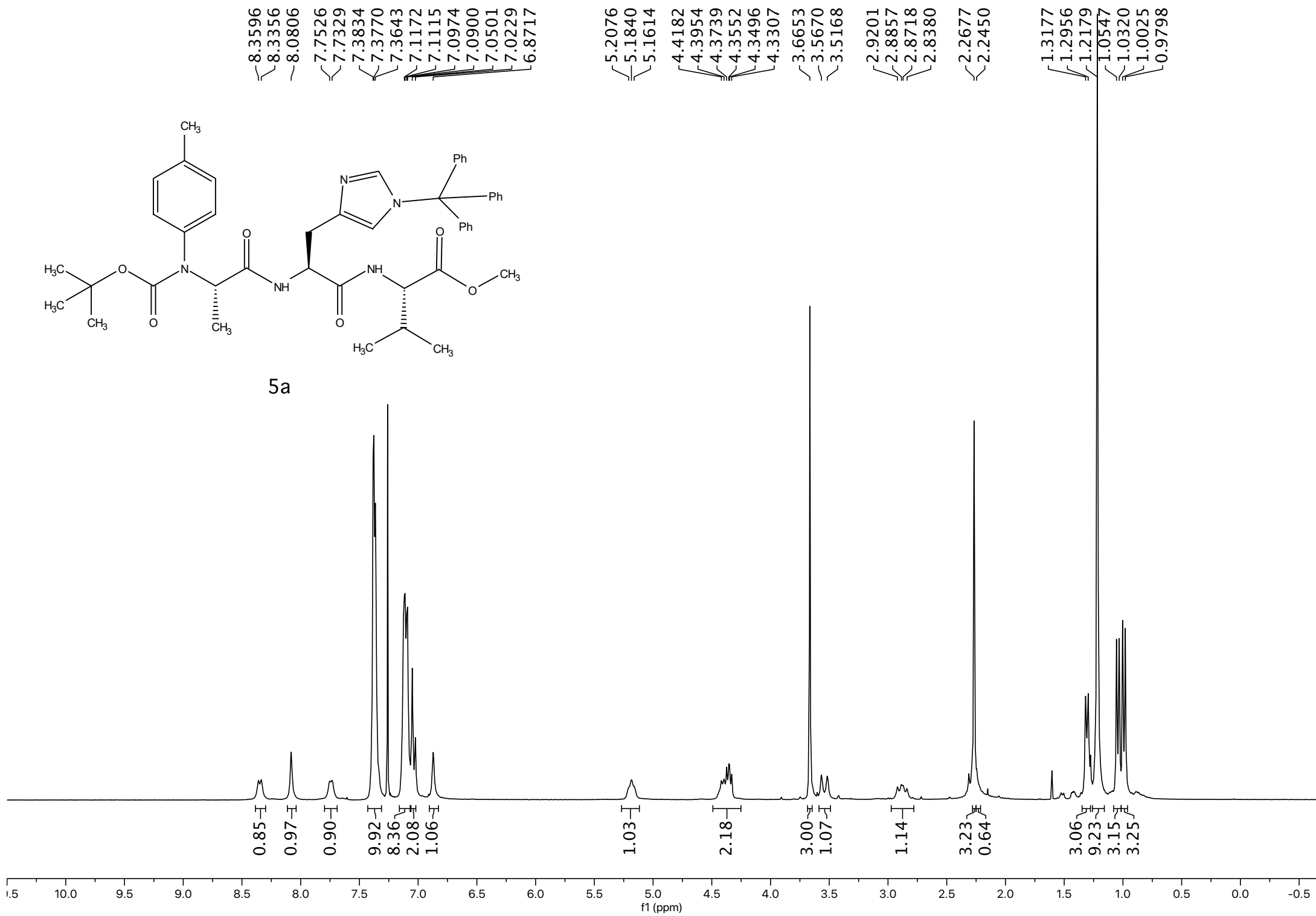




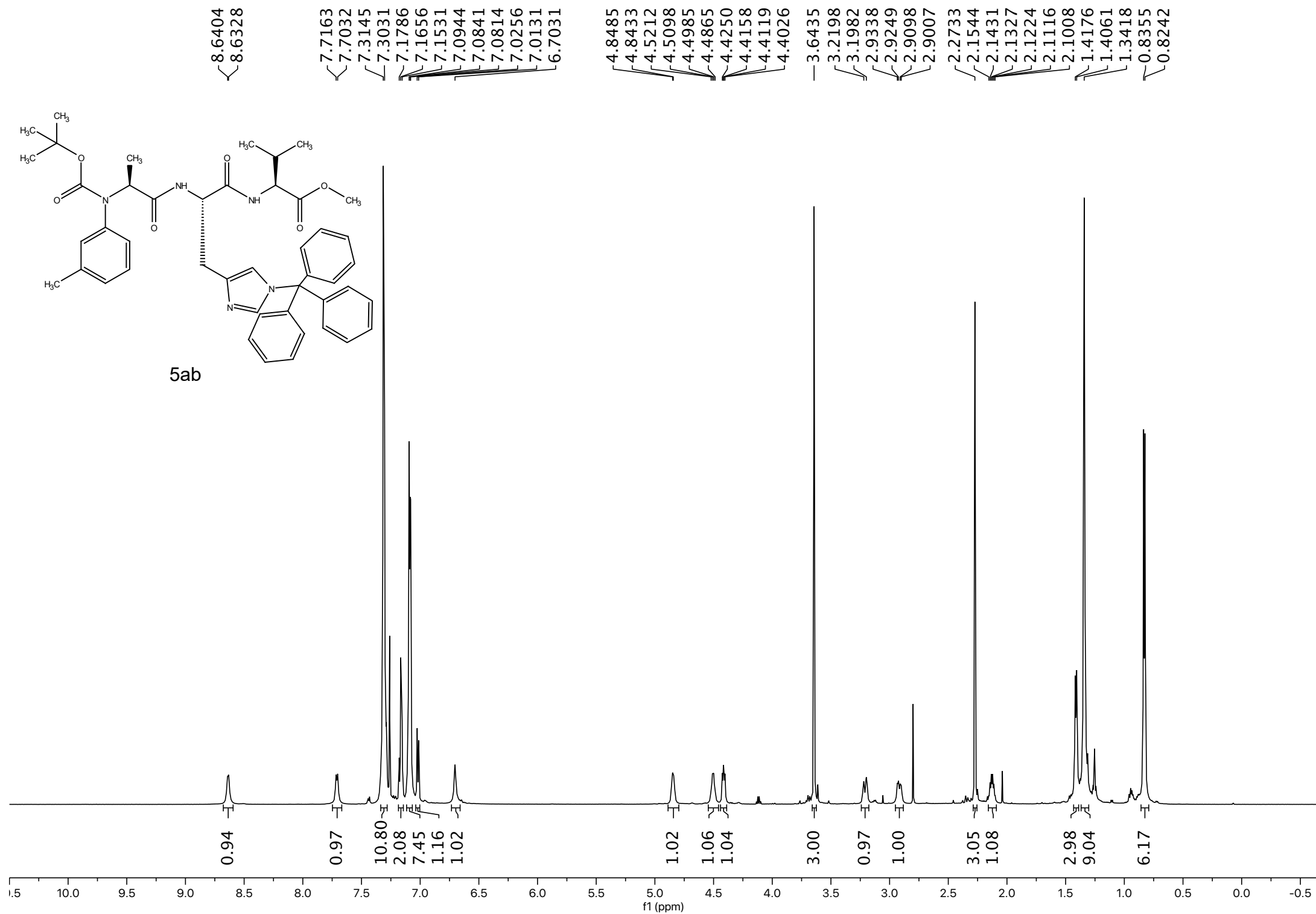
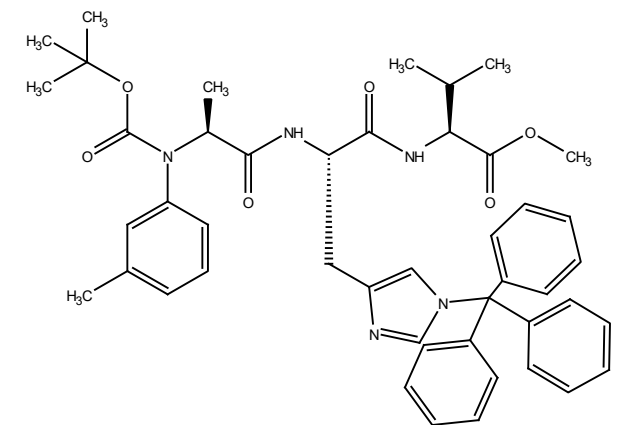








5ab



172.0989
172.0655
171.3948

154.7062

142.4150
141.3130
138.5670
138.1657
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129.8668
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128.5622
128.1808
128.1541
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125.2298
119.6418

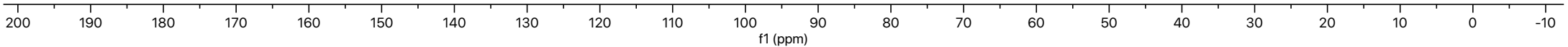
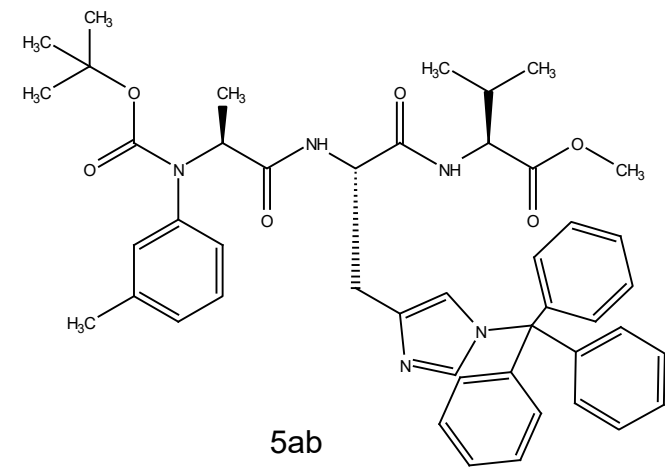
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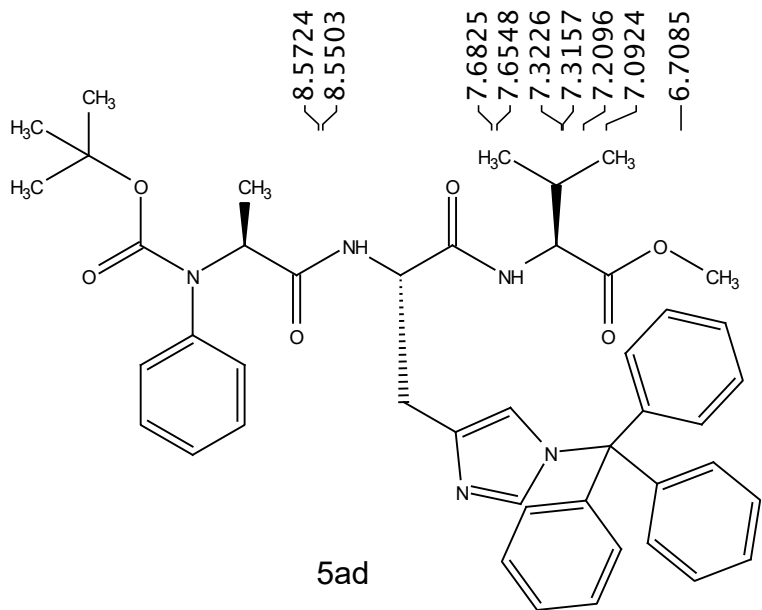
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57.7178
53.3322
52.0243

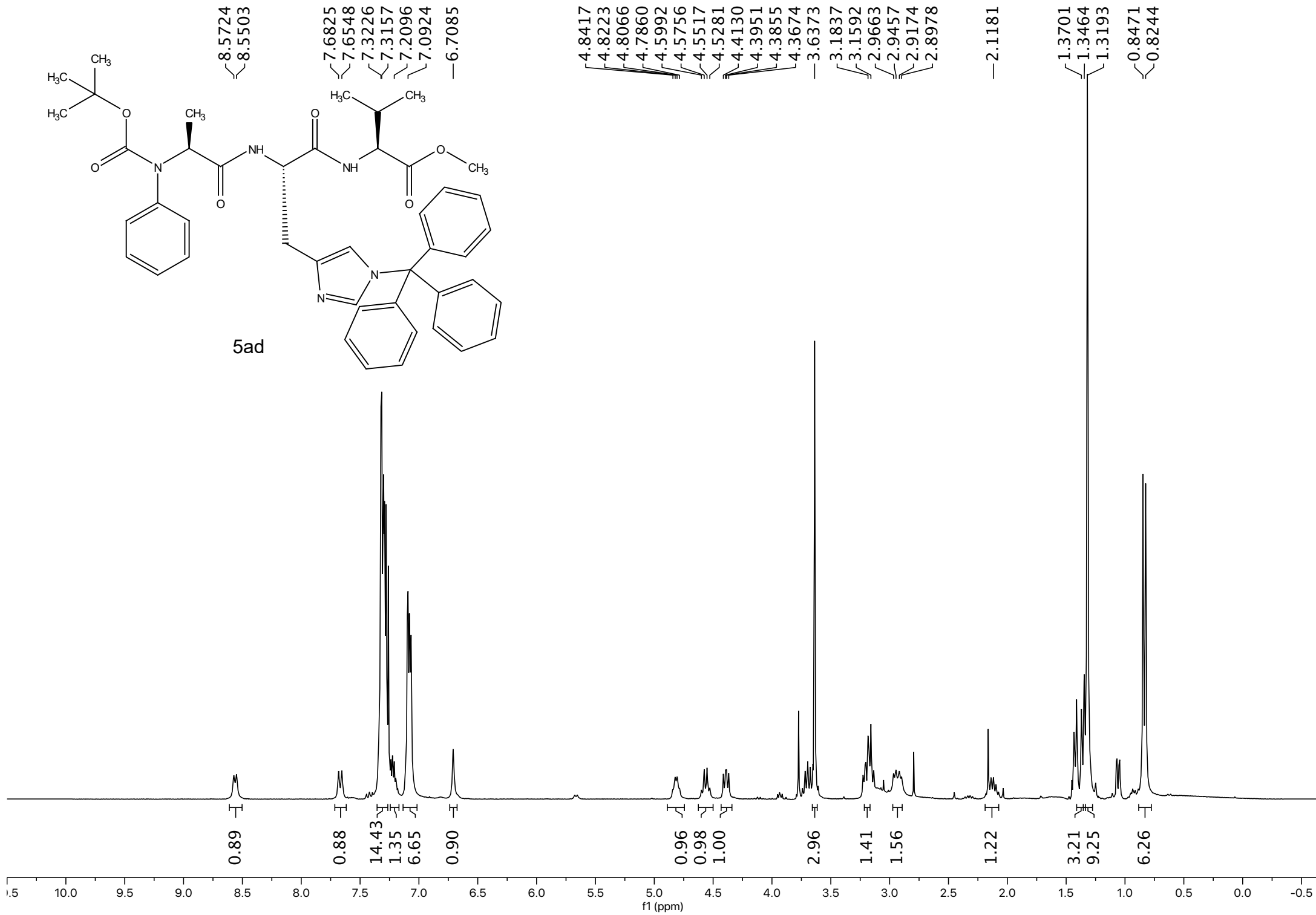
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21.4057
19.1830
18.0710
15.7118

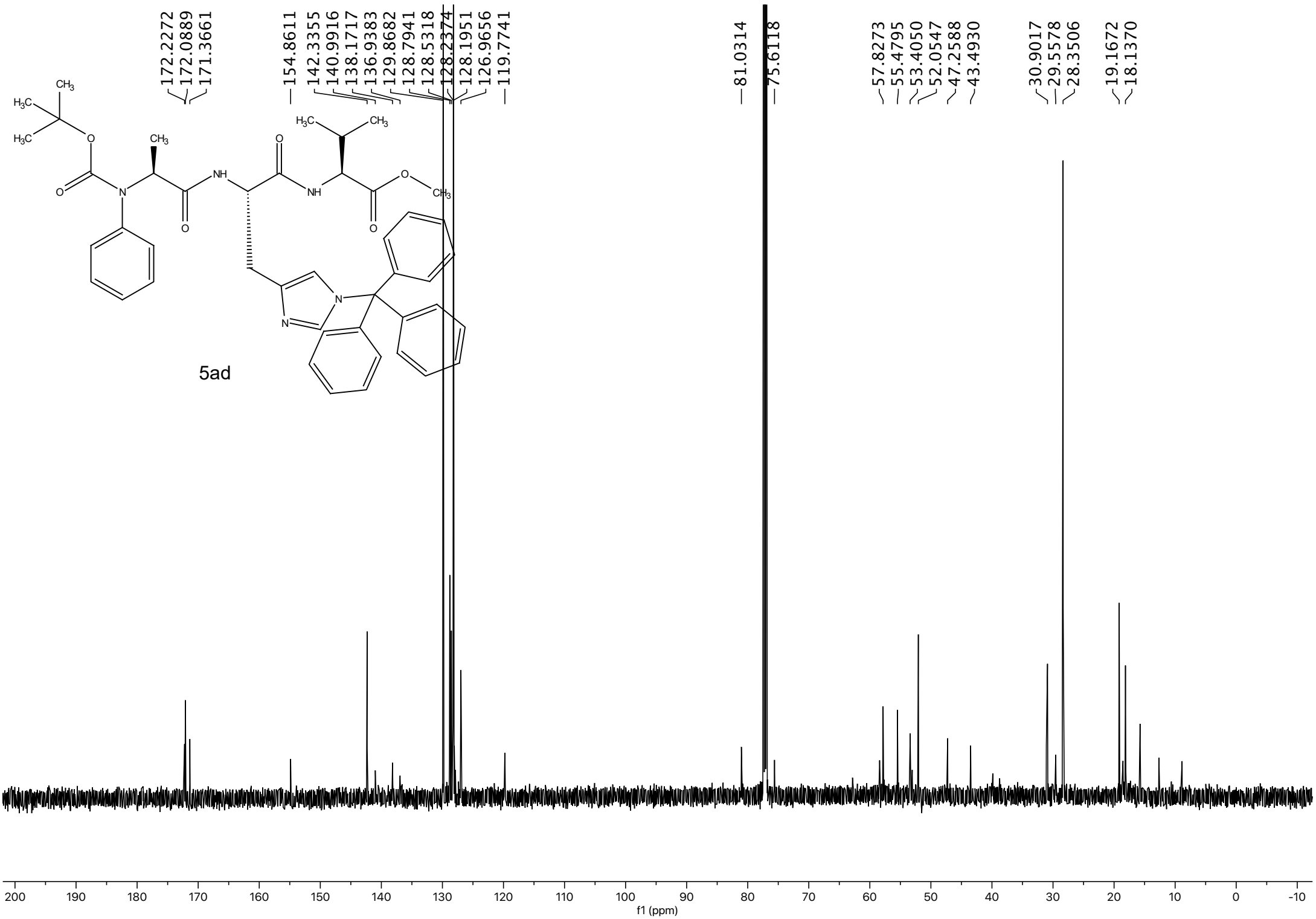
5ab

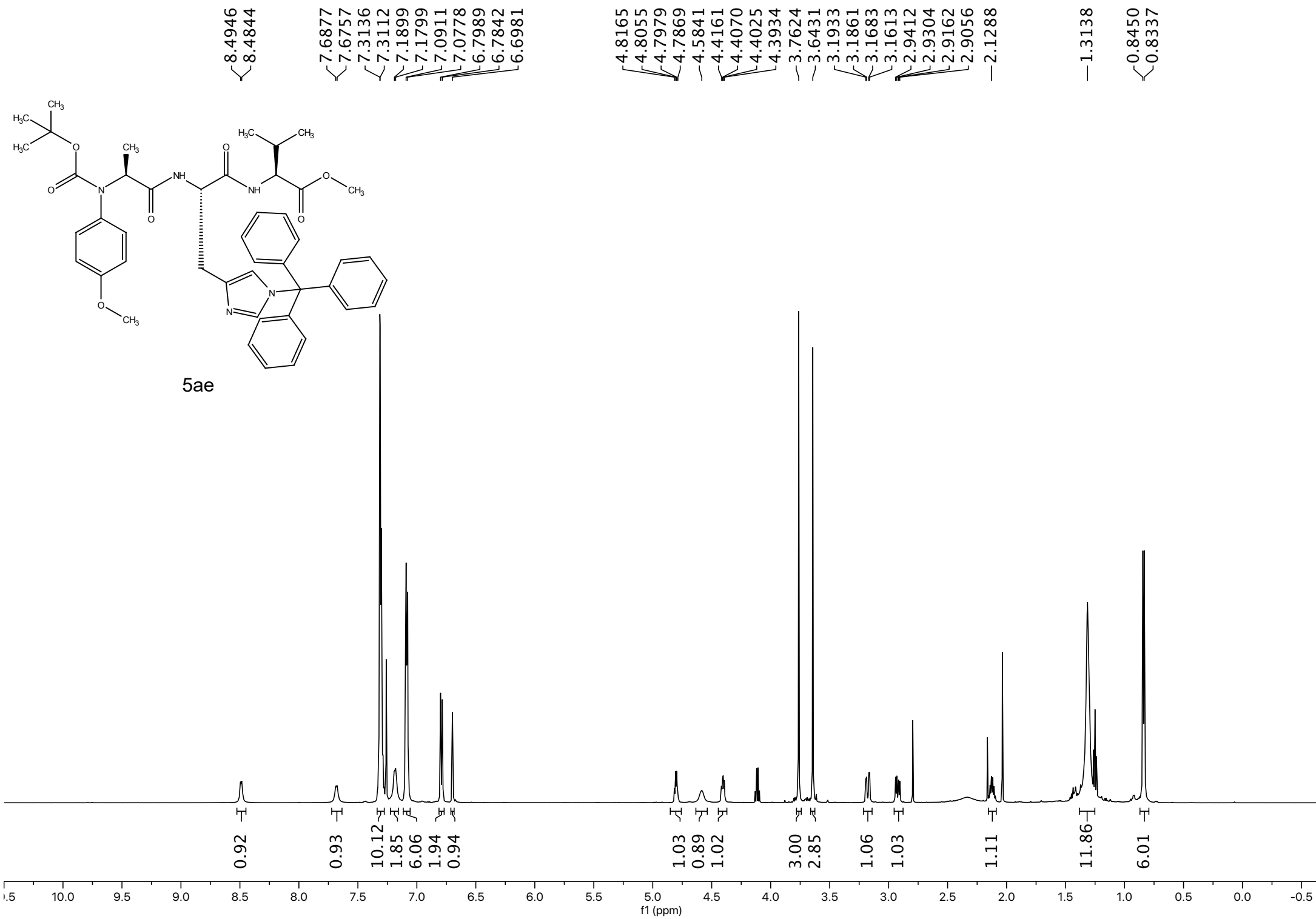


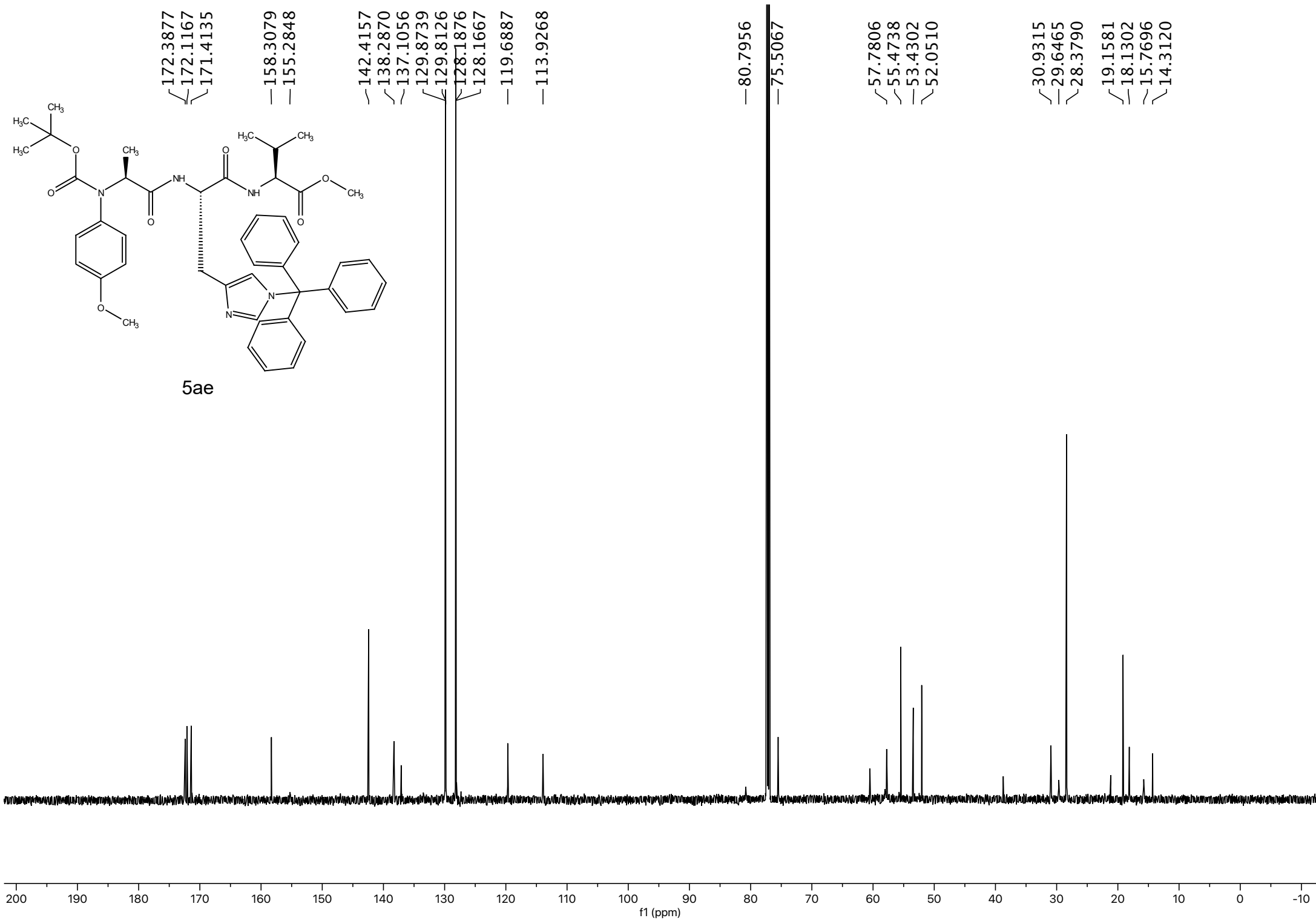


5ad

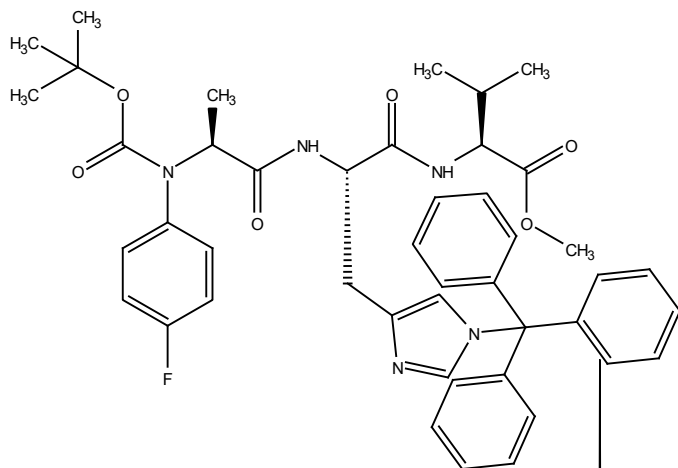




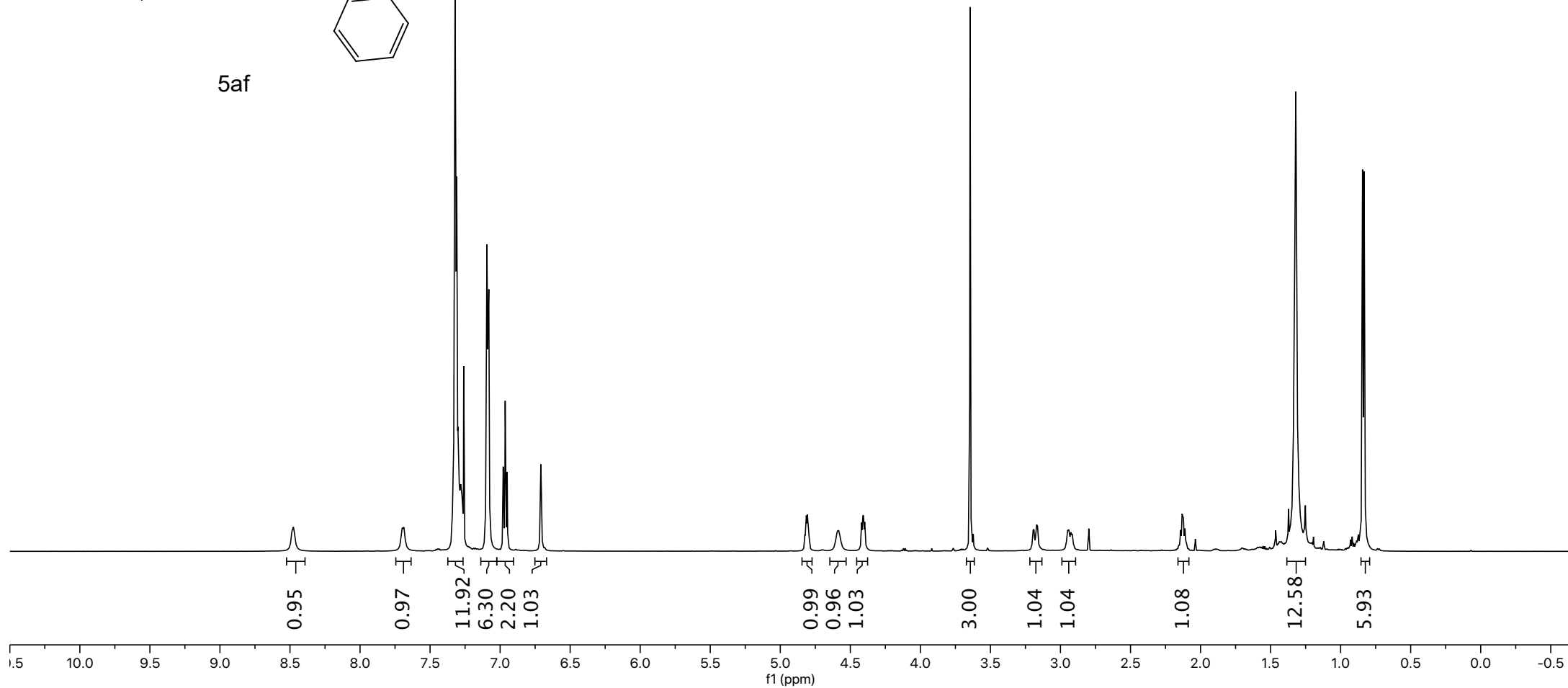


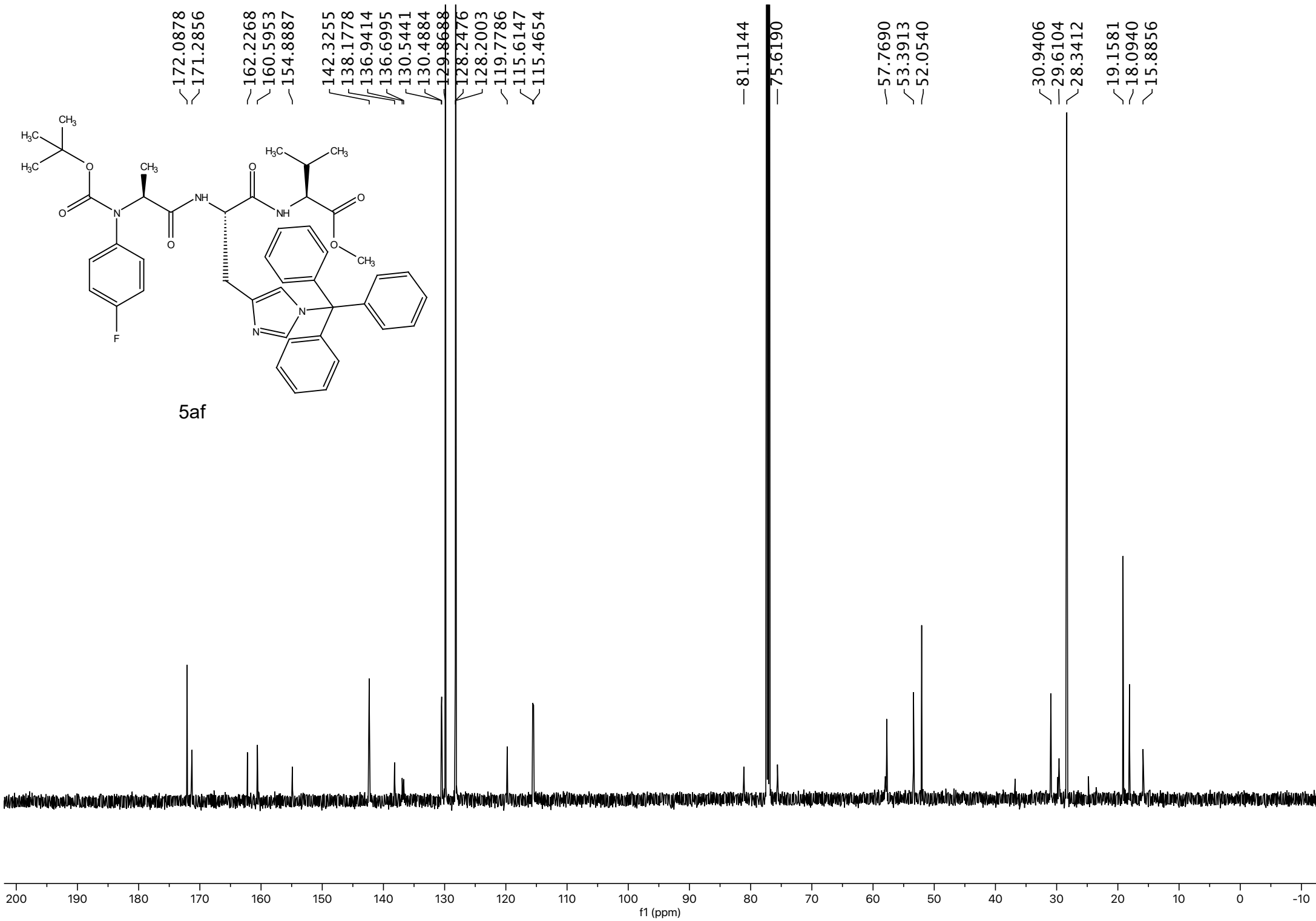


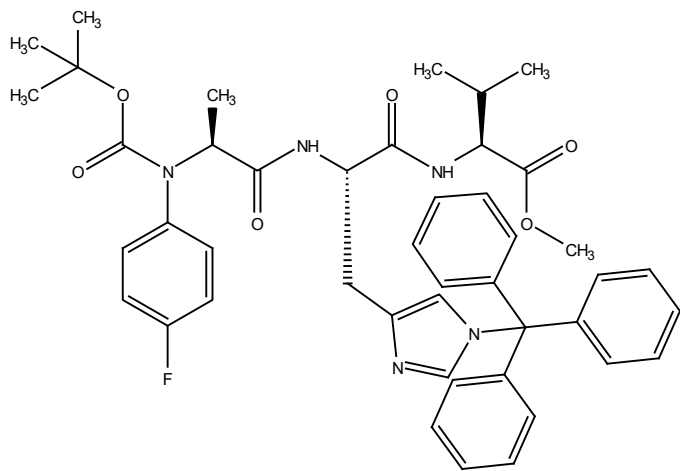
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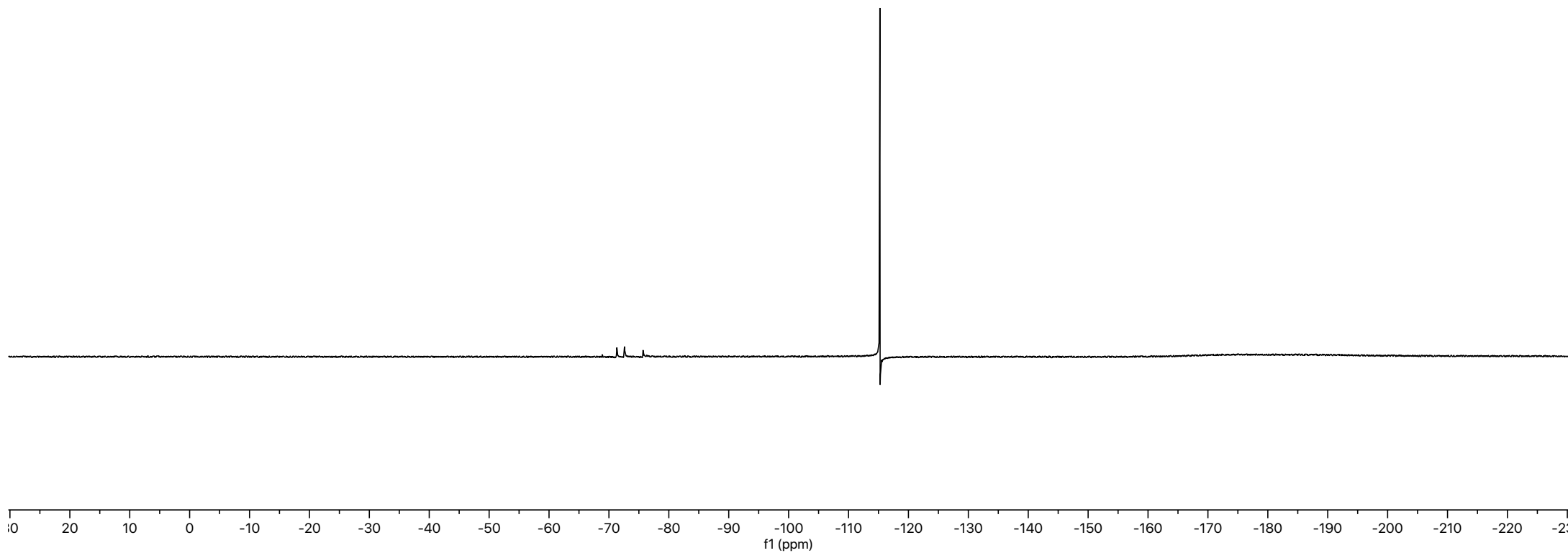
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7.6876
7.3214
7.3192
7.0939
6.9779
6.9637
6.9493
6.7092
4.8243
4.8142
4.8061
4.7958
4.5828
4.4194
4.4102
4.4062
4.3971
3.6450
3.1949
3.1895
3.1703
3.1649
2.9490
2.9400
2.9251
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2.1431
2.1319
2.1221
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2.1000
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0.8436
0.8323

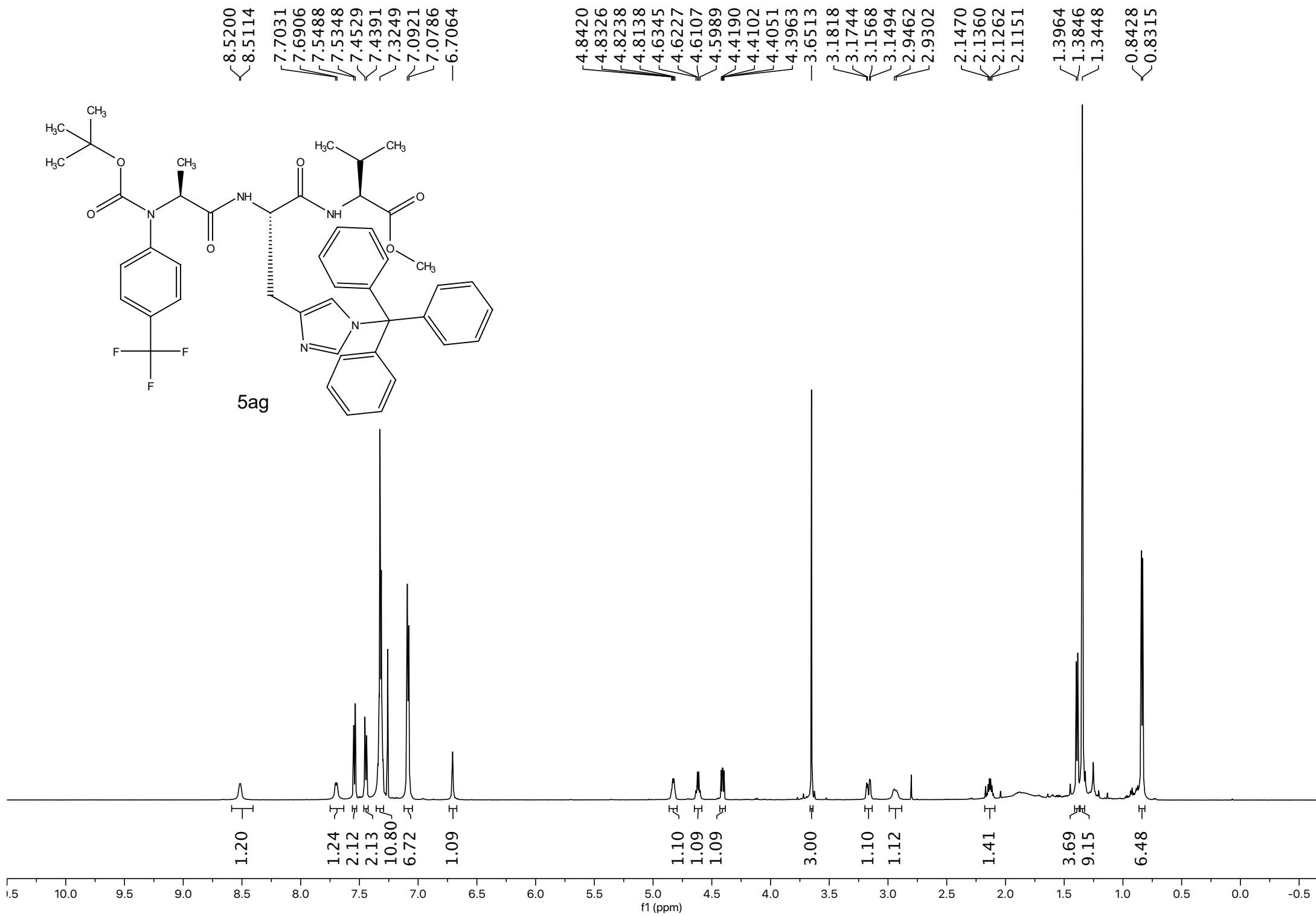


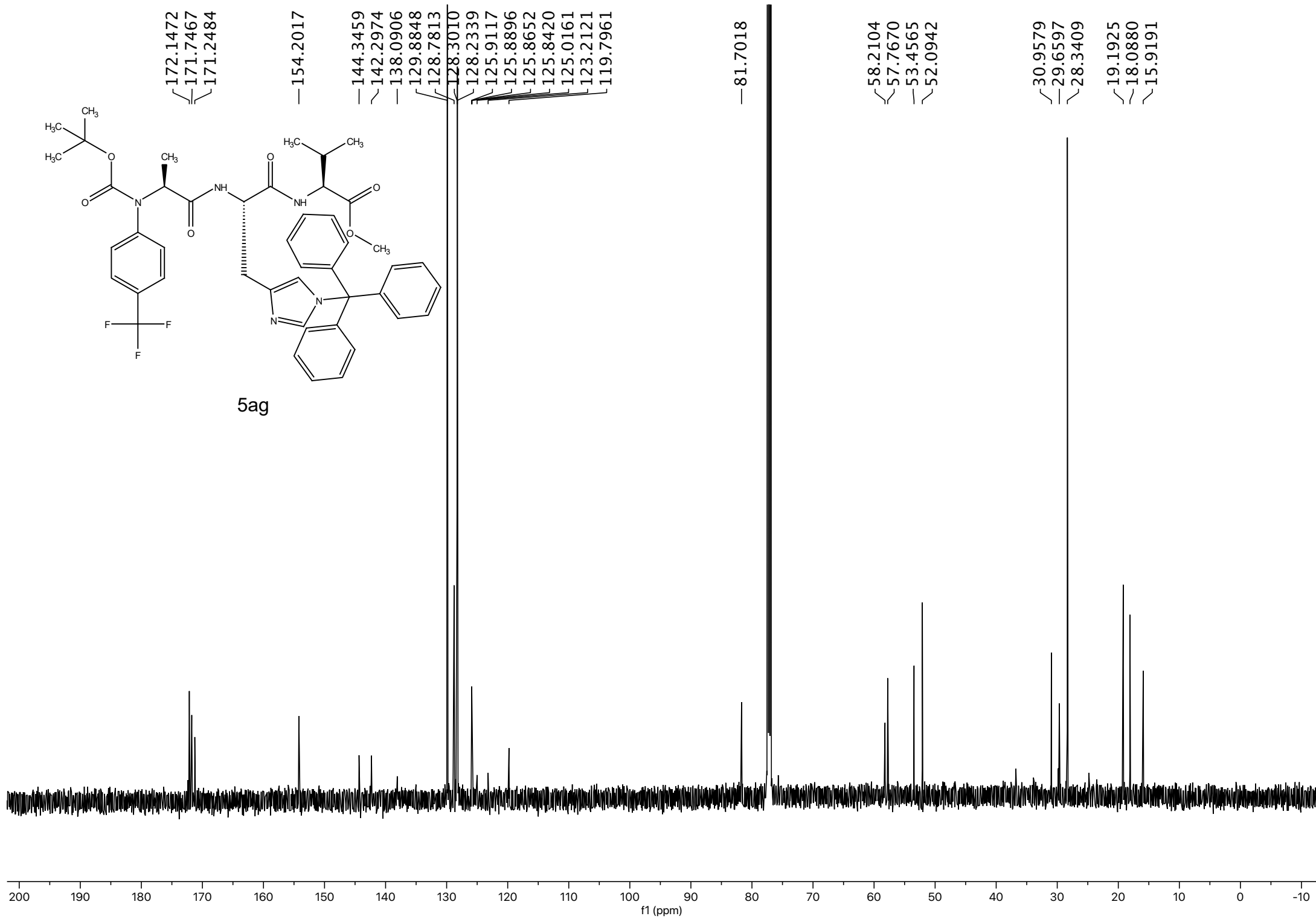


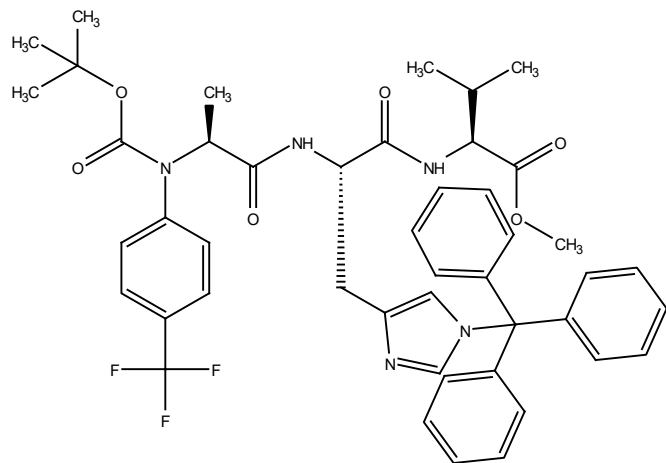


5af



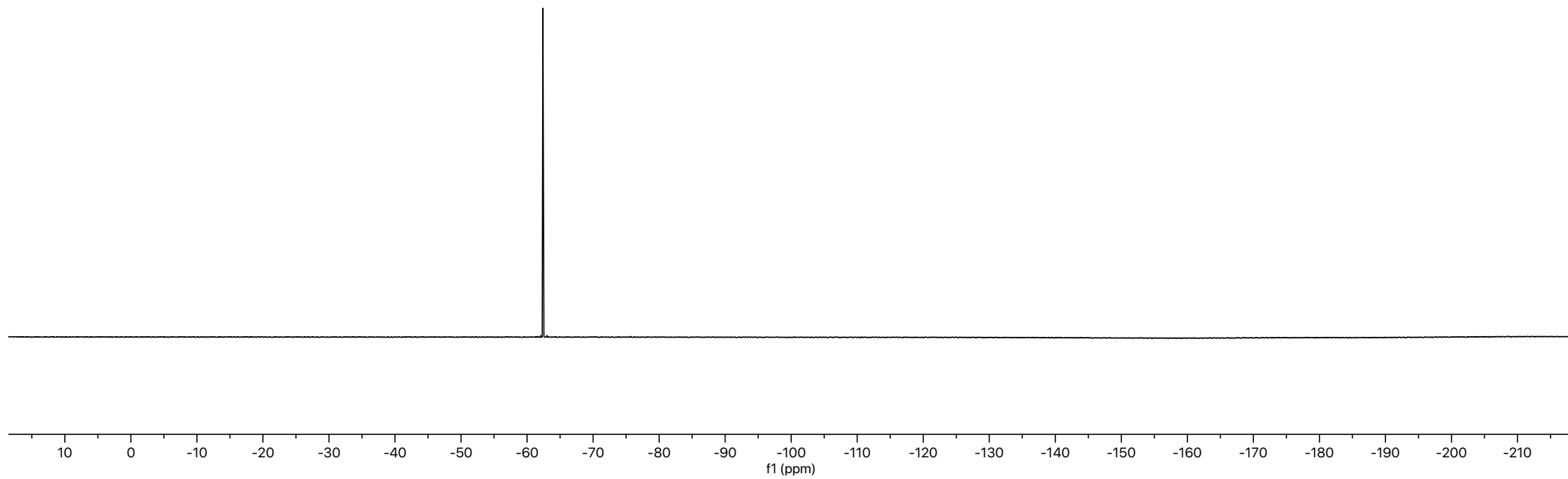




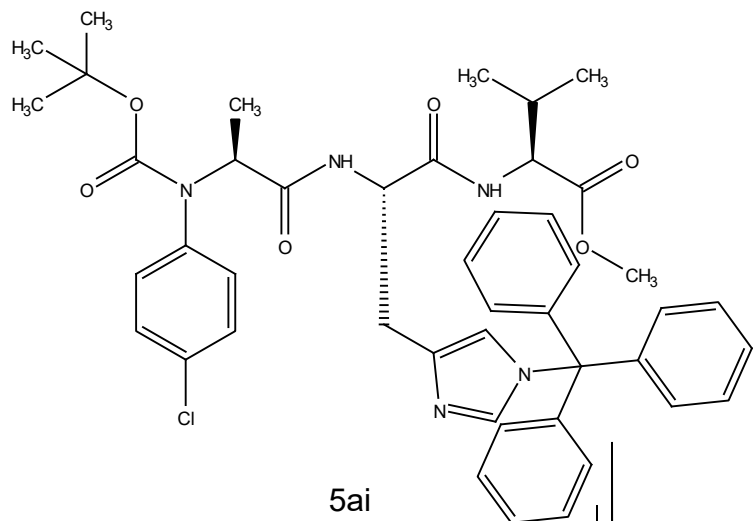


5ah

--62.3989



5ai



8.4635
8.4544

7.6821
7.6685
7.3253
7.3236
7.2477
7.0939
7.0842
7.0806
6.7114

4.8289
4.8184
4.8099
4.7994
4.6038
4.5929
4.5812
4.5705
4.4181
4.4092
4.4043
4.3954
3.6481
3.1910
3.1837
3.1660
3.1588
2.9547
2.9449
2.9300
2.9201
2.1344
2.1244
2.1133

1.3436
1.3256

0.8461
0.8347

1.01

1.13

10.82

3.94

6.56

1.06

1.06

1.02

1.08

3.00

1.06

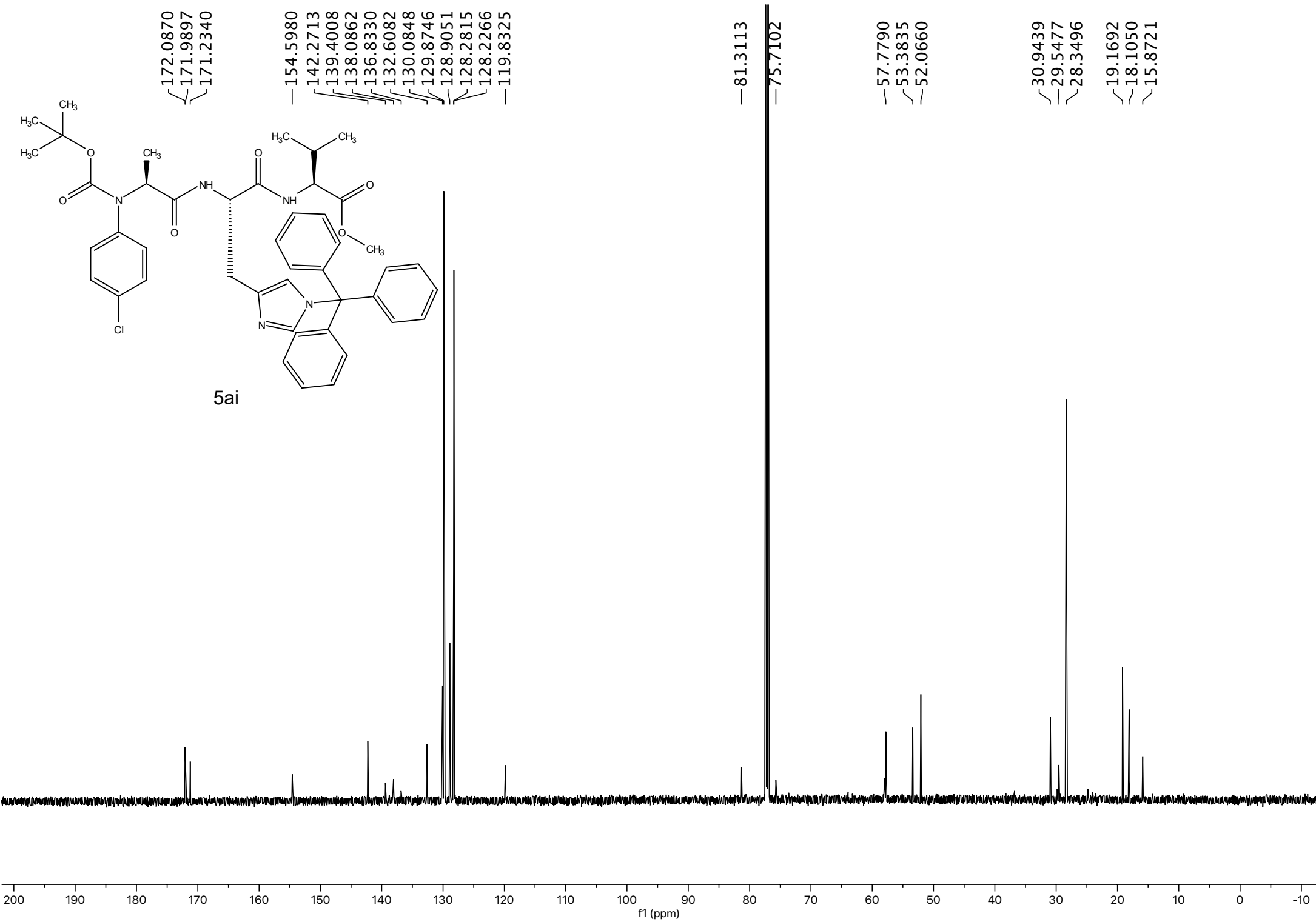
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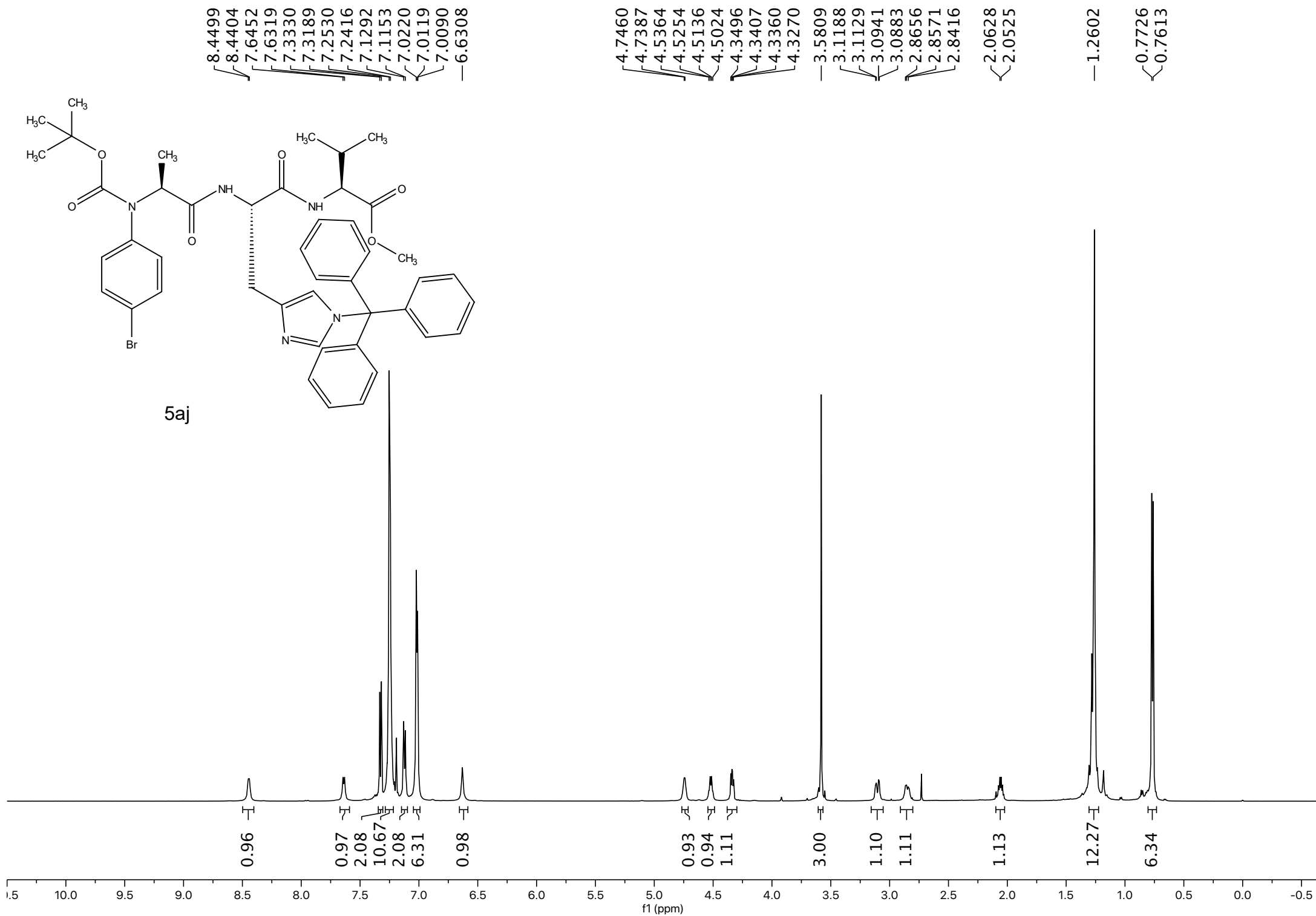
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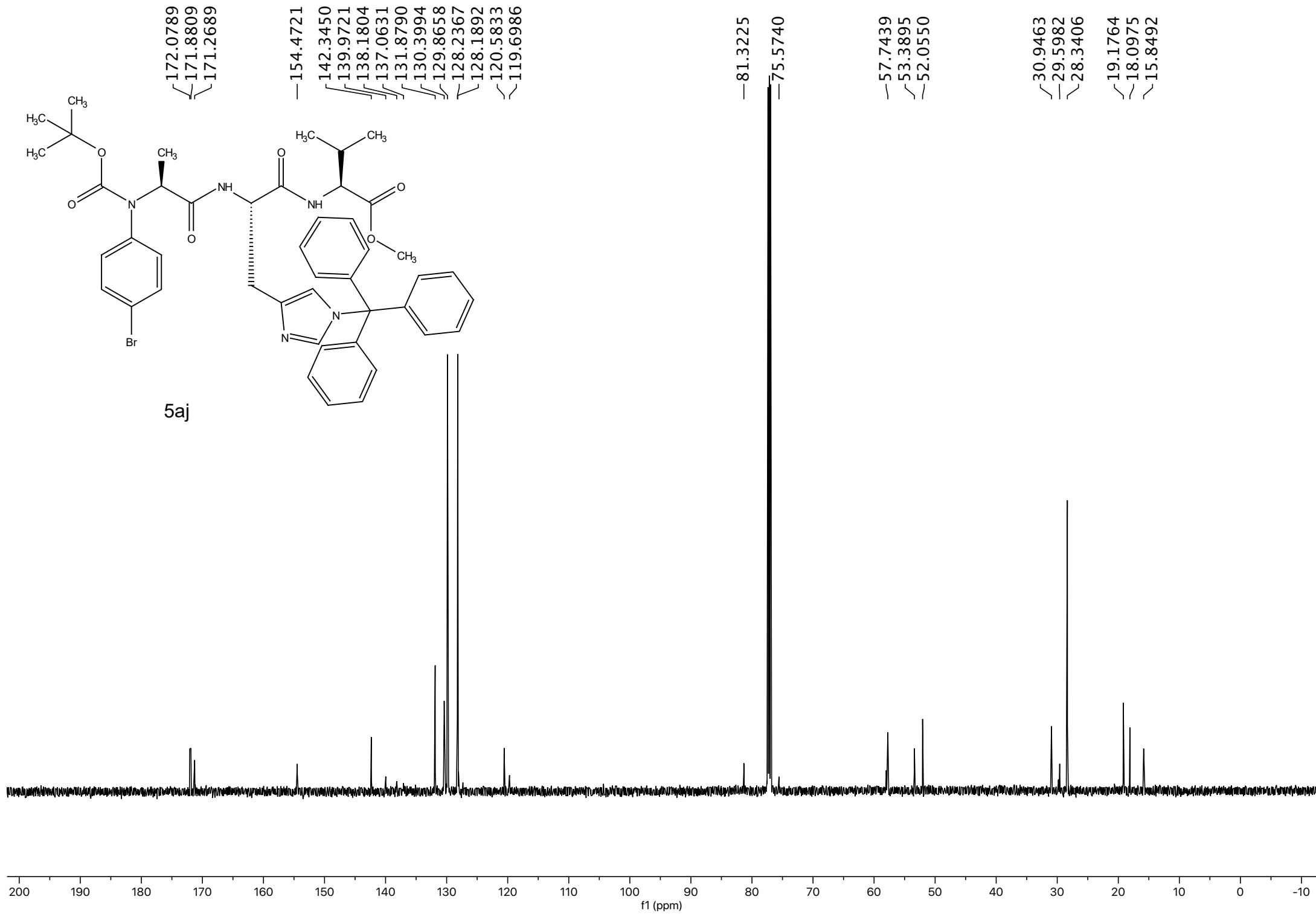
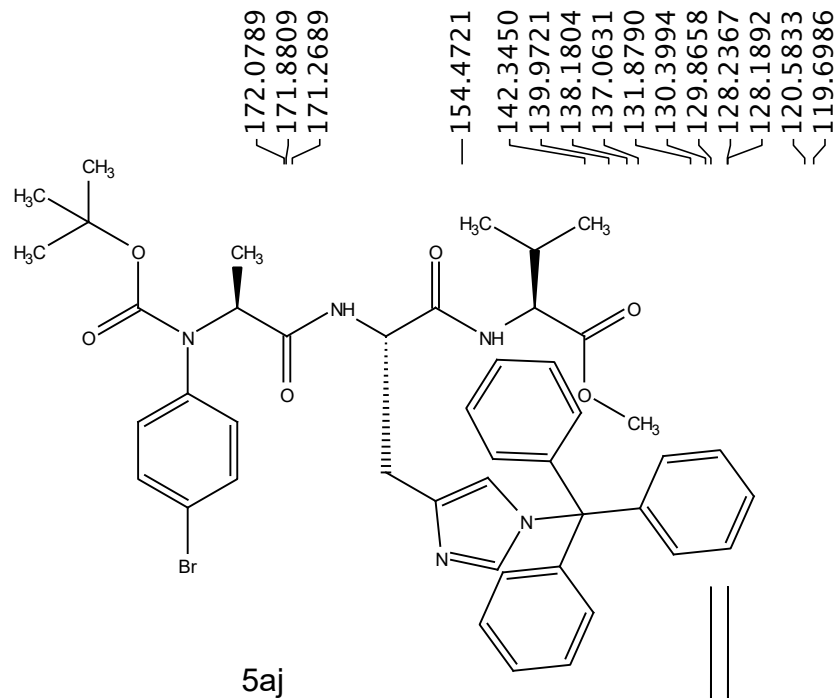
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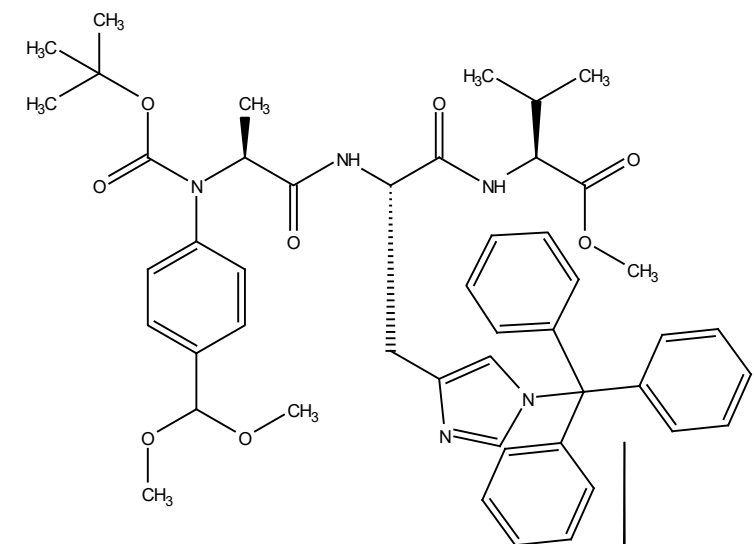
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f1 (ppm)

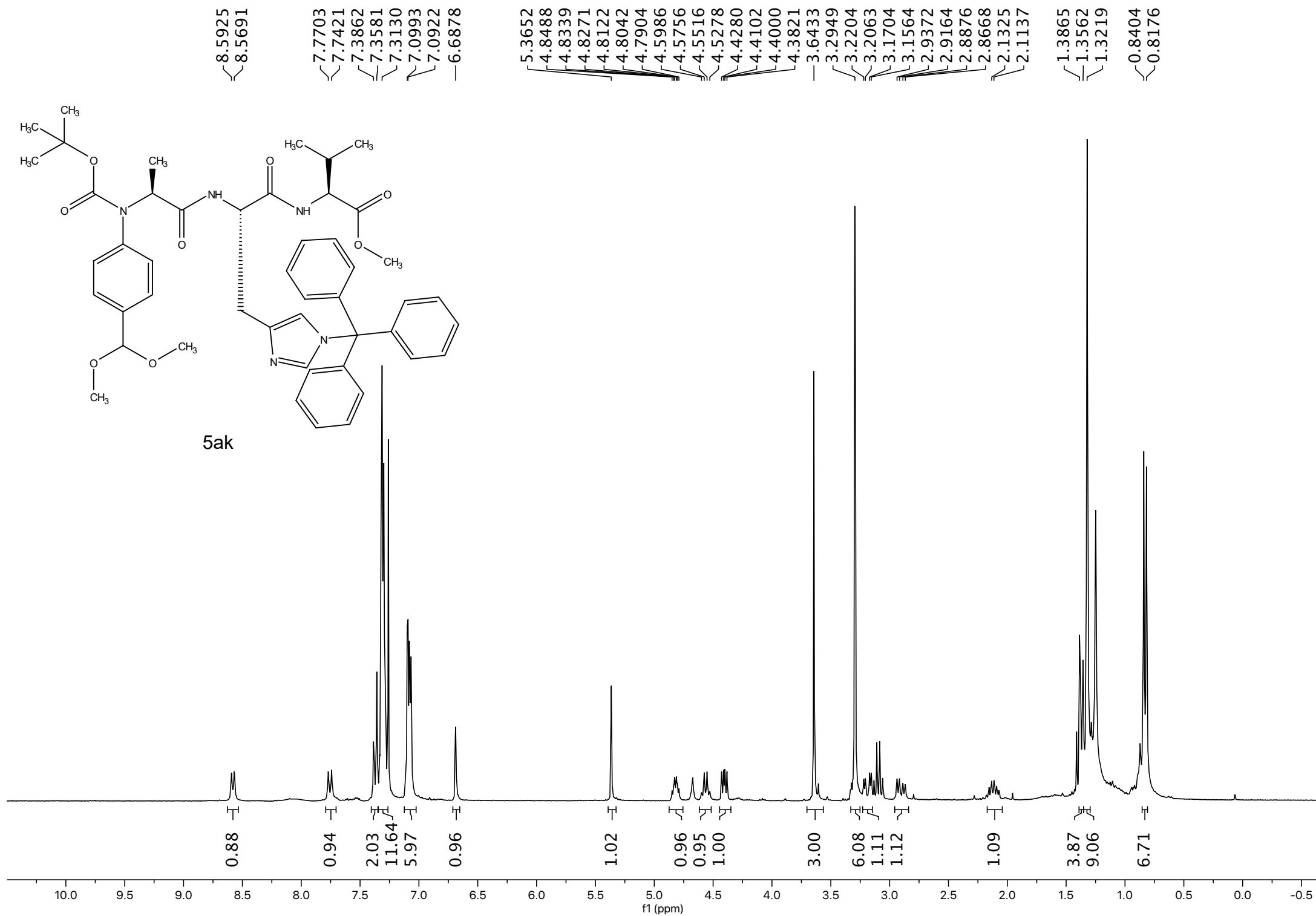


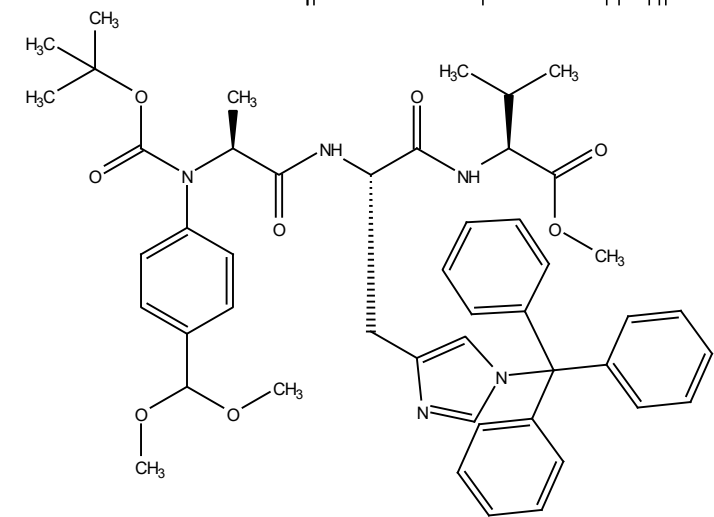






5ak





5ak

172.0728
172.0082
171.4011

154.6916

142.4448
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128.1649
127.1981
119.6354

102.7790

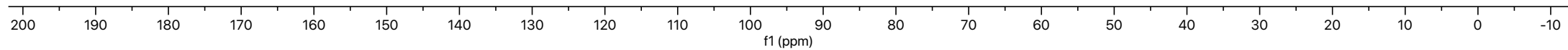
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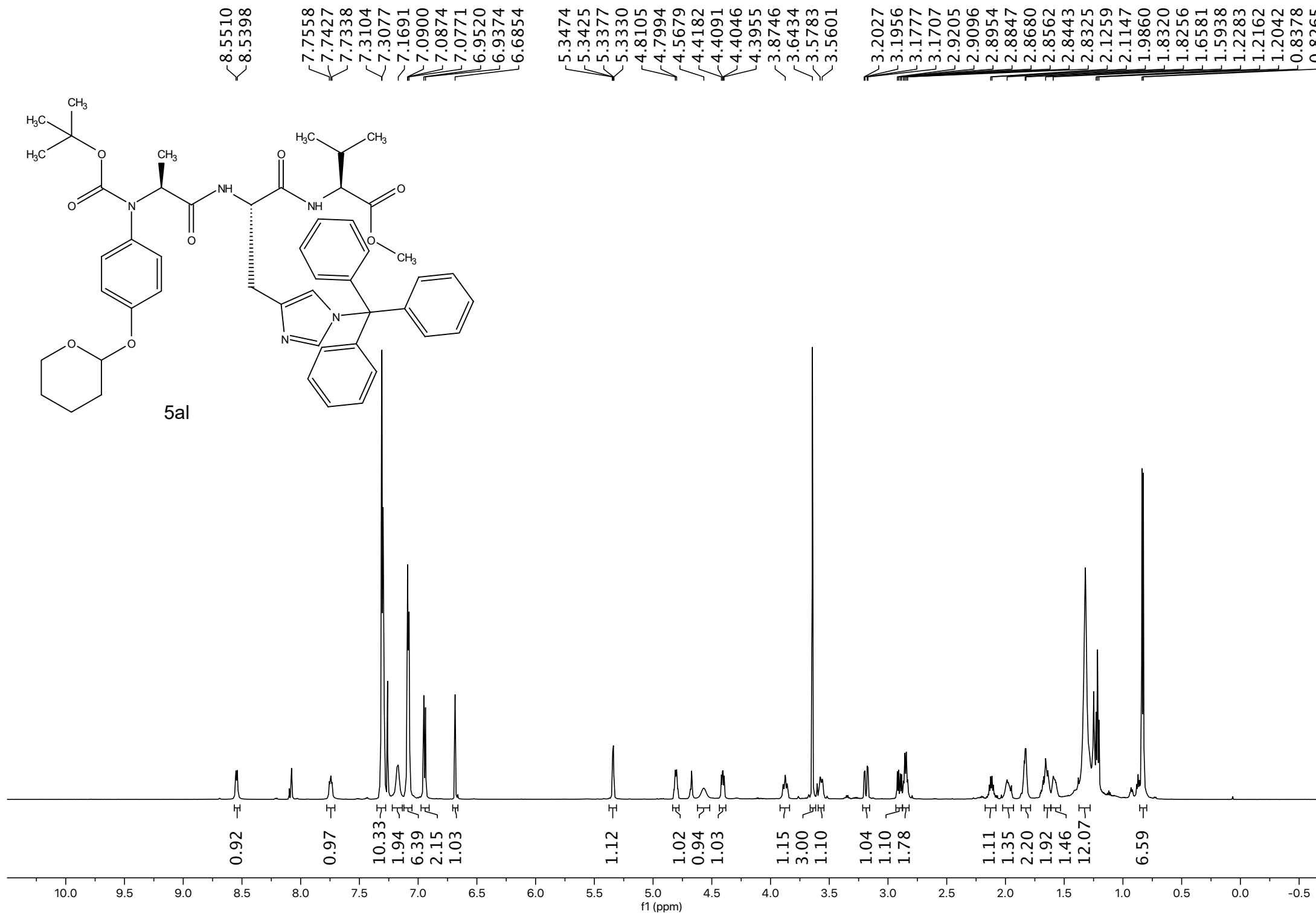
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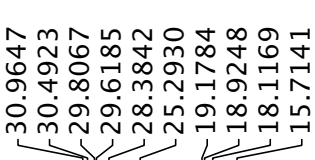
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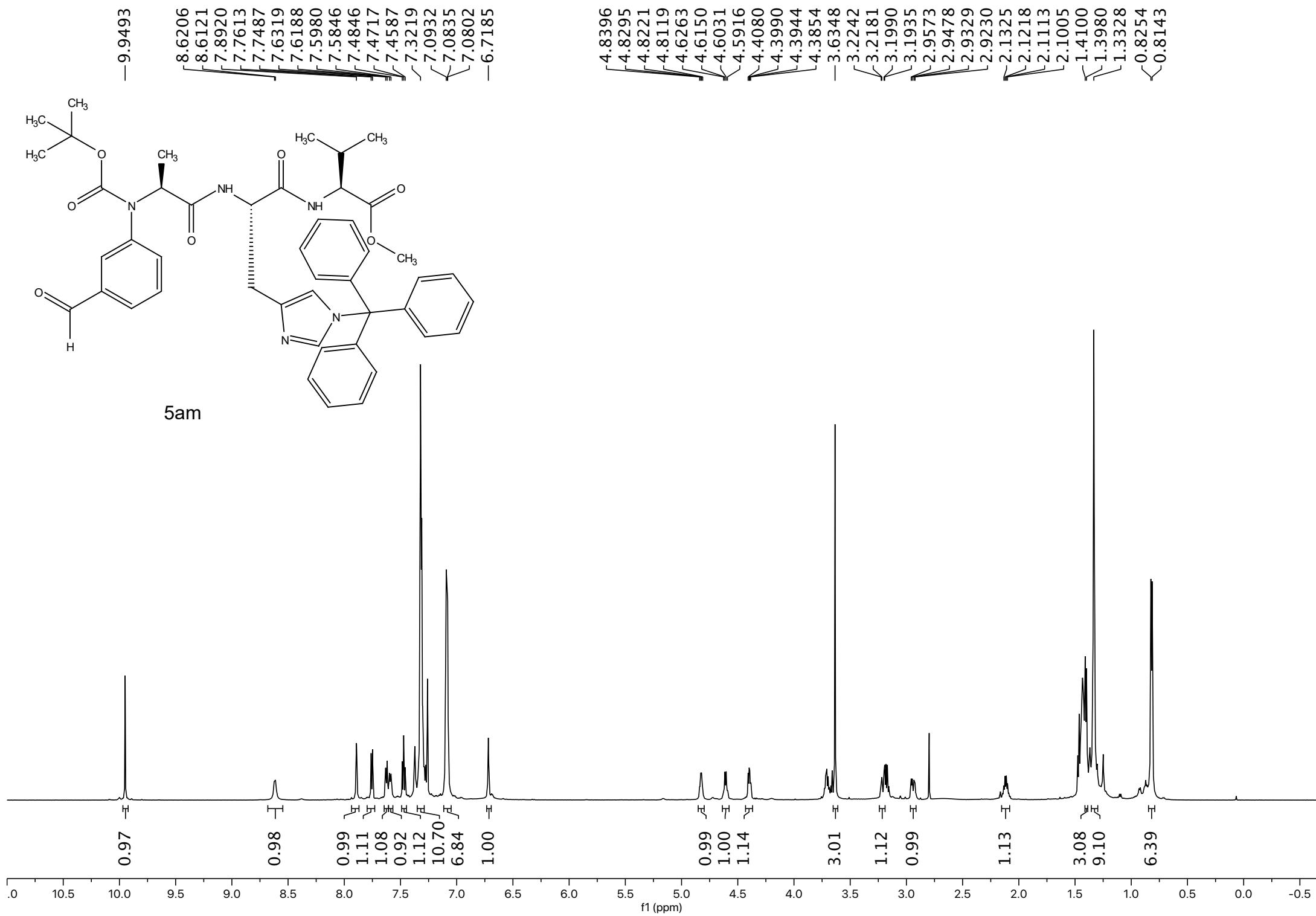
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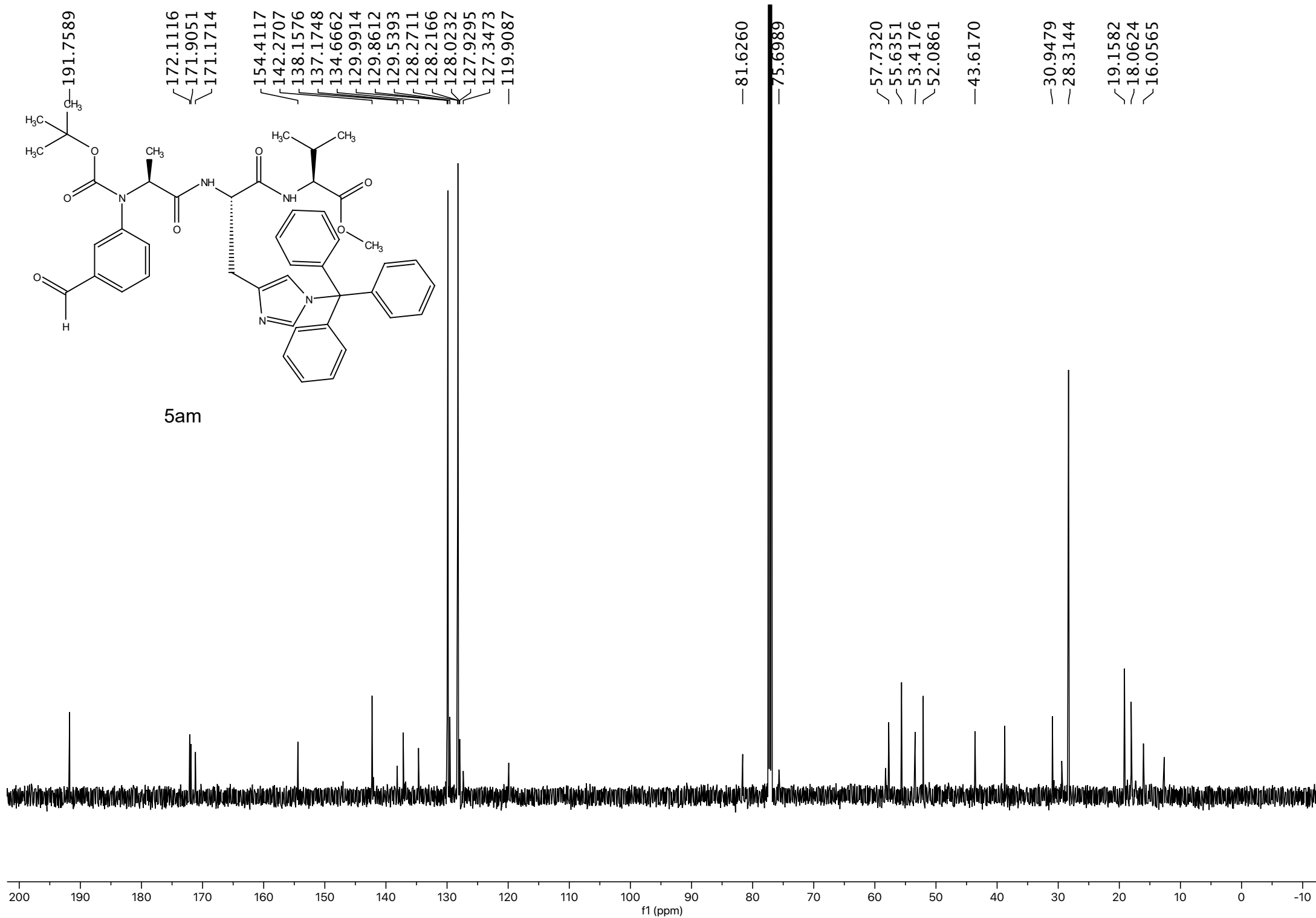
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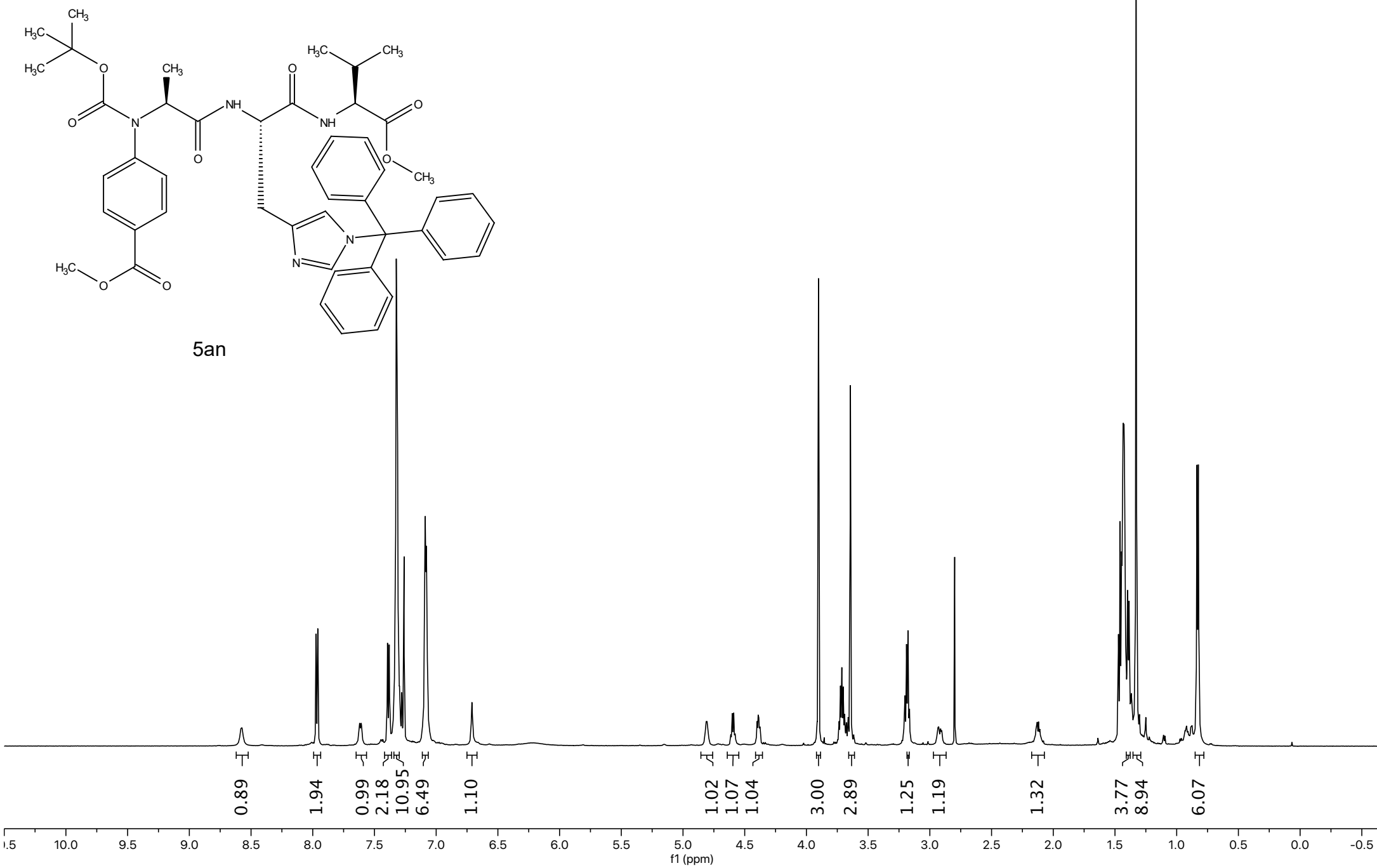


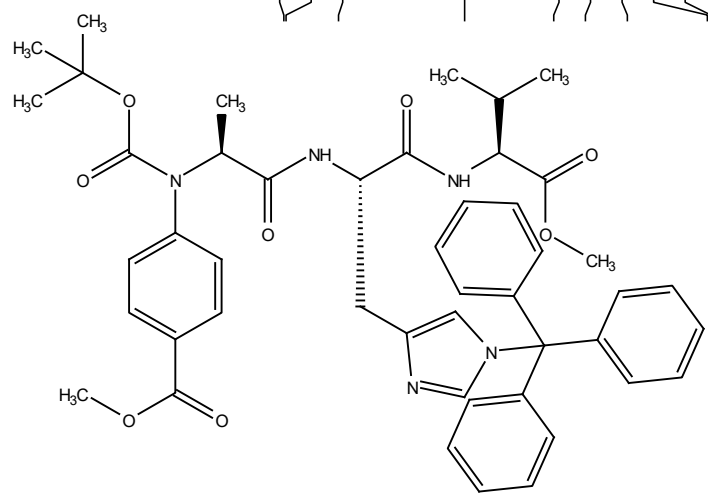












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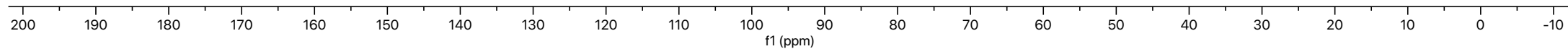
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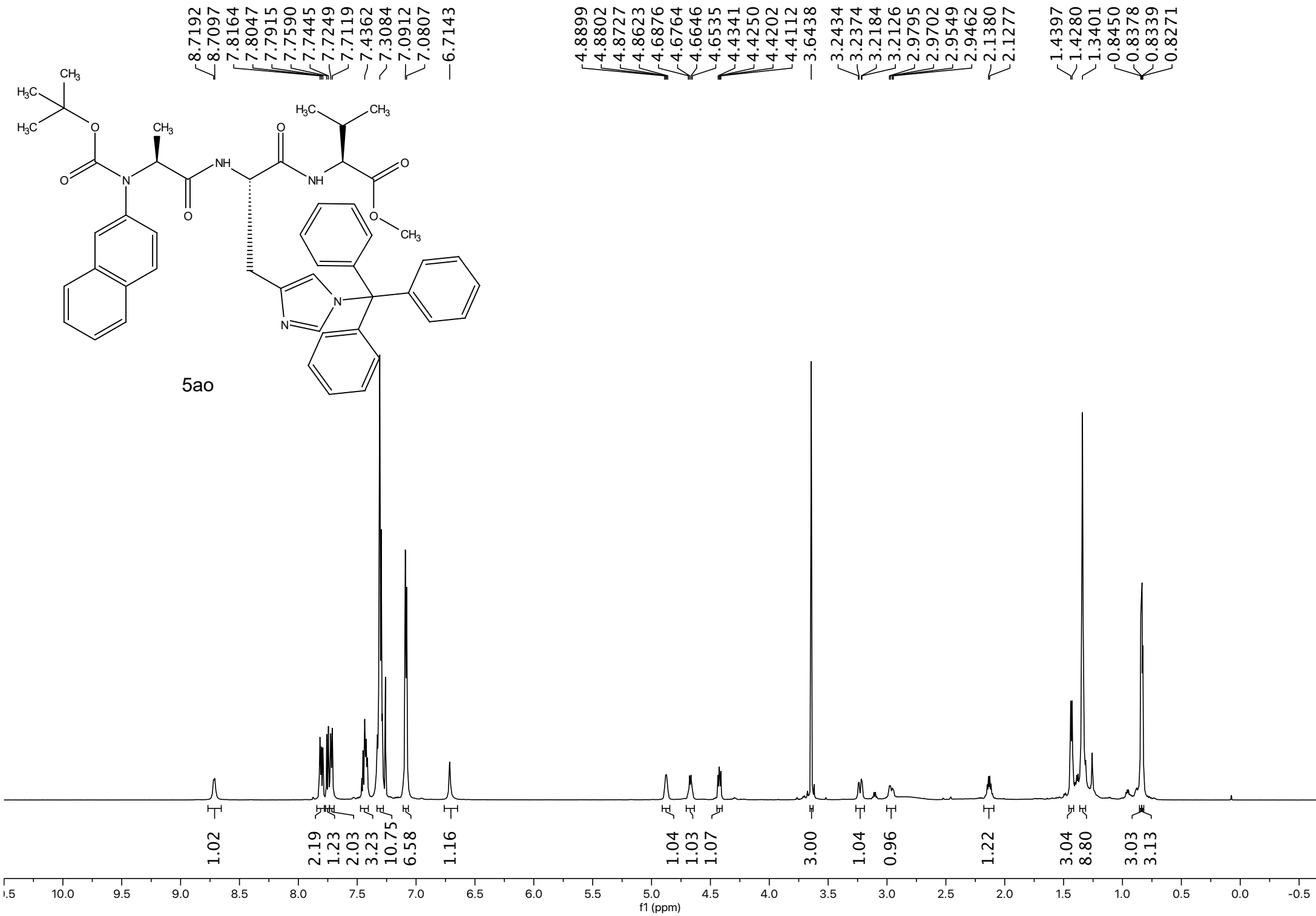
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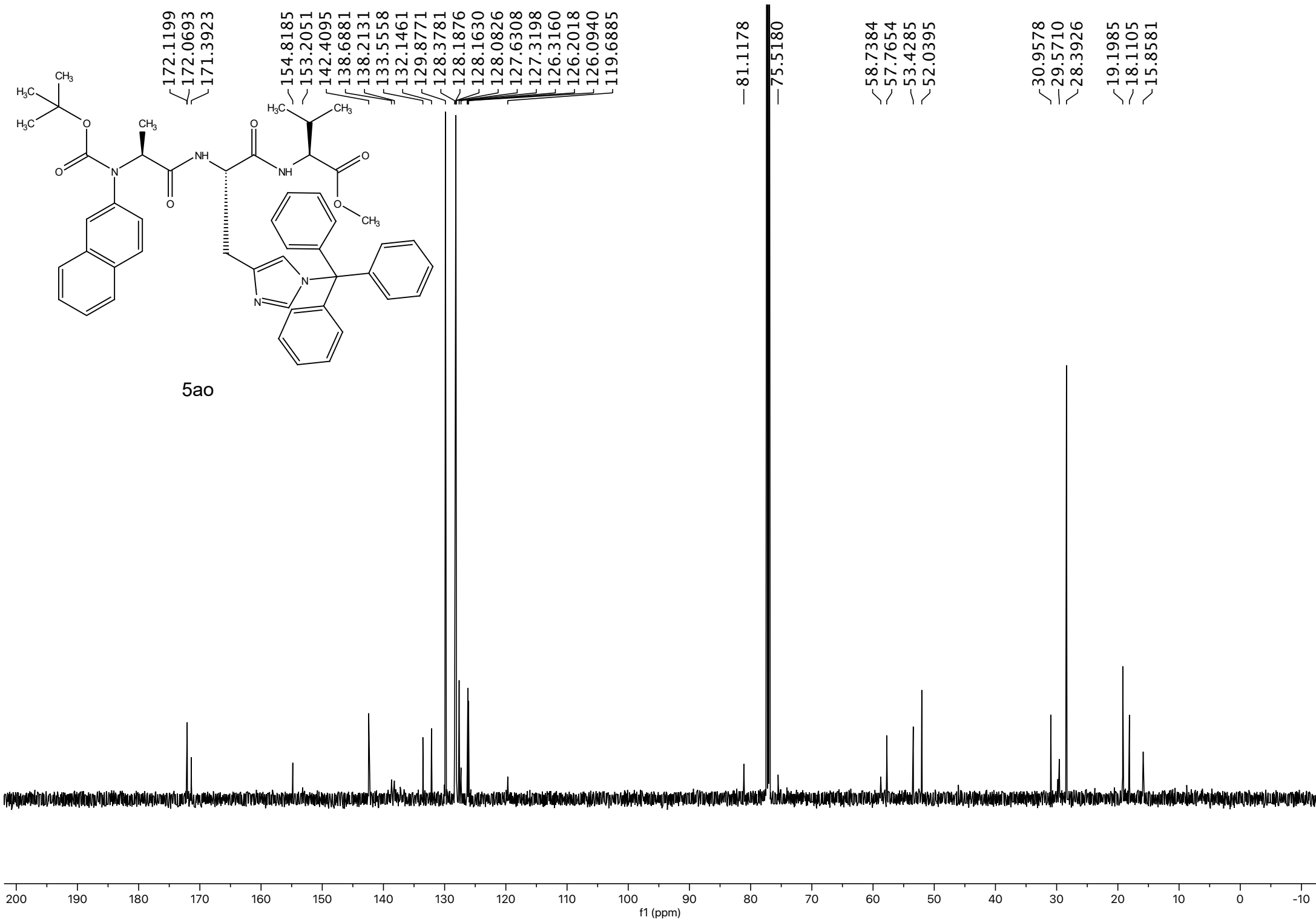
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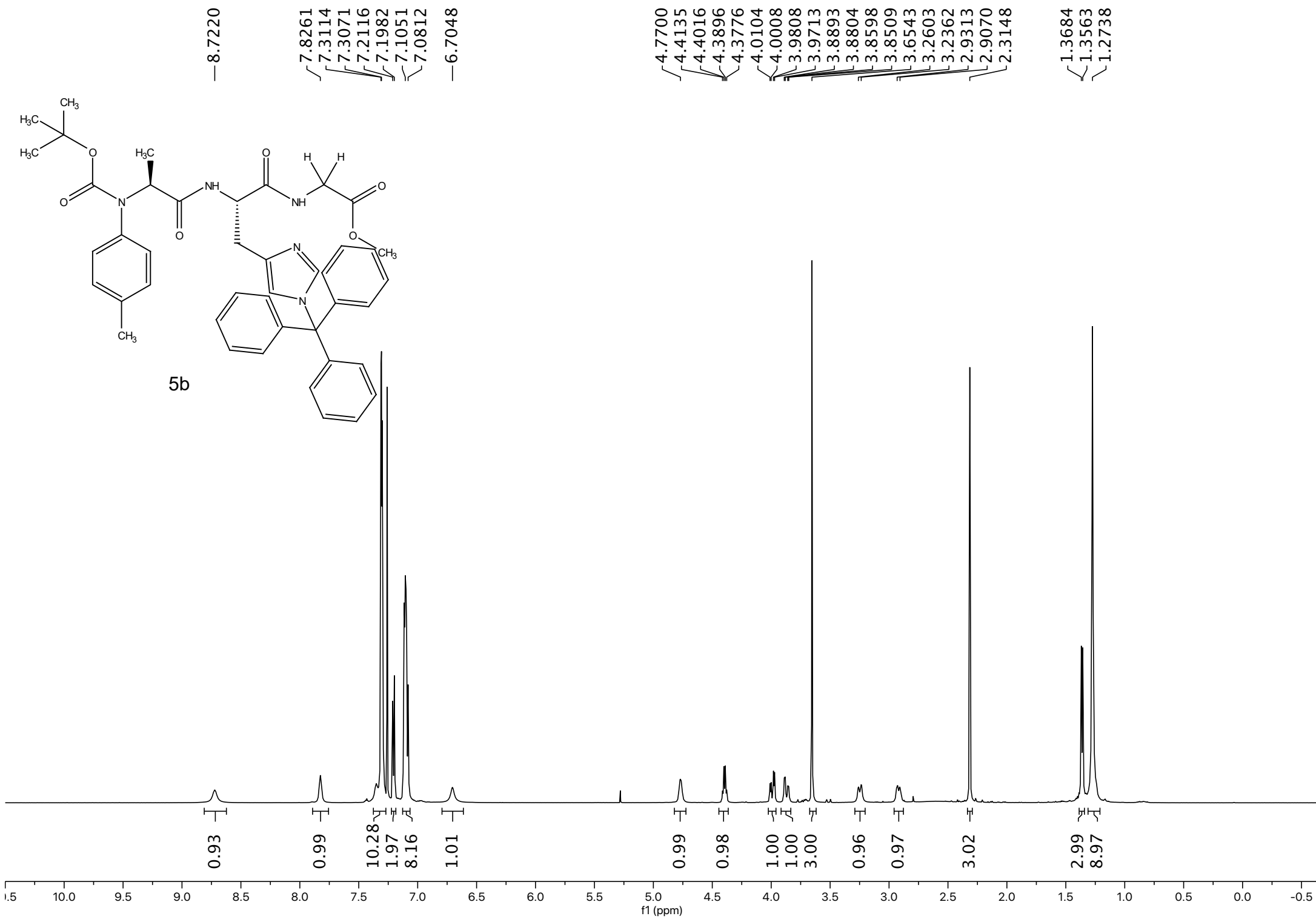
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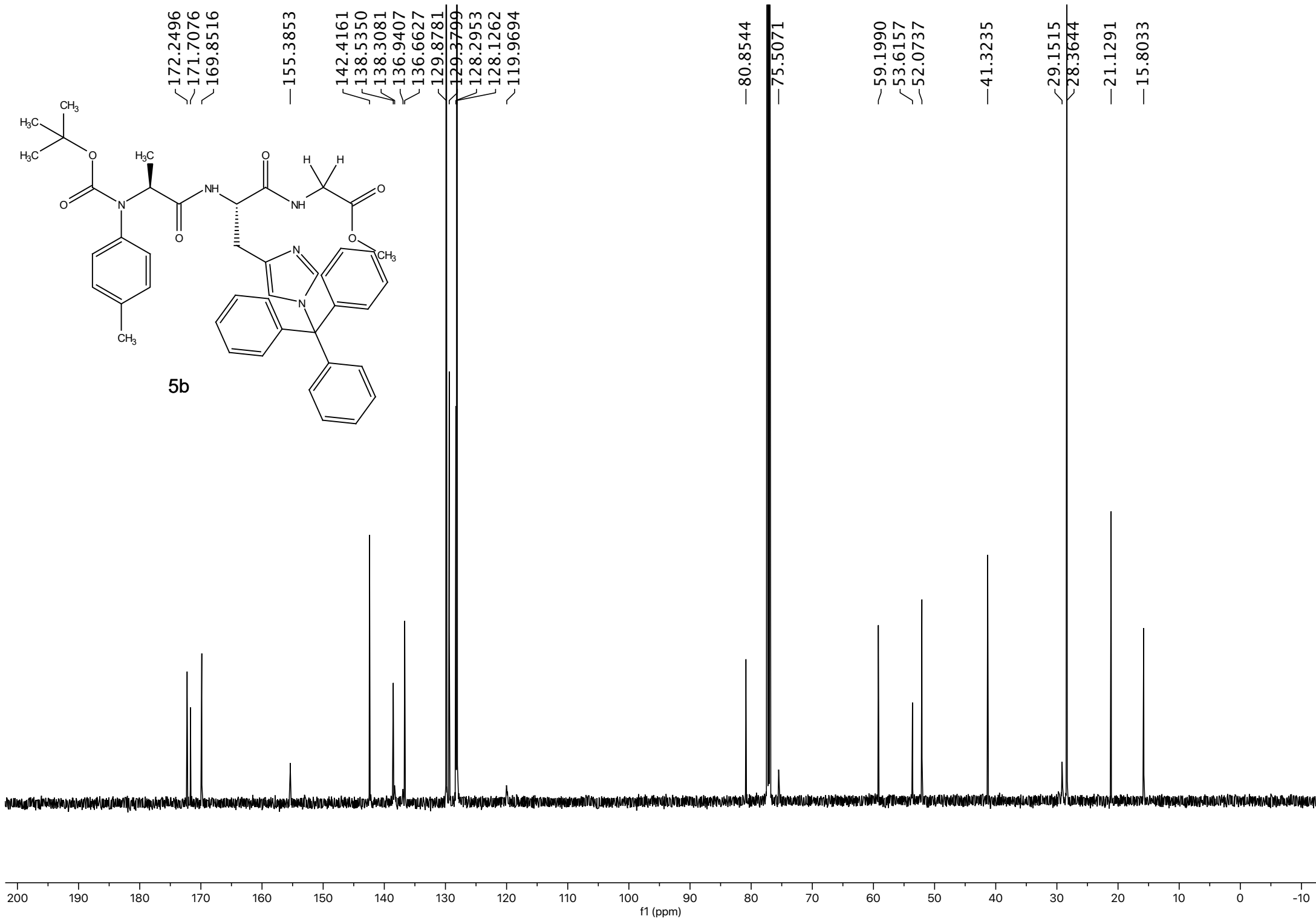
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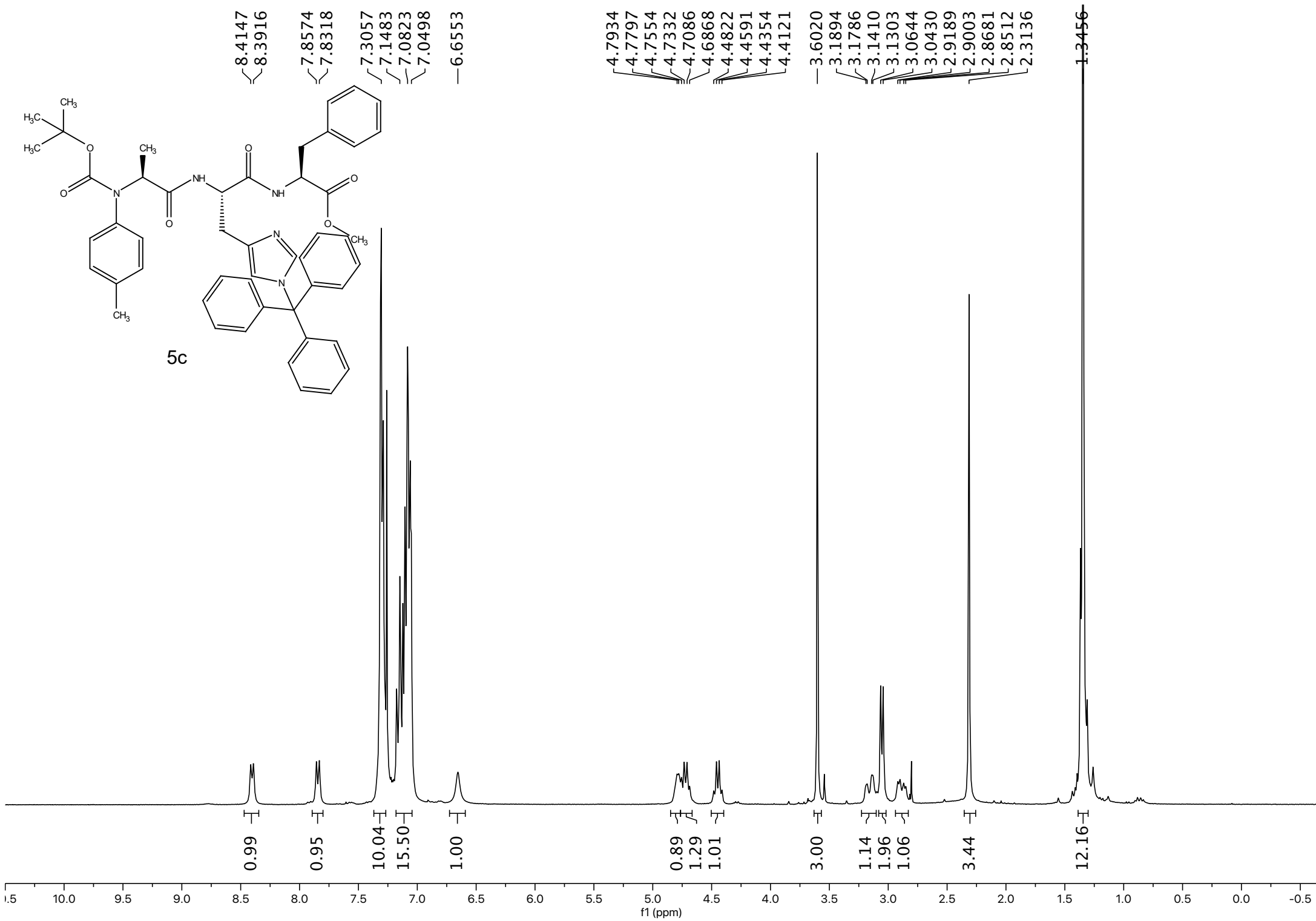


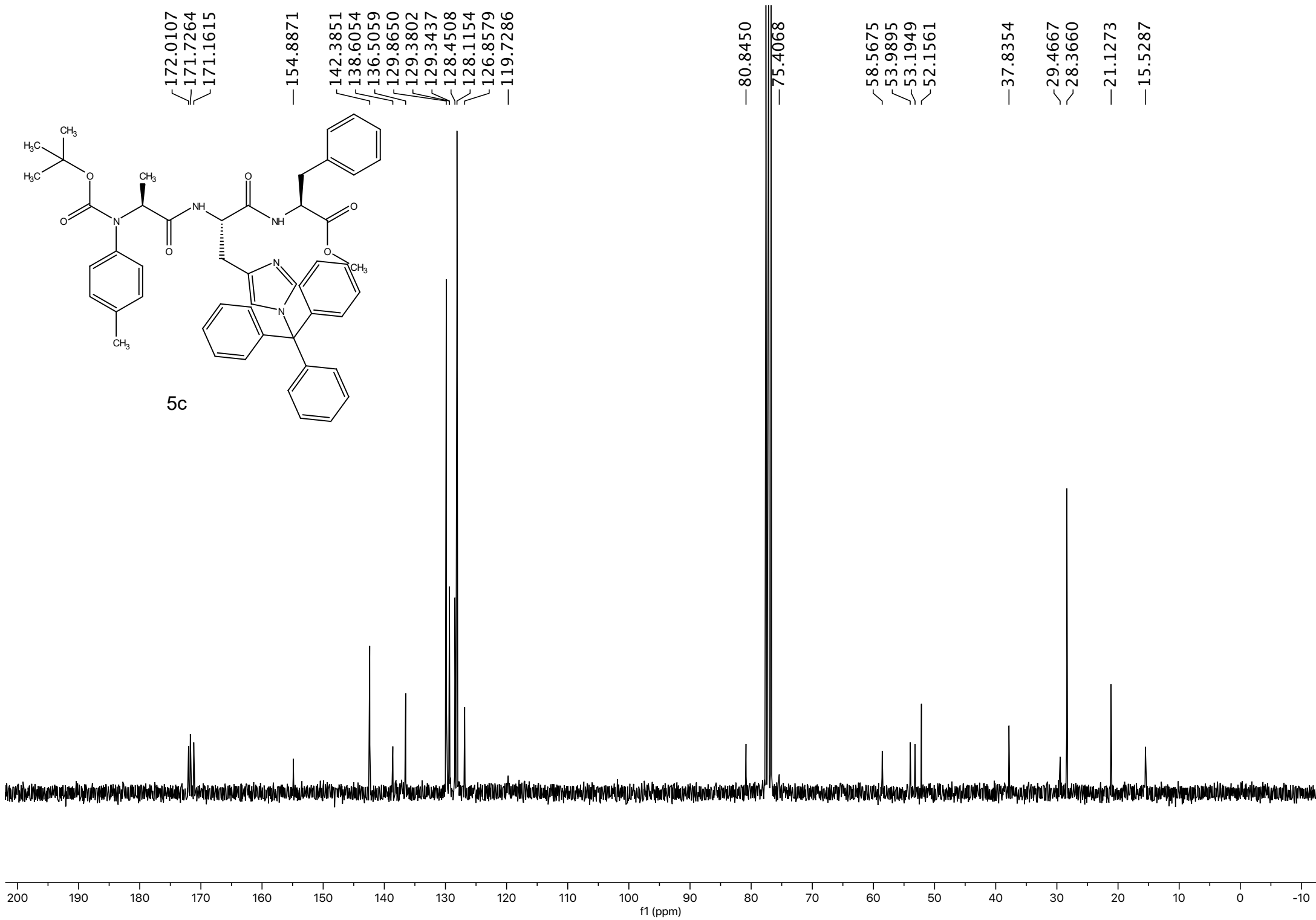


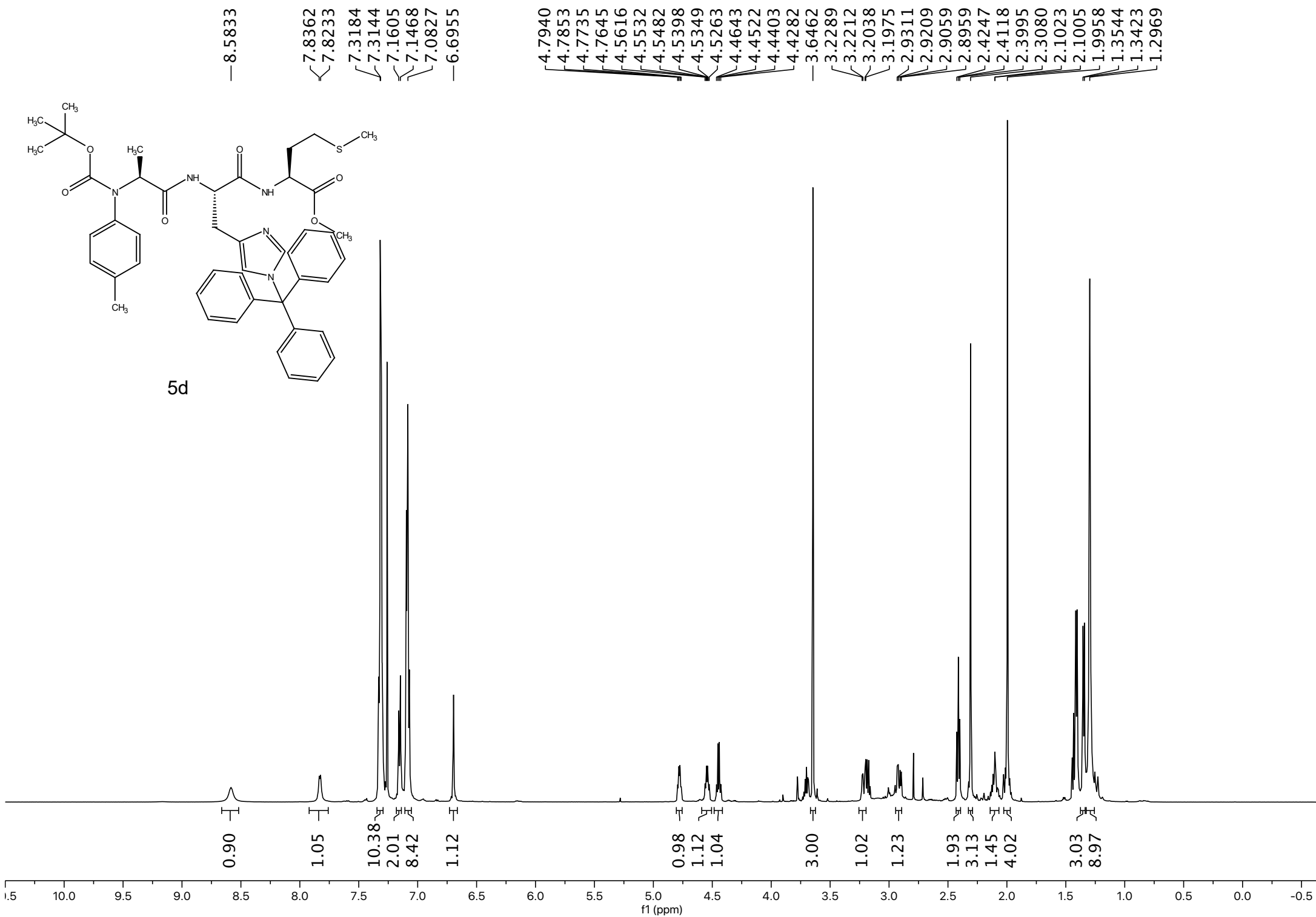


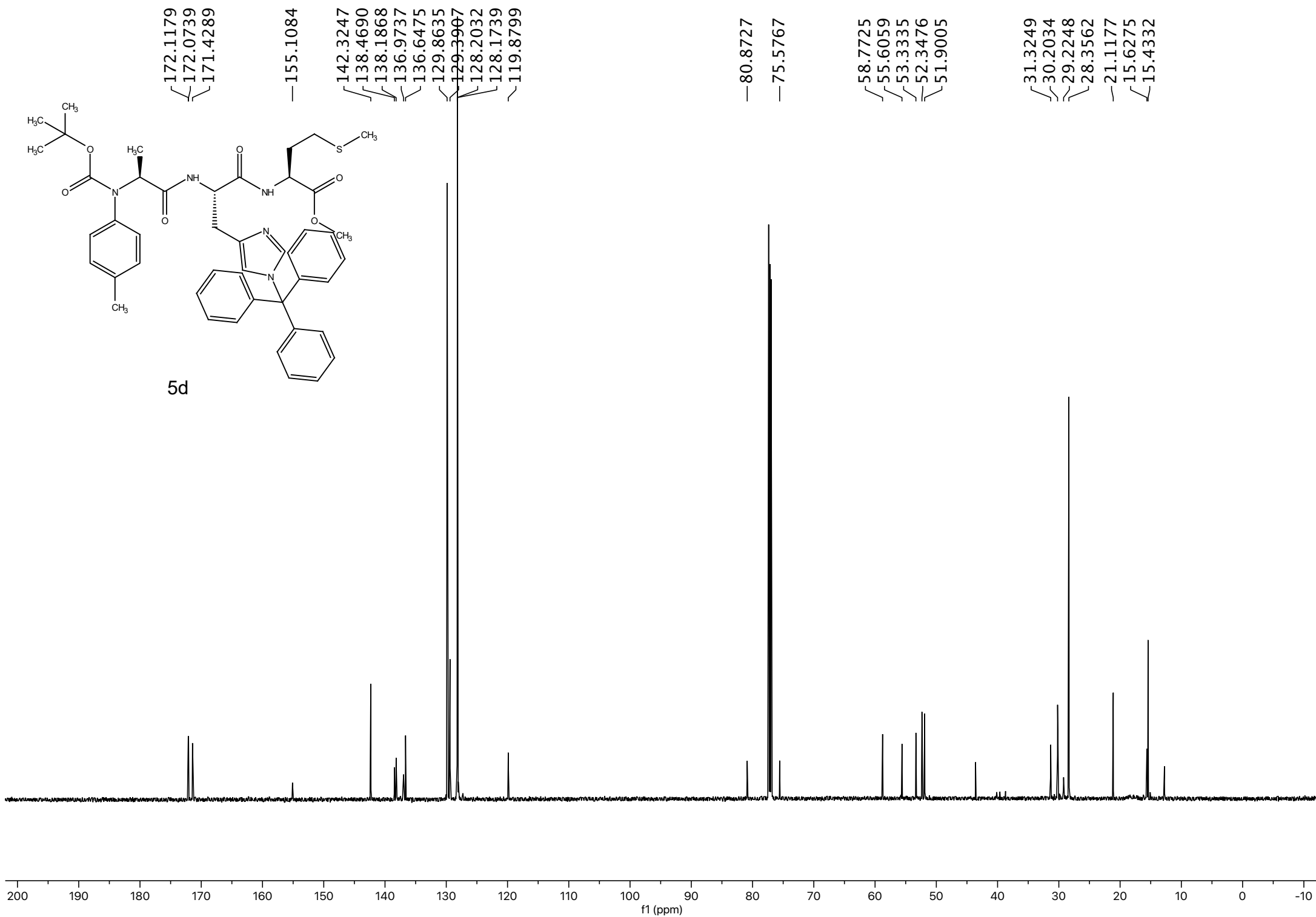


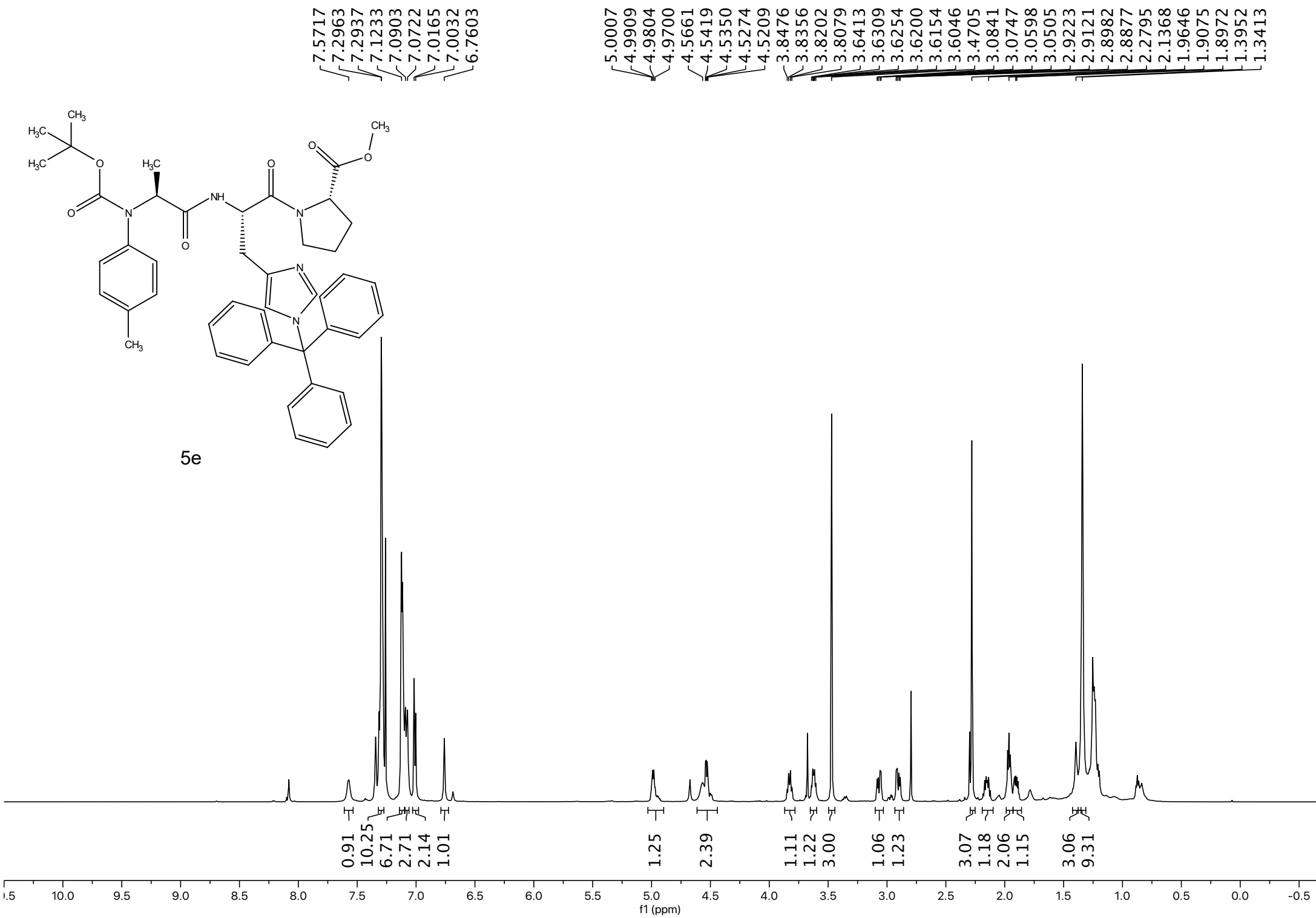


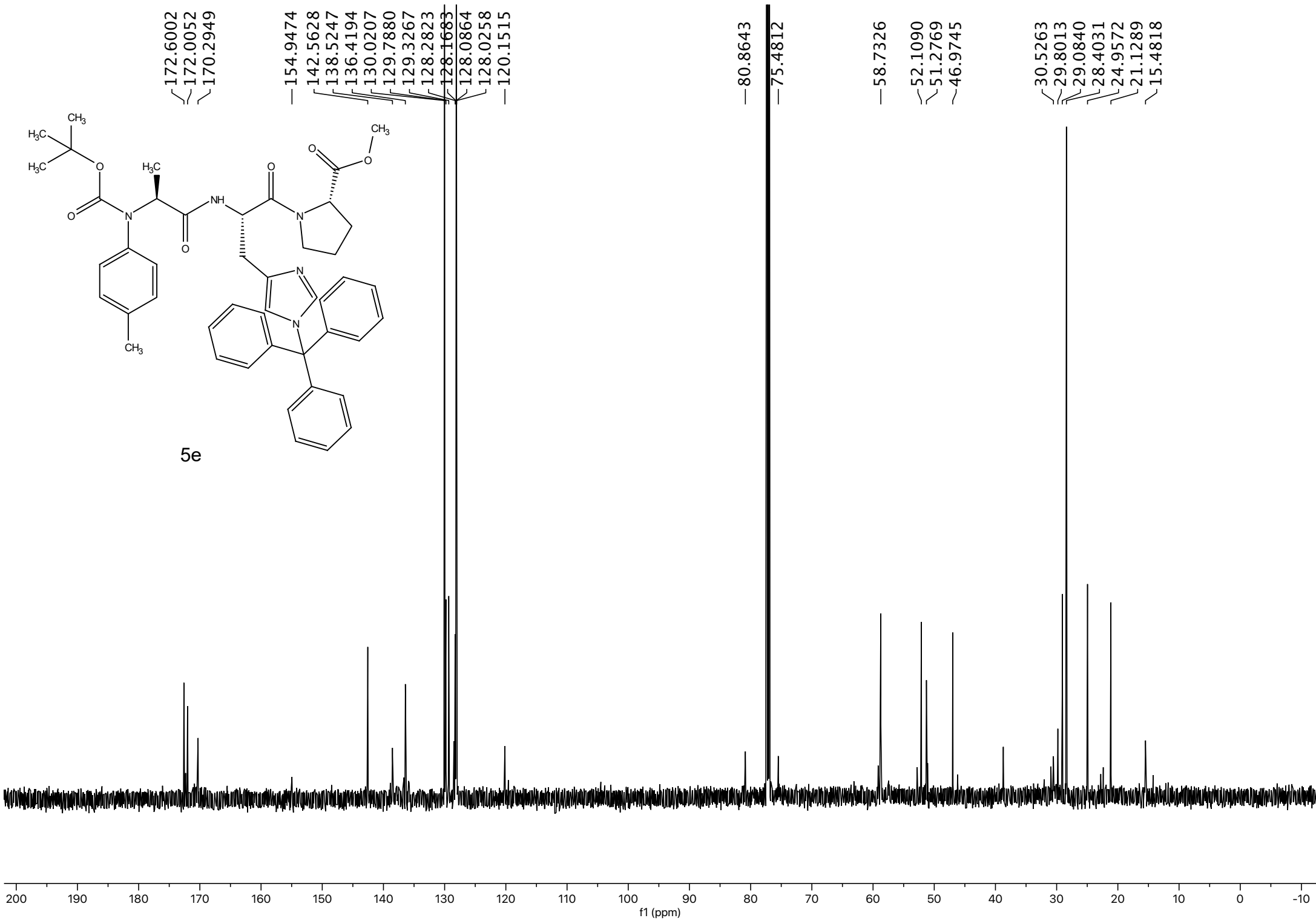


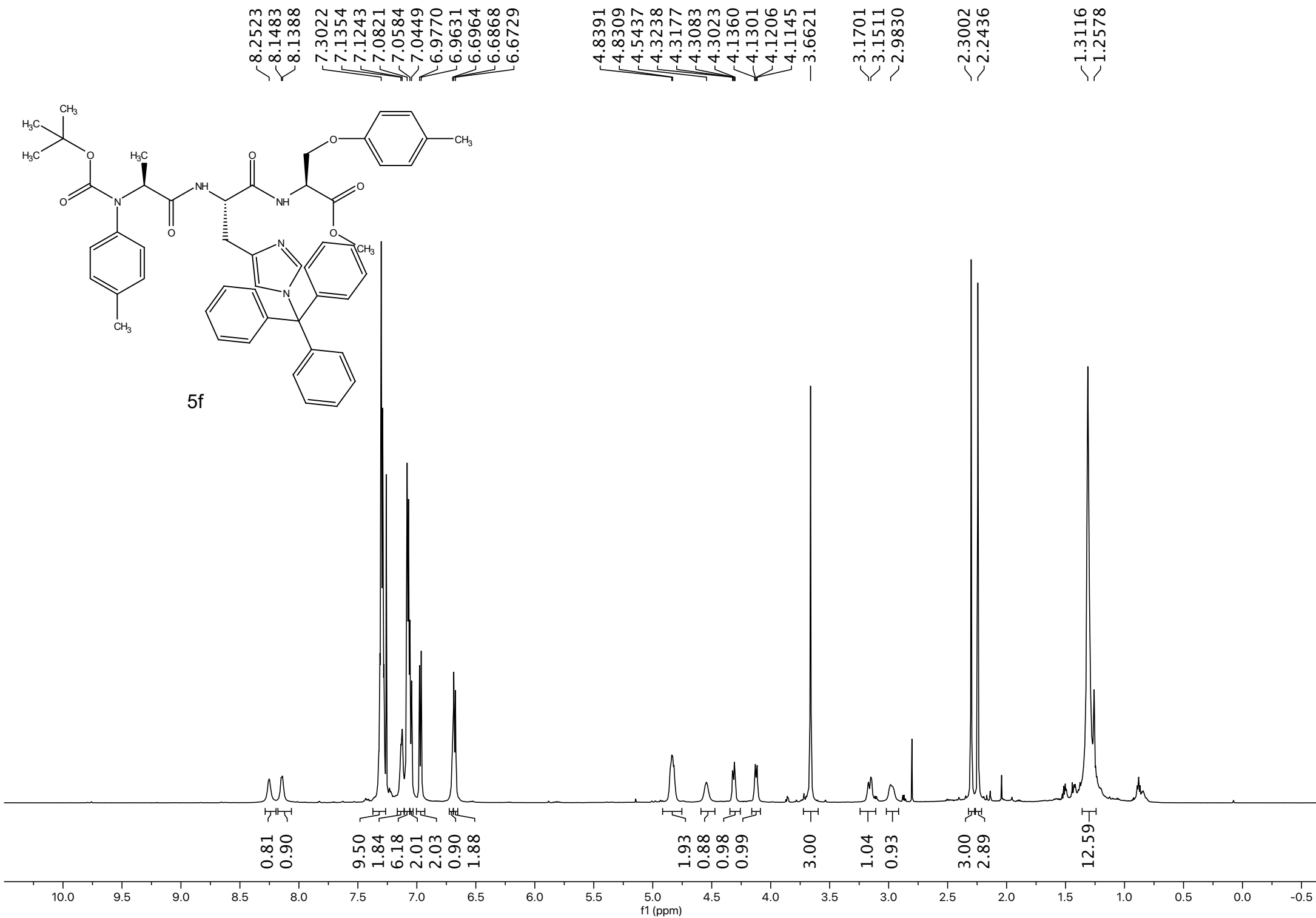


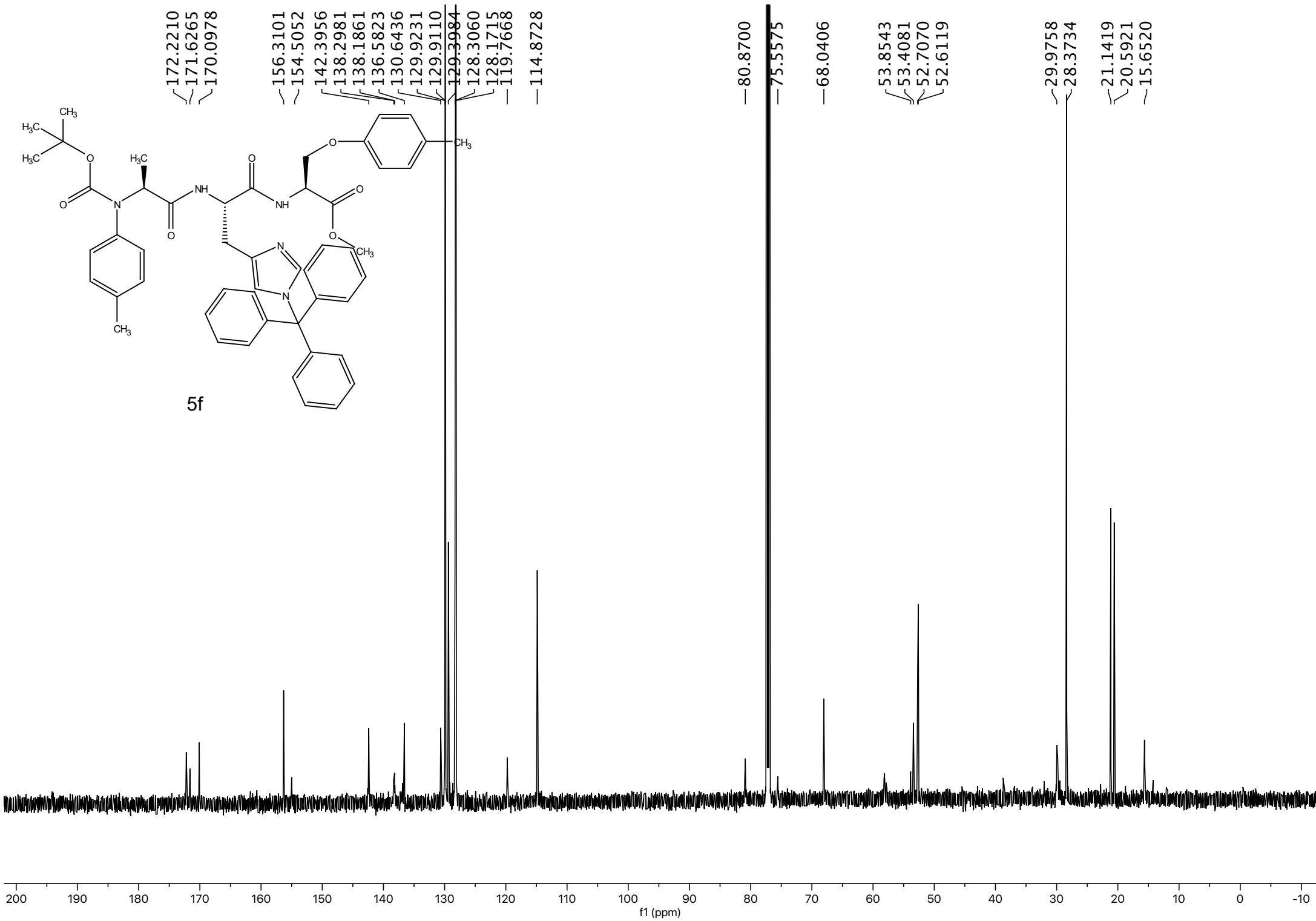




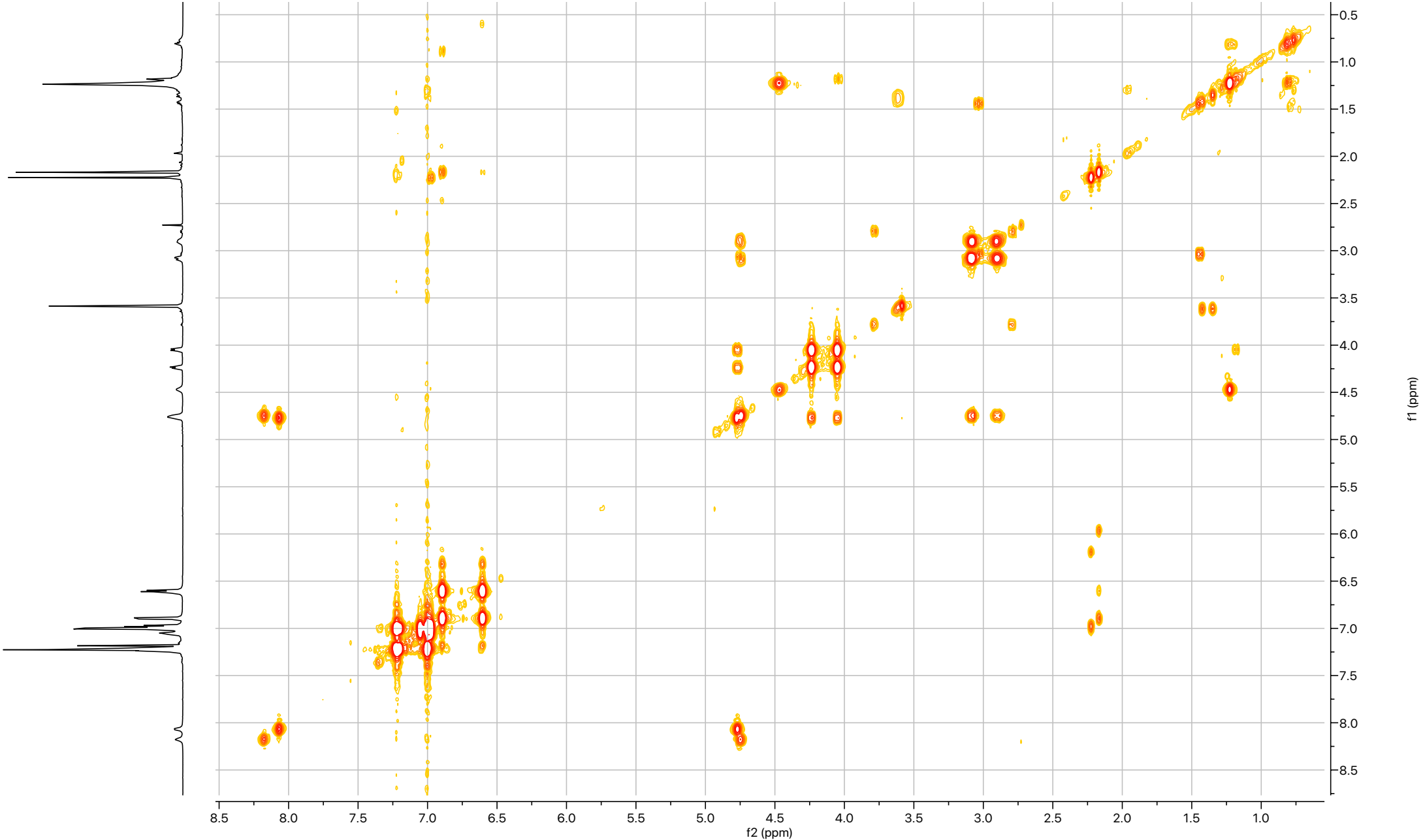


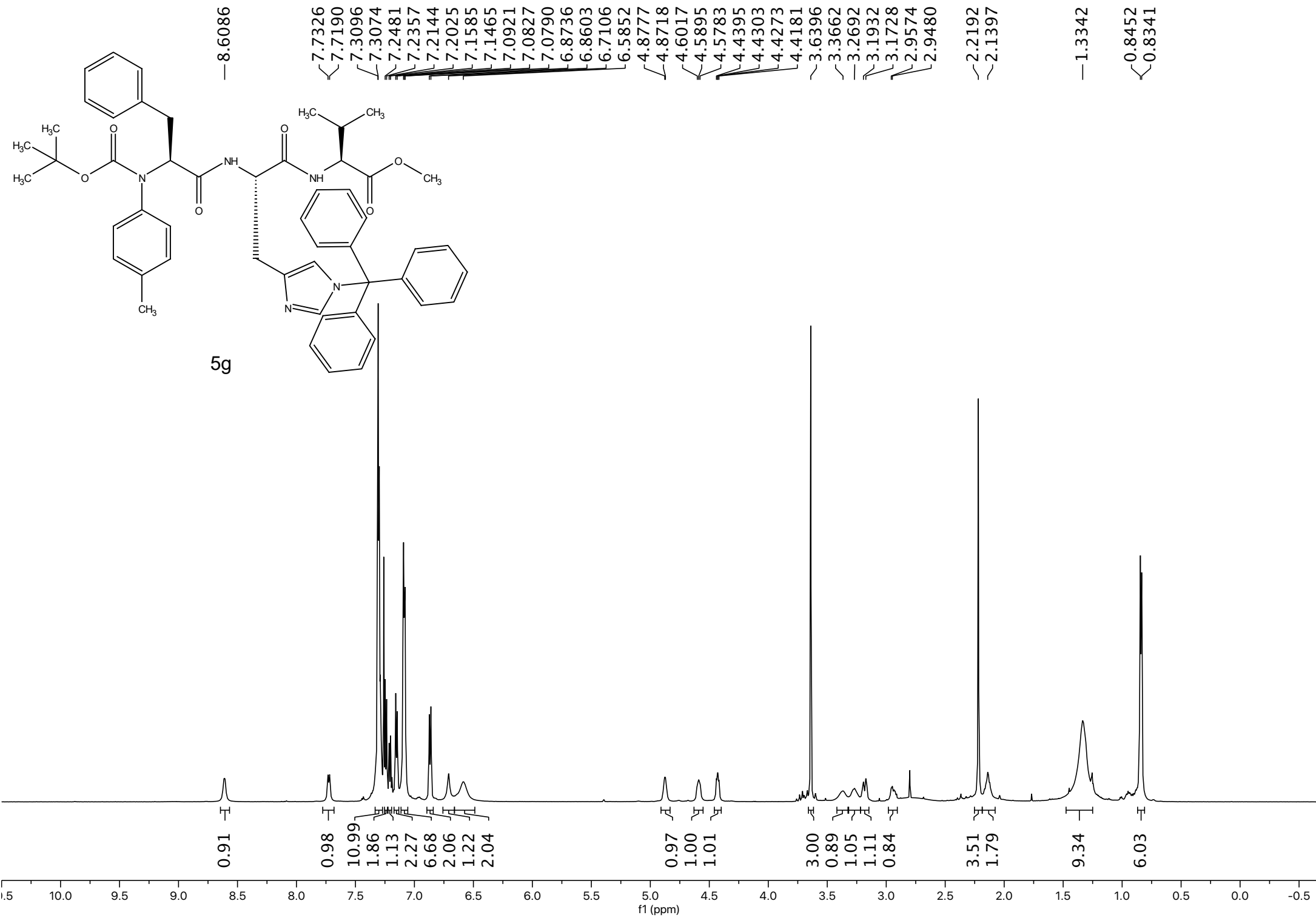


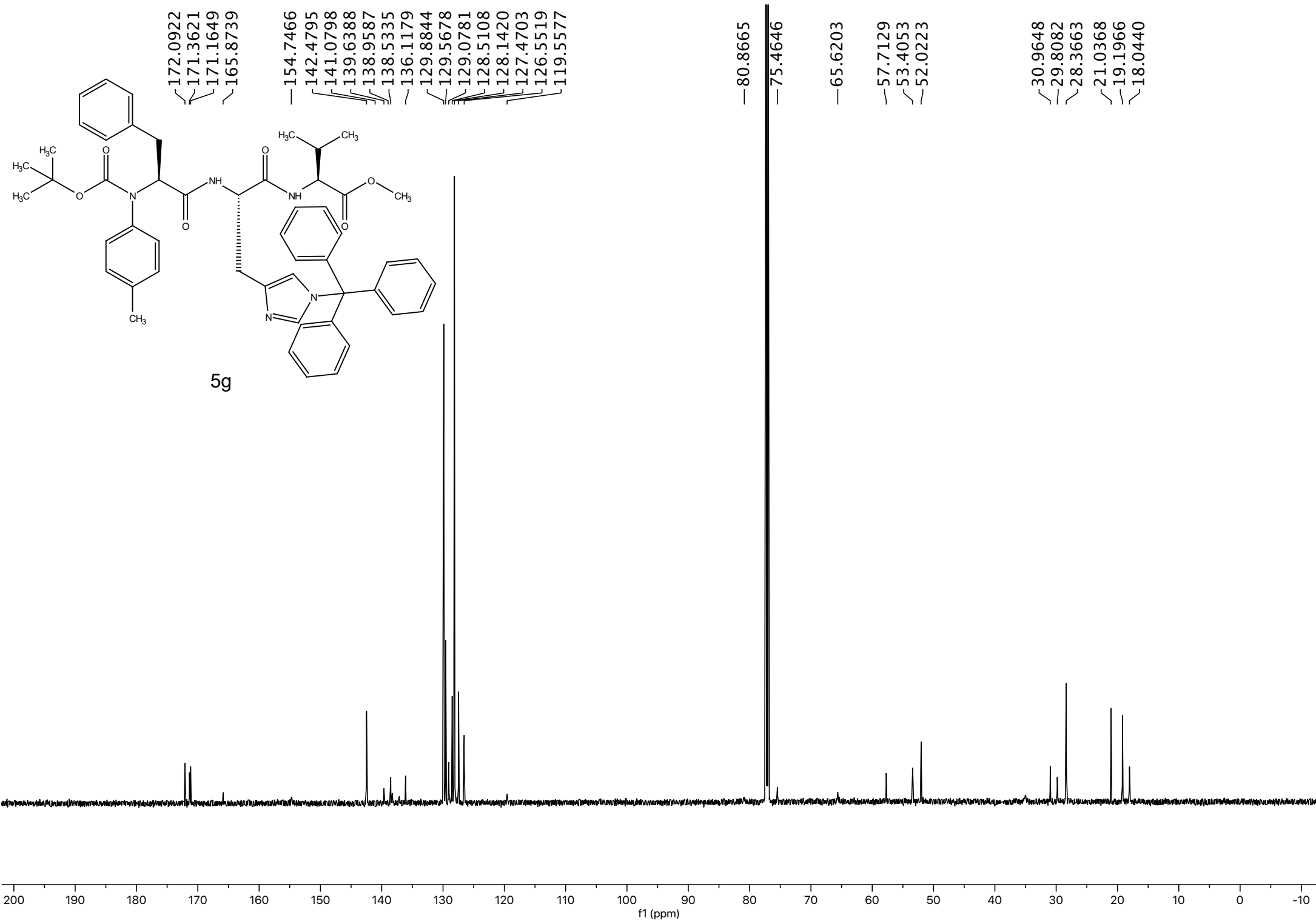


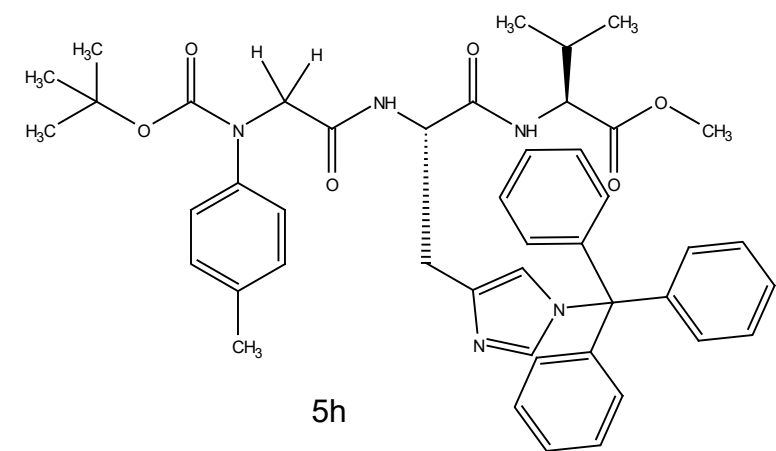


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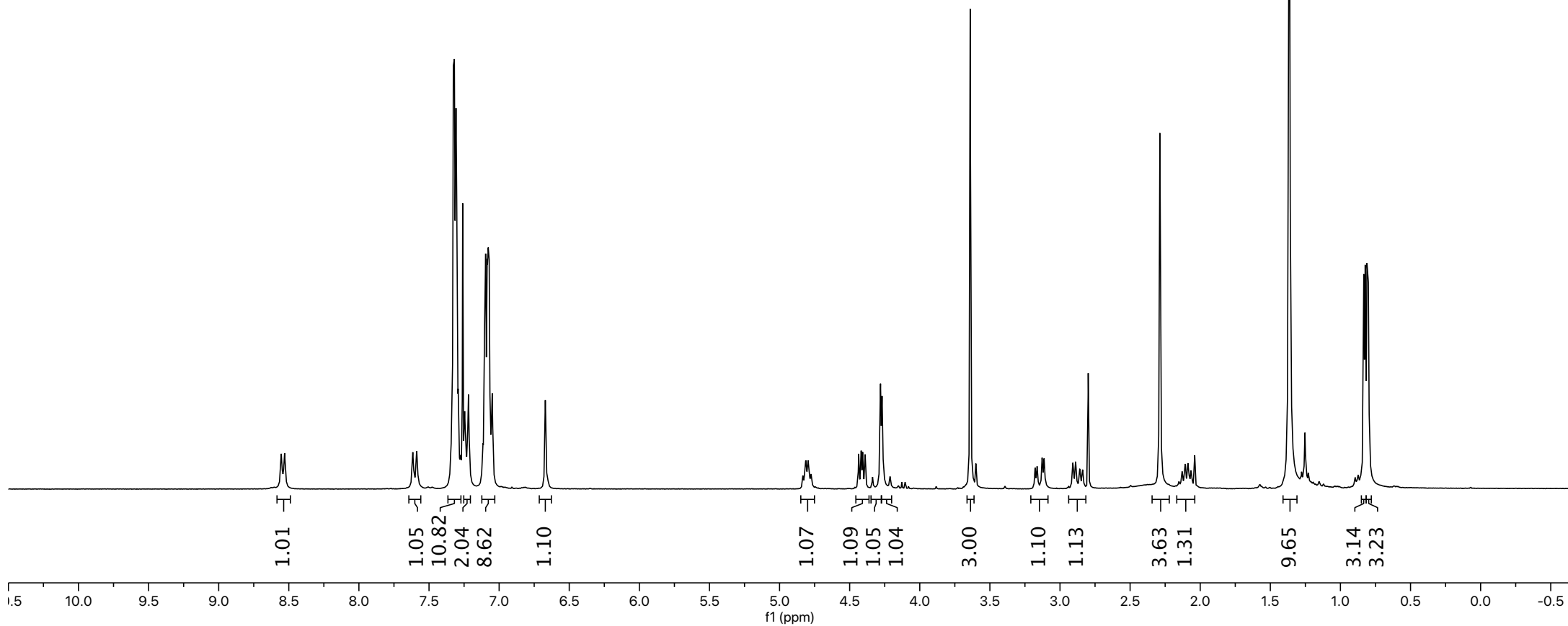


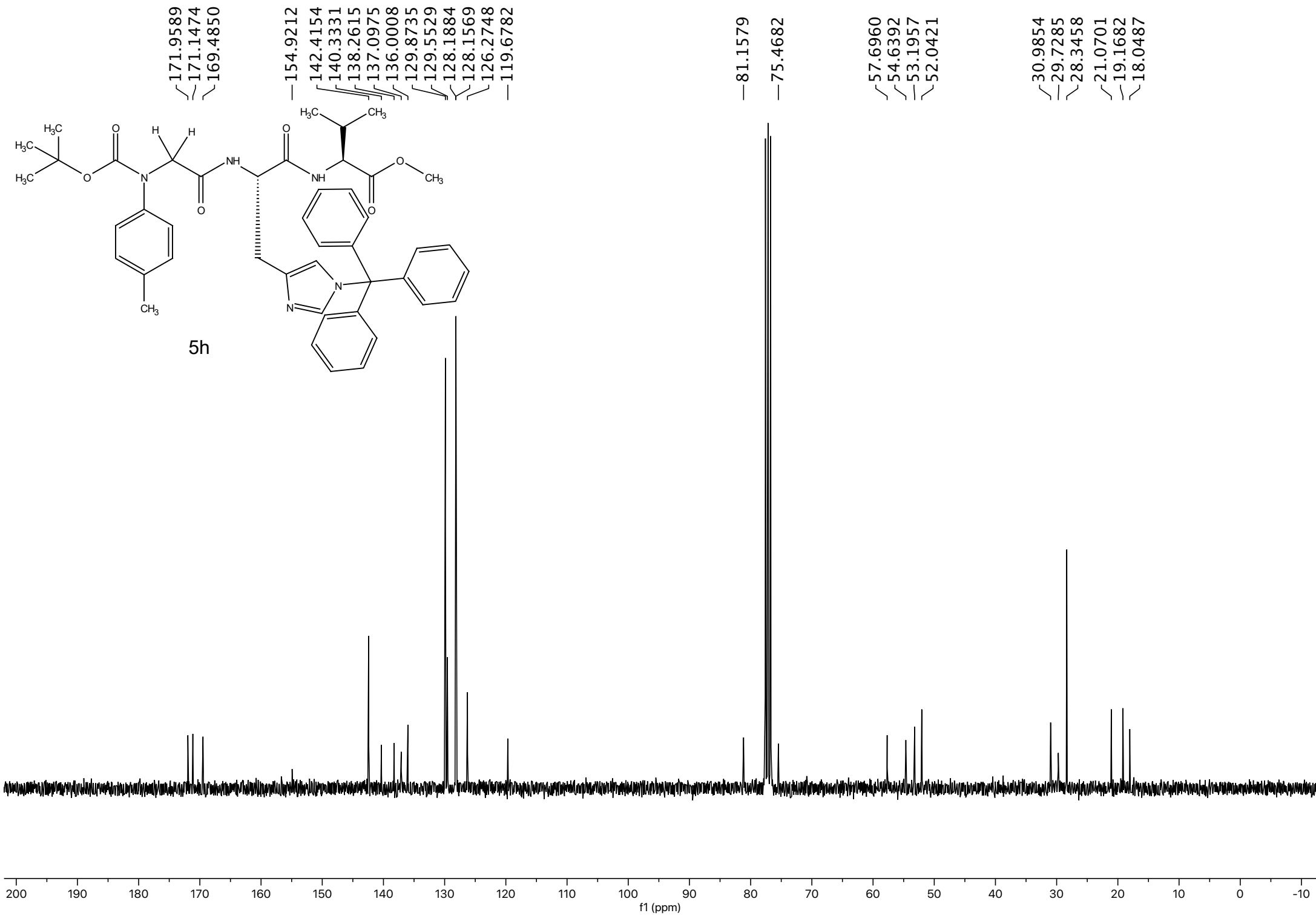


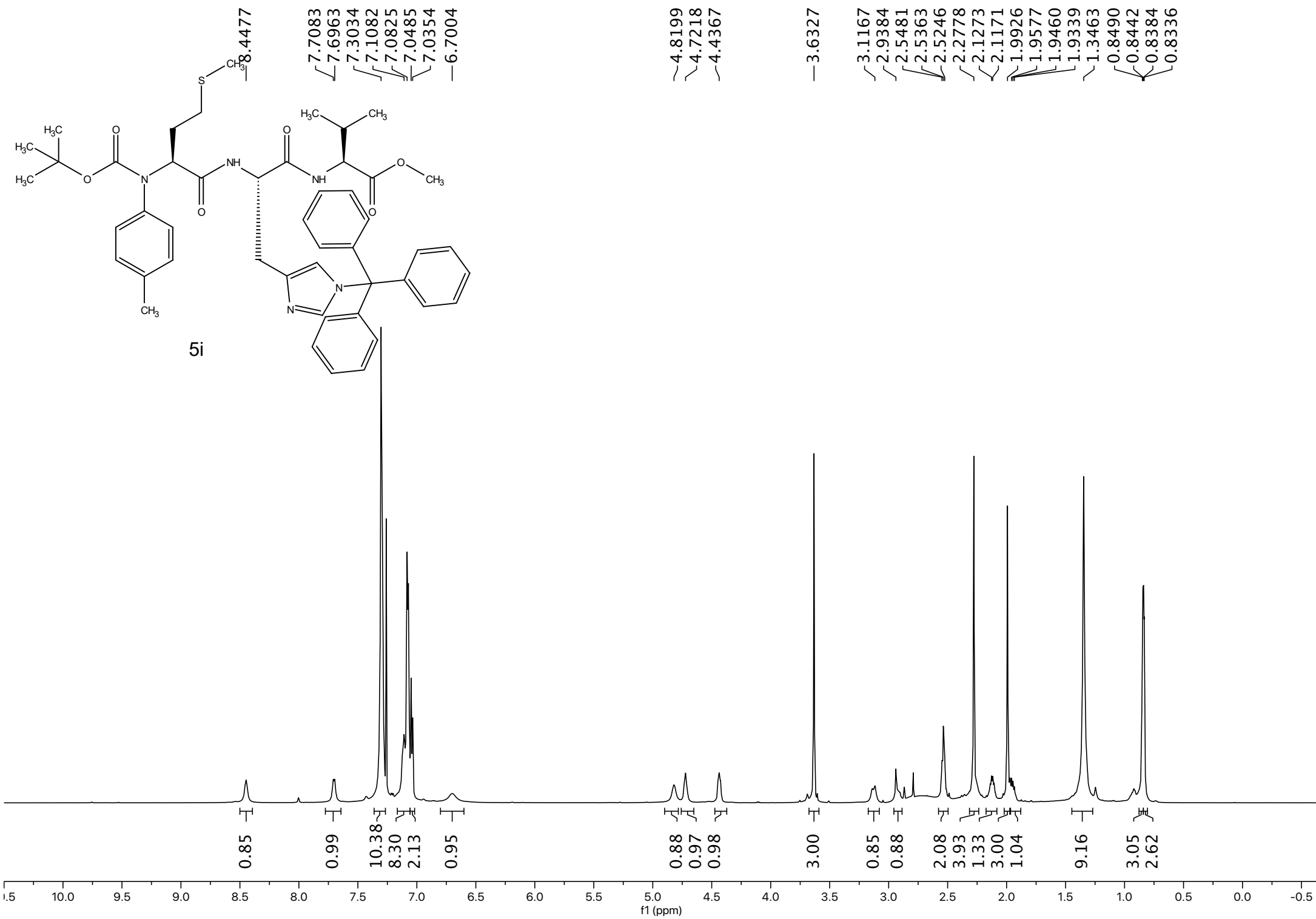


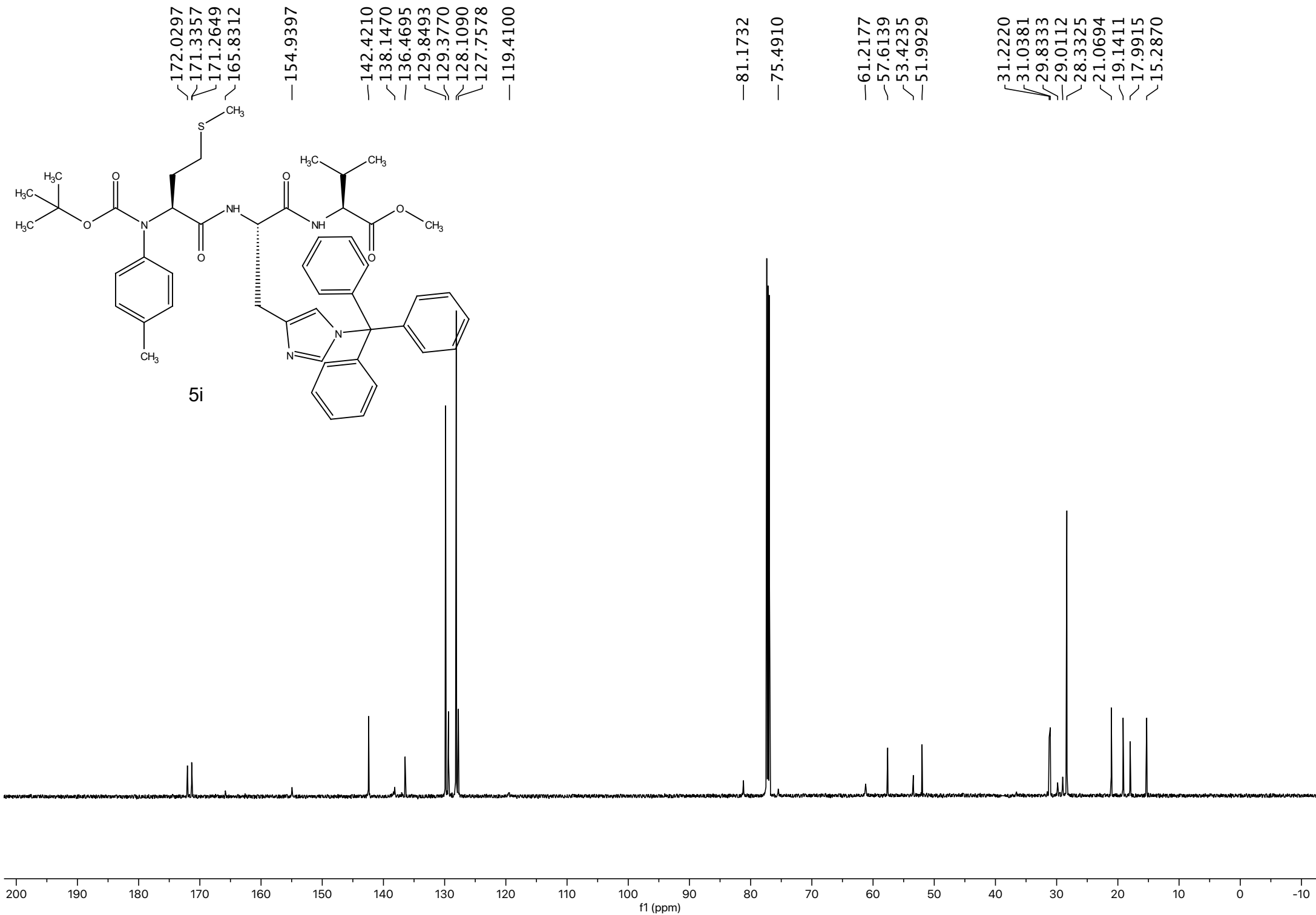
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— 6.6702

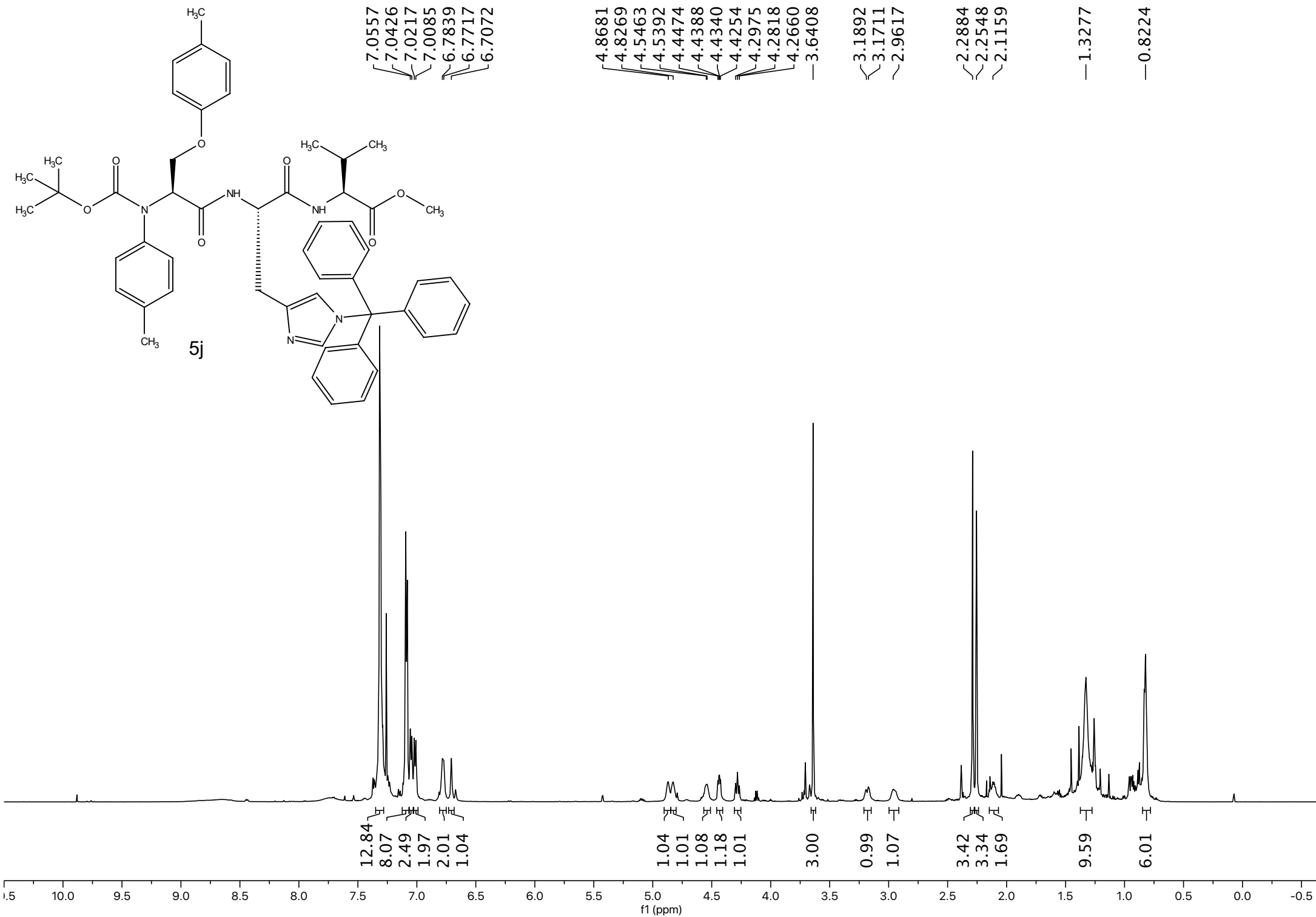
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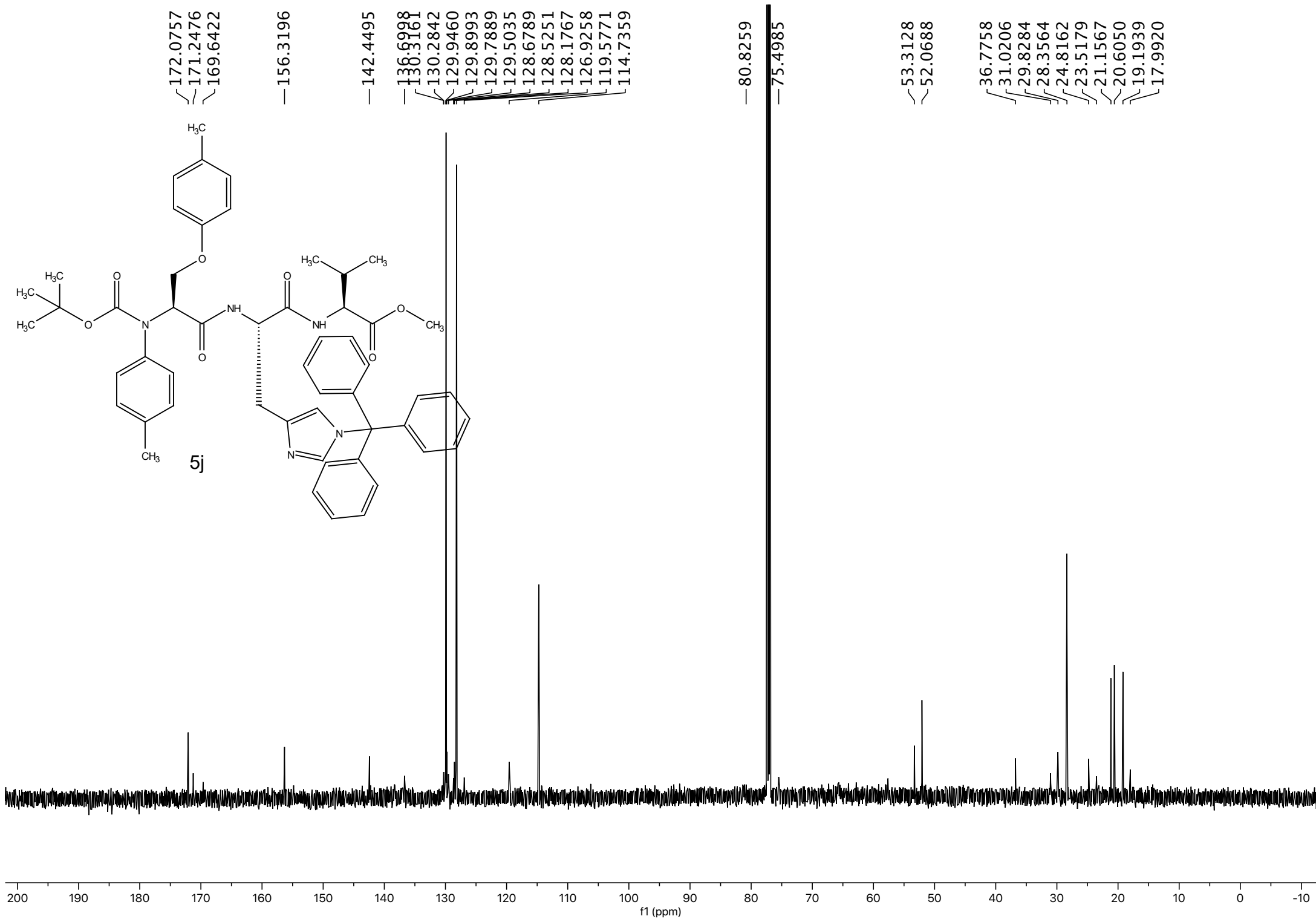


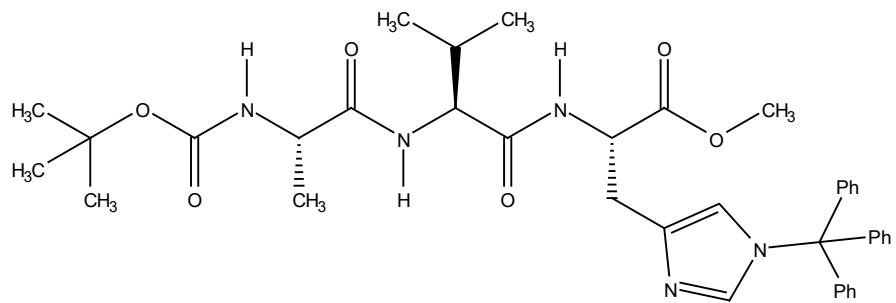










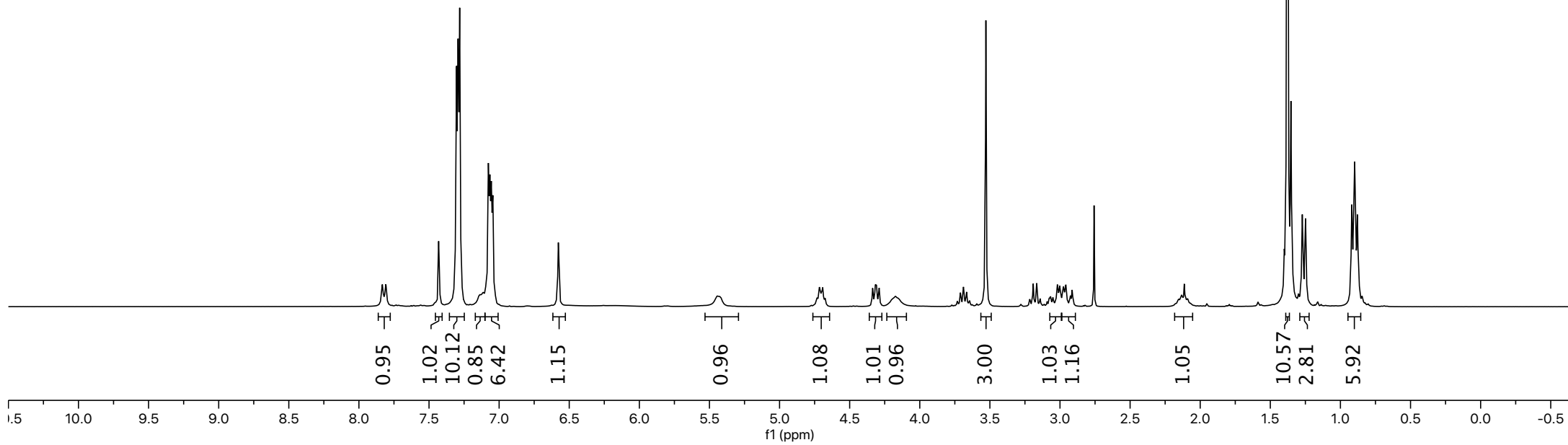


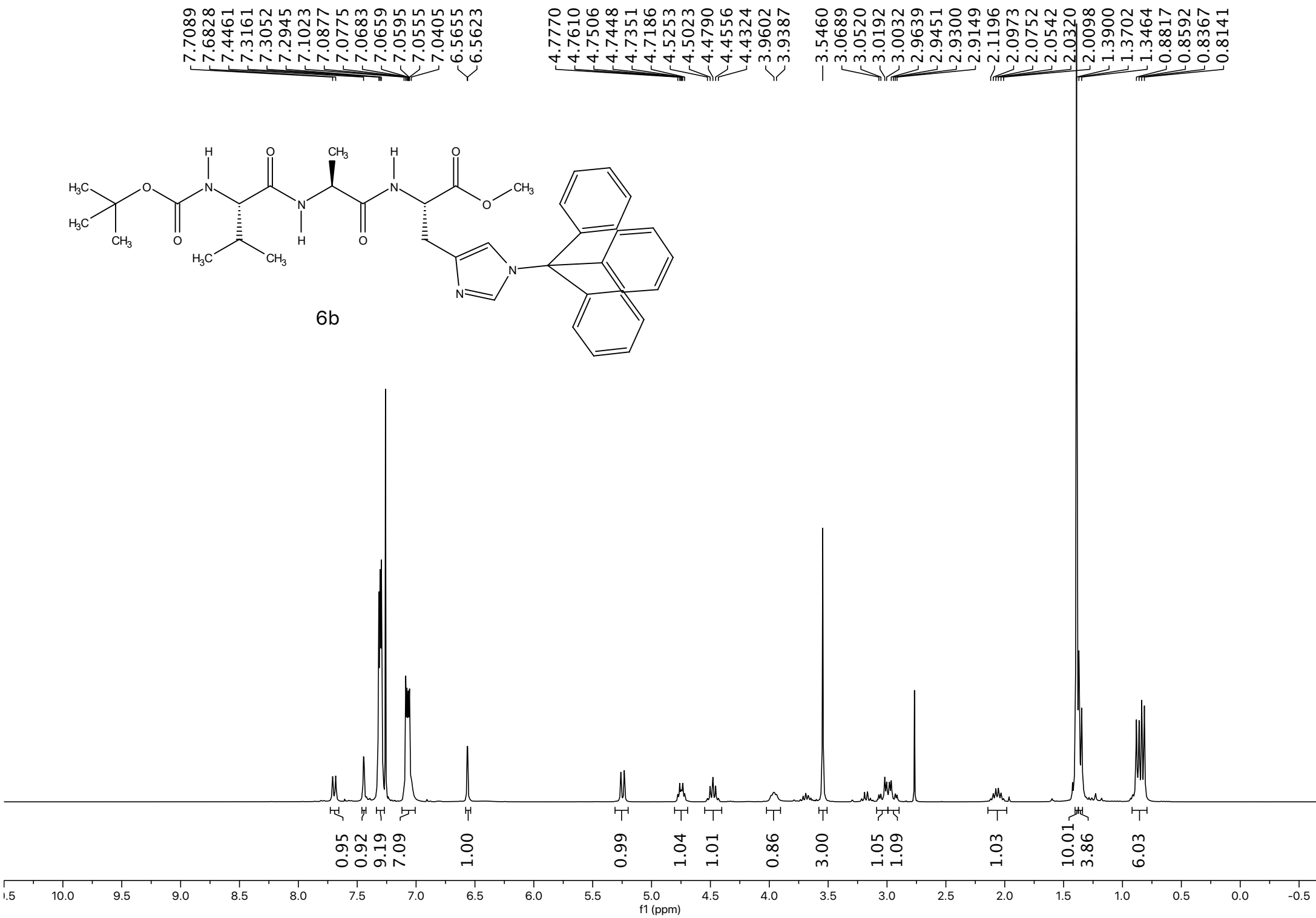
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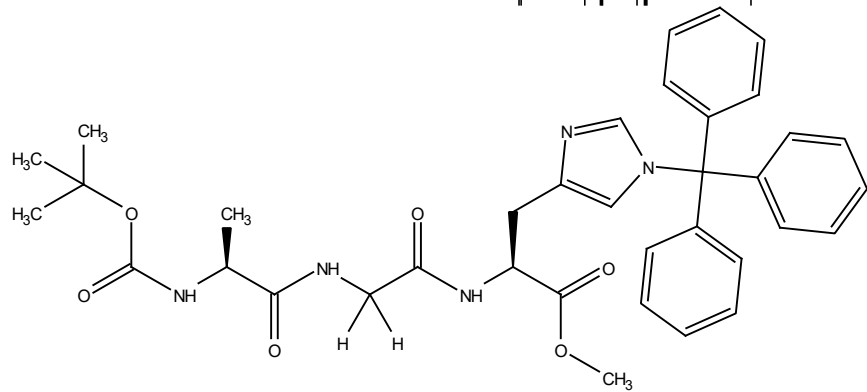
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7.2929
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7.0558
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— 6.5776

— 5.4433

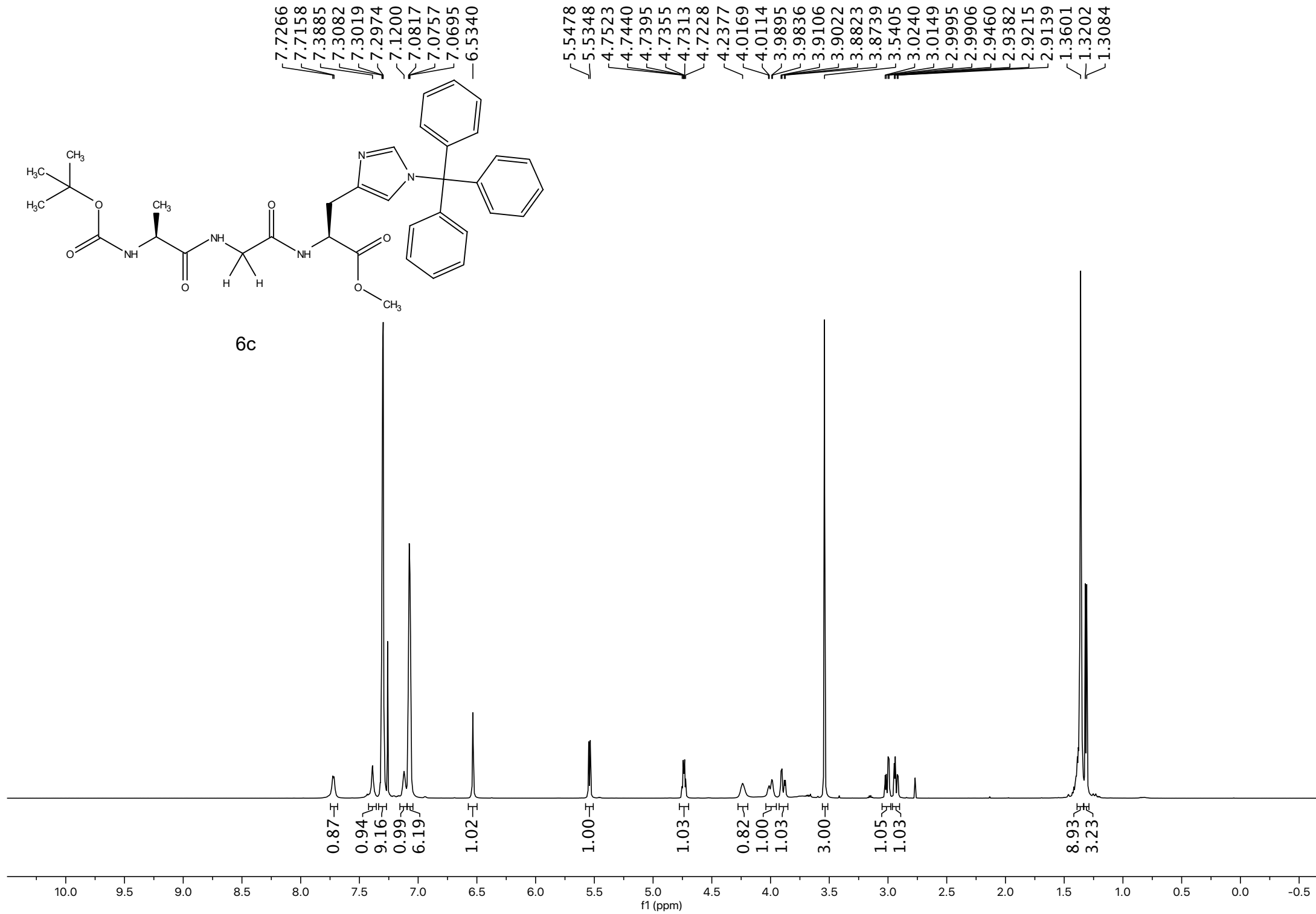
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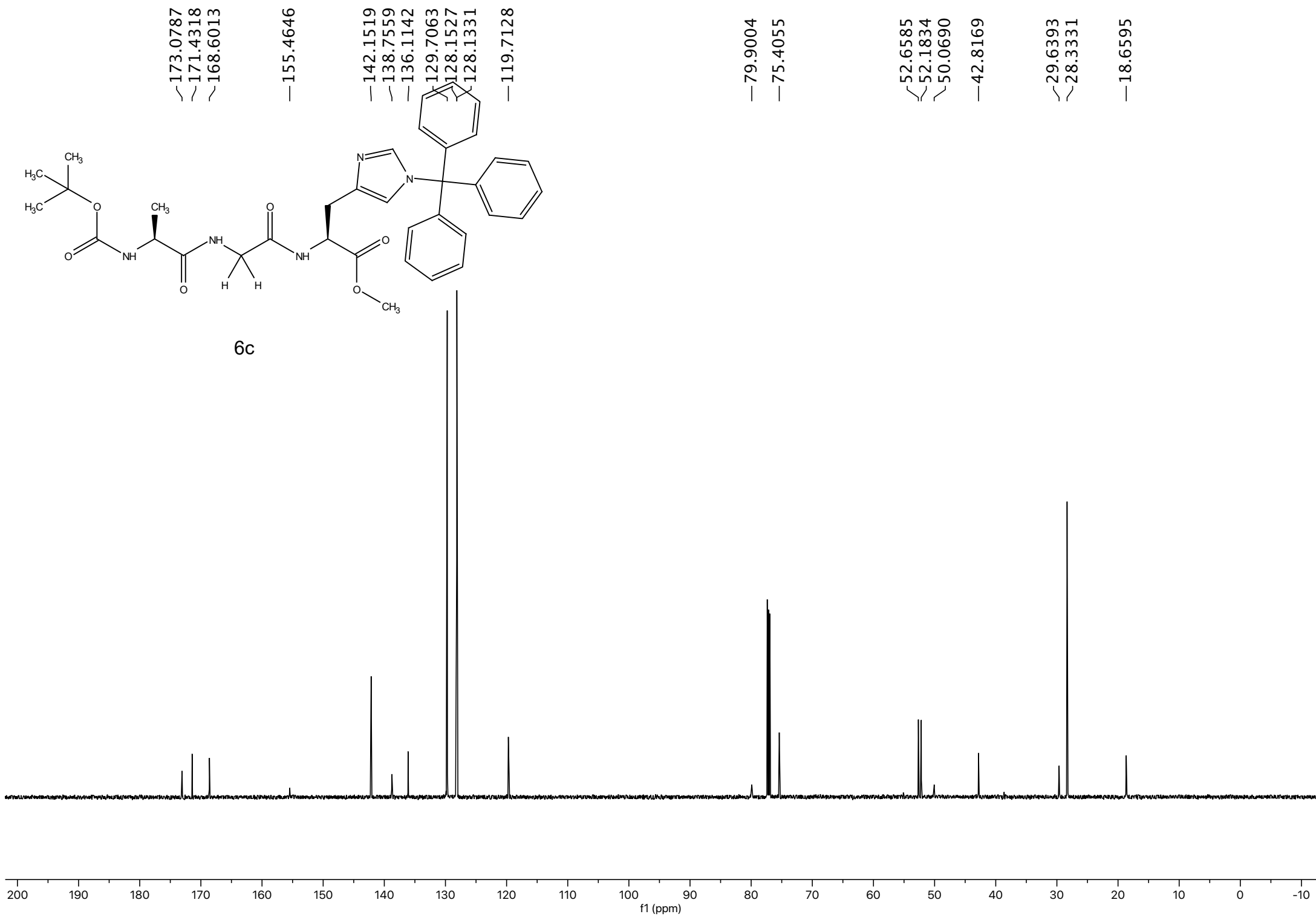


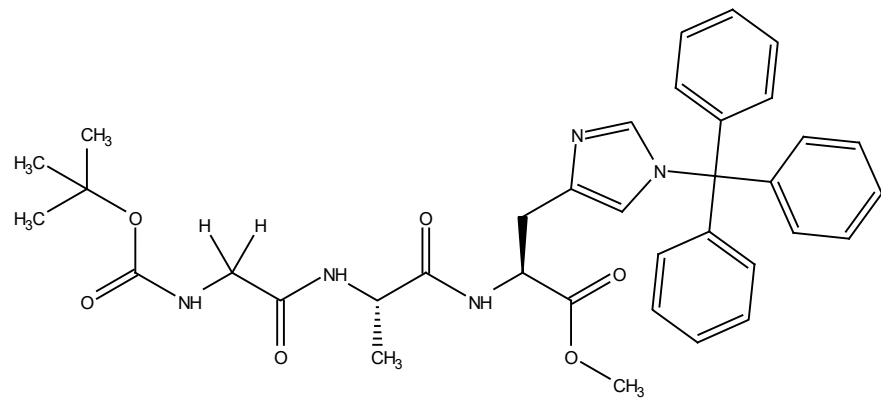




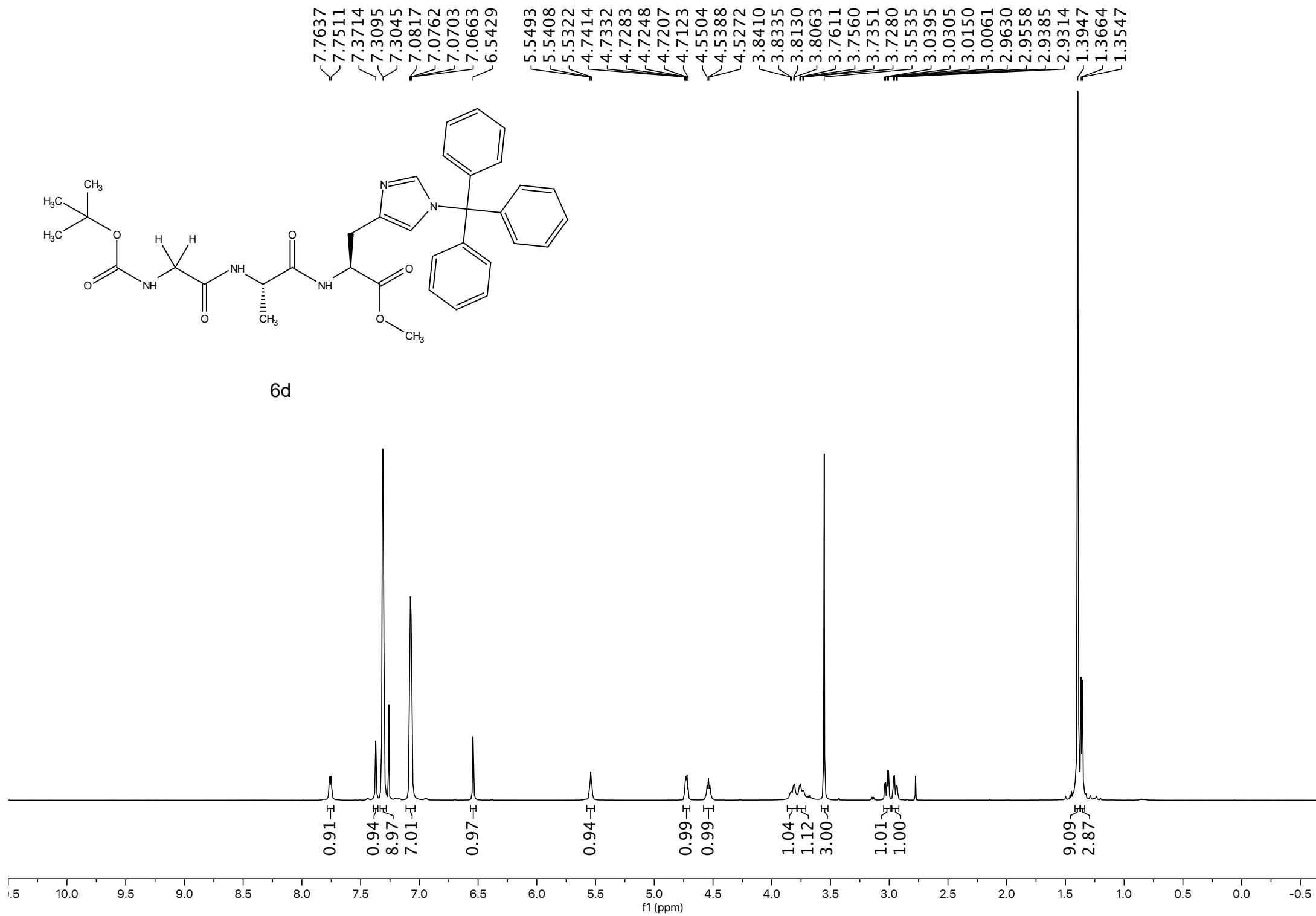
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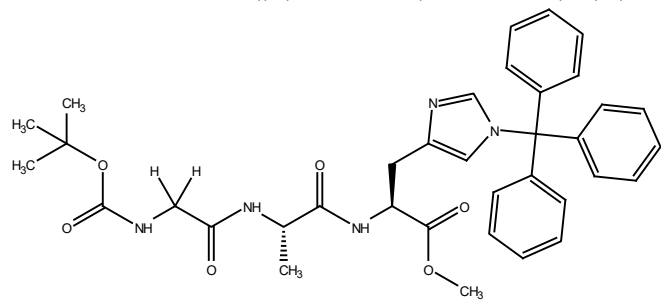




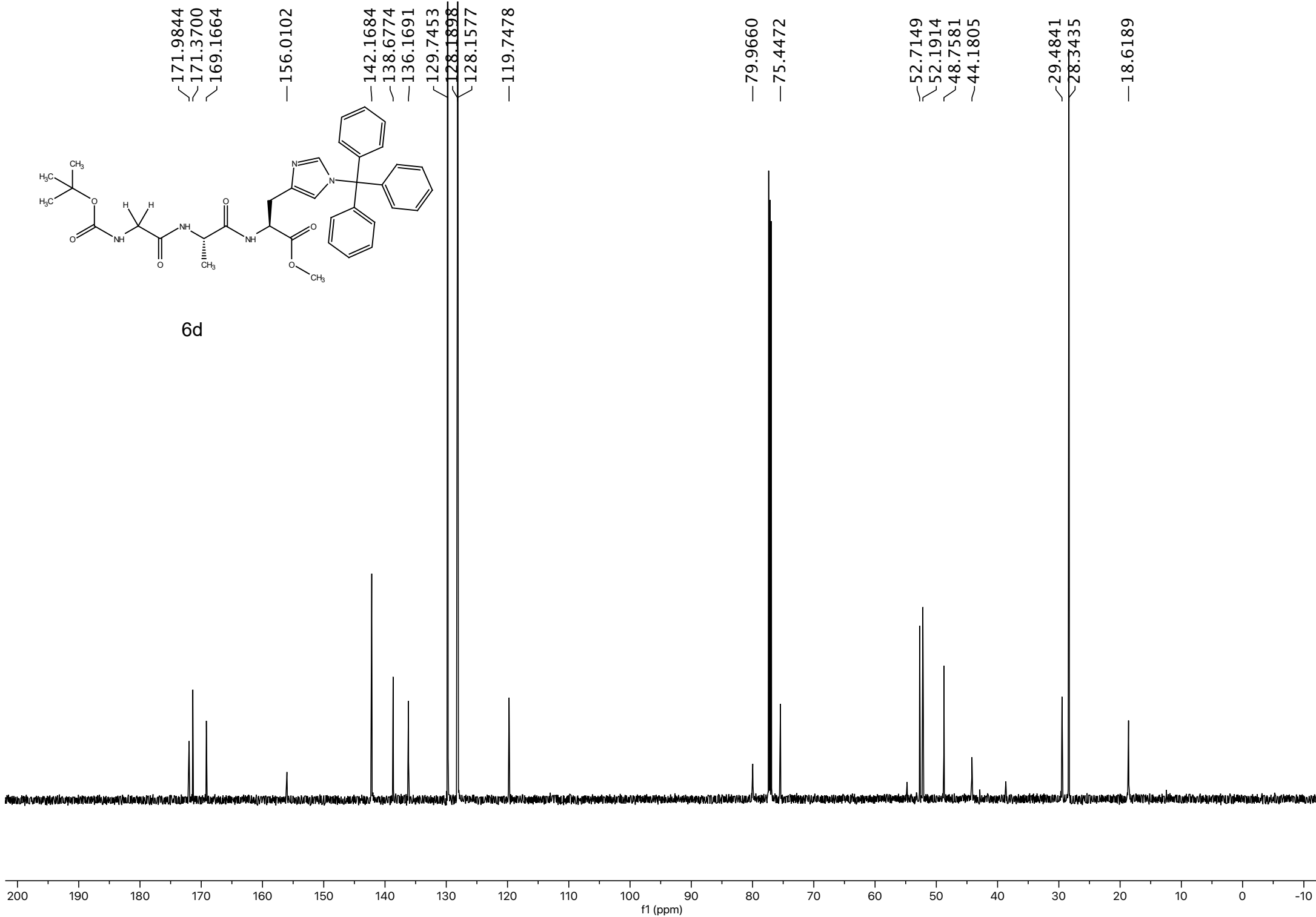


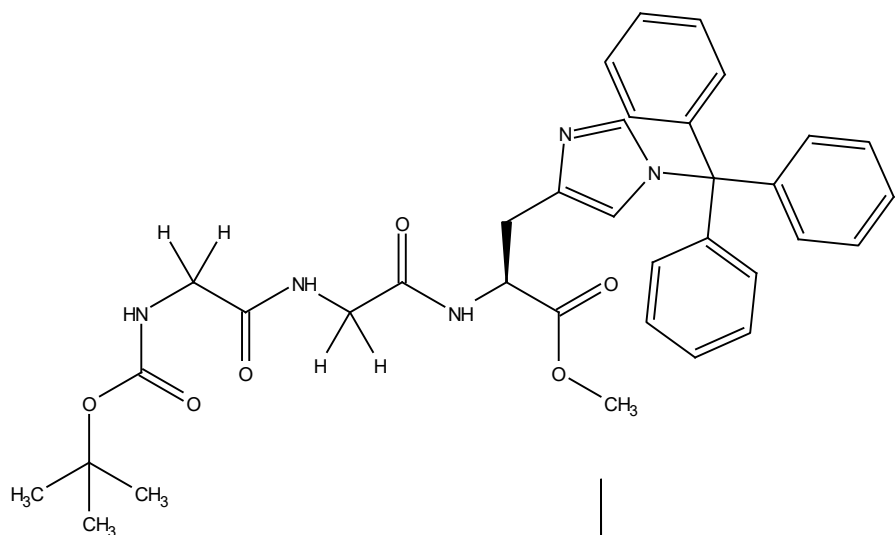
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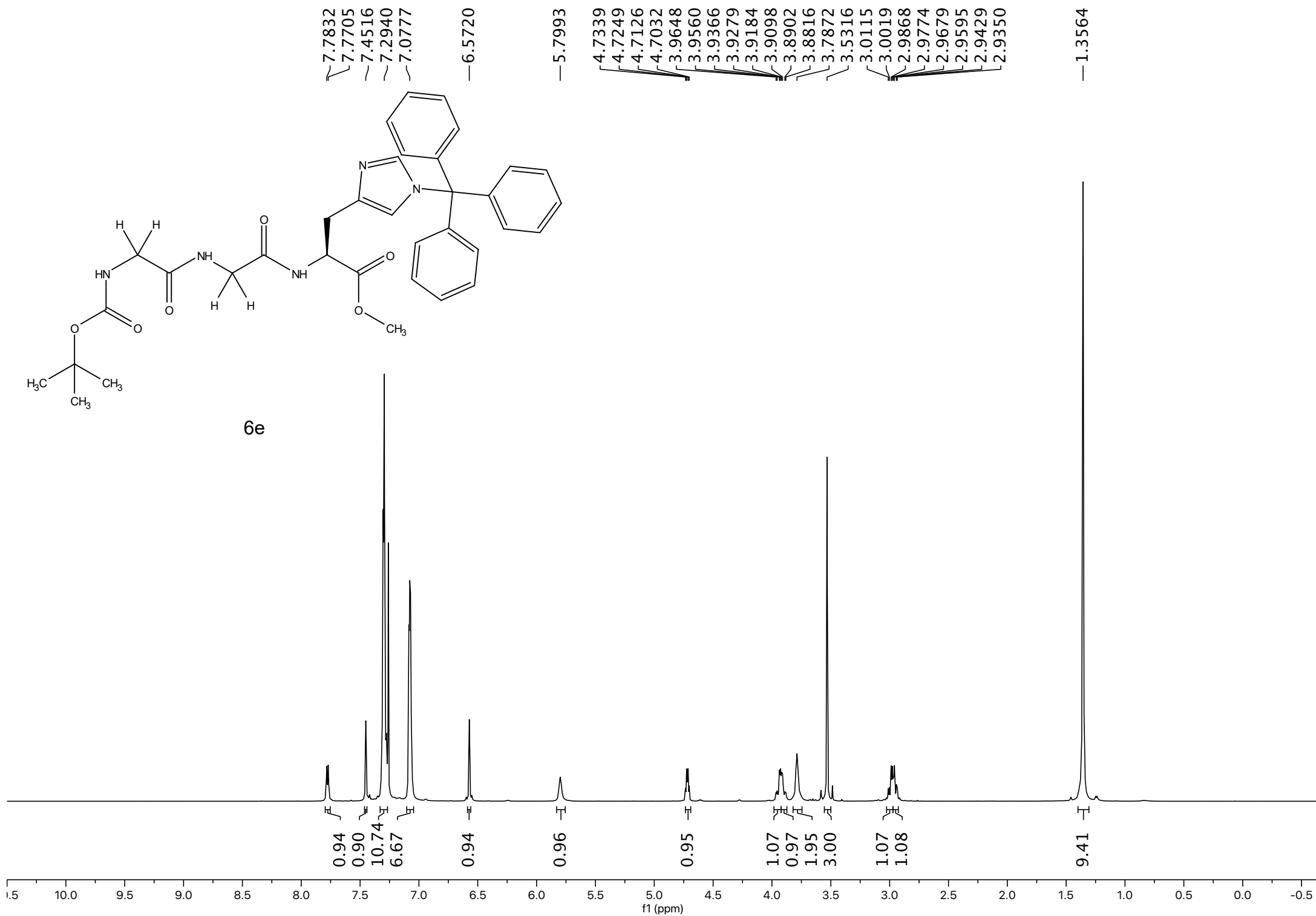


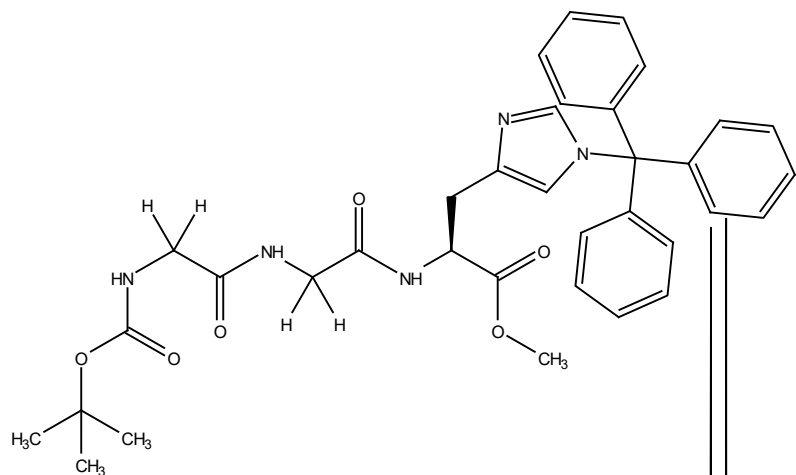
6d





6e





6e

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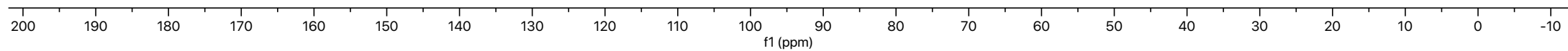
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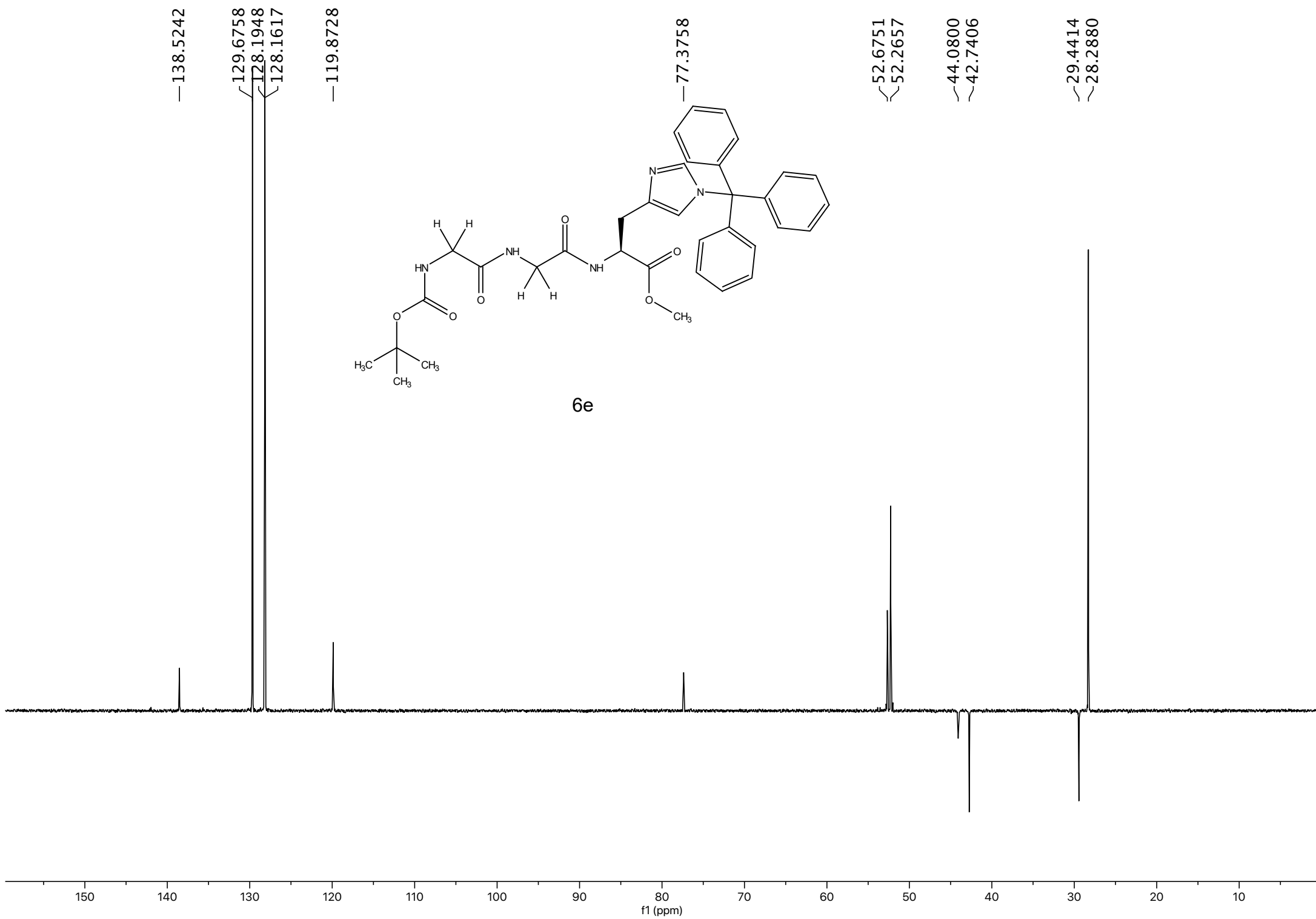
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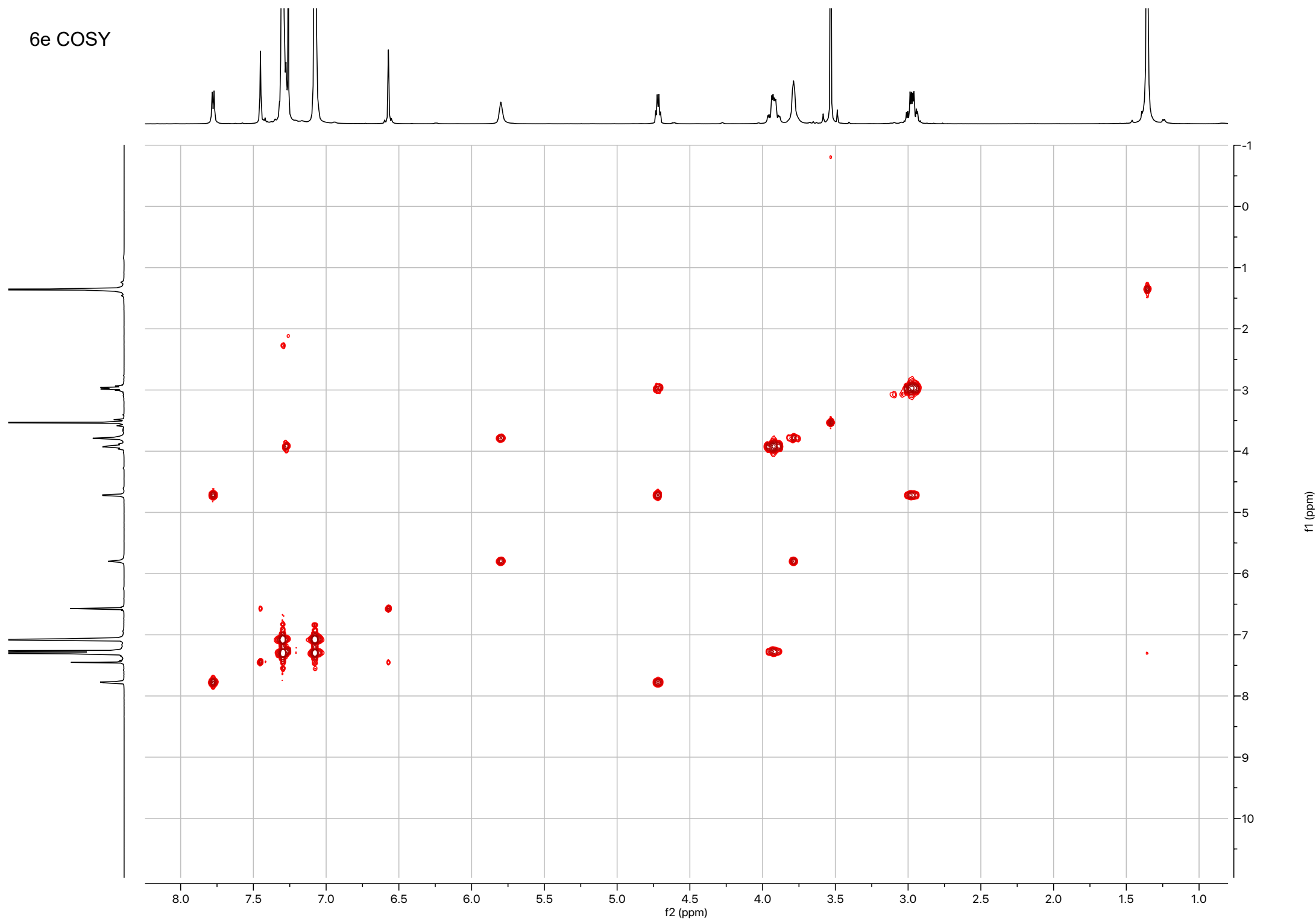
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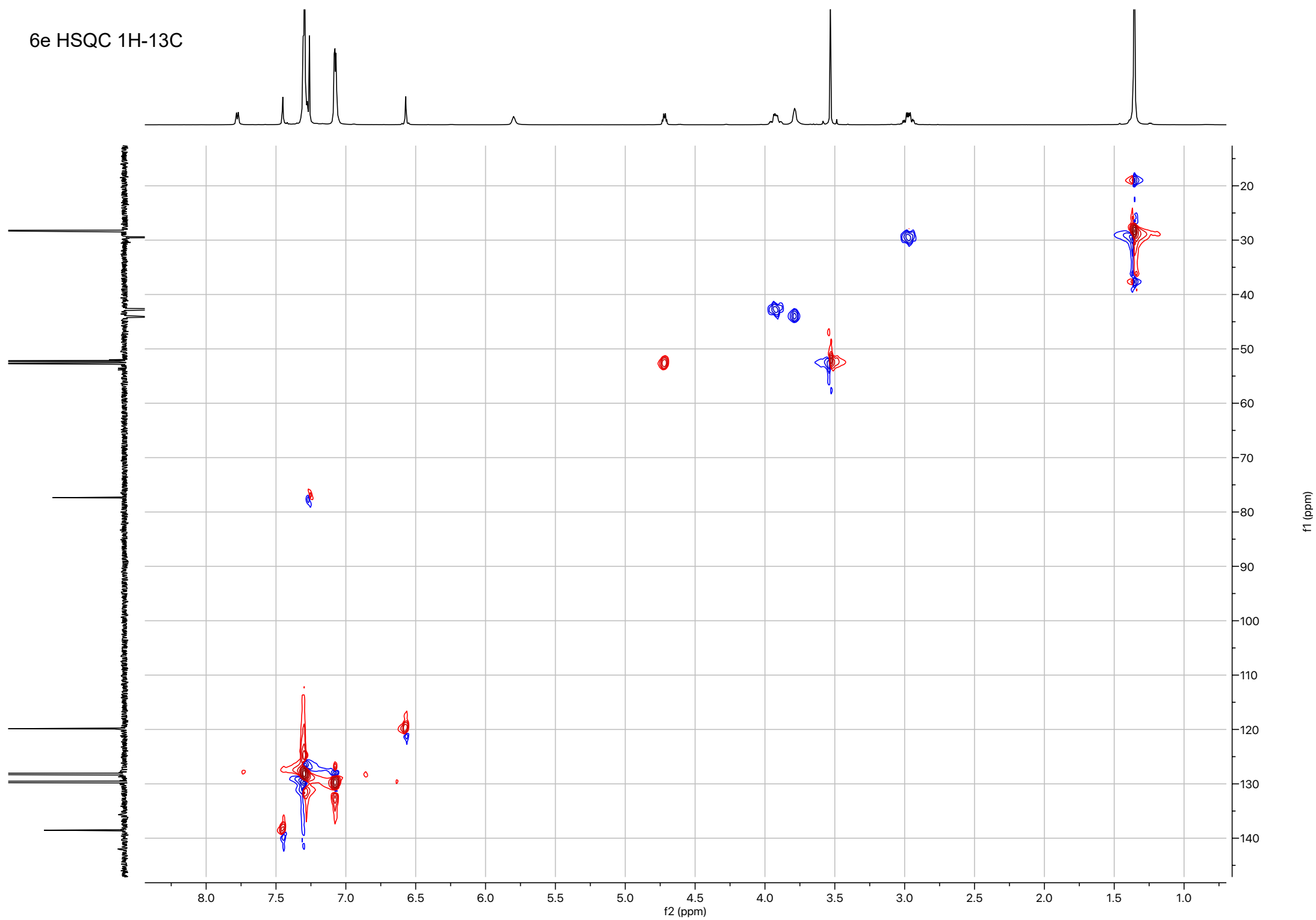




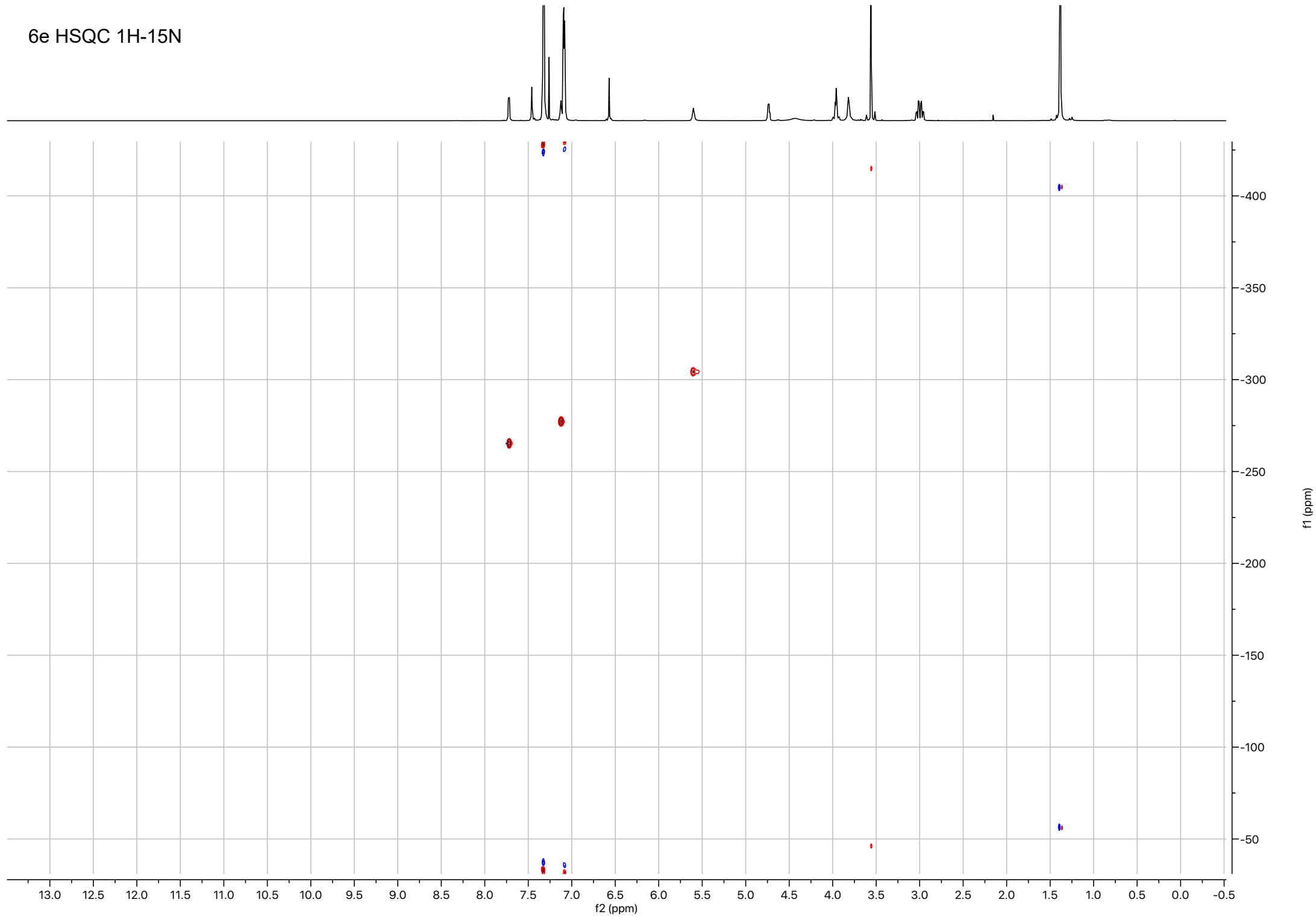
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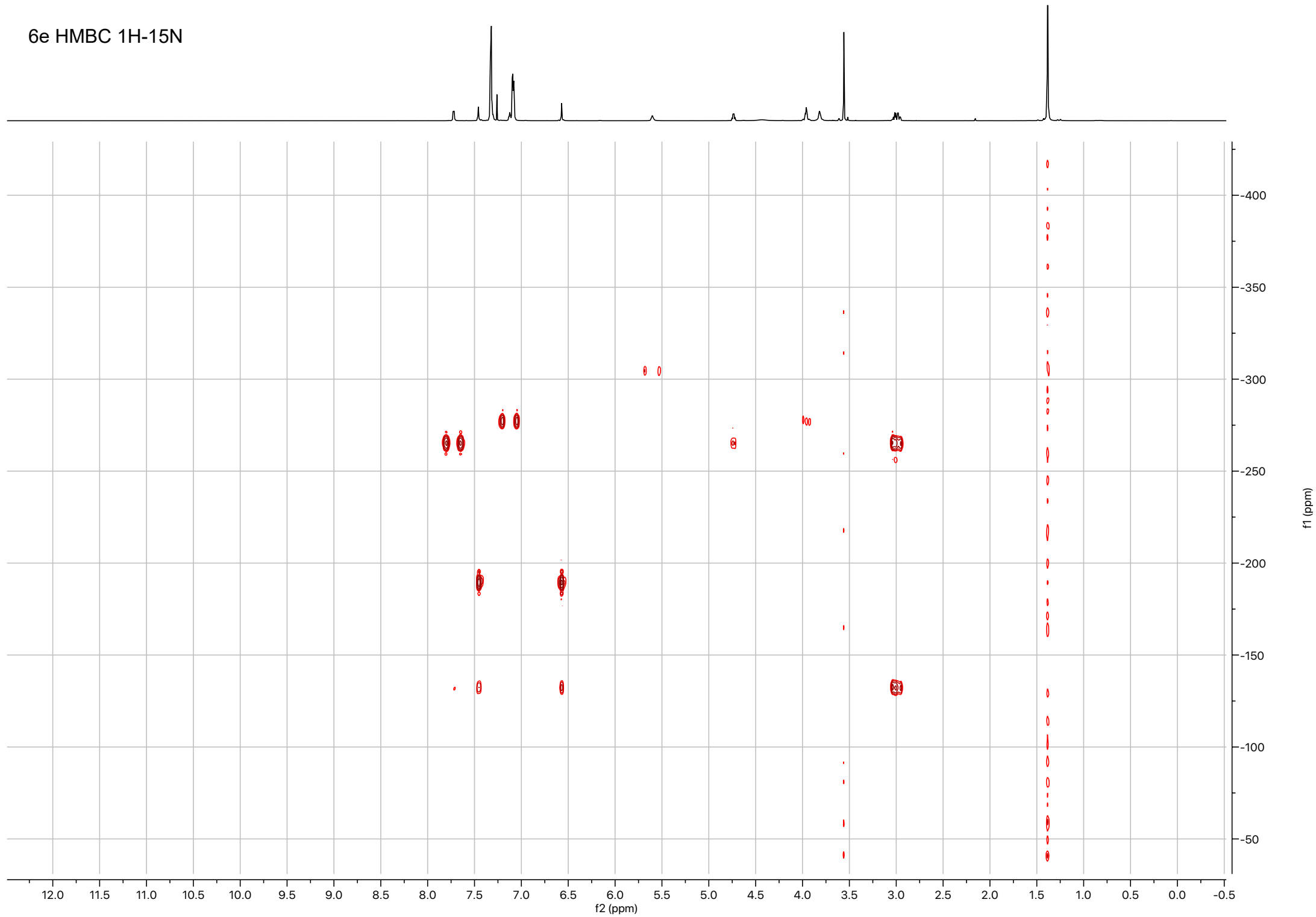
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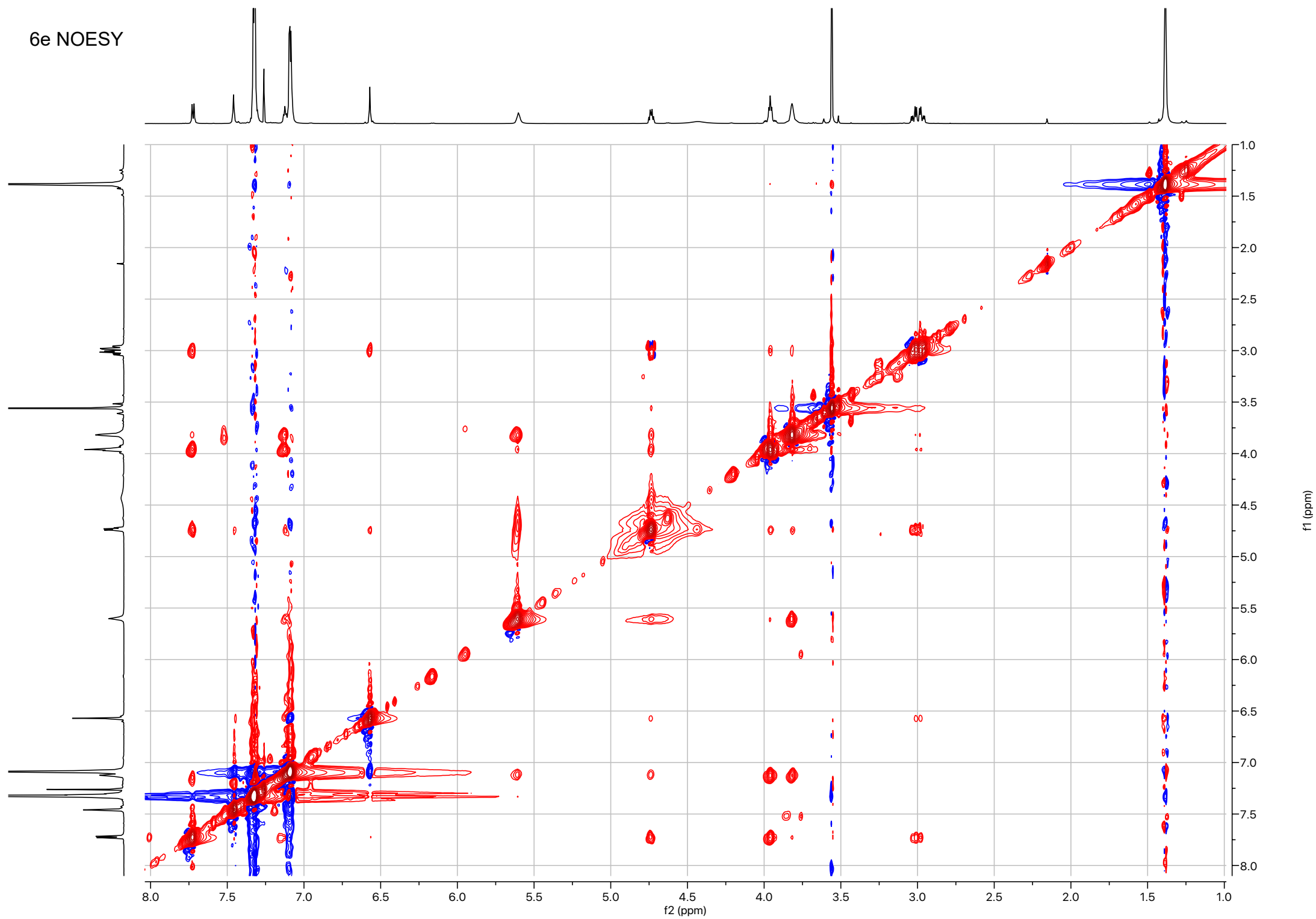
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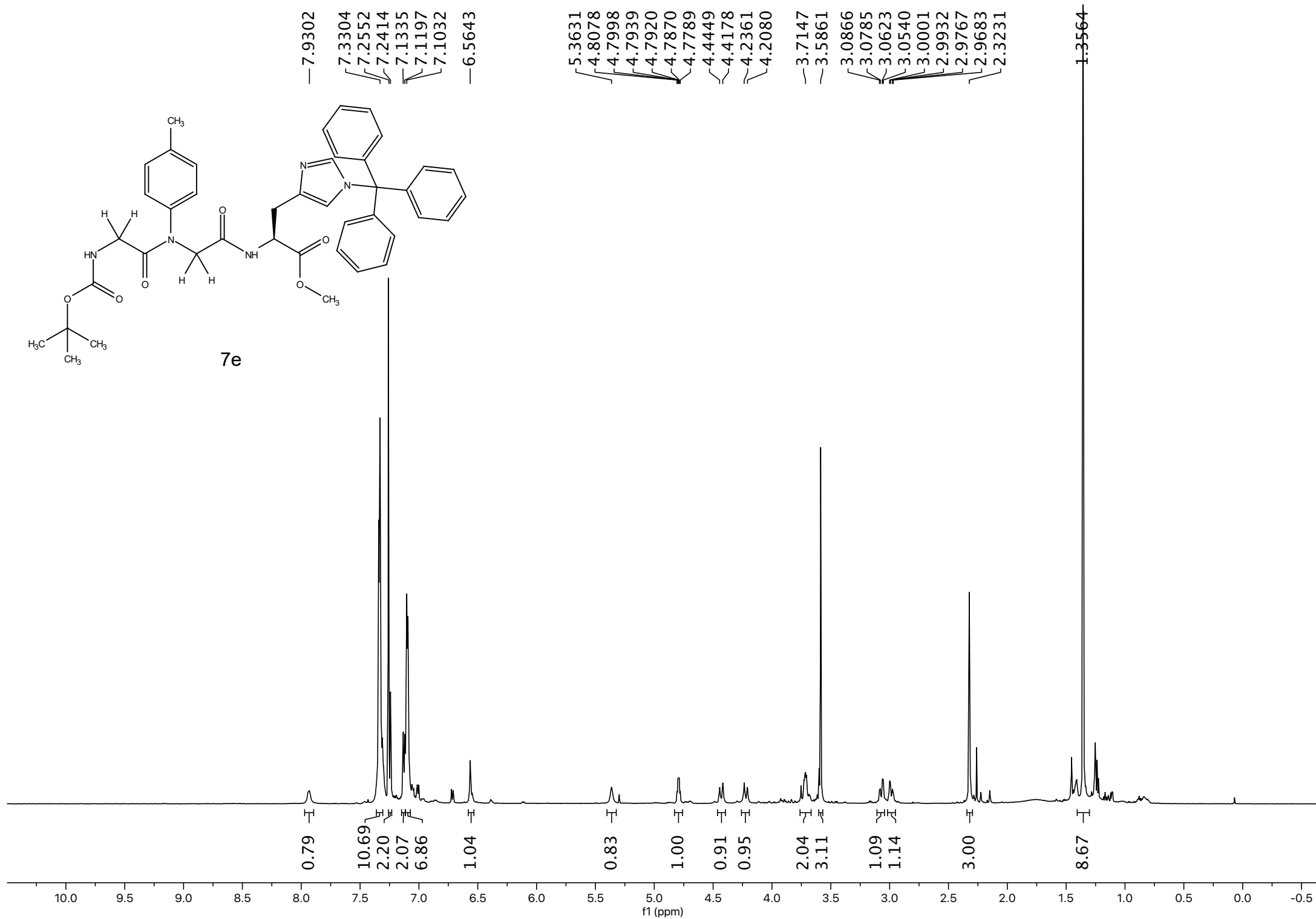
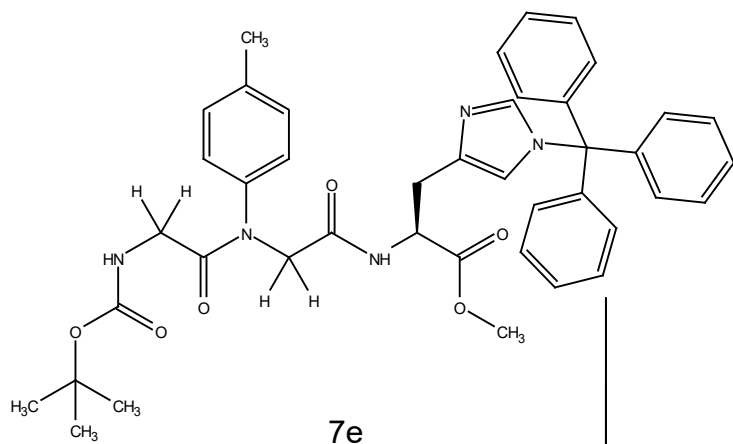


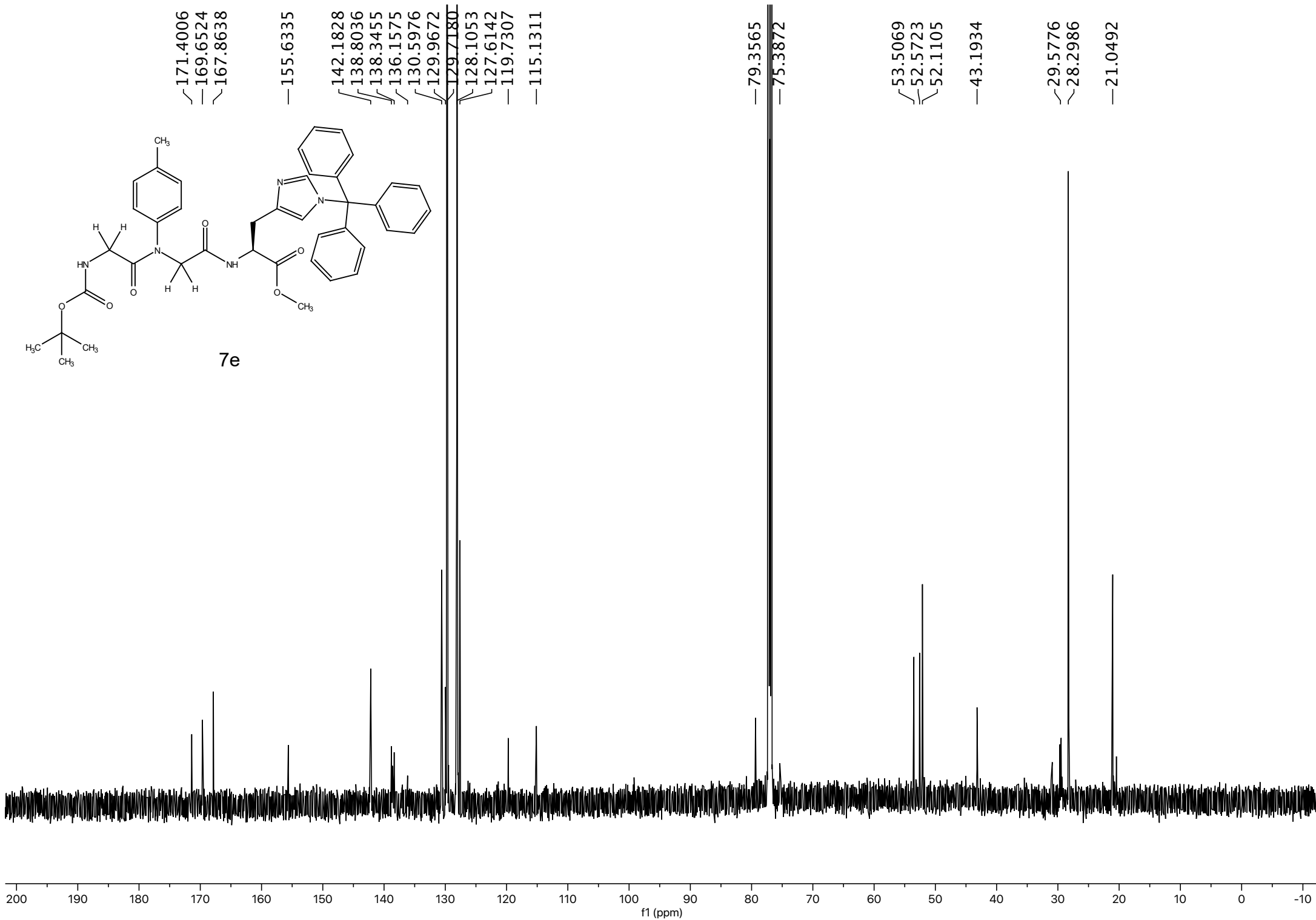
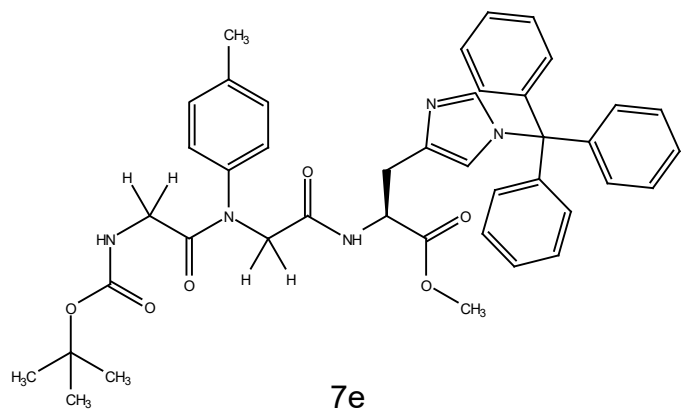
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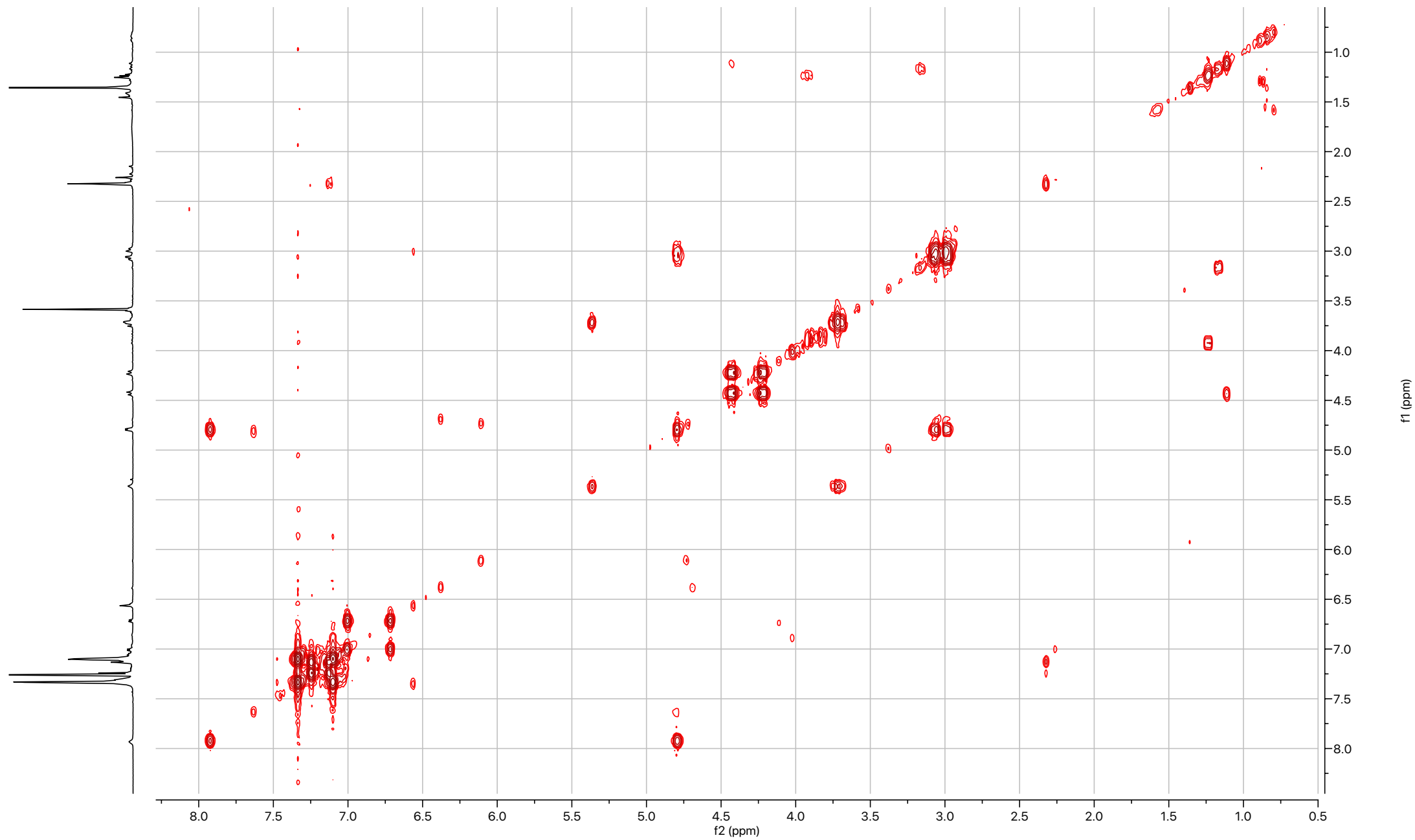
6e NOESY



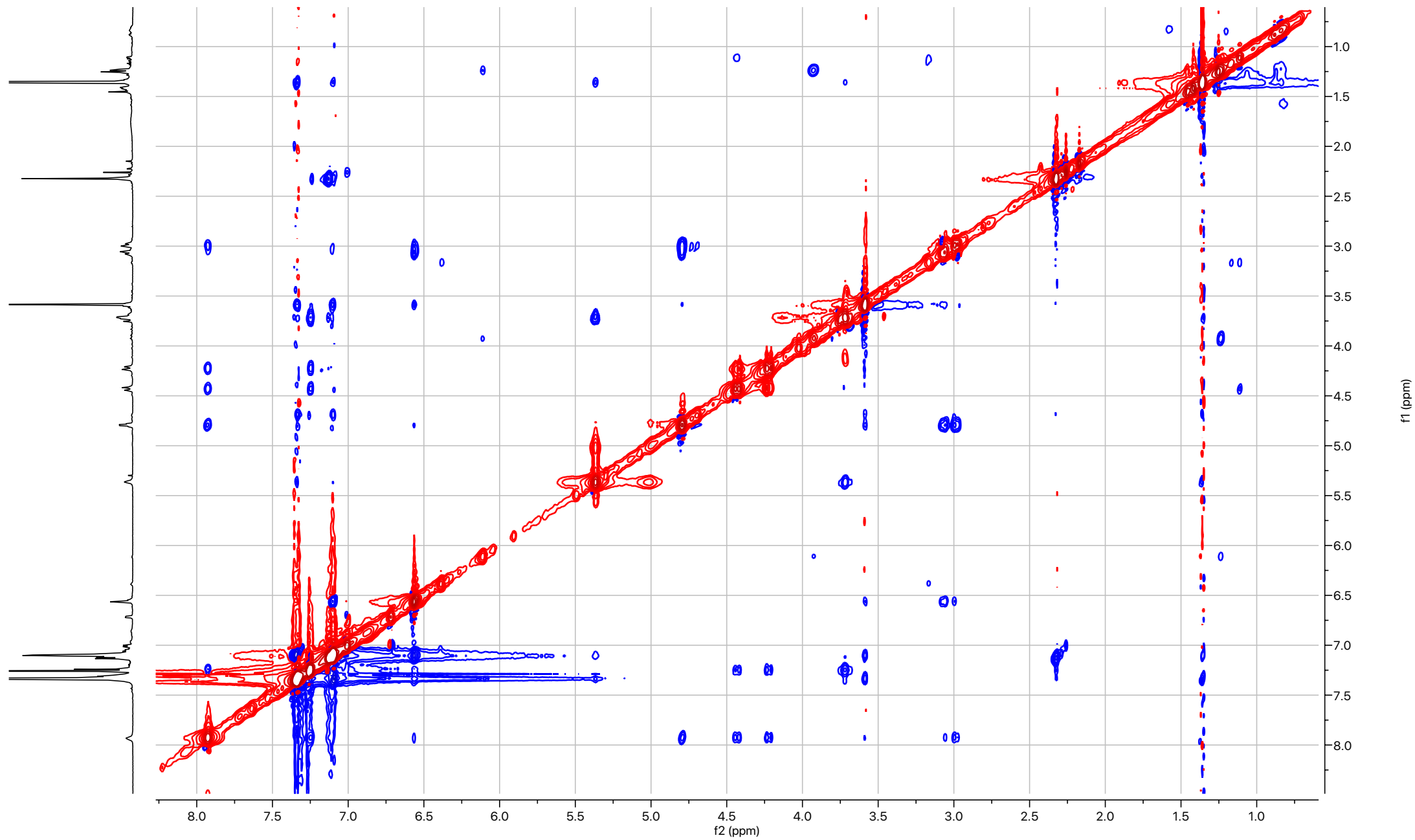




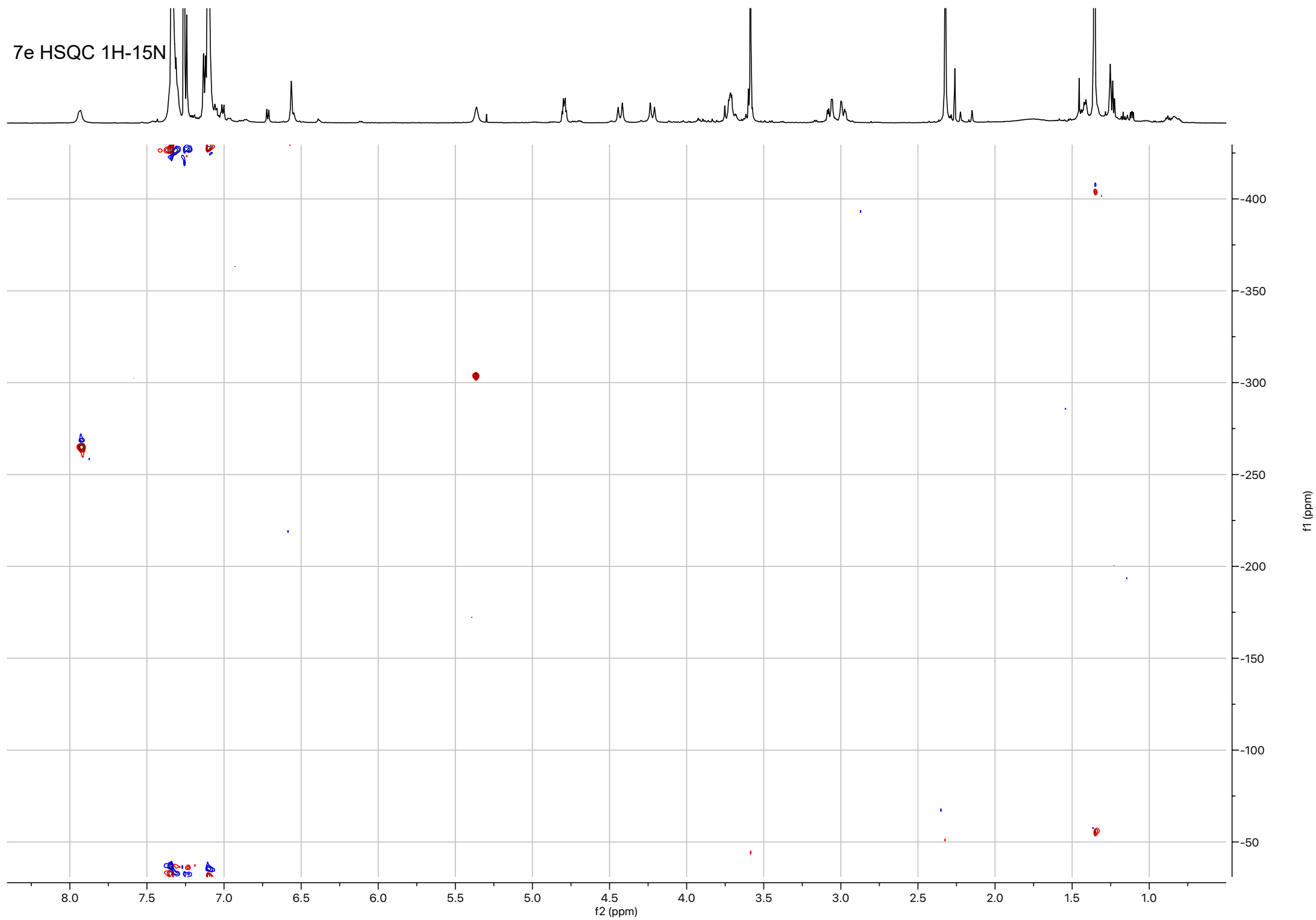
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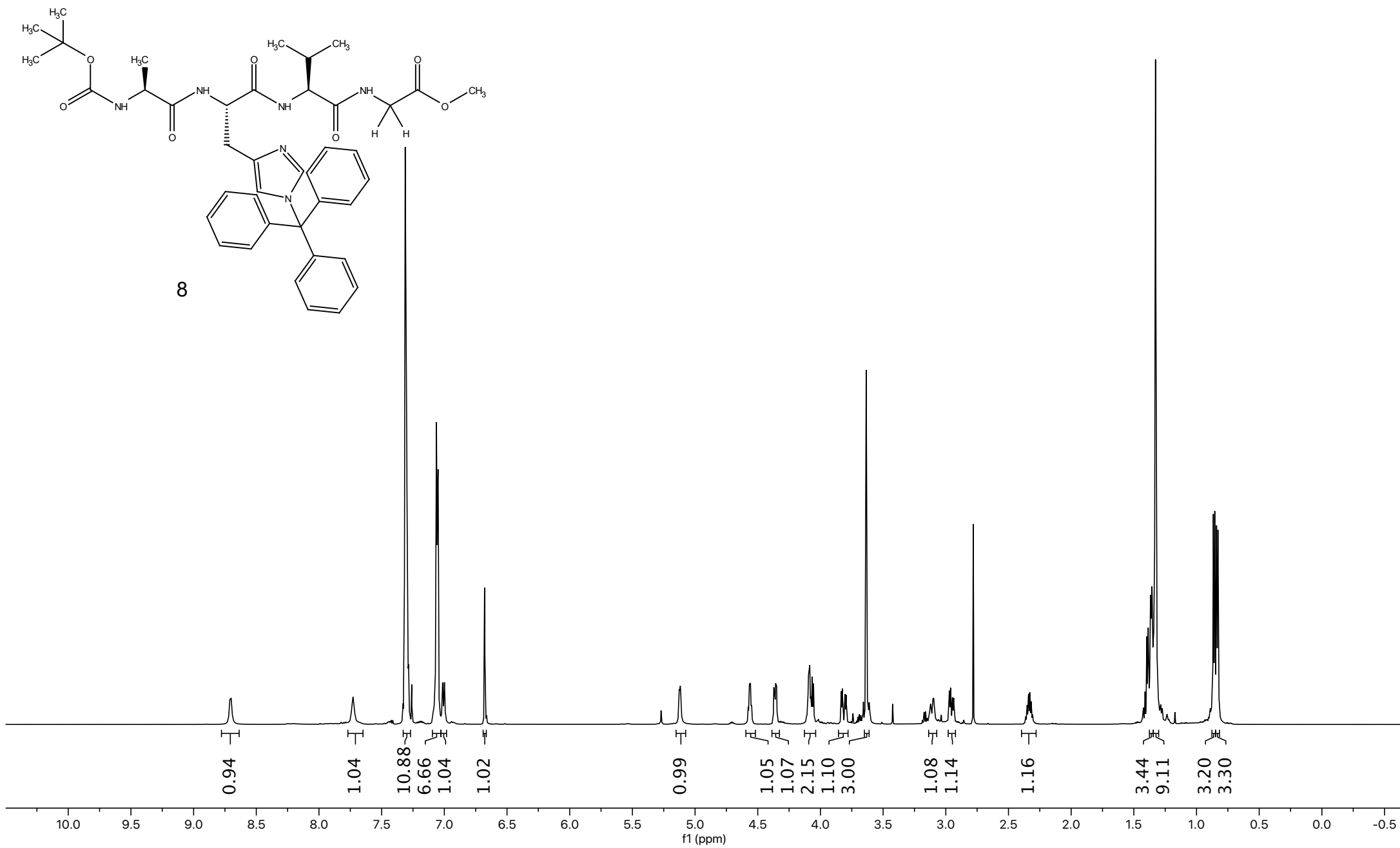


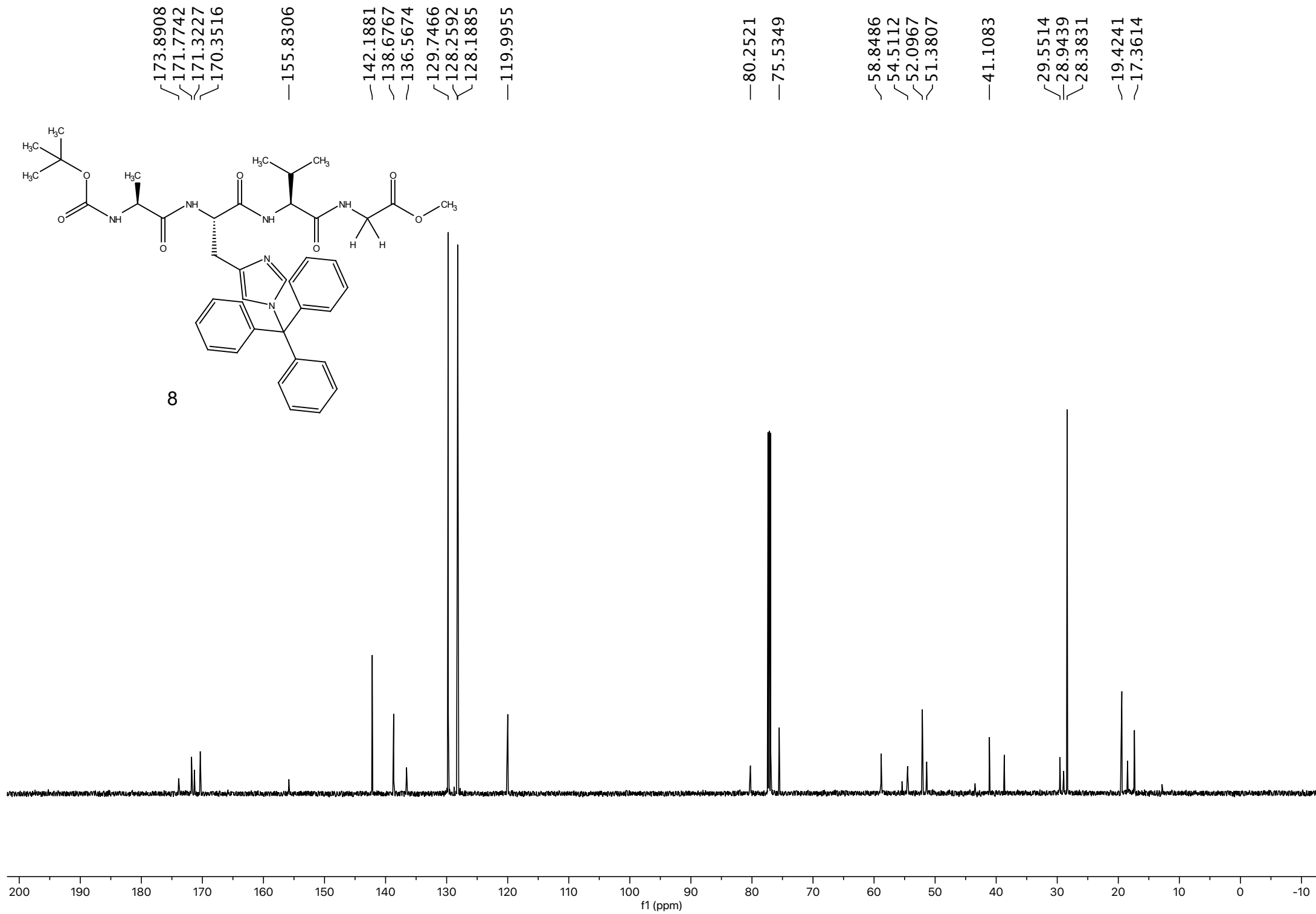
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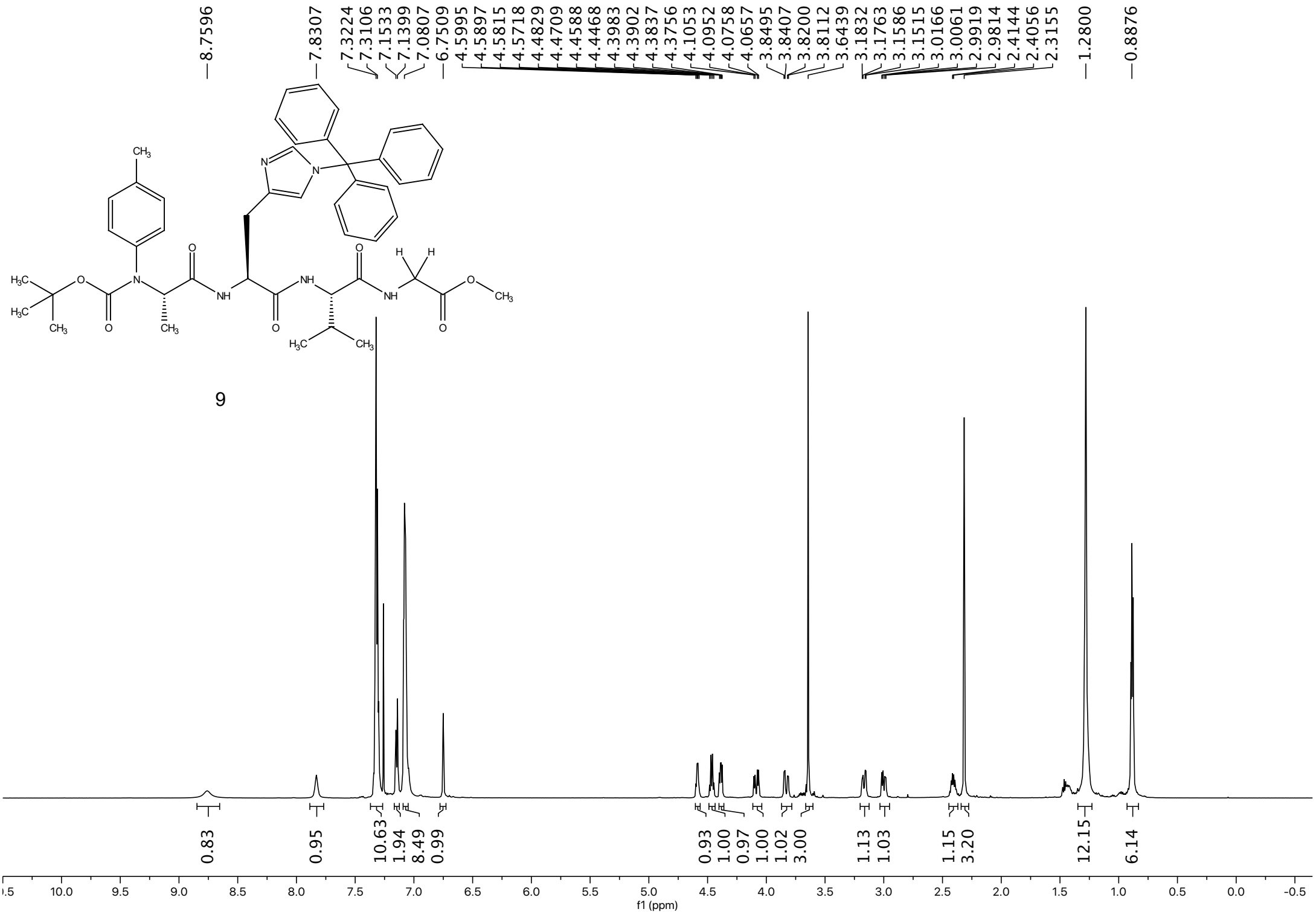


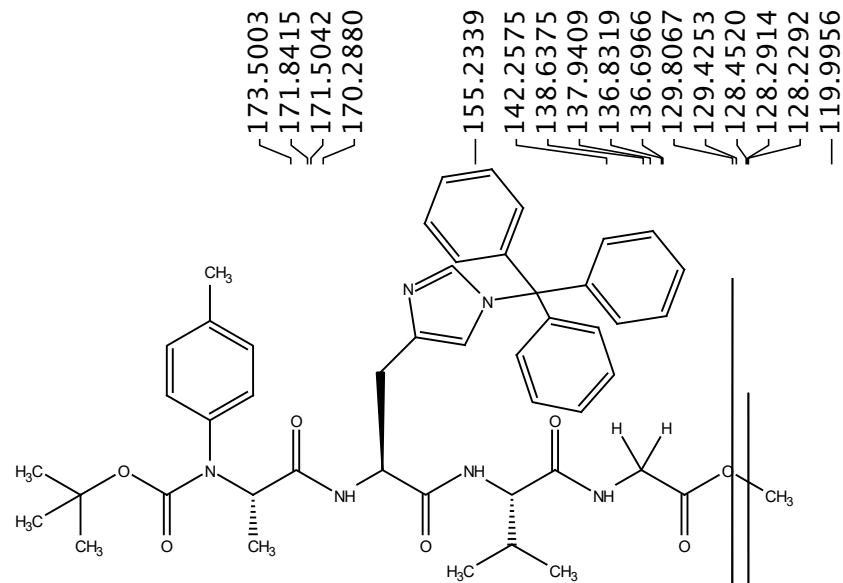
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9

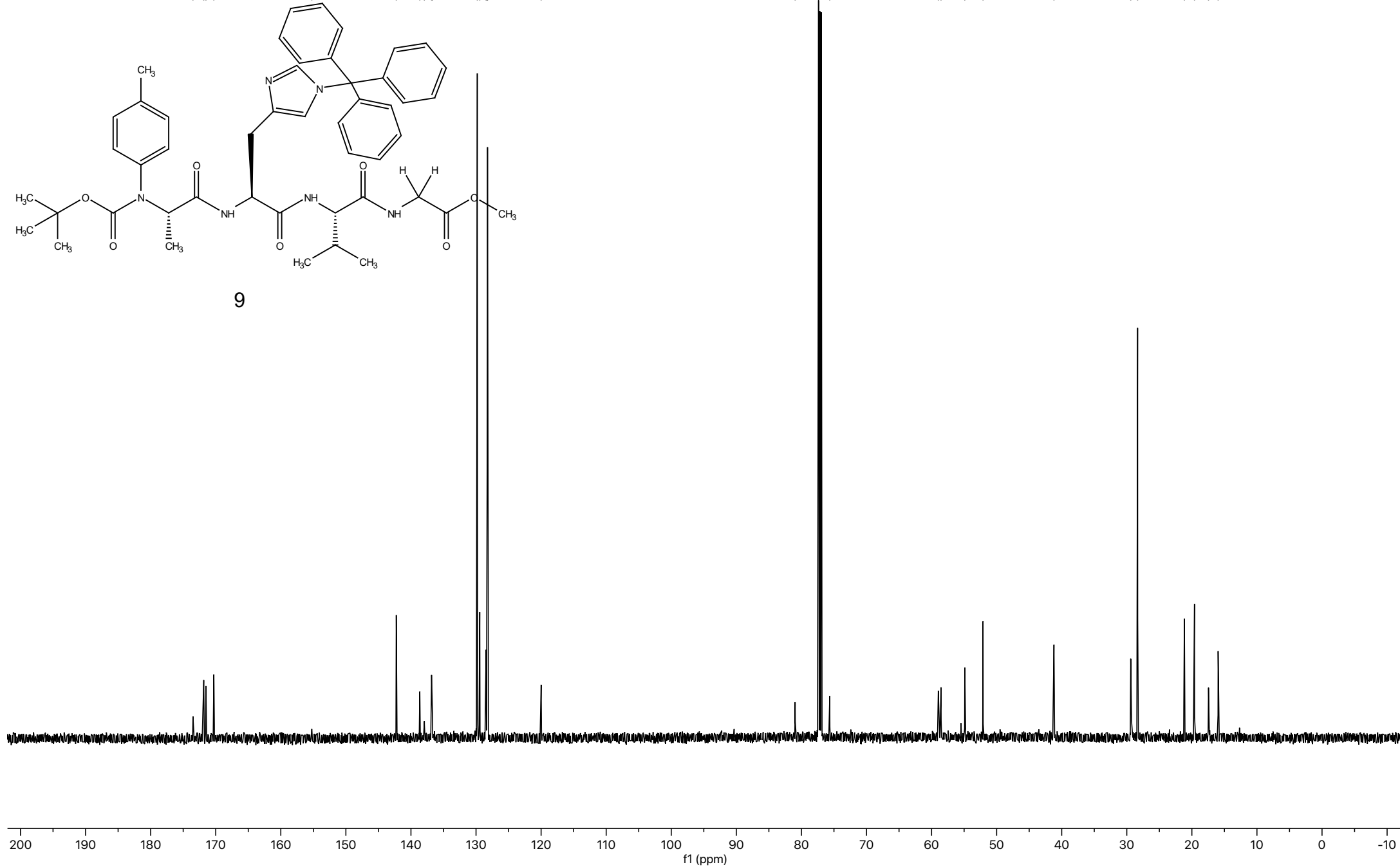
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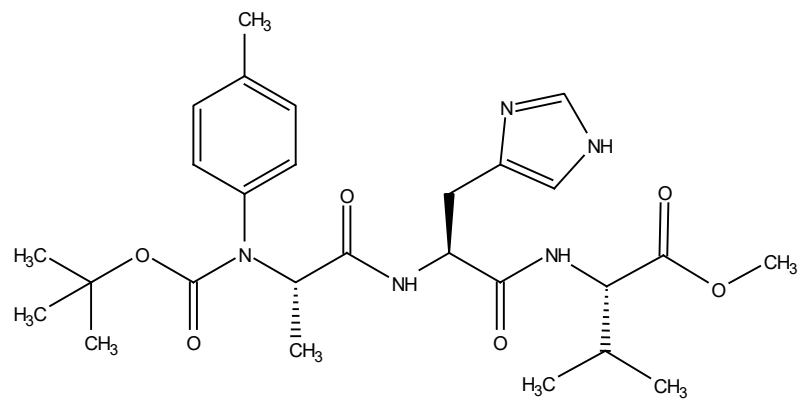
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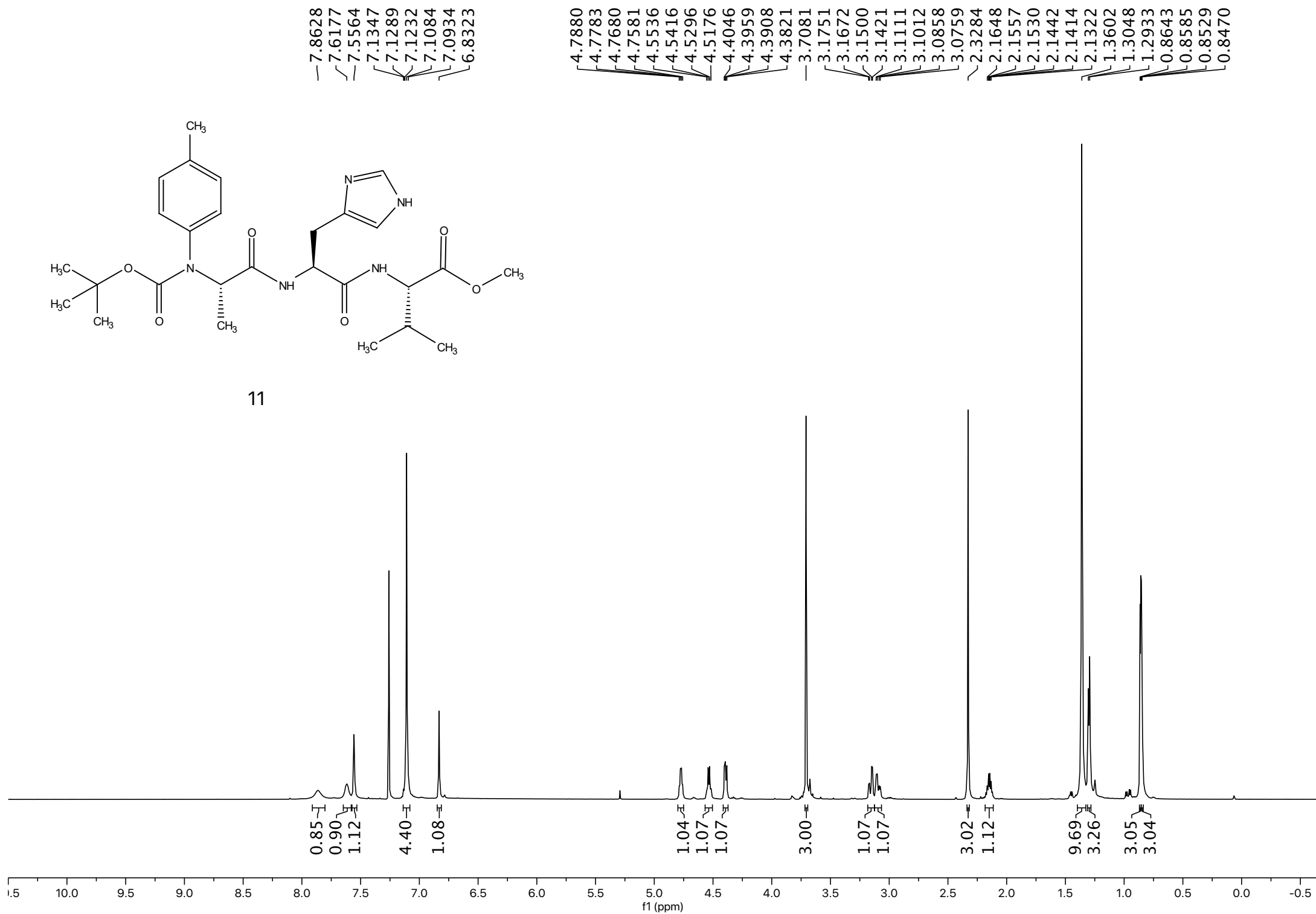
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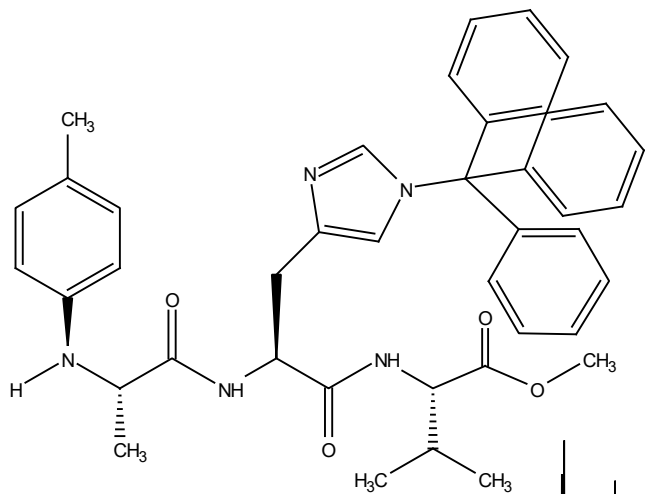
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11





12

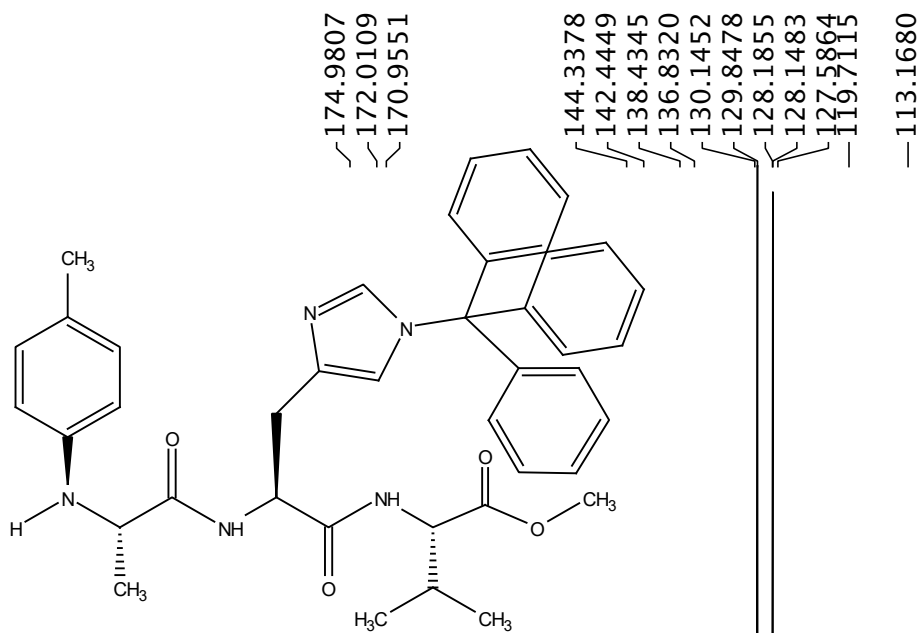
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6.07
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f1 (ppm)



12

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